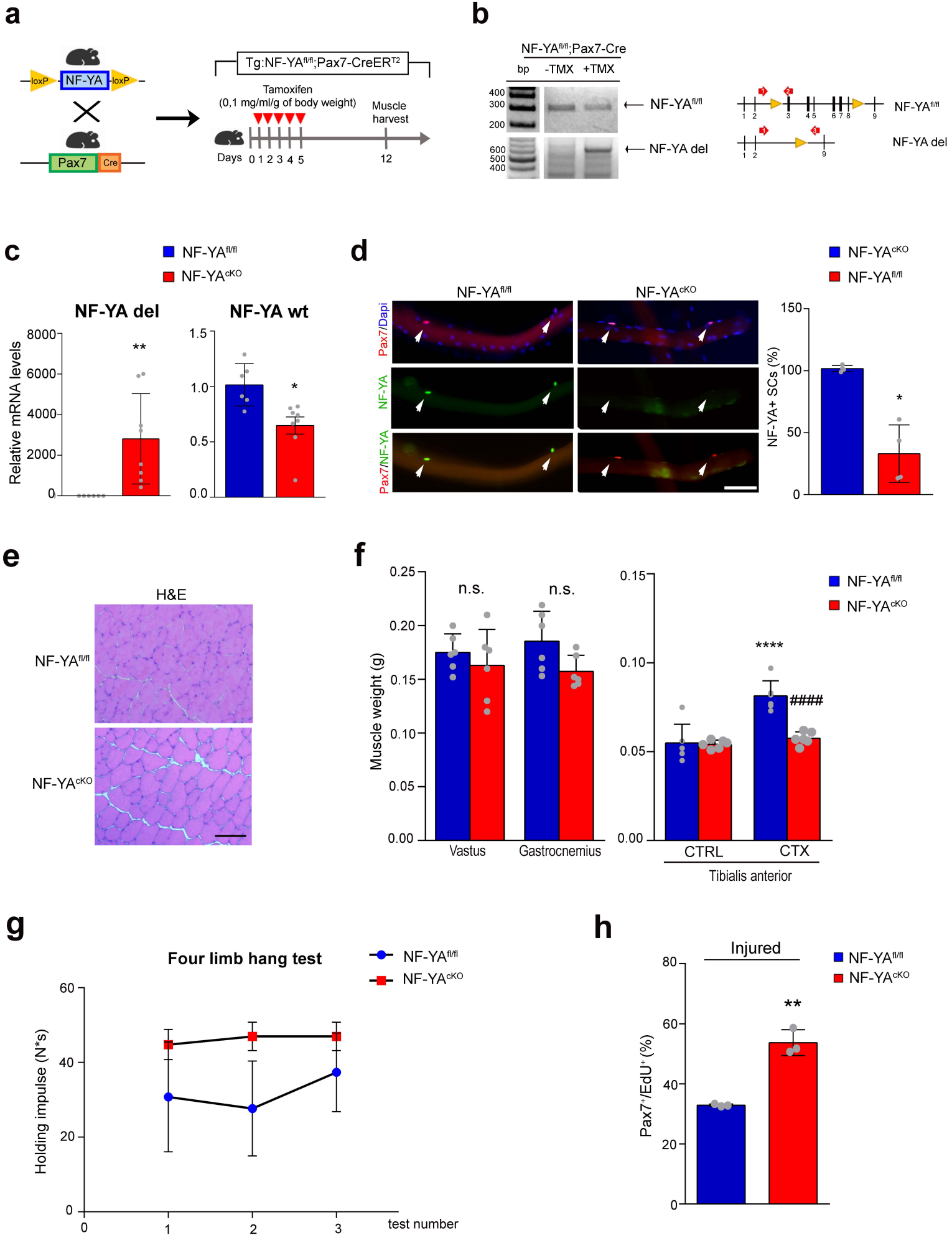
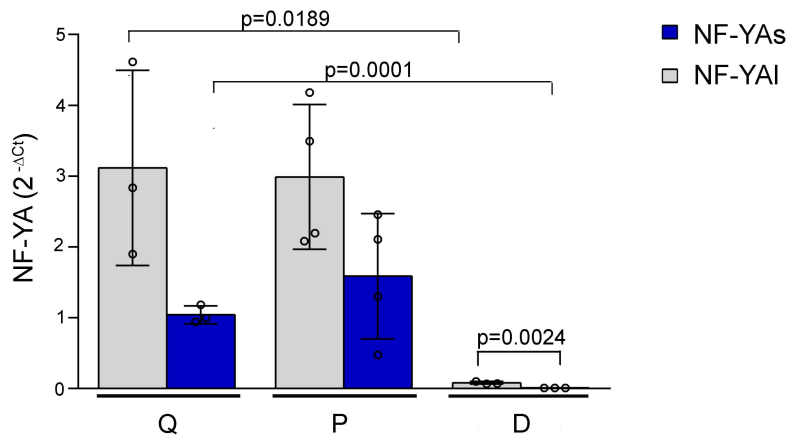


