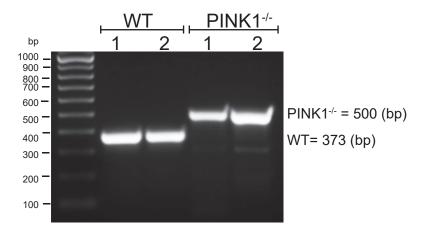
Title: Cleaved PINK1 induces neuronal plasticity through PKA mediated BDNF functional regulation

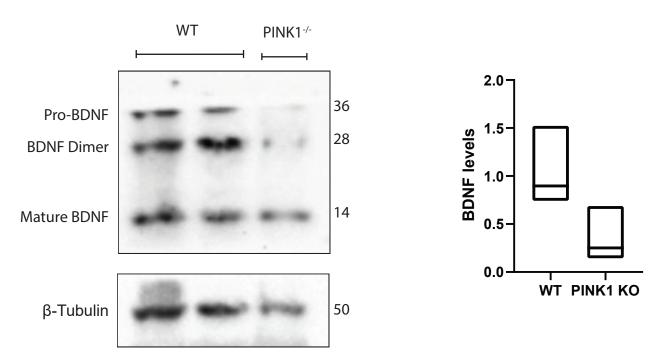
Authors: Smijin K. Soman, David Tingle, Raul Y. Dagda, Mariana Torres, Marisela Dagda, Ruben K. Dagda

Supplementary figures

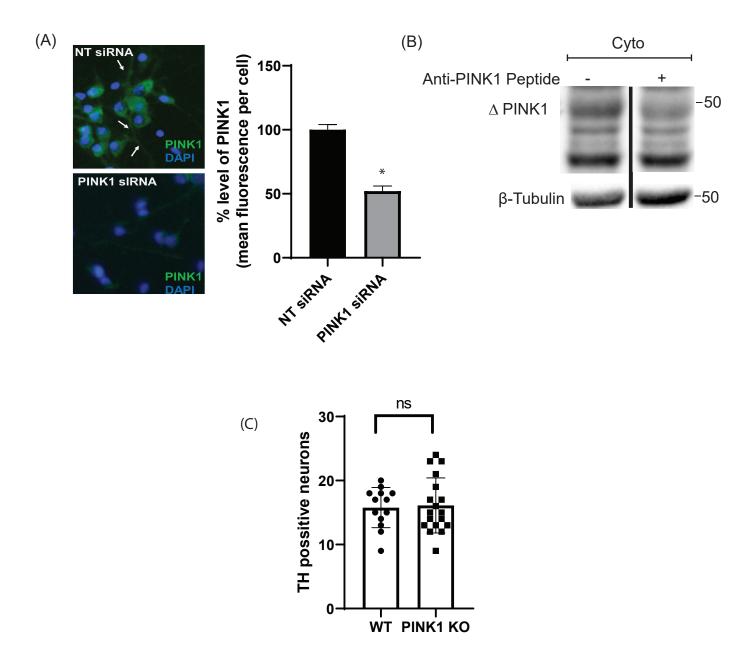


1A: RT-PCR data showing wt Pink1 allele (373bp) in the first and second column; the third and fourth column shows the PINK1 mutant allele (500bp) in homozygous PINK1-/- mouse.

1B

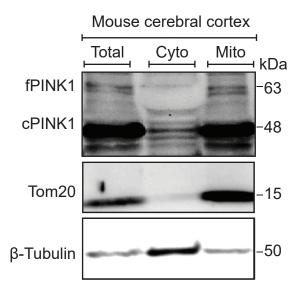


1B: Representative Western blotting for the indicated proteins in lysates extracted 10 month old WT and PINK1KO mouse cerebral cortex. Densitometry analysis of WT and PINK1 KO BDNF levels. Values were normalized to β-Tubulin levels. N=3 (3 mouse per experimental group).



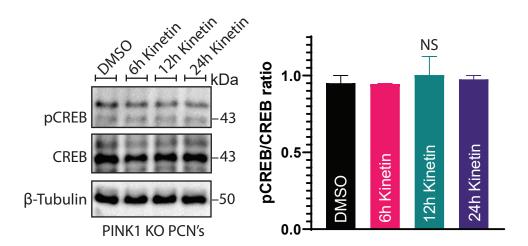
A, Representative immunofluorescence (IF) images of mouse PCNs transfected with a non-targeted siRNA control (NTsiRNA) or with a pre-validated mouse PINK1 siRNA (Life Technologies). 3 days post transfection neurons were immunostained for PINK1 (green) and counterstained with DAPI (blue) to visualize nuclei. Quantification on the right shows IF-based quantification of mean levels (±SEM) of endogenous PINK1 for each condition (*:p<0.05 vs. NTsiRNA, 80 neurons per condition, t-test). **B**, Representative Western blotting showing that the immunoreactivity of cleaved PINK1 in the cytosol (cyto) is reduced in the presence of a PINK1 blocking peptide (5 μg peptide :1μg antibody). Discontinuity in the western blot images (shown by a line) indicate that certain lanes have been cropped to maintain visual clarity. The above mentioned data confirms the specificity of anti-PINK1 antibody (; BC100-494; Novus Biologicals, Littleton, CO). **C**, Graphical representation of Tyrosine hydroxylase possitive neurons in substantia nigra of the midbrain section of WT and PINK1 KO 10 month old mice.

