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Supplemental information

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Extracellular PKM2 Facilitates Organ-tissue Fibrosis Progression

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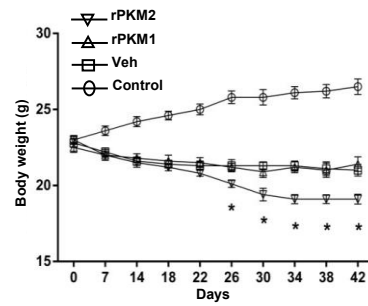
Supplementary Figure S1

A

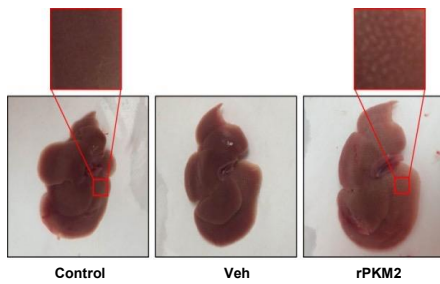
Serum PKM2 levels in liver disease patients

Serum PKM2 ng/ml	Range	Mean
Normal (n=10)	0.08 – 0.35	0.24
Viral hepatitis (n=10)	0.89 – 2.07	1.59
Cirrhosis (n=10)	1.8 – 7.6	3.9

B



C

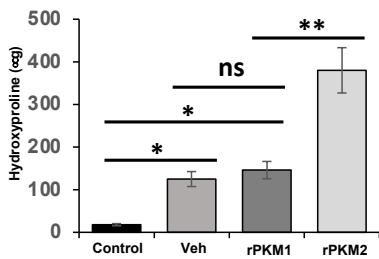


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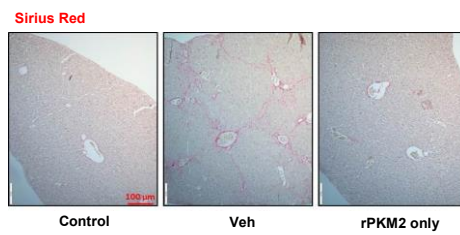
Serum markers in TAA fibrosis mice

	ALT (U/L)	AST (U/L)
Control	34.2±7.3	29.7±6.4
Veh	93.5±27	97.7±44
rPKM1	92.4±23	98.4±36
rPKM2	169±22.8	207.9±51

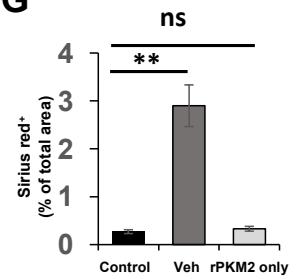
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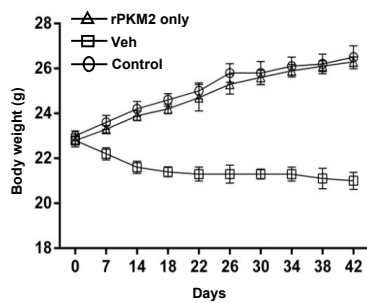
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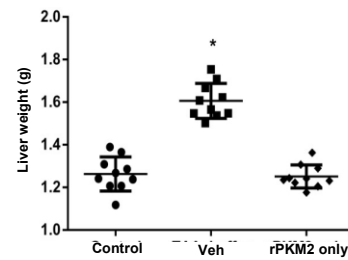
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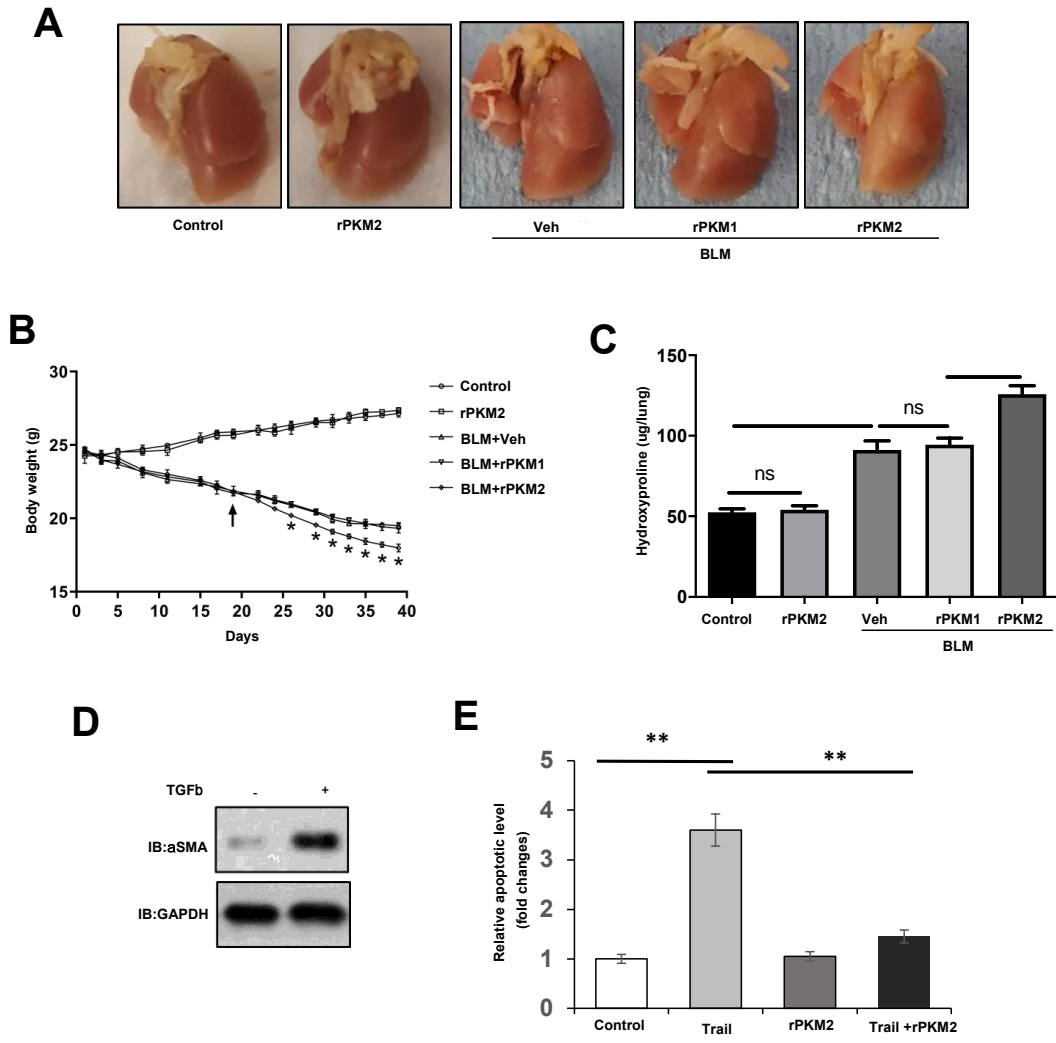


