

Figure S1: Definition of ILC populations in the FGT.

FGTs were isolated from WT mice infected with *C. muridarum* for 30 days and leucocytes were analysed via immunostaining and subsequent flow cytometry.







Wt and *vav-bcl-2* mice were infected intravaginally with $5x10^5$ IFUs of *C. muridarum*. Single cell suspensions of the genital tracts or its parts were prepared and stained for flow cytometry. **A**, cells were isolated from the uterine horns and stained intracellularly for human (transgenic) Bcl-2. Bcl-2-expression of scatter-gated (see Fig. 1), lineage (CD3-CD5-CD19-Ly6G-TCR β -TCR γ δ -F4/80-Fc ϵ R1a)-negative cells (including all ILCs) is shown (31 days p.-i.; dark gray, wt, light gray, *vav-bcl-2*). **B-D**, absolute cell numbers of ILC2 (**B**), cNK (**C**) and trNK (**D**) cells were determined as in Fig. 1, 2 (**B**, total FGT, **C** and **D**, uterine horns and oviducts separately). Analyses were conducted on days 30 (**B**) or 31 (**C**, **D**). Data are from three mice per group.





Figure S3: Division of FGT parts.

Mouse FGTs were dissected into different parts. Following procedures were performed on FGT parts: **A** DNA isolation, **B** flow cytometry analysis and **C** RNA isolation.





qPCR-data showing absolute numbers of *C. muridarum* genome copies/mg tissue. FGTs of WT mice infected with $5x10^5$ IFU *C. muridarum* were removed at indicated time points and dissected into designated parts. DNA was extracted and qPCR performed. DNA from $1x10^7$ IFUs of *C. muridarum* was isolated, diluted and subsequently used to create a standard curve. Data show means/SEM of 2-14 mice. Significance between means was tested by Turkey's multiple comparison test (* p=0.05, ** p<0.01, **** p>0.001, **** p<0.0001, n.s., p>0.05).



Figure S5: Absolute numbers of ILCs in FGT uterine horns comparing WT and *Ncr1*^{GFP/GFP} mice upon infection with *C. muridarum*.

WT and *Ncr1*^{GFP/GFP} mice were infected intravaginally with $5x10^5$ IFUs of *C. muridarum*. FGTs were isolated at indicated time points and leucocytes of FGT uterine horns were analysed via immunostaining and subsequent flow cytometry. Absolute cell numbers were obtained including a defined number of reference beads. **A** cNK cells were defined as CD45⁺, Lin⁻, NK1.1⁺, Eomes⁺, CD49a⁻ cells. **B** trNK cells were defined as CD45⁺, Lin⁻, NK1.1⁺, Eomes⁺, CD49a⁻ cells. **B** trNK cells were defined as CD45⁺, Lin⁻, NK1.1⁺, Eomes⁻, CD49a⁺ cells. **D** ILC2 cells were defined as CD45⁺, Lin(CD3, CD5, CD19, TCR β , TCR $\gamma\delta$, F4/80, Fc κ R1 α , Ly6G)⁻, CD127⁺, ST2⁺, GATA3⁺ cells. Data show means/SEM of 5-9 mice. Significance between means was tested by unpaired t test (* p=0.05, n.s., p>0.05).

Figure S5



Figure S6



WT and *Ccr2^{-/-}* mice were infected intravaginally with 5x10⁵ IFUs of *C. muridarum*. FGTs were isolated at indicated time points and leucocytes of whole FGTs were analysed via immunostaining and subsequent flow cytometry. Absolute cell numbers were obtained including a defined number of reference beads. **A** Definition of myeloid populations in the FGT. **B** Monocytes were defined as CD45⁺, CD11b⁺, CD64⁺, Ly6C⁺, CD11c⁻ cells. **C** Neutrophils were defined as CD45⁺, CD11b⁺, CD64⁻, Ly6C⁺, Ly6C⁺, Ly6C⁺, CD11c⁻ cells. **C** Neutrophils were defined as CD45⁺, CD11b⁺, CD64⁻, Ly6C⁺, Ly6C⁺, Ly6C⁺, Ly6C⁺, CD11b⁺, CD64⁻, Ly6C⁺, Ly6C⁺, Ly6C⁺, CD11b⁺, CD64⁻, Ly6C⁺, Ly6C⁺, Ly6C⁺, CD11b⁺, CD64⁻, Ly6C⁺, Ly6C⁺, Ly6C⁺, Ly6C⁺, CD11b⁺, CD64⁻, Ly6C⁺, Ly6C⁺, Ly6C⁺, CD11b⁺, CD64⁻, Ly6C⁺, Ly6C





Figure S7: Correlation of other immune cell numbers and the corresponding oviduct weight.

WT mice were infected intravaginally with $5x10^5$ IFUs of *C. muridarum*. FGTs were isolated 30 dpi and leucocytes of FGT oviducts were analysed via immunostaining and subsequent flow cytometry. Data points represent the absolute number of cells in the oviduct which were obtained including a defined number of reference beads, and corresponding oviduct weights. **A** Neutrophils were defined as CD45⁺, CD11b⁺, CD64⁻, Ly6C⁺, Ly6G⁺ cells. **B** Monocytes were defined as CD45⁺, CD11b⁺, CD64⁺, Ly6C⁺, Ly6C⁺, Ly6C⁺, CD11c⁻ cells. **C** T cells were defined as CD45⁺, TCRβ⁺ cells. Significance was tested via the Pearson correlation coefficient "r" (*p=0.05; **p<0.01, n.s., p>0.05).





WT mice were infected intravaginally with $5x10^5$ IFUs of *C. muridarum*. FGTs were isolated 30 dpi and leucocytes of FGT uterine horns were analysed via immunostaining and subsequent flow cytometry. Data points represent the absolute number of ILCs in the uterine horns which were obtained including a defined number of reference beads, and corresponding uterine horn weights. **A** cNK cells were defined as CD45⁺, Lin(CD3, CD5, CD19, TCR β , TCR $\gamma\delta$, F4/80, Fc ϵ R1 α , Ly6G)⁻, NK1.1⁺, Eomes⁺, CD49a⁻ cells. **B** trNK cells were defined as CD45⁺, Lin⁻, NK1.1⁺, Eomes⁺, CD49a⁺ cells. **C** ILC1 cells were defined as CD45⁺, Lin⁻, NK1.1⁺, Eomes⁻, CD49a⁺ cells. **D** ILC2 cells were defined as CD45⁺, Lin⁻, CD127⁺, ST2⁺, GATA3⁺ cells. Significance was tested via the Pearson correlation coefficient "r" (*p=0.05; **p<0.01, **** p<0.001, n.s., p>0.05).

Figure S8