

Supplementary Data 1. Alignment of HSP83 coding sequences

The ten copies of *HSP83* in TriTrypDb (Tb927.10.10890, 10900, 10910, 10920, 10930, 10940, 10950, 10960, 10970, 10980) were aligned using Clustal O (1.2.1). Synonymous variations are highlighted in green, nonsynonymous variations are in yellow, RNAi sequence is in red.

CLUSTAL O(1.2.1) multiple sequence alignment

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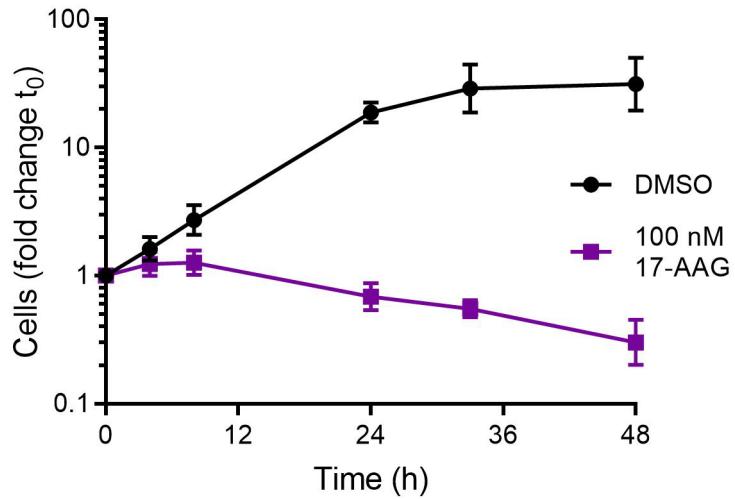
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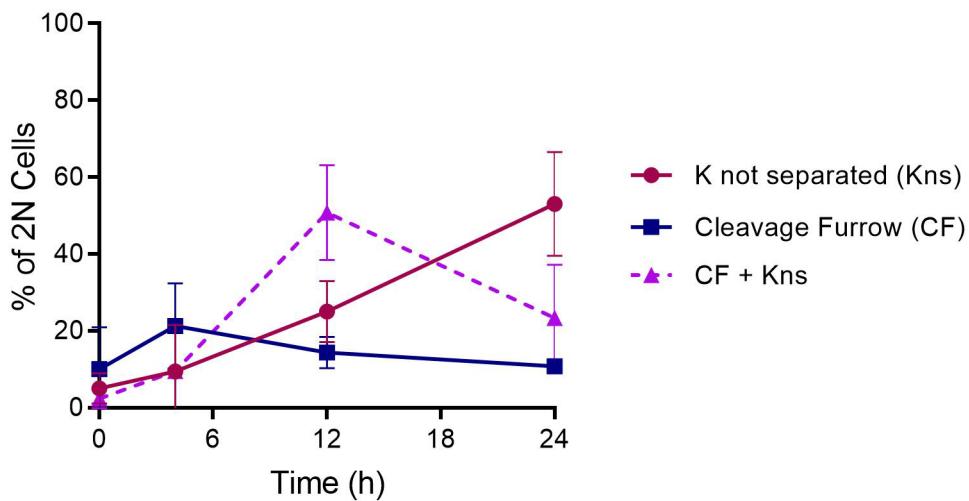
Supplementary Table 1. Primers used in this study