I



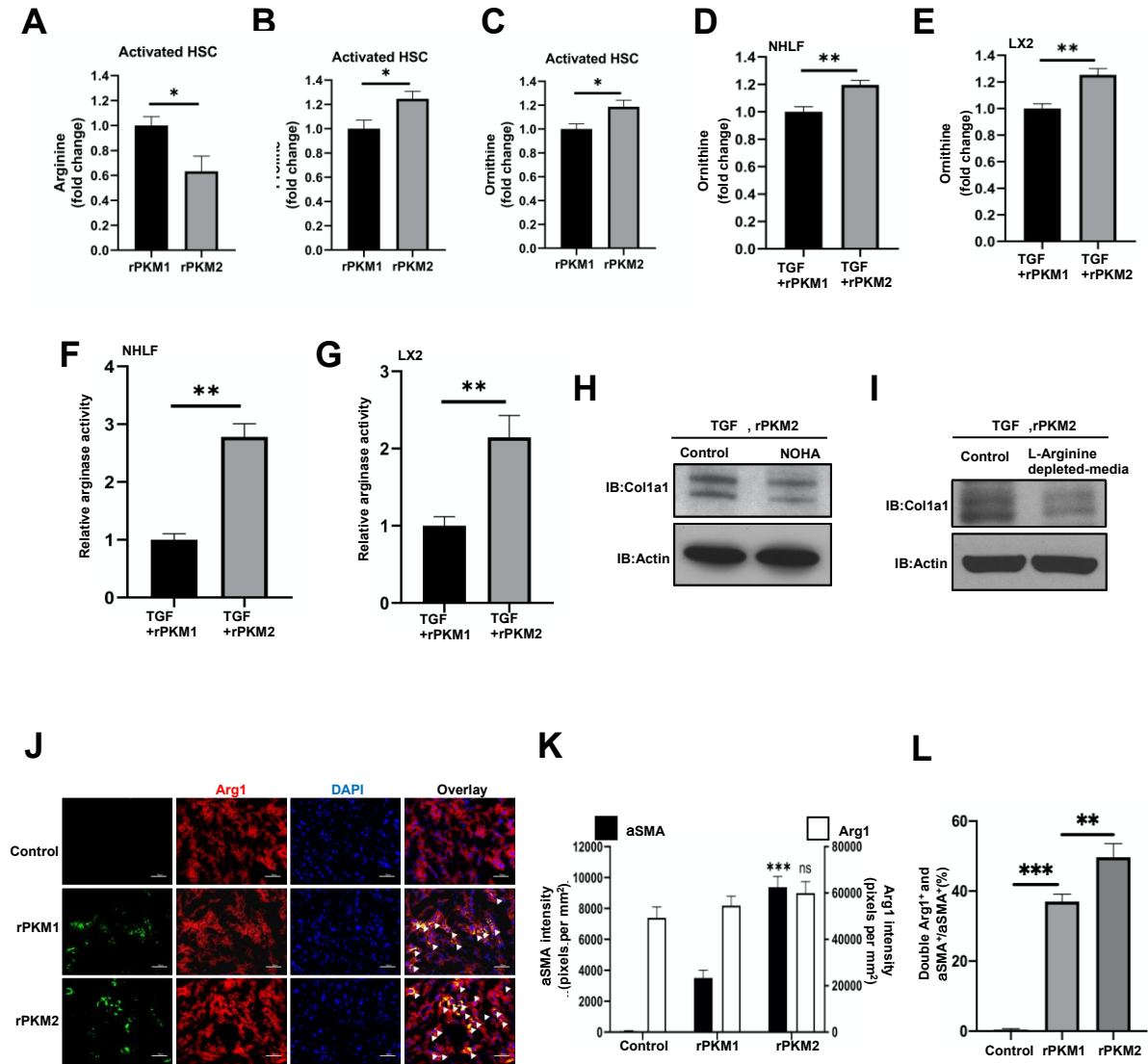
Supplementary Figure 1 EcPKM2 facilitates liver fibrosis progression. Related to Figure 2.

(A) PKM2 levels (ng/ml) in serum of healthy individuals and liver disease patients were measured by ELISA. The range is low and high PKM2 levels in serum. Means is the mean value of PKM2 levels calculated from 10 healthy individuals and patients (n=10). Each patient sample was independently measured five times in the ELISA assay. (B) - (E), liver fibrosis was induced by TAA/alcohol and the animals were subsequently treated with addition of rPKM1, rPKM2 or vehicle into TAA as shown in Fig. 2A. (B) Body weight changes during treatment course. Arrow indicates addition of rPKM2/rPKM1/vehicle to the TAA fibrosis induction. (C) Images of representative collected livers with enlarged call-out to show surface features. (D) Serum levels of ALT and AST in mice treated with indicated agents were measured by a commercial service (CPath). (E) Collagen levels in fibrotic livers were measured by hydroxyproline assay and presented as hydroxyproline (μg). (F) - (I) Mice were treated with TAA/alcohol plus vehicle as shown in Fig. 2A (Veh) or rPKM2 without TAA/alcohol (rPKM2 only). Control is the animal without treatment. (F), (G) Representative images (F) and Quantitation (G) of Sirius red staining of liver sections from mice with indicated treatment. (H) Body weight changes during treatment course. (I) Liver weight of the animals at end point of the experiments. Error bars in (B), (E), (G), and (H) represent mean \pm S.E.M. Scale bars, 100 μm . * P <0.05, ** P <0.01. ns: statistically no significance.



