Supplement Methods

Flow cytometry

Whole FGTs or FGT parts were mechanically disrupted and enzyme-digested with 0.5 mg/ml collagenase D (Roche; catalogue no. 11088866001), 0.5 U/ml dispase (Corning; catalogue no. 354235), 0.5 mg/ml DNAse I (Sigma-Aldrich; catalogue no. DN25-1G) at the indicated time points. Leucocytes were isolated using a 40% and 80% discontinuous Percoll (GE Healthcare; catalogue no. 17-0891-01) gradient. Surface immunostaining was performed and cells were fixed in 4% formaldehyde (CHEMSOLUTE; Perkin Elmer, catalogue no. 2137-1L-PE). For intracellular staining, cells were permeabilized using the permeabilization buffer provided in the Foxp3 kit (eBioscience; catalogue no. 00-5523-00) and ILCs, myeloid cells and T cells were subsequently analysed by flow cytometry on a LSR Fortessa (BD Bioscience). The following antibodies were used (antigen[conjugate-clone-supplier]): Biotin (V500-BD Biosciences); CD11b (AF700-M1/70-BioLegend); CD11c(PE-Cy7-N418-eBioscience); CD127 (PE-Cy7-A7R34-BioLegend); CD16/CD32 (2.4G2 BD Biosciences); CD19 (Biotin_MB19-1-eBioscience); CD3 (Biotin-145-2C11-BioLegend); CD45.2 (FITC-104-BioLegend); CD49a (PE-H31/8-BD Biosciences); CD5 (Biotin-53-7.3-eBioscience); CD64 (PE-X54-517.1-BioLegend); EOMES (ef660-Dan11mag-eBioscience); F4/80 (Biotin-BM8-BioLegend); FcεR1α (Biotin-MAR-1-BioLegend); GATA3 (APC-REA174-Miltenyi Biotec); IFNγ (BV785-XMG1.2-BioLegend); Ly6G (Biotin-1A8-BioLegend); NK1.1 (PE-Cy7-PK136-eBioscience); RORγt (PE-Q31-378-BD Biosciences); ST2 (BV421-429-93-BD Biosciences); TCRβ (Biotin-H57-597-BioLegend); TCRβ (PerCP-Cy5.5-H57-597-BioLegend); TCRγδ (Biotin-eBIOGL3-BioLegend); TNF (BV421-MP6-XT22-BioLegend); anti-hBCL-2 (PE, 51-65115X, BD); Fixable Viability Dye (ef780-eBioscience). For cytokine staining, cells were incubated 4 h at 37 °C with GolgiPlug containing Brefeldin A (BD Biosciences; catalogue no. 555029) before immunostaining.

Quantitative PCR

FGT parts were homogenized in TRI Reagent® using a tissue homogenizer. For RNA isolation, the RNA isolation kit by Zymo Research (Direct-zol RNA Miniprep; catalogue no. R2053) was used. The Transcriptor First Strand cDNA Kit (Roche; catalogue no. 04897030001) was used to transcribe isolated RNA into cDNA with anchored-oligo$(dt)_{18}$ primers. For the qPCR, 50 ng cDNA was used and mixed with SYBR selected master mix (Thermo Fisher Scientific; catalogue no. 4472918). Specific primers for *IL-15* (fw: ACATCCATCTCGTGCTACTTGT; rev: GCCTCTGTTTTAGGGAGACCT), *IL-18* (fw: GTGAACCCCAGACCAGACTG; rev: CCTGGAACACGTTTCTGAAAGA), *IL-33* (fw: TCCAACTCCAAGATTTCCCCG; rev: CATGCAGTAGACATGGCAGAA), *TWEAK* (fw: TGCCTTGGCCTCCTGCTGGTCGT; rev: GCCGGACTAGTTGTTCCAAGAAA), *Fn14* (fw: AGGCTACTGTGGCCCATTCTG; rev: CCCTCTCCACCAGTCTCCTCTA) were used. Specific primers for the reference gene *β-Aktin* (fw: CTAAGGCCAACCGTGAAAAG; rev: ACCAGAGGCATACAGGGACA) were used. qPCR was performed using the QuantStudio™ 5 Real-Time PCR System (Thermo Fisher Scientific) and fold-change expression levels were determined relative to uninfected mice calculating ΔΔCt-values after normalization of gene expression to the reference gene *β-actin*.

Detection of C. muridarum *genome copies in FGT tissue*

Tissue digestion and DNA isolation were performed using the DNeasy Blood and Tissue Kit (Qiagen, catalogue no. 69506). 100 ng of isolated DNA was used for qPCR and mixed with SYBR selected master mix (Thermo Fisher Scientific; catalogue no. 4472918). Specific primers for *C. muridarum* (fw: GCCGTTTTGGGTTCTGCTT; rev: CGAGACGTAGGCTGATGGC) were used. qPCR was performed using the QuantStudio™ 5 Real-Time PCR System (Thermo Fisher Scientific) and Ct values and corresponding tissue weights were used to determine chlamydial genome copies per mg tissue using the equation of the standard curve. The standard curve was generated via DNA isolation (DNeasy Blood

and Tissue Kit) of a defined number of IFUs of *C. muridarum*, subsequent dilution series and qPCR.