Name	Purpose	Sequence ^a
MX-F	Maxicircle probe for Southern blots	CTAACATACCCACATAAGACAG ACACGACTCAATCAAAGCC
H83Xb-F	Incorporating restriction sites (Xb – XbaI, Nd – NdeI) on RNAi sequence for <i>HSP83</i>	CCC <u>C</u> T <u>C</u> T <u>A</u> G <u>A</u> TATTGTGAAGAAGGCCCTGG
H83Nd-R		CCCC <u>C</u> AT <u>A</u> T <u>G</u> C <u>T</u> CTTCATTGCCTTGACAA
H83Xh-F	Incorporating restriction sites (Xh – XhoI, As – AscI) on RNAi sequence for <i>HSP83</i>	CTCT <u>C</u> TC <u>G</u> AG <u>T</u> ATTGTGAAGAAGGCCCTGG CTTT <u>G</u> G <u>C</u> G <u>C</u> CC <u>C</u> CTTTCATTGCCTTGACAA
H84Xb-F	Incorporating restriction sites (Xb – XbaI, Nd – NdeI) on RNAi sequence for <i>HSP84</i>	CCC <u>C</u> T <u>C</u> T <u>A</u> G <u>A</u> G <u>G</u> C <u>T</u> CC <u>C</u> TTTGACACC
H84Nd-R		CCCC <u>C</u> AT <u>A</u> T <u>G</u> T <u>G</u> TGAAC <u>C</u> TGGTCGGTACAA
H84Xh-F	Incorporating restriction sites (Xh – XhoI, As – AscI) on RNAi sequence for <i>HSP84</i>	CTTC <u>C</u> TC <u>G</u> AG <u>A</u> G <u>G</u> C <u>T</u> CC <u>C</u> TTTGACACC GTAT <u>G</u> G <u>C</u> G <u>C</u> CC <u>T</u> GTGAAC <u>C</u> TGGTCGGTACAA
H84Kp-F	Incorporating restriction sites (Kp – KpnI, Xh – XhoI) on terminal 400 bp of <i>HSP84</i>	CTAT <u>GG</u> T <u>AC</u> CC <u>GG</u> TT <u>GG</u> CG <u>TC</u> GTCAAATC
H84Xh-R		CTT <u>A</u> CT <u>CG</u> AG <u>G</u> CTT <u>A</u> T <u>CG</u> GC <u>AG</u> T <u>GG</u> CT <u>CC</u> T
3UH84Bm-F	Incorporating restriction sites (Bm – BamHI, Sc – SacI) on 400 bp of 3'UTR of <i>HSP84</i>	CATT <u>GG</u> G <u>AT</u> CC <u>GG</u> T <u>GC</u> AG <u>AG</u> TAA <u>AC</u> AT <u>TT</u> TT <u>TC</u>
3UH84Sc-R		CAAT <u>G</u> AG <u>G</u> CT <u>CC</u> AA <u>CG</u> C <u>AC</u> AG <u>TAA</u> AG <u>GG</u> T <u>AT</u>
H83m-F	<i>HSP83</i> probe for mRNA	CCAGAGTCTGACGAACCAGTC
H83m-R	northern blots	CCAGGTATT <u>C</u> CTG <u>C</u> TGG <u>T</u> CT
H84m-F	<i>HSP84</i> probe for mRNA	CTAAAGGGCAATGCAGGGGA
H84m-R	northern blots	CCGGCTGTACAC <u>CT</u> TGACAT