Supplementary Figure 2 EcPKM2 facilitates liver fibrosis progression. Related to Figure 2.

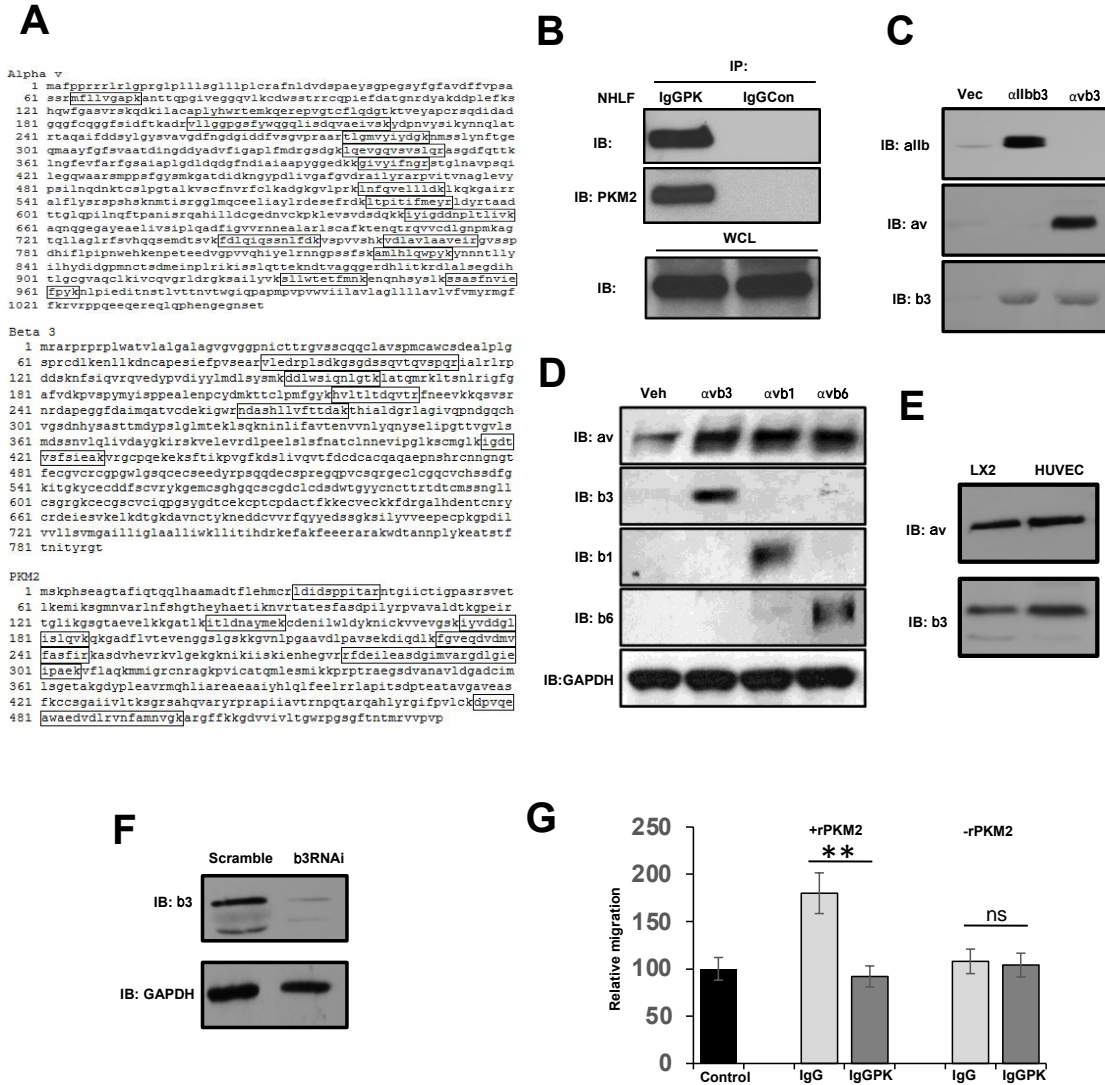
(A) - (C), lung fibrosis was induced by bleomycin and the animals were subsequently treated with addition of rPKM1, rPKM2 or vehicle into bleomycin as shown in Fig. 2G. (A) Representative images of collected lungs from animals that underwent indicated treatment. (B) Body weight changes during treatment course. Arrow indicates addition of rPKM2/rPKM1/buffer to the bleomycin fibrosis induction. (C) Collagen levels in fibrotic lungs were measured by hydroxyproline assay and presented as hydroxyproline (μg). (D) Cellular levels of αSMA in human primary HSC cells that were treated (+) or untreated (-) with TGF β were analyzed by immunoblot (IB: αSMA). Immunoblot of GAPDH (IB:GAPDH) is a loading control. (E) Apoptosis of TGF β treated LX-2 cells that underwent additional indicated treatment was measured by Annexin v-FACS assay. The apoptosis is presented as relative apoptosis by defining the untreated LX-2 cells as reference 1. Error bars in (B), (C), and (E) represent mean \pm S.E.M. * P <0.05, ** P <0.01. ns: statistically no significance.



Supplementary Figure 3 EcPKM2 facilitates collagen synthesis in myofibroblasts by upregulating Arg-1. Related to Figure 4.