The transcription factor NF-Y participates to stem cell fate decision and regeneration in adult skeletal muscle

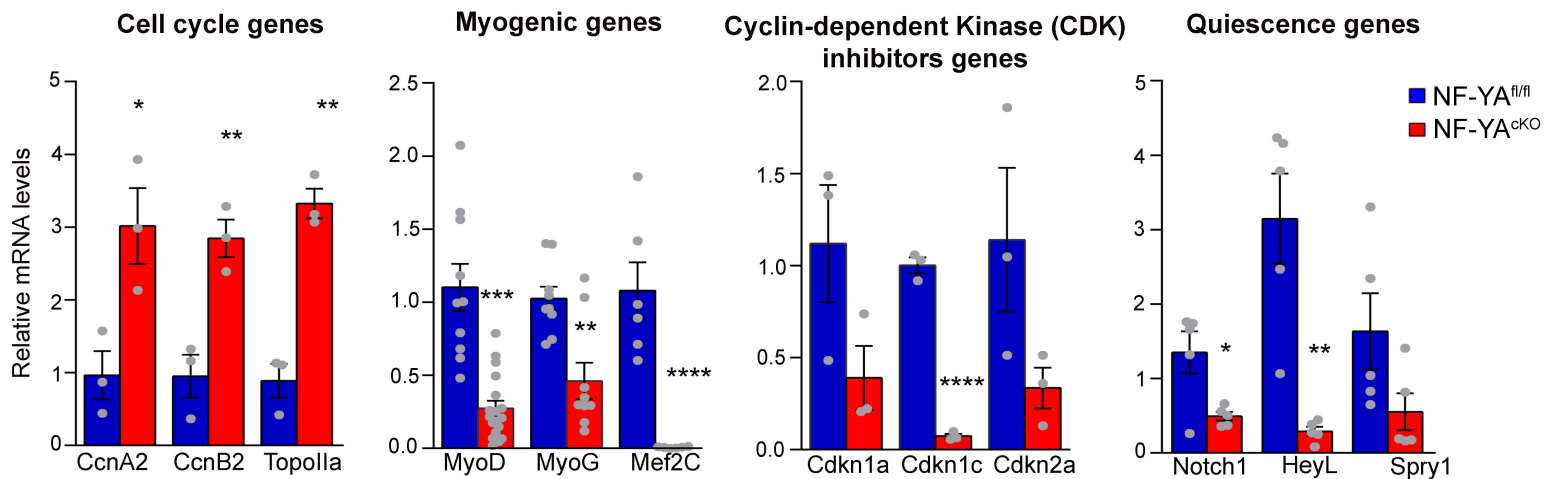
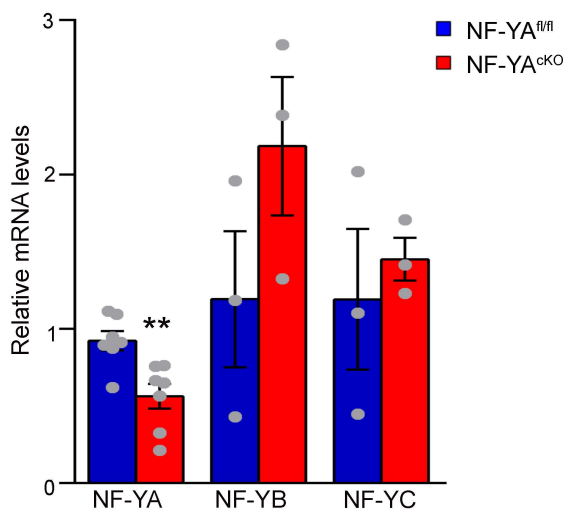
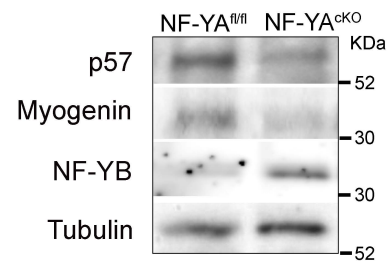
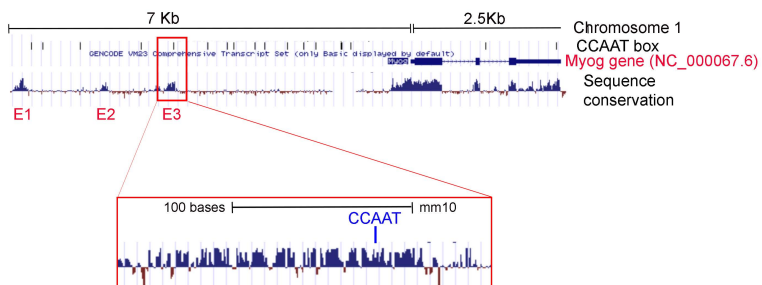
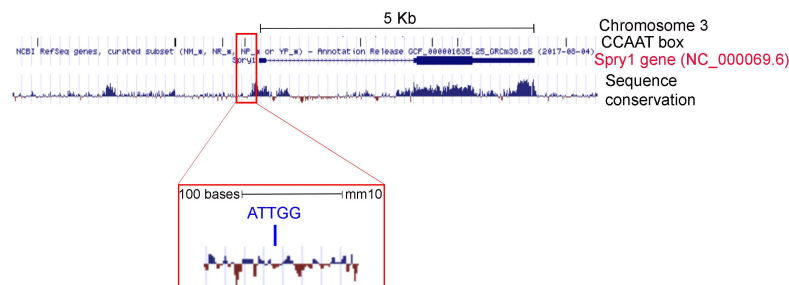
Supplementary Information