Supplemetary figure 3

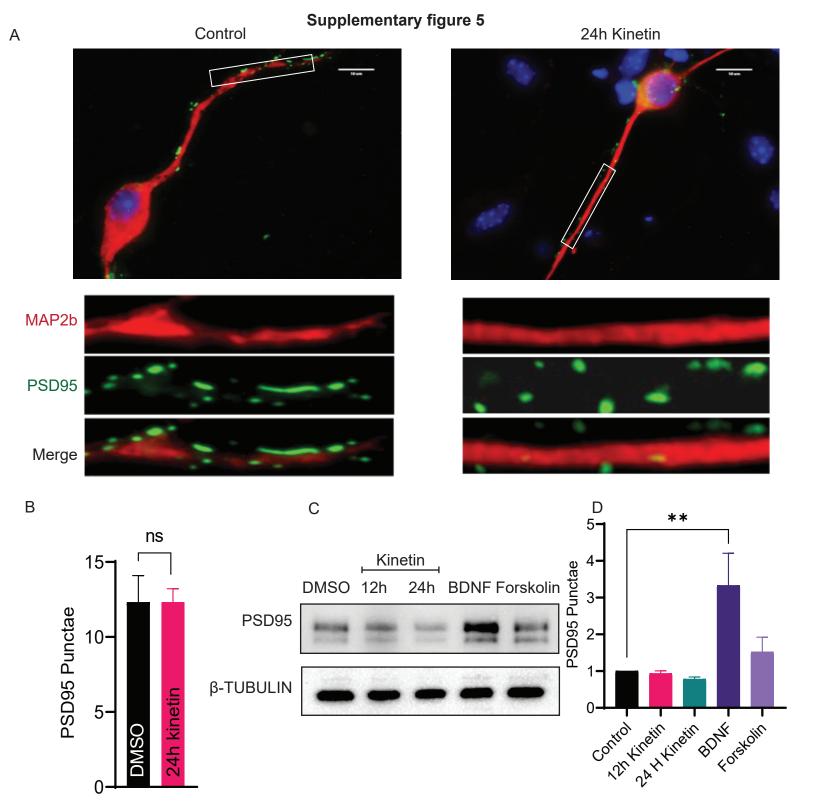


Representative Western blotting for the indicated proteins in lysates extracted after subcellular fractionation of mouse cerebral cortex. Note that 75 µg of total cell lysate, cytosolic and mitochondrial protein was loaded into the gel, which accounts for 12.45%, 1.93%, and 24.79% of the total, cytosolic and mitochondrial lysate, respectively. Densitometry with correction for the relative fraction of total mitochondrial and cytosolic proteins loaded on the gel reveals that 75.48% of the 48kDa cPINK1 is localized to the cytosolic fraction whereas the smaller remnant of cPINK1 is localized to mitochondria.

Supplemetary figure 4

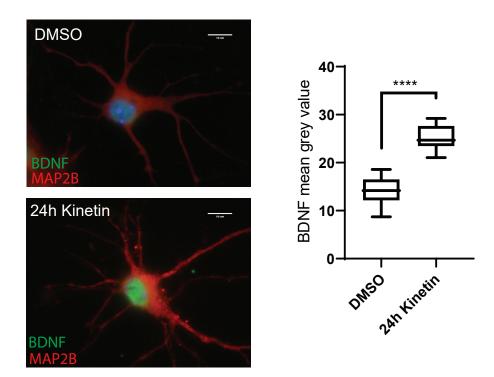


Representative Western blotting for the indicated proteins in cell lysates extracted at different time-points from DMSO, 6h kinetin, 12h kinetin, and 24h kinetin treated PINK1-/- PCNs. Densitometric analysis of Phospho-CREB and CREB expression levels in from DMSO, 6h kinetin, 12h kinetin, and 24h kinetin treated PINK1-/- PCNs. Values were normalized to β -Tubulin.