(A), (B) Relative abundance of Arginine (A) and Proline (B) in activated hHSC cells were analyzed by HPLC-MS. The cells were treated by TGF β + rPKM1 (black bars) or TGF β + rPKM2 (grey bars). The amino acid levels in the cells that were treated with TGF β + rPKM1 are set as reference 1. (C) - (E) Cellular ornithine levels in activated hHSC (C), NHFL (D) and LX-2 (E) cells were analyzed using a commercially available ornithine kit. (F), (G) Arginase activities in extracts of NHFL (F) and LX-2 (G) cells were measured using a commercially available arginase activity assay kit. (H), (I) Cellular levels of collagen 1A1 in NHFL cells were analyzed by immunoblot (IB:Coll1a1). In (A) – (I), the cells were treated with TGF β plus rPKM1 or rPKM2. In (H), the cells were treated with Arg-1 inhibitor (MOHA). In (I), the cells were cultured in L-Arginine-depleted media. In (H) and (I), control is the cells without MOHA treatment (in H) and without L-Arg depletion (in I). Immunoblot of Actin (IB:Actin) is a loading control. (J) – (L) Representative images of immunofluorescence (IF) staining (J) and quantitation of α SMA and Arg1 staining (K) and α SMA and Arg1 double staining over α SMA staining (%) (L) in sections from liver of mice that were treated by the indicated agents. In (J), green is IF staining of α SMA. red is Arg1 staining. Blue is DAPI staining. Arrows indicate α SMA and Arg1 double staining. In (K), quantities of α SMA and Arg1 are presented as staining intensity pixels per mm². Error bars in (A), (B), (C), (D), (E), (F), (G), (K), and (L) represent mean \pm S.E.M. Scale bars, 100 μ m. * P <0.05, ** P <0.01, *** P <0.001. ns: statistically no significance.

Supplementary Figure S4



Supplementary Figure 4 EcPKM2 interacts with integrin $\alpha_v\beta_3$ and activates the integrin signaling.

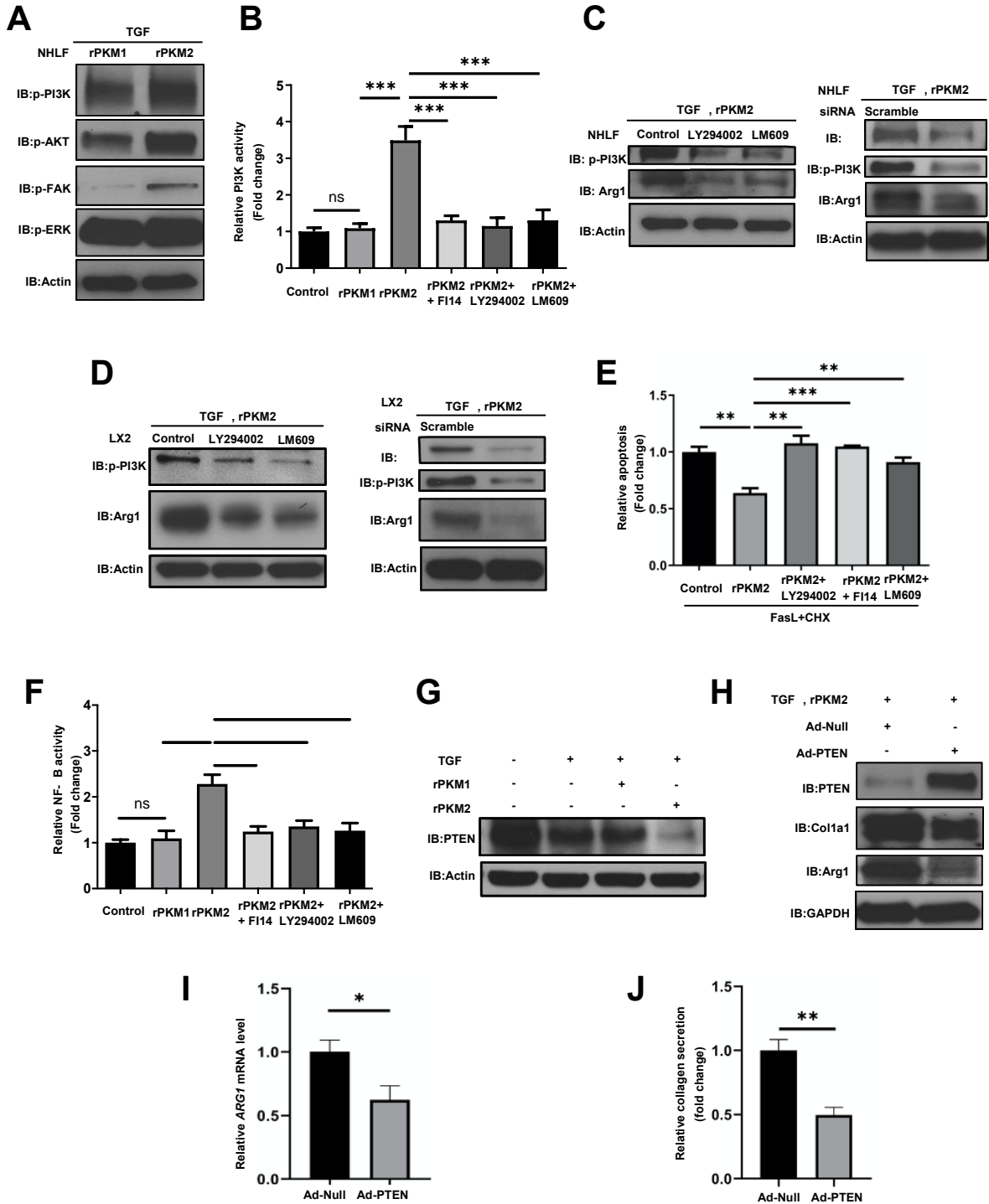
Related to Figure 5.