Supplementary Fig.1: Characterization of NF-YA^{ckO} mouse model. (a) Schematic representation of mice crossed to obtain NF-YA^{fl/fl};Pax7-CreER^{T2} F1 generation and Tamoxifen administration to induce NF-YA deletion in NF-YA^{ckO} mice. (b) Left panel: PCR analysis of genomic DNA from NF-YA^{fl/fl};Pax7Cre Tamoxifen-treated and untreated mice. The arrows indicate NF-YA^{fl/fl} amplicon, generated by primers 1 and 2, and NF-YA deleted (del) amplicon, generated by primers 1 and 3. PCR analysis was tested for each animal used for subsequent analyses. Right panel: Schematic representation of NF-YA coding gene, loxP elements (yellow triangles) and primers used for PCR analysis (red arrows). (c) qRT-PCR analysis of NF-YA deleted (del) (p=0.0100) and wt (p=0.0066) transcripts in SCs isolated from NF-YA^{ckO} versus NF-YA^{fl/fl} mice, arbitrarily set at 1. Data represent mean ± s.d. (two-tailed unpaired t-test: *p<0.05, **p< 0.01; n=3 mice). (d) Left panel: Representative immunofluorescence images of NF-YA and Pax7 in EDL myofibers at day 0 (d0) following isolation from NF-YA^{fl/fl} and NF-YA^{ckO} mice. Right panel: Quantification of NF-YA+ SCs identified by Pax7 positivity in NF-YA^{ckO} versus NF-YA^{fl/fl} myofibers, arbitrarily set at 100%. Data represent mean ± s.d. (two-tailed unpaired t-test: p=0.011; *p<0.05; n=4 mice). (e) Representative images of NF-YA^{ckO} and NF-YA^{fl/fl} TA cross-sections stained with H&E. Scale bar: 50 μm. (f) Left panel: weight of vastus and gastrocnemius uninjured muscles of NF-YA^{ckO} and NF-YA^{fl/fl} mice. Right panel: weight of tibialis anterior muscle after 60 days from CTX injury of NF-YA^{ckO} and NF-YA^{fl/fl} mice. Data represent means ± s.d. (two-tailed unpaired t-test: *p<0.05, **p< 0.01; n=6 mice). (g) Four limb grip test performed on adult NF-YA^{ckO} and NF-YA^{fl/fl} mice 60 days after CTX injury. Three independent tests were performed. Data represent means ± s.d. (two-tailed unpaired t-test; n.s.= not significant. n=5 mice). (h) Percentage of EdU+/Pax7+ cells in TA muscle sections following 5 days from CTX-injury. Data represent mean ± s.d. (two-tailed unpaired t-test: **p<0.01; n=3 mice).