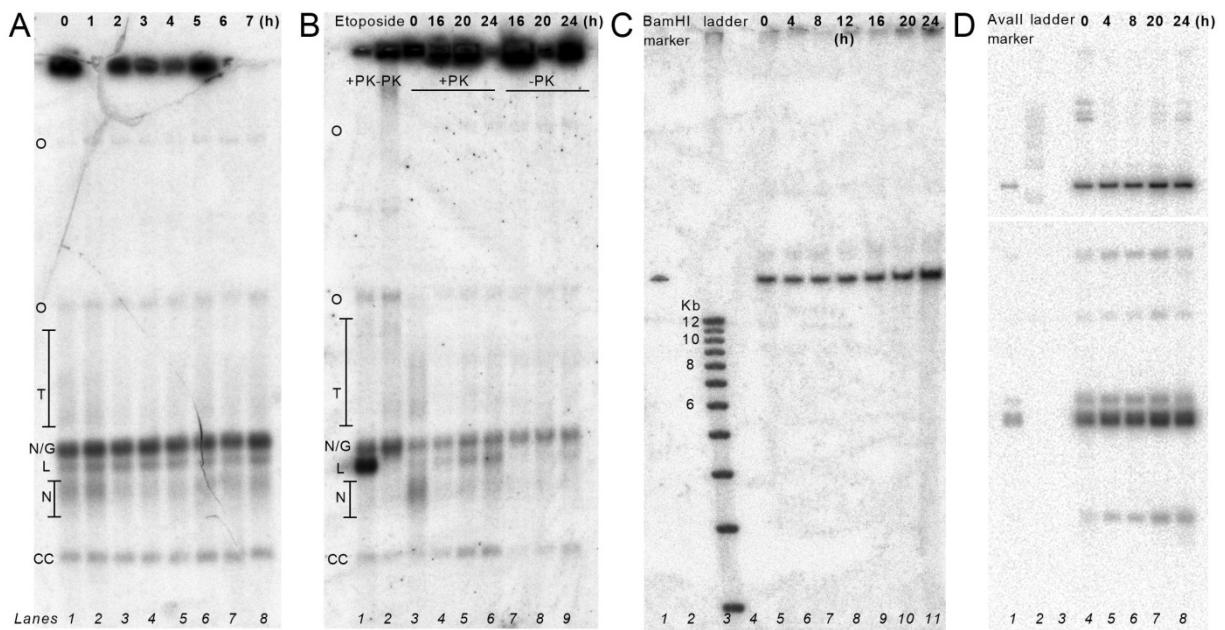
^aRestriction sites underlined



Supplementary Figure 1. 17-AAG treatment halts trypanosome growth followed by a gradual decline in cell number. 2×10^5 Cells were treated with either DMSO or 100 nM 17-AAG and incubated at 37°C. Aliquots were taken across 48 h and live cells counted by hemocytometer and microscopy. Data are from 8 independent growth curves, geometric M ± SD, n = 3-8 for each point.

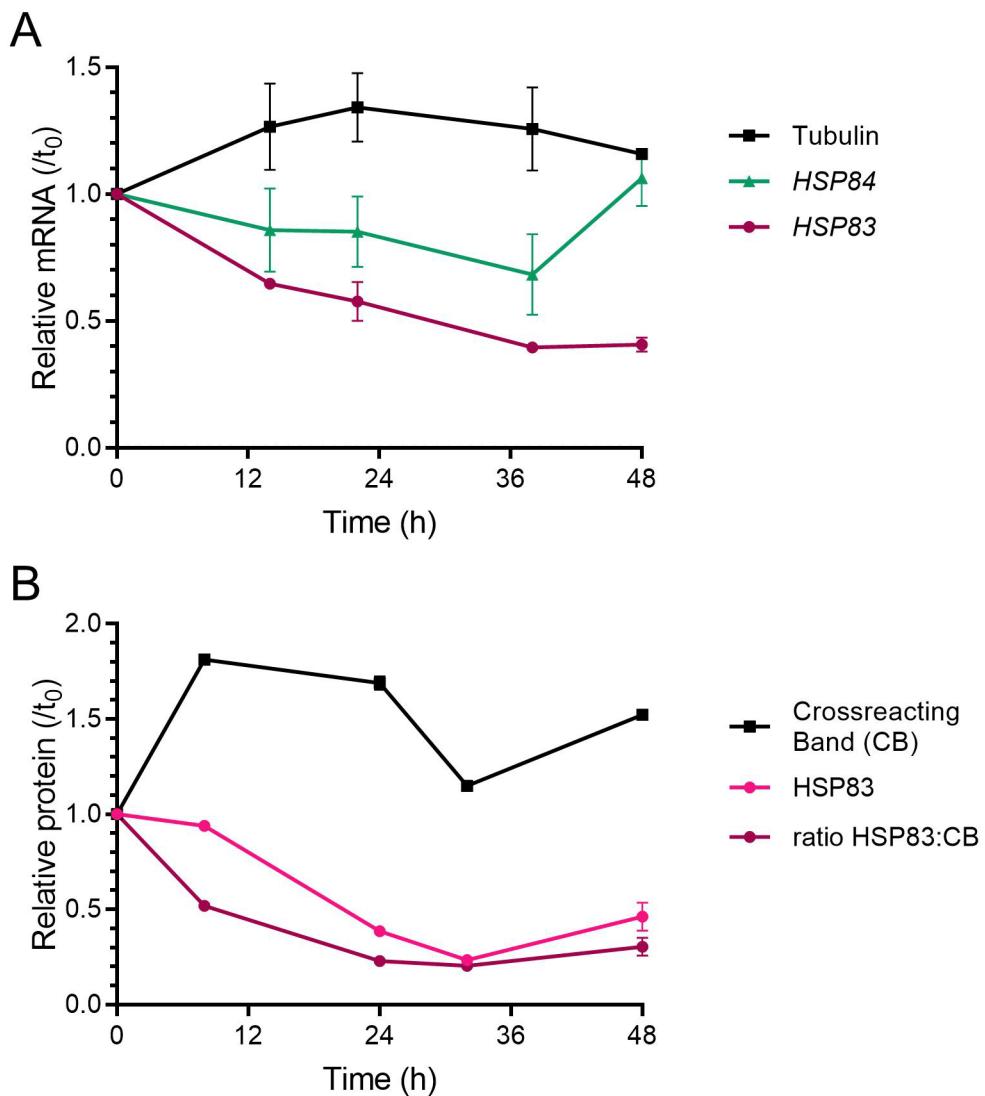


Supplementary Figure 2. Abnormalities in kinetoplast segregation and cleavage furrow ingression after treatment with an Hsp90 inhibitor. Cells treated with 100 nM 17-AAG were harvested at indicated intervals, stained with DAPI, examined by fluorescence microscopy, and 2N cells were scored for presence of a visible cleavage furrow but normal kinetoplasts (blue, CF), no cleavage furrow but kinetoplasts not normally separated (red, Kns), or presence of both abnormalities (purple, CF + Kns). Values are M ± SD; >175 2N cells observed at each time point.