(A) The peptide fragments (boxed) from trypsin digestion of the His-tag pulled-down crosslinking band of His-rPKM2-LX-2 crosslinks (see Figure 5A) and analyzed by MALDI-tof/tof that match the aa sequence of integrin α_v , β_3 , and PKM2. **(B)** Co-immunoprecipitation of integrin β_3 with PKM2 (IP:IgGPK) in extracts of NHLF cells treated with TGF β was examined by immunoblot of integrin β_3 (IB: β_3). Immunoblot of PKM2 (IB:PKM2) indicates amounts of PKM2 that are precipitated down. Immunoprecipitation using IgG purified from pre-immune serum (IP:IgGCon) is control immunoprecipitation. Immunoblot of integrin β_3 (IB: β_3) in whole cell lysate (WCL) is a control indicating equal amounts of cell extracts used in the IPs. **(C)** Expression of integrin α_{IIb} , α_v , and β_3 (IB: α_v , IB: α_{IIb} , IB: β_3) in CHO cells that exogenously express integrin $\alpha_v\beta_3$ and $\alpha_{IIb}\beta_3$ pairs was probed by immunoblot analyses. **(D)** Expression of integrin α_v , β_1 , β_3 , and β_6 (IB: α_v , IB: β_3 , IB: β_1 , IB: β_6) in CHO cells that exogenously express integrin $\alpha_v\beta_3$, $\alpha_v\beta_1$, and $\alpha_v\beta_6$ pairs was probed by immunoblot analyses. In (C) and (D), Veh the cells were transfected with empty vector. **(E)** Expression of integrin α_v (IB: α_v) and β_3 (IB: β_3) in LX-2 and HUVEC cells was probed by immunoblot analyses. **(F)** Expression of integrin β_3 (IB: β_3) in LX-2 cells in which integrin β_3 was knocked down by β_3 RNAi or scrambled RNAi was probed by immunoblot analyses. Immunoblot of GAPDH in (D) and (F) is a loading control. **(G)** Migration of LX-2 cells treated with TGF β in the presence of rPKM2 (+rPKM2) absence of rPKM2 (-rPKM2) and with addition of antibodies IgGPK or IgGCon in the culture media were analyzed. The migration was assayed in plats with vitronectin (VN) coating. The cell migrations are presented as relative migration by defining the cell migration of control in the absence of rPKM2 and without antibody treatment as

100. Error bars in (A), (B), (C), (D), (H) and (I) represent mean \pm S.E.M. Scale bars, 100 μ m.

** $P < 0.01$, *** $P < 0.001$. ns: statistically no significance.

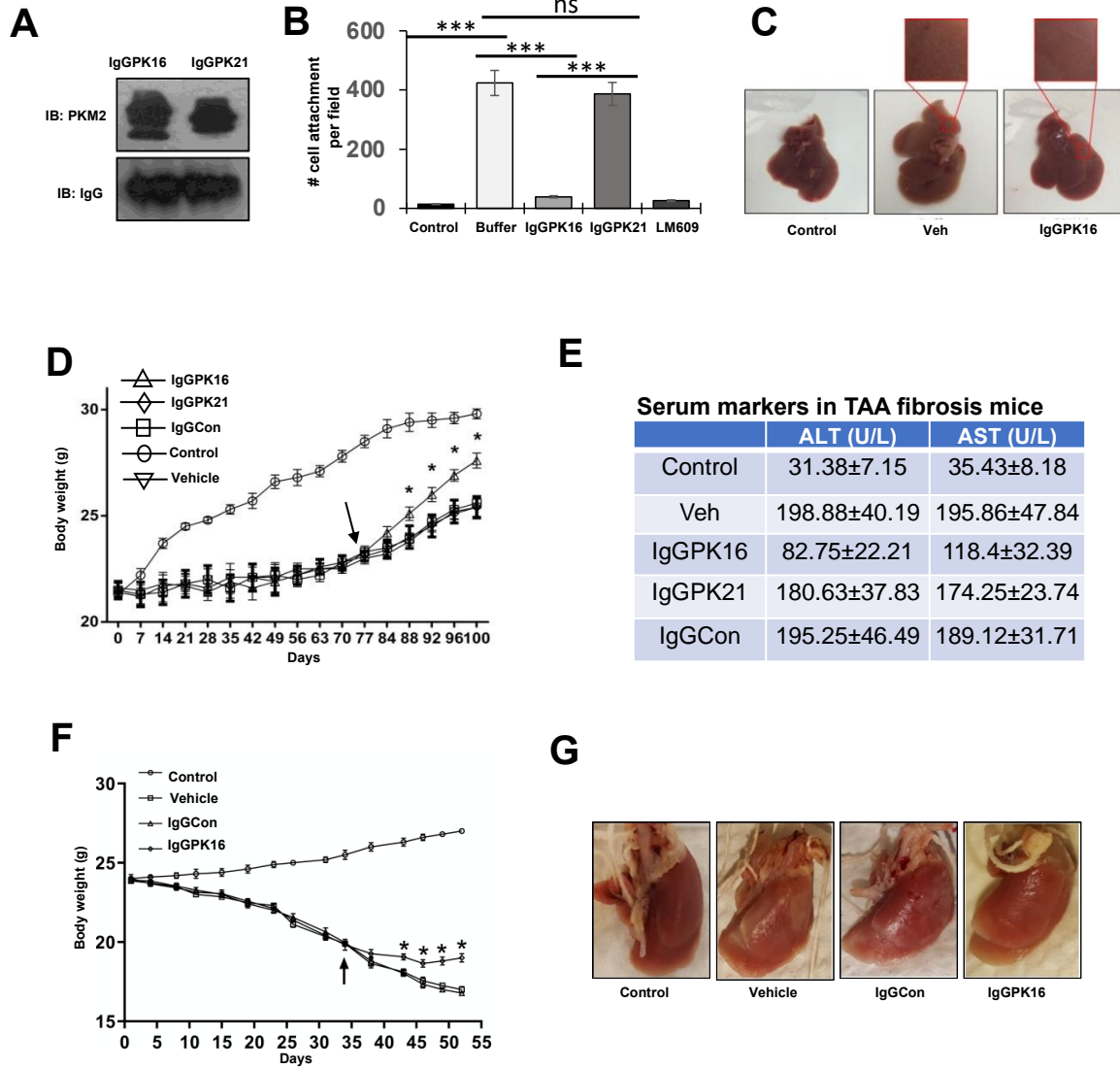
Supplementary Figure S5



Supplementary Figure 5 EcPKM2 Activates FAK-PI3K signaling axis. Related to Figure 6.