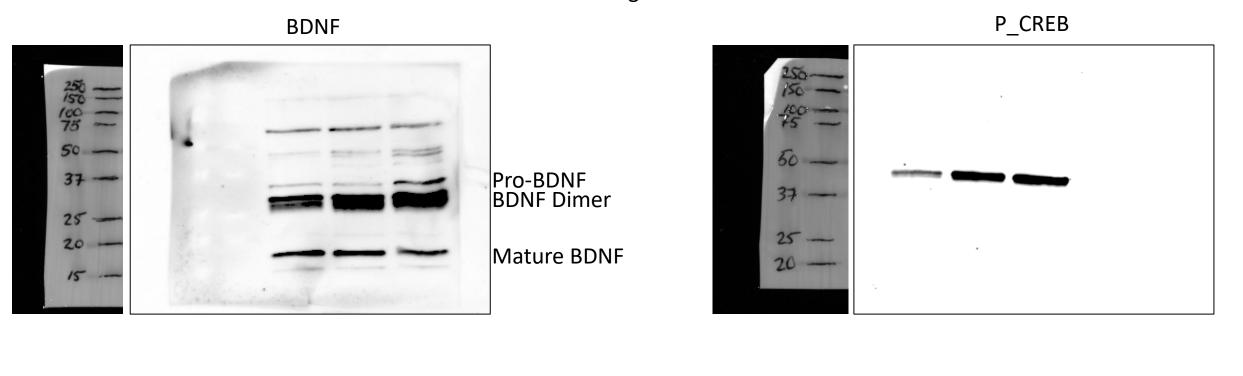
A, Representative epifluorescence merged images of PSD-95 (green) immunostaining in PINK1-KO PCNs treated with DMSO (Control) and 24h kinetin. Nucleus is counterstained by DAPI (blue). Magnified images of PSD-95 (green) punctae on dendrites (MAP2b-red). Scale bar represents 10µM. B, Quantification of PSD-95 expression in PINK1-KO PCNs treated with DMSO (Control) and 24h kinetin. An unpaired t-test was used for statistical analysis. Bars denote the average ratio \pm SEM and are representative of two independent experiments (N=20; 10 neurons per genotype per experiment). C, Representative Western blot for PSD95 immunoblotting in lysates extracted at differential time-points from DMSO (Control), 4h kinetin, 8h kinetin, 12h kinetin, and 24h kinetin treated PINK1-/- PCNs. F, Densitometry analysis of PSD95 expression levels in DMSO (Control), 4h kinetin, 8h kinetin, 12h kinetin, and 24h kinetin treated PINK1-/- PCNs. Values were normalized to β -Tubulin. Ordinary one-way ANOVA and post-hoc analysis using Dunnett's multiple comparison test were used for statistical analysis. Bars denote the average ratio \pm SEM and are representative of two independent experiments.

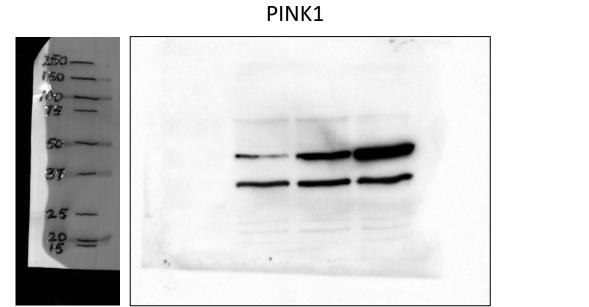
Supplementary figure 6



Representative immunofluorescence (IF) images of WT mouse PCNs treated with Kinetin for 24h. Neurons were immunostained for BDNF (green), MAP2B (Red) and counterstained with DAPI (blue) to visualize nuclei. Quantification on the right shows IF-based quantification of mean levels (±SEM) of endogenous BDNF for each condition (****:p<0.001: 10 Neurons per experimental group, t-test).

Fig1A





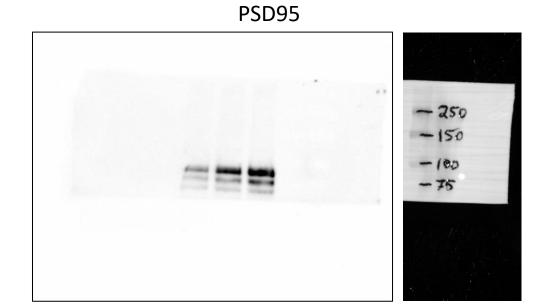
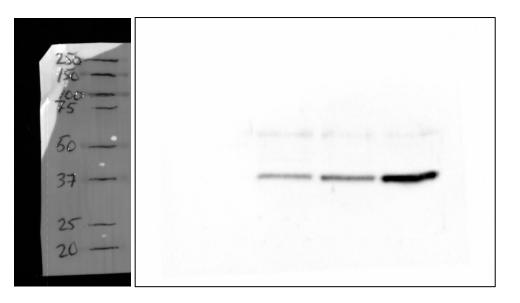