Supplementary Fig. 2: Transcriptional analysis of enriched SC-cultures. mRNA levels of NF-YA isoforms (NF-YAI and NF-YAs) in Pax3/GFP+ sorted SCs quiescent (Q), proliferating (P) and differentiating (D). Data represent mean \pm s.d. (two-tailed unpaired t-test: p values are shown in figure; n=3 mice).

a**b****c****d****e**

Supplementary Fig. 3: Validation of transcriptional modulation of direct target genes in NF-YA^{CKO} mouse. (a) qRT-PCR analysis of the indicated genes in NF-YA^{fl/fl} and NF-YA^{CKO} SCs. Data represent mean \pm s.d. (two-tailed unpaired t-test: *CcnA2* p=0.0290, *CcnB2* p=0.0085, *Topolla* p=0.0014, *MyoD* p=0.0003, *MyoG* p=0.0016, *Mef2C* p<0.0001, *Cdkn1c* p<0.0001, *Notch* p=0.0179, *HeyL* p=0.0016; *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001, n=5 mice). (b) Relative mRNA levels measured by qRT-PCR of NF-YA, NF-YB and NF-YC genes in NF-YA^{fl/fl} and NF-YA^{CKO} SCs. Data represent mean \pm s.d. (two-tailed unpaired t-test: *NF-YA* p=0.0041; **p<0.05, n=5 mice). (c) Western blot of whole extracts from SCs isolated from NF-YA^{fl/fl} and NF-YA^{CKO} mice with anti-p57, anti-Myogenin and anti-NF-YB antibodies. Tubulin expression was used as loading control. n= 3 experiments. (d) Schematic representation (UCSC Genome browser) of *Myogenin* gene and upstream enhancer elements (E1, E2, E3). The enlargement shows the NF-Y binding element (CCAAT) within the E3 region. (e) Schematic representation (UCSC Genome browser) of *Sprouty1* gene and upstream promoter region. The enlargement shows the NF-Y binding element (CCAAT) at about 200 bp upstream of TSS.

Supplementary Tables

Supplementary Table 1: Sequence of each primer used for mice genotyping.

Genomic qPCR	FORWARD	REVERSE
<i>NF-YA^{fl/fl}</i>	GTAAGTCAGGCTCCAGGG	AGGCAAGGCAGATTTAGGAAGGTC
<i>NF-YA^{del}</i>	GTAAGTCAGGCTCCAGGG	GGTTGTCAGGATGTTCCGACG
<i>Pax7-Cre^{ERT2}</i>	CCACACCTCCCCCTGAACCTGAAACATAAA	GAATCCCCGGGGAGTCGCATCCTGCGG

Supplementary Table 2: Sequence of each primer used in real time qPCR for mRNA analysis.