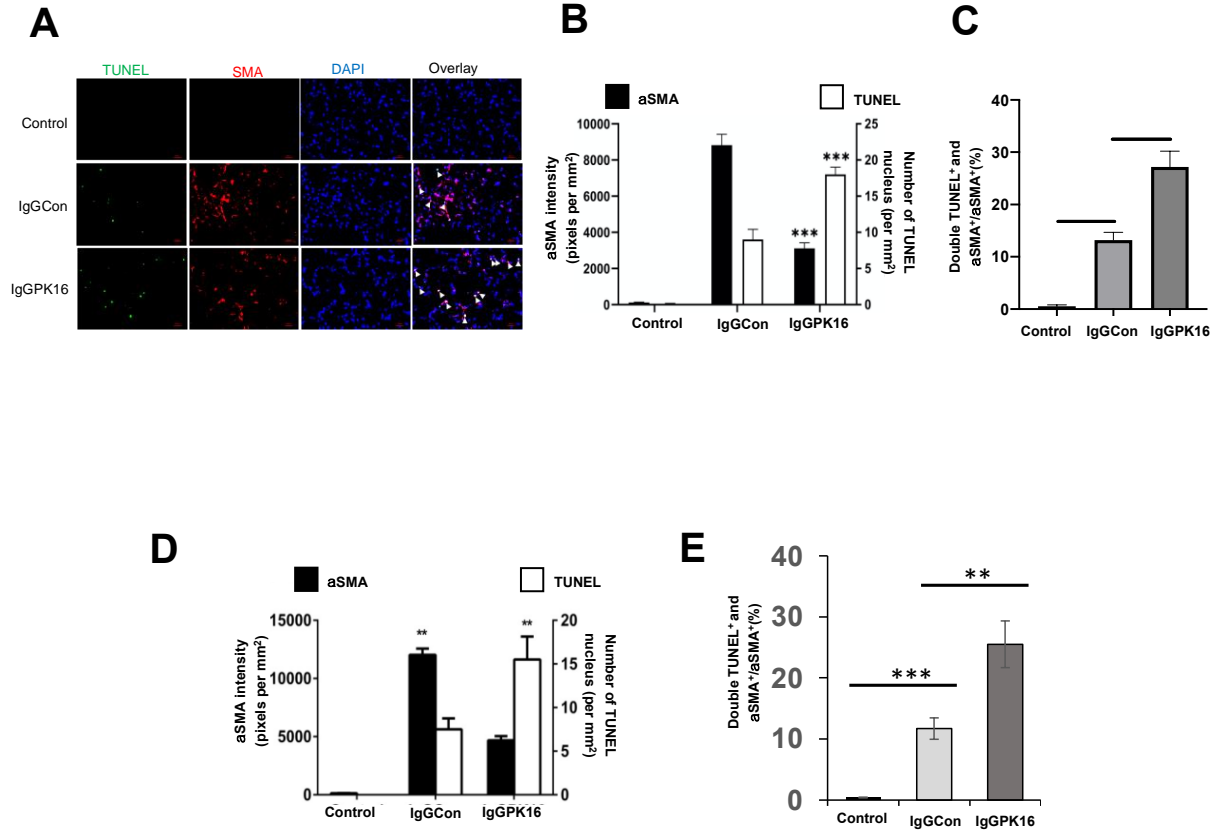
(A) Cellular levels of phospho-PI3K (IB:p-PI3K), phospho-AKT (IB:p-AKT), phospho-FAK (IB:p-FAK), and phospho-ERK (IB:p-ERK) in NHLF cells were analyzed by immunoblot. The cells were treated by TGF β plus rPKM1 or rPKM2. (B) Activation of PI3K in NHLF cells in the presence of indicated agents was measured by ELISA PI3K kit. The PI3K activities are presented as relative activity by defining the activities of PI3K in buffer treated cells as 1 (Control). (C), (D) Cellular levels of phospho-PI3K (IB:p-PI3K) and Arg-1 (IB:Arg1) in NHLF (C) and LX-2 (D) cells were analyzed by immunoblot. The cells were treated by TGF β and rPKM2. (Left) The TGF β and rPKM2 treated cells were further treated by PI3K inhibitor (LY294002), antibody LM609, or no treatment (Control). (Right) Integrin β_3 was knocked down (β_3) or knockdown by scrambled siRNA (Scramble). Immunoblot of integrin β_3 (IB: β_3) indicate cellular level of the integrin. (E) Apoptosis of NHLF cells under treatment of indicated agents were measured by apoptosis kit. Cell apoptosis is presented as Relative apoptosis by defining apoptosis of cell treated with buffer (control) as 1. (F) NF-kB activity in NHLF cells in the presence of indicated agents was measured using transcription assay kit. The NF-kB activity is present as Relative NF-kB activity by defining the measured NF-kB activity in buffer treated NHLF cells as 1. (G) Cellular levels of PTEN (IB:PTEN) in NHLF cells treated with indicated agents were analyzed by immunoblot. (H) Cellular levels of PTEN (IB:PTEN), collagen 1A1 (IB:collagen1a1), and Arg-1 (IB:Arg1) in NHLF cells were analyzed by immunoblot. The cells were treated with TGF β and rPKM2. PTEN was exogenously expressed in the cells using adenoviral vector (Ad-PTEN). Ad-Null is empty viral vector. (I) Cellular levels of Arg-1 mRNA in NHLF cells were analyzed by qRT-PCR. (J) Secreted collagen in culture medium of NHLF cells were analyzed by precipitation (as in Fig. 4 C&D). The cells were treated with TGF β and rPKM2. PTEN was exogenously expressed in the

cells using adenoviral vector. Immunoblots of Actin (IB:Actin) in (A), (C), (D), and (G) and GAPDH (IB:GAPDH) in (H) and are loading controls. Error bars in (B), (E), (F), (I), and (J) represent mean \pm S.E.M. * P <0.05, ** P <0.01, *** P <0.001. ns: statistically no significance.