Fig1A Synaptophysin β-Tubulin



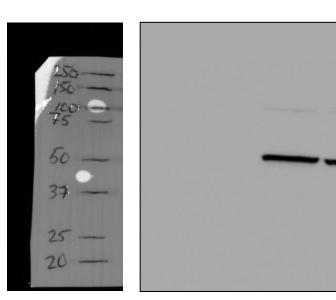
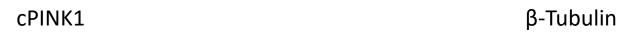
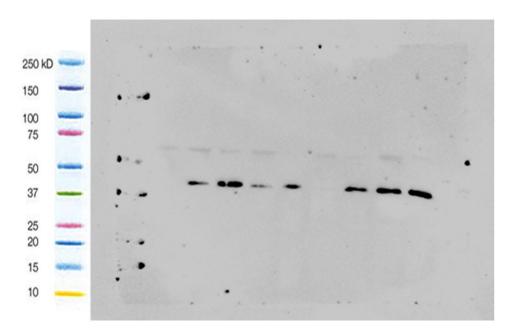


Fig 1D





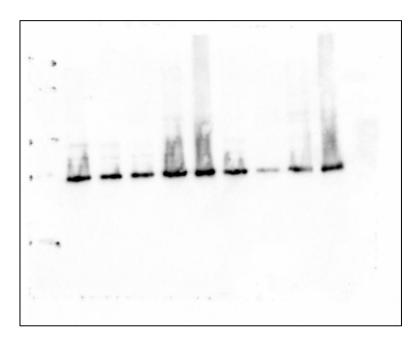
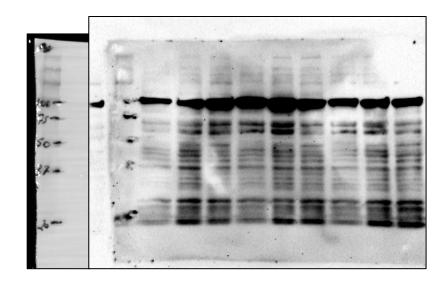


Fig1F

PINK1



β-Tubulin

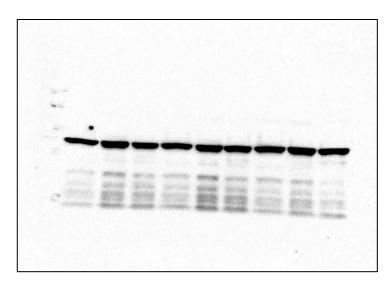
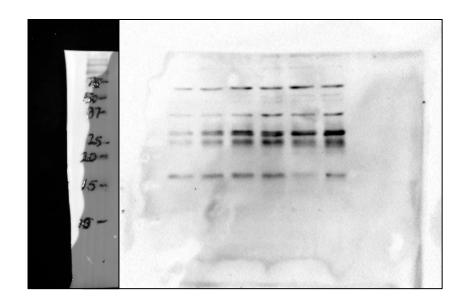


Fig2E

BDNF



β-Tubulin

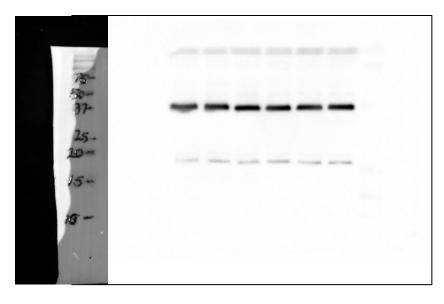
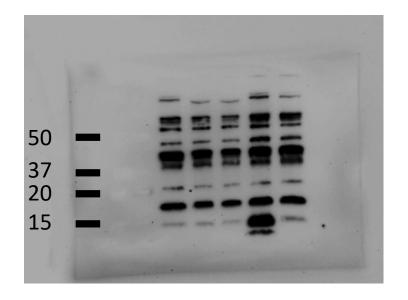
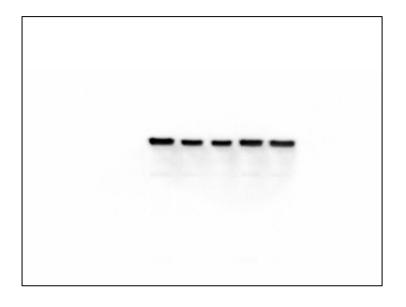