RT-qPCR	FORWARD	REVERSE
<i>Rps15</i>	ACCTACAAACCCGTGAAGCA	CCAGGGACCAAACCAGTC
<i>NF-YA wt</i>	ACGAAGGAAATACCTCCATGAGTC	CTTCCCCACGTTCCGT
<i>NF-YA deleted</i>	CAGACCCTCCAGGTAGATCCAA	TGTGGTTAGGAACTCGGATGA
<i>Pax7</i>	GCTACCAGTACAGCCAGTATG	GTCACTAAGCATGGGTAGATG
<i>eMyHC</i>	TTGATGCCAAAACCTACTGCT	GGGTCCTGCTGTCTTCTGTC
<i>Myf5</i>	CACCACCAACCCTAACCAGA	GTTCTCCACCTGTTCCCTCA
<i>Myogenin</i>	GAGCCCCACTTCTATGATGG	GTCCCCAGTCCCTTTTCTTC
<i>Mef2c</i>	AGTACACCGAGTACAACGAGC	GCCTGTGTTACCTGCACTTGG
<i>Ccnb2</i>	TCTCTGATGCTCTGCTCTGC	CCGAAACTTGGAATGGACTT
<i>Ccna2</i>	CAGAACTCATTGGCTCTCA	GCCAAGGGAAAAGGAAGAAG
<i>Topo-IIa</i>	ACGGAATGACAAGCGAGAAG	AAACAACAACCGAGCCAAAG
<i>Cdkn1a</i>	CAAGGAGACCCCAAAGTCC	GAGGCAGATTTCTATCACTCC
<i>Cdkn1c</i>	TCCACCTCCATCCACTGC	AGAACCGCTGGGACTTAAC
<i>Cdkn2a</i>	CGCTTTTGTTCGGTTTTGTT	TCCAGGGGCTTATGATTCTG
<i>NF-YA</i>	ACAGTATCACCGCATCCTTAAGA	CCTTCGTTCCCTTTGGGATTT
<i>NF-YB</i>	AAGCGGAAGACAATCAATGG	ATCTGTGGCGGAGACTGCT
<i>NF-YC</i>	GCTACCAATGCCAACAGAT	CCCTCAGTCTCCAGTCACCT
<i>Mre11a</i>	CTTATCCGACTACGGGTGGA	TTTCCCTTTTGTCCCTGTG
<i>Brca1</i>	GAAACACGCCAAATGTCTGA	GATACGCTGGTGCTCTCCTC
<i>Rad51</i>	GGTTAGAGCAGTGTGGCATAA	TAGTTCCTTCTTCGGTGCATAAG
<i>Notch1</i>	GCAACTGTCTCTGCCATATAC	GTCTTCAGACTCCTTGCATACC
<i>HeyL</i>	CATCACTCCCTGAAGACGAAAG	GGAAGGGTTGTAGCCTTAGATG
<i>Spry1</i>	CACACTTCGCTAGTGGTGATT	TGGTCTAGGGACAGAATCGTAG
<i>Birc5</i>	CCGAGAACGAGCCTGATTT	GAGTGCTTTCTATGCTCCTCTATC
<i>Bcl2l2</i>	GTGGGTAGAAGCTTTGGTAGTT	GCTGGATAGAGAGACCCTAGAA

Supplementary Table 3: Sequence of each primer used in real time qPCR for ChIP analysis.

<i>RT-qPCR</i>	<i>FORWARD</i>	<i>REVERSE</i>
<i>Chrm7</i>	ATAAAGGCTTGGCACTCGTC	CAGTTCCTTTGCTTGATCC
<i>Ccnb1</i>	CAGGCATAGAGCCTGACCTC	GTCTGCCGGGCTTAGGTTTA
<i>Ccnb2</i>	AAATACAAGCCAGCCAATCAA	GACGAGGCACAGCCACTC
<i>Topo-IIa</i>	CCTTCCTCATTGGTCAGATTTT	GACTCGCTCTCATTGGCTCT
<i>Cdkn1c promoter</i>	CCAAGCTGGACAGGACAAG	AAGCGTTCCATCGCTGTT
<i>Cdkn1c 5'UTR</i>	GATCTGACCTCAGACCCAATTC	TGCTCAGAGACCTGCTCA
<i>Myogenin enhancer</i>	CAGGTCAGAGCTGATGGAT	CACTAGCTGCCCTCTGATG
<i>Spry1</i>	AAACTCAACTCTAAGGGTGGCT	GAGGAGCGGGCATTCCA