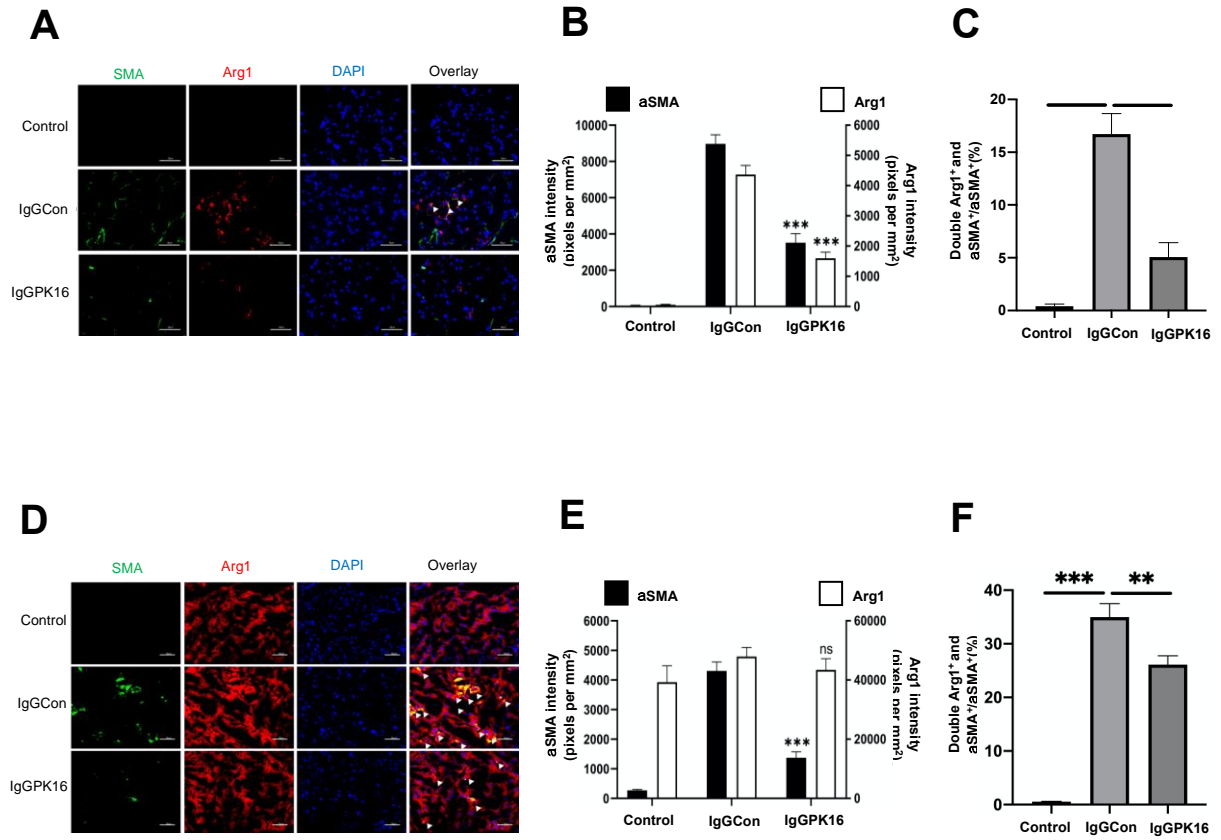
Supplementary Figure 6 Antibody against PKM2 reversed organ/tissue fibrosis. Related to Figure 7.

(A) Recognition of rPKM2 (IB:PKM2) by two clones of rabbit monoclonal antibody IgGPK21 and IgGPK16 is illustrated by immunoblot. IB:IgG is a loading controls to show amount of purified IgG used in each immunoblot. (B) Attachment of LX-2 cells to rPKM2 coated plate in the presence of buffer, PKM2 antibody clone 16 (IgGPK16), PKM2 antibody clone 21 (IgGPK21), and LM609 (α v β 3Ab). The cells were treated with TGF β prior to attachment assay. The attachment is presented as total number of cells attached to the plate per view field. Control is the cell attachment to the plate without rPKM2 coat. (C) - (E) Liver fibrosis is induced by TAA/alcohol and the animals were subsequently treated with indicated agents as shown in figure 7A. (C) Representative images of collected livers with enlarged call-out to show surface features. (D) Body weight changes during treatment course. Arrow indicates starting time point of antibody or vehicle treatment. (E) Serum levels of ALT and AST in blood of mice treated with indicated agents were measured by a commercial service (CPath). (F), (G) Lung fibrosis was induced by bleomycin and the animals were subsequently treated with IgGPK16, IgGCon, or vehicle as shown in Fig. 7G. (F) Body weight changes during treatment course. Arrow indicates starting time point of antibody or vehicle treatment. (G) Representative images of collected lungs from animals that underwent indicated treatment. Control in (C) - (G) is animals without fibrosis induction and subsequent treatment. Error bars in (B), (D), and (E) represent mean \pm S.E.M. * P <0.05, *** P <0.001. ns: statistically no significance.



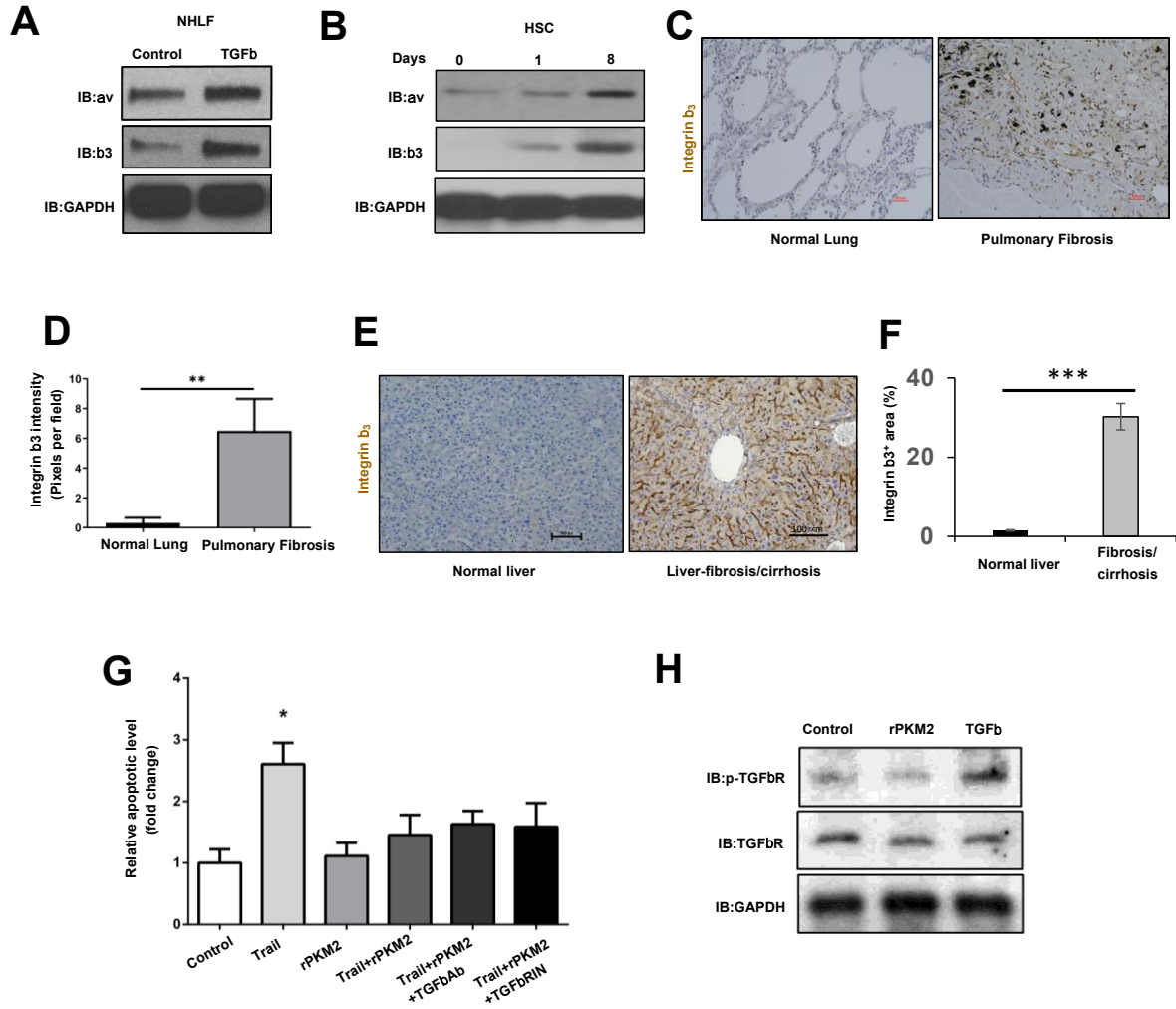
Supplementary Figure 7 PKM2 antibody increases myofibroblasts apoptosis in fibrotic lung and liver. Related to Figure 3 and Figure 7.