Fig2G

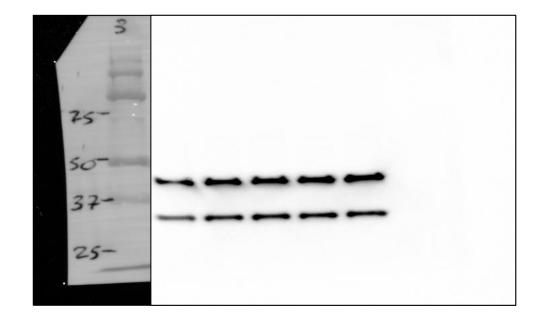
BDNF



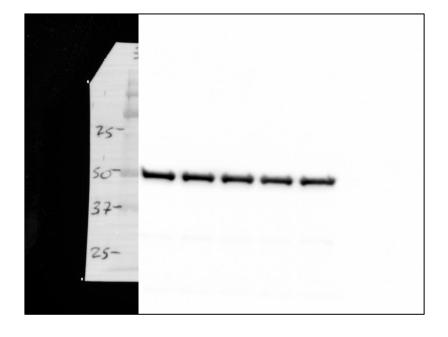
β-Tubulin



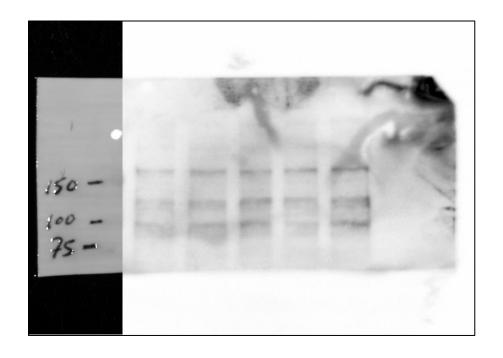
cPINK1



β-Tubulin



 $Phospho\text{-}TRK\beta$



 $\beta\text{-Tubulin}$

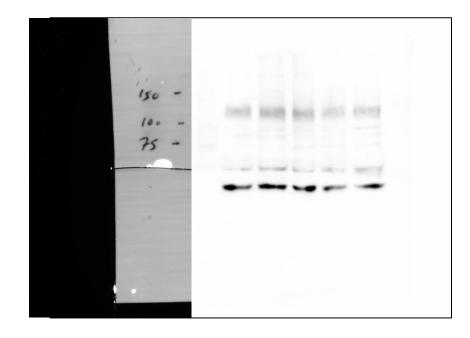
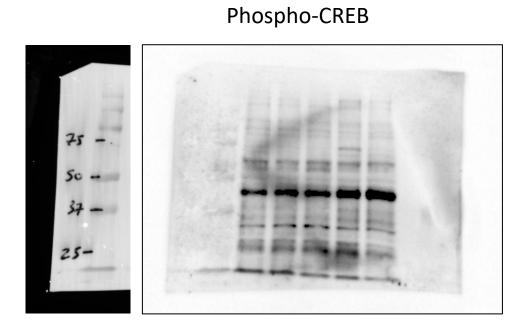
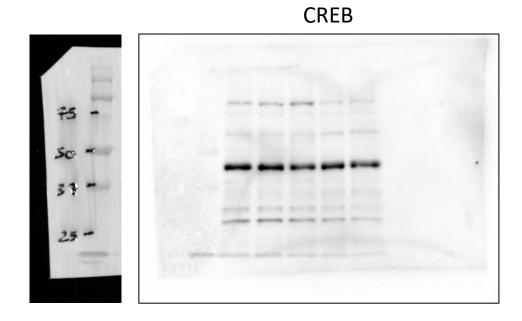


Fig4E





β-Tubulin

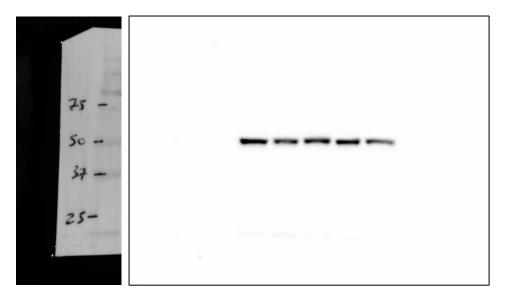
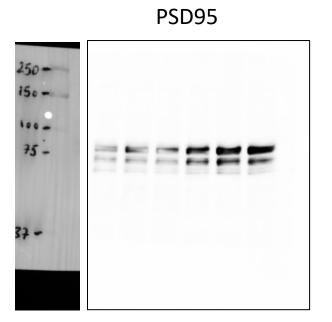
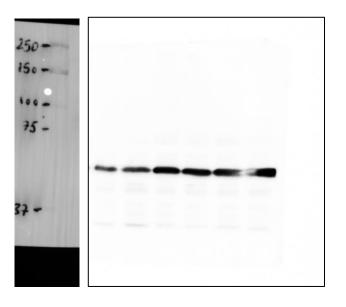