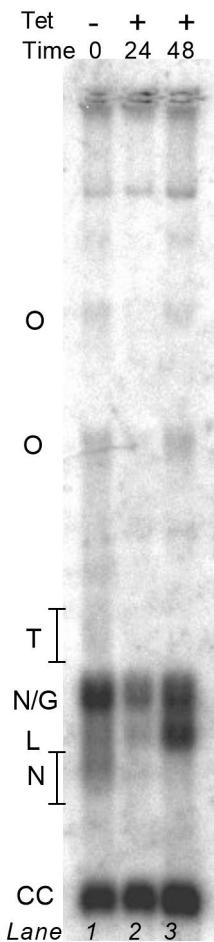


Supplementary Figure 3. 17-AAG treatment leads to changes in free minicircle topology and in the cell content of mini- and maxicircle DNA. (A) Southern blot analysis of free minicircles isolated from trypanosomes treated for short periods with 100 nM 17-AAG. *Lane 1*, DNA from untreated cells; *Lanes 2-8*, increasing duration of 100 nM 17-AAG, as indicated (**bold**, top of lanes). (B) Southern blot analysis of free minicircles isolated from trypanosome lysates treated with or without proteinase K (\pm PK) prior to electrophoresis. *Lanes 1 and 2*, DNA from trypanosomes treated with topoisomerase II poison etoposide; *Lanes 3-9*, DNA from cells treated with 100 nM 17-AAG. Without protease digestion the linear forms are selectively lost from analysis, indicating their covalent linkage to topoisomerase protein [1]. *O*, Oligomers of two or more interlocked circles; *T*, theta structures; *N/G*, mature nicked, or gapped daughters; *L*, Lines, *N*, immature nicked daughters; *CC*, covalently closed template circles. (C) Southern blot analysis of free maxicircles (0.6% gel). *Lane 1*, BamHI-linearized maxicircle marker; *Lane 3*, DNA size ladder; *Lanes 5-11*, increasing duration of 17-AAG (100 nM) treatment. (D) Southern blot analysis in 1.5% gel of total minicircles and total maxicircles after digestion of kDNA (ethanol precipitated from lysates) by Avall restriction enzyme (overnight). The blot was

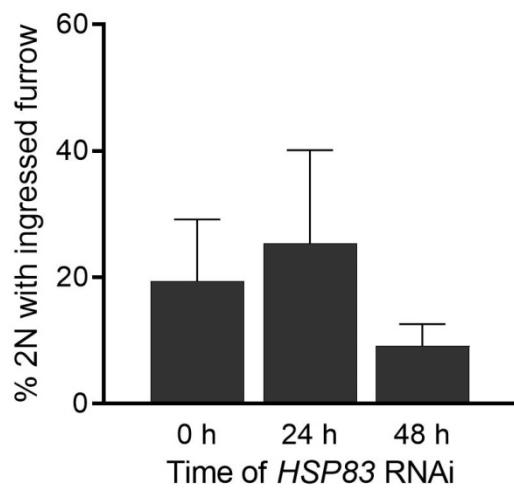
first probed for minicircles (bottom panel), then stripped and reprobed for maxicircles (top panel). *Lane 1*, Avall-cut network marker; *Lane 2*, DNA size ladder; *Lanes 4-8*, increasing duration of 17-AAG (100 nM) treatment. Samples loaded by equal cell equivalents.