(A) – (C) Representative images of immunofluorescence (IF) staining (A) and quantitation of α SMA and TUNEL staining (B) and α SMA and TUNEL double staining over α SMA staining (%) (C) in sections from lung of mice that were treated by the indicated agents. In (A), Red is IF staining of α SMA. Green is TUNEL staining. Blue is DAPI staining. Arrows in overlay indicate examples of α SMA and TUNEL double staining. (D), (E) Quantitation of α SMA and TUNEL staining (D) and α SMA and TUNEL double staining over α SMA staining (%) (E) in sections from liver of mice that were treated by the indicated agents. In (B) and (D), quantity of α SMA is presented as staining intensity pixels per mm^2 , quantity of TUNEL staining is presented as number of positive nucleus per mm^2 . Control in (A) – (E) is animals without fibrosis induction and subsequent treatment. Error bars in (B), (D), and (E) represent mean \pm S.E.M. ** $P < 0.01$, *** $P < 0.001$.



Supplementary Figure 8 PKM2 antibody decreases Arg-1 expression in myofibroblasts in fibrotic lung and liver. Related to Figure 4 and Figure 7.

(A) – (C) Representative images of immunofluorescence (IF) staining (A) and quantitation of α SMA and Arg1 staining (B) and α SMA and Arg1 double staining over α SMA staining (%) (C) in sections from lung of mice that were treated by the indicated agents. (D) – (F) Representative images of immunofluorescence (IF) staining (D) and quantitation of α SMA and Arg1 staining (E) and α SMA and Arg1 double staining over α SMA staining (%) (F) in sections from liver of mice that were treated by the indicated agents. In (A) and (D), green is IF staining of α SMA. red is Arg1 staining. Blue is DAPI staining. In (B) and (E), quantities of α SMA and Arg1 are presented as staining intensity pixels per mm². Control in (A) – (F) is animals without fibrosis induction and subsequent treatment. Error bars in (B), (C), (E), and (F) represent mean \pm S.E.M. ** P <0.01, *** P <0.001. ns: statistically no significance.



Supplementary Figure 9 Myofibroblasts express integrin $\alpha_v\beta_3$. EcPKM2 does not activate TGF β signaling. Related to Figure 5.

(A), (B) Cellular levels of integrin α_v (IB: α_v) β_3 (IB: β_3) in NHLF (A) and human primary HSC (B) cells were analyzed by immunoblot. The cells were treated with TGF β for 0 (control in (A)), 1, and 8 days before cell extracts preparation. **(C) – (F)** IHC staining of integrin β_3 in tissue array sections from lung (C and D) and liver (E and F) fibrosis patients. (C) and (E) Representative images of IHC staining of integrin β_3 . (D) and (F) Quantitation of integrin β_3 staining (integrin β_3 stain intensity (pixels per field) in (D) and integrin β_3^+ area (%) in (E)) of lung and liver diseases tissue array. Quantitation were calculated from randomly selected four view fields from each spot of the array that were analyzed. The integrin β_3 staining intensity and integrin β_3^+ area were calculation of all tissue spots in the array (Liver disease tissue array: Total 9 normal n=9, 15 viral hepatitis n=15, 40 cirrhosis liver n=40 spots. Lung disease tissue array: Total 7 normal n=7, 25 lung fibrosis n=25 spots). **(G)** Apoptosis of LX-2 cells under treatment of indicated agents were measured by apoptosis kit. TGF β Ab is a TGF β neutralizing antibody. TGF β RIN is an inhibitor that inhibits TGF β receptor activity. Cell apoptosis is presented as Relative apoptosis by defining apoptosis of cell treated with buffer (control) as 1. **(H)** Cellular levels of phospho-TGF β R in LX-2 cells that were treated by the indicated agents were analyzed by the immunoblots (IB:p-TGF β R). Immunoblot of TGF β R (IB:TGF β R) is a loading control indicates the total TGF receptor levels. Control, the LX-2 cells treated by buffer. Immunoblot of GAPDH (IB:GAPDH) in (A), (B), and (H) is a loading control. Error bars in (D), (F), and (G) represent mean \pm S.E.M. * P <0.05, ** P <0.01, *** P <0.001