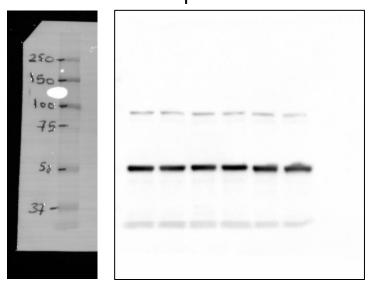
Fig5E

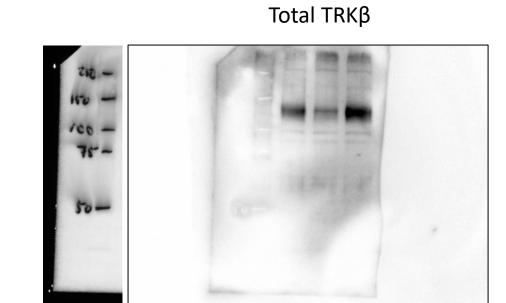


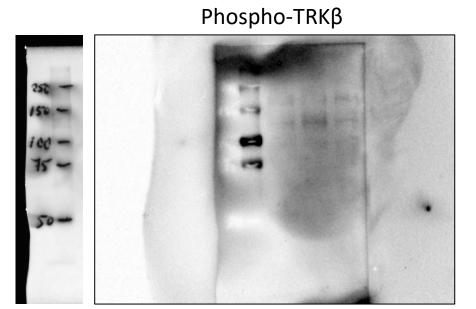
Synaptophysin

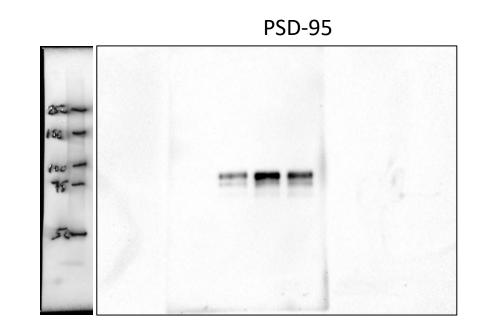


β-Tubulin





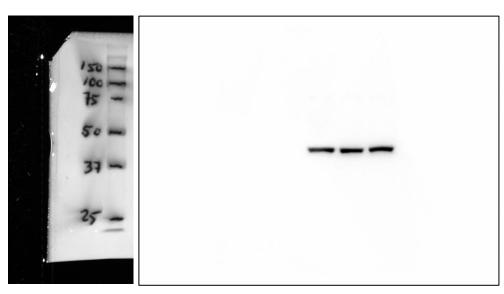


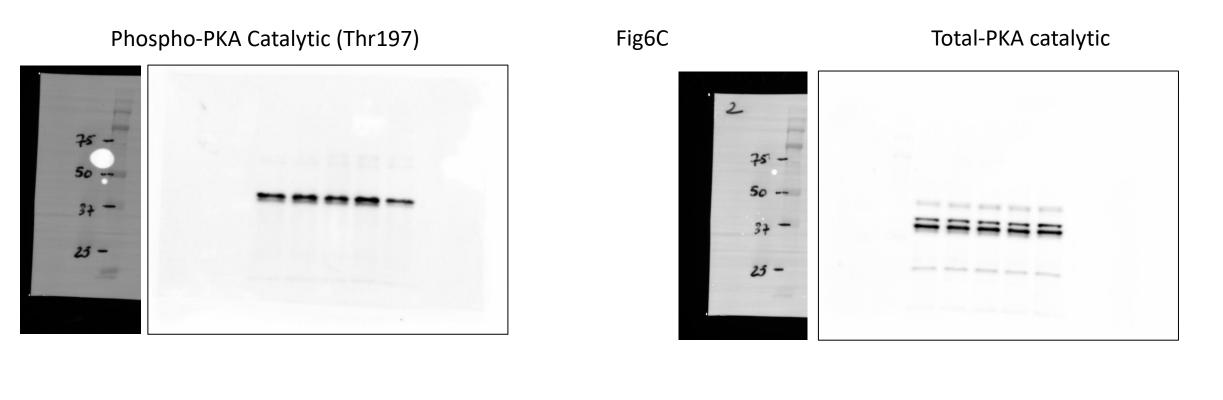


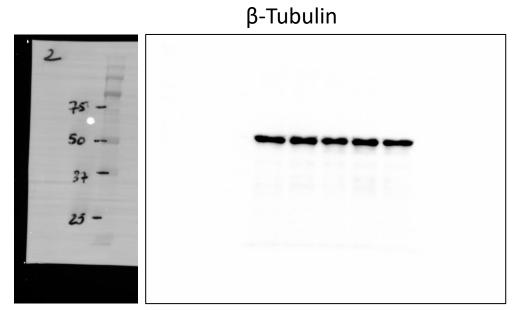
BDNF

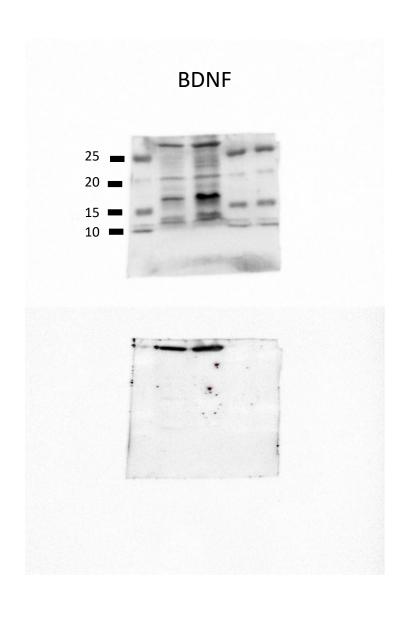