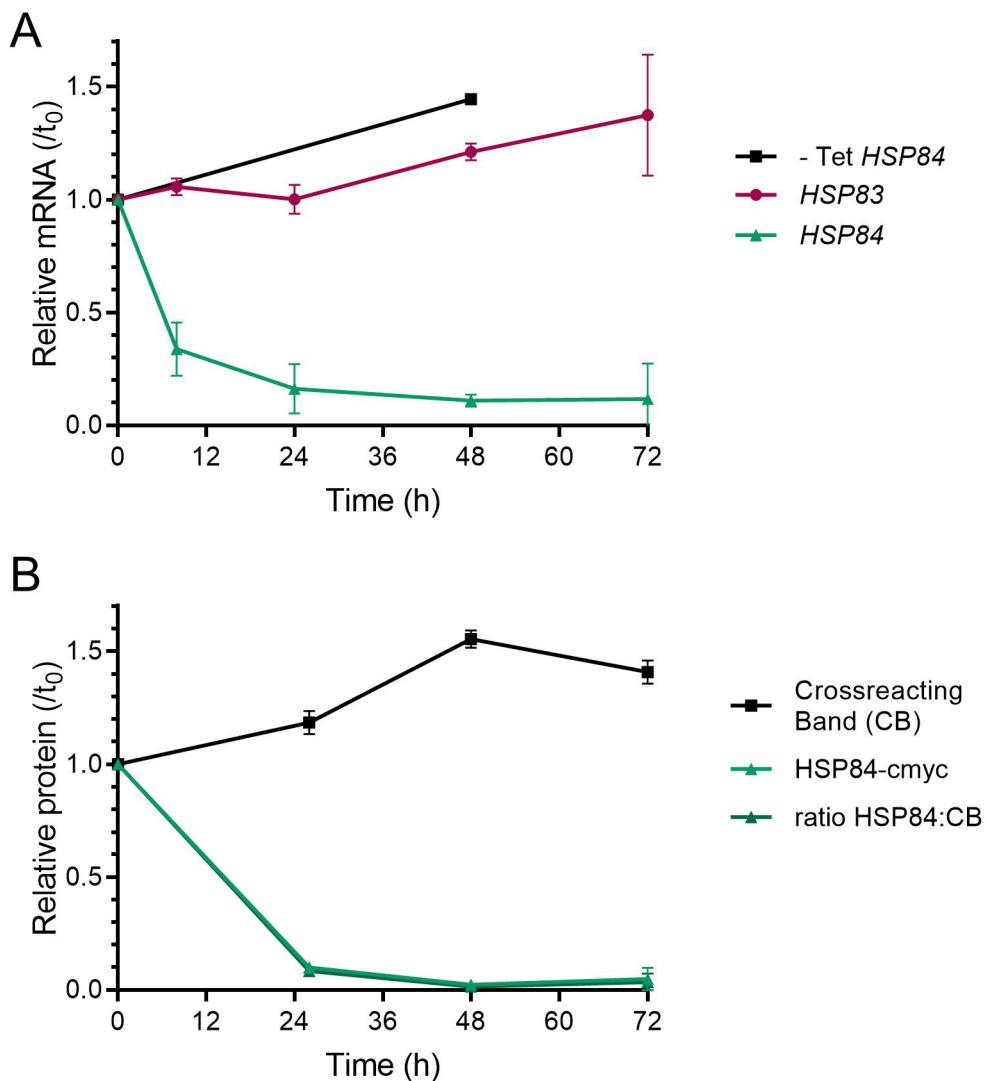
Supplementary Figure 4. Densitometry quantification of *HSP83* mRNA and protein from Figure 4, by mean grey values. (A) Time course of mRNA in northern blots from cells induced for RNAi of *HSP83*. *HSP83* (red), tubulin (black), *HSP84* (green), shown relative to time 0 h. (B) *HSP83* (pink) and a cross-reacting protein (50 kDa, black) in western blots from cells induced for *HSP83* RNAi over time, shown relative to uninduced cells, and ratioed to each other (red). Symbols are M ± SD of three densitometry measurements of the same blot.



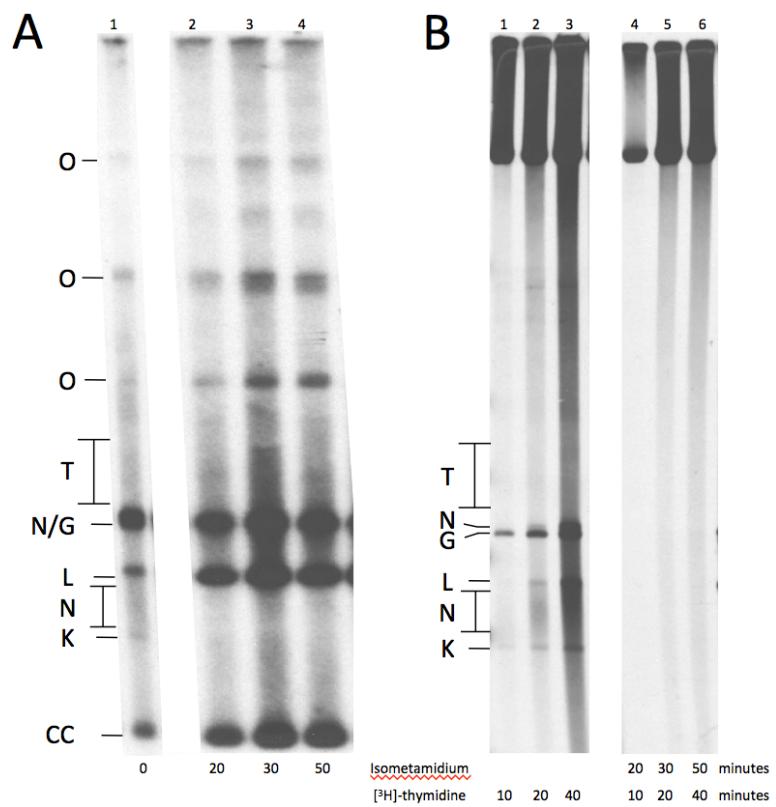
Supplementary Figure 5. Southern blot of free minicircles isolated at indicated times from *HSP83* RNAi-induced trypanosomes and separated in ethidium agarose (biological replicate of Fig. 5A to show theta region). *Lane 1*, uninduced cells at t_{0h} ; *Lanes 2 and 3*, increasing hours of RNAi. Lanes loaded by equal cell equivalents. *O*, oligomers of two or more interlocked circles; *T*, theta structures; *N/G*, mature nicked, or gapped daughters; *L*, linears, *N*, immature nicked daughters; *CC*, covalently closed template circles.



Supplementary Figure 6. *Hsp83* RNAi induction does not significantly change the percent of 2N cells with a visibly ingressed cleavage furrow. Merged phase contrast and DAPI fluorescence images of trypanosomes before and after induction of *HSP83* RNAi. $M \pm SD$, $n \geq 3$.



Supplementary Figure 7. Densitometry quantification of mRNA and protein in cells with inducible constructs for RNAi of *HSP84*, by mean grey values (Figure 6 data). (A) mRNA bands of *HSP84*, from uninduced (*black*) or induced (*green*) cells, and of *HSP83* (*red*) in induced cells, by northern blot, shown relative to 0 h. (B) *HSP84-cmcy* (*light green*) and a cross-reacting protein (60 kDa, *black*) bands, and their ratio (*dark green*), by western blot, across time in induced cells (shown relative to uninduced, time 0 h). Symbols are M ± SD of three densitometry measurements of the same blot.



Supplementary Figure 8. Impact of antitrypanosomal drug isometamidium chloride (Samorin) on free minicircles. African trypanosomes *T. equiperdum* were treated in vitro with 10 μ M isometamidium, and minicircle DNA was isolated and imaged as described previously [2,3]. (A) Southern blot of free minicircles obtained before (Lane 1) and at indicated intervals after addition of isometamidium to cell culture (Lanes 3-4). (B) Fluorographs to detect incorporation of [³H]thymidine into nascent DNA. Lanes 1-3, free minicircles isolated from control cells metabolically labeled for increasing times with [³H]thymidine. Lanes 4-6, minicircle DNA from cells treated with 10 μ M isometamidium for 10 min prior to the addition of [³H]thymidine. All samples obtained in the same experiment; lanes loaded by equal cell equivalents. O, Oligomers of two or more interlocked circles; T, theta structures; N/G, mature nicked, and CC, circular circles.

gapped daughters; L , Linears, N , immature nicked daughters; CC, covalently closed template circles.

References Cited in Supplement

1. Shapiro TA, Klein VA, Englund PT. Drug-promoted cleavage of kinetoplast DNA minicircles: Evidence for type II topoisomerase activity in trypanosome mitochondria. *J Biol Chem.* **1989**;264: 4173-4178.
2. Shapiro TA, Englund PT. Selective cleavage of kinetoplast DNA minicircles promoted by antitrypanosomal drugs. *Proc Natl Acad Sci U S A.* **1990**;87: 950-954. doi: 10.1073/pnas.87.3.950
3. Shapiro TA, Showalter AF. In vivo inhibition of trypanosome mitochondrial topoisomerase II: Effects on kinetoplast DNA maxicircles. *Mol Cell Biol.* **1994**;14: 5891-5897.