Fig6A

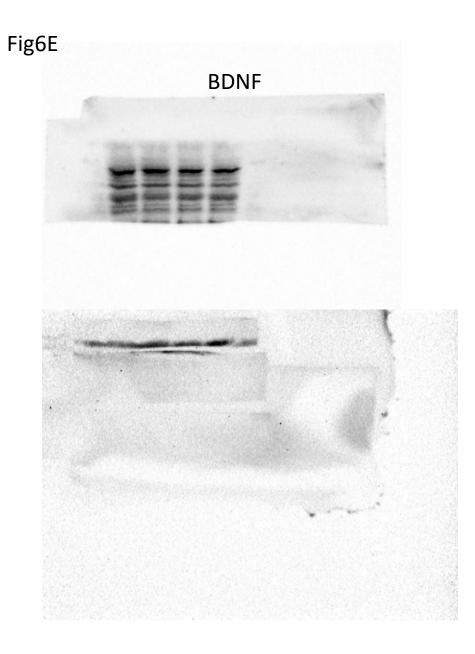
β-Tubulin





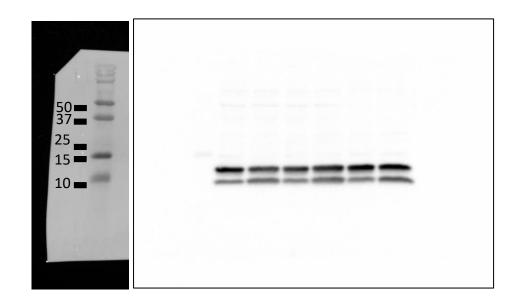


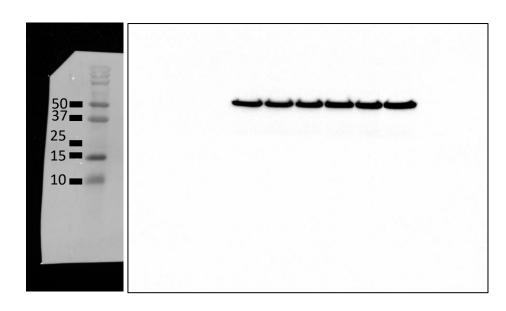




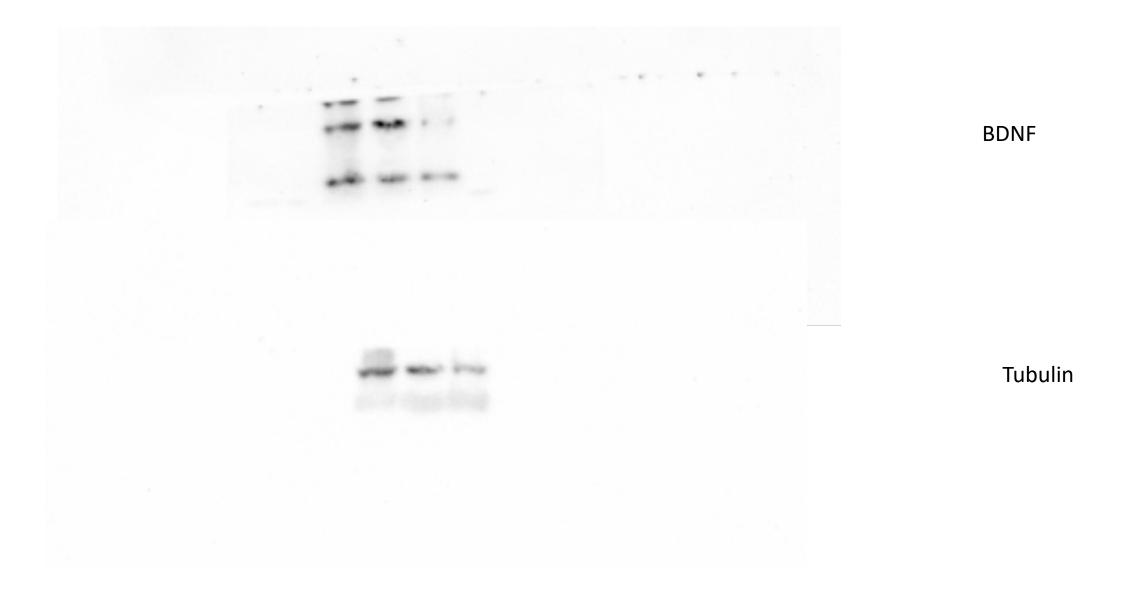
Tubulin



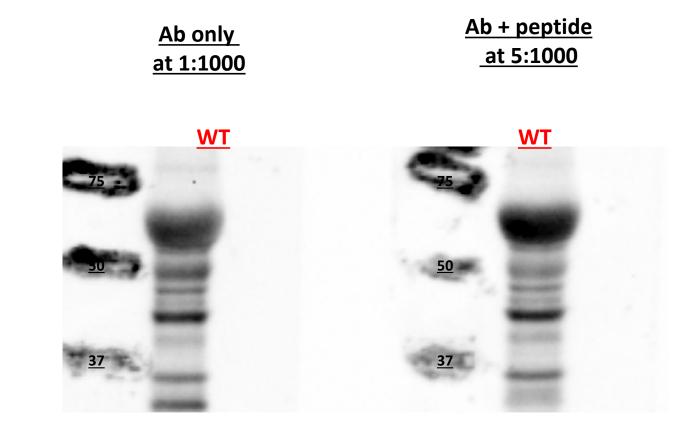




Supplementary Fig. 1B

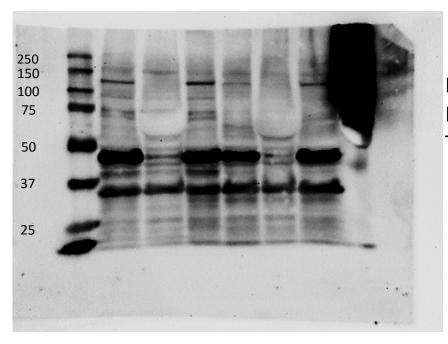


Supplementary Fig. 2b

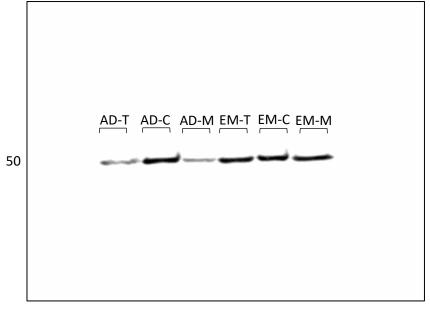


Exposure 20 seconds

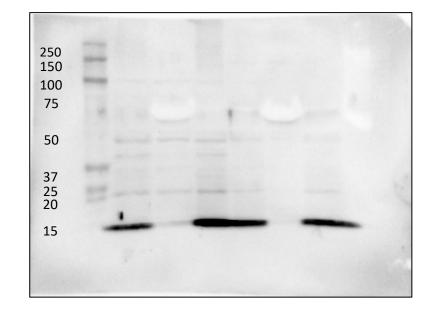
Supplementary Fig. 3



Pink1 FL-63KDa Truncated-48Kda

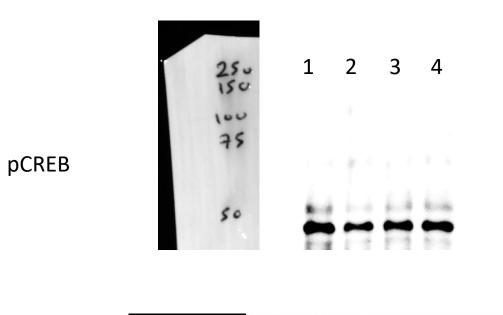


B-Tubulin=50kDa



Tom20=15KDa

Supplementary Fig. 4



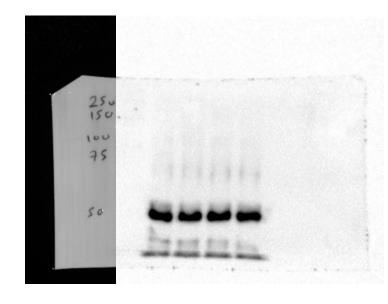
1: DMSO

2: 6h Kinetin

3: 12h Kinetin

4: 24h Kinetin

β-Tubulin



CREB 250 150 75

Supplementary Fig. 5C

