

1 **SUPPLEMENTAL FILES**
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3 **SUPPLEMENTAL METHODS**
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5 **Patients and Treatment Protocol**

6 The detailed treatment protocols and eligibility criteria for HOVON trials were described previously.^{1,2} All
7 HOVON patients received intensive chemotherapy induction followed by risk-based assignment to either
8 chemotherapy or stem cell transplantation as consolidation treatment. Patients from the MLL cohort
9 were treated either with intensive chemotherapy, non-intensive chemotherapy or were given supportive
10 care, all according to the standard treatment protocols in Germany. For survival analyses only MLL
11 patients treated with intensive chemotherapy were included, in combination with all HOVON patients.
12 All three HOVON protocols (H42A, H92 and H102) included the same therapeutic backbone with high
13 dose chemotherapeutics and investigational drugs. Specifically, H102 study, included patients 18 to 65
14 years of age, investigated Clofarabine, H92 randomly assigned Laromustine as investigational drug
15 while in H42A, patients 18 to 60 years of age, received similar induction chemotherapy but were
16 randomly administered G-SCF. The MLL patients were treated with the same 7+3 backbone as the
17 HOVON patients without the addition of an investigational drugs. No significant differences in overall
18 survival were found between HOVON (N=889) and intensively treated MLL patients (N=334; Figure
19 S9A), which may be expected since, the investigational drugs did not render survival benefits. In
20 addition, we included an analysis of patients who achieved complete remission only since for both H42A
21 (N=133) and H92 (N=43) only patients with complete remission were included in the current study (due
22 to lack of availability of data on the presence of SF mutations in patients who did not reach CR from
23 H42A and H92 studies). It should be noted that the majority of patients in our study were treated on the
24 H102 protocol (N=889, including both CR and non-CR patients) and therefore the relatively small
25 number of patients from H42A and H92 result in only a minor overrepresentation of CR patients in our
26 cohort, which is not expected to substantially affect the conclusions. Accordingly, all patients with
27 complete remission presented comparable outcome for both HOVON and MLL patients (Figure S9B).
28 A specific subgroup of patients was defined as secondary AML and included patients with antecedent
29 myeloid disorder (N=61). Since this is supposed to be a subgroup with substantially different
30 characteristics, they were analysed separately together with MDS (N=72) and treatment related AML
31 (N= 43; Supplemental Table S6).

32

33 **Sample preparation**

34 Mononuclear cell fraction was isolated from bone marrow (BM) or peripheral blood (PB) by Ficoll-
35 Hypaque (Nygaard, Oslo, Norway) density centrifugation and cryopreserved for further processing. For
36 targeted NGS (see below), following thawing, cells were lysed in RLT buffer with the addition of DTT
37 (Qiagen, Venlo, The Netherlands). Subsequently DNA was isolated as described previously.³

38

39 **Mutational profile**

40 The mutational status was determined for 41 commonly mutated genes in AML, including 7 splicing
41 factors (ASXL1, BCOR, BCORL1, BRAF, CBL, CEBPA, CSF3R, CUX1, DNMT3A, ETV6, EZH2, FLT3-
42 ITD, FLT3-TKD, GATA2, IDH1, IDH2, IKZF1, JAK2, KDM6A, KIT, KRAS, NOTCH1, NPM1, NRAS,
43 PHF6, PTPN11, RAD21, RUNX1, SETBP1, SF1, SF3A1, SF3B1, SMC1A, SMC3, SRSF2, STAG2,
44 TET2, TP53, U2AF1, U2AF2, WT1, ZRSR2).

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46 **Whole genome sequencing**

47 Whole genome sequencing (WGS) libraries were generated from 1µg of DNA extracted from bone
48 marrow or peripheral blood samples using the TruSeq PCR free library prep kit, following the
49 manufacturer's recommendations (Illumina, San Diego, CA, USA) and sequenced on a NovaSeq6000
50 or HiSeqX Illumina instruments following a 2x150bp paired-end reads standard protocol at a mean depth
51 of coverage of >100x. Bioinformatic analysis of WGS data was performed using Illumina's BaseSpace
52 Sequence Hub and in-house pipelines. Reads were aligned against human genome build GRCh37/hg19
53 with the tool Isaac3.⁴ Variant calling was performed using Strelka2⁵ and additional variant annotation
54 was performed using Ensembl VEP.⁶ Only exonic (non-synonymous single nucleotide variants (SNVs)
55 and small insertions/deletions (indels)) and variants at splicing acceptor/donor sites were considered in
56 this study. Because matched normal samples were not available, tumor-unmatched normal variant
57 calling was performed using a pool gender-matched DNA (Promega, Madison, WI, USA). In order to
58 remove germline and benign variants as well as technical artifacts from the dataset a strict filtering
59 strategy was applied as described below.

60

61 **Filtering whole genome sequencing data**

62 Upon variant calling the following in-house filtering strategy⁷ was applied to remove sequencing artifacts
63 as well as germline and likely benign variants, resulting in a dataset consisting of reliable and likely
64 somatic, pathogenic variants.

65

66 *Sequencing artifacts*

67 Repetitive regions and regions potentially troublesome for variant calling were annotated based on
68 Ensembl repeat database and the Global Alliance for Genomics and Health (GA4GH) database. Firstly,
69 variants located in regions of established low confidence variant calling (as specified in Genome in a
70 Bottle Consortium using sample NA12878/HG001, <https://github.com/genome-in-a-bottle>).
71 Subsequently, we discarded variants with low frequency (VAF < 15%) located in low complexity regions
72 (homopolymers), tandem repeats (i.e. microsatellites), segmental duplications or repetitive regions
73 interspersed throughout the genome (i.e. transposable elements). Finally, variants supported by ≤ 3
74 reads for the alternative allele as well as with the total depth of coverage ≤ 5 were removed from
75 downstream analysis.

76

77 *Germline variants*

78 First, variants with the global population frequency ≥ 0.001 (based on the genome aggregation
79 database; gnomAD) were discarded. Next, variants labelled as germline in ClinVar or COSMIC
80 databases were filtered out. Finally, variants not well annotated in hematological malignancies with a
81 VAF between 40-60% or > 90% in all samples were eliminated. This strategy was applied in order to
82 remove as many of the germline variants as possible although it should be noted that due to the lack of
83 matched germline control some residual germline variants are likely to be retained in our dataset.
84 However, since this study focuses on genes recurrently mutated in AML these residual germline variants
85 are not expected to affect the analysis.

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Likely benign variants

First, variants listed as benign/likely benign in ClinVar database were removed. Subsequently, we used an in-house developed tool HePPy (Hematological Predictor of Pathogenicity)⁸ to remove missense variants with HePPy score < 0.75 (indicating low pathogenicity).

Statistical Analysis

In our study, OS was defined as time from the initial diagnosis to death or last follow-up. EFS was defined as time from initial diagnosis to an event (refractoriness, relapse, death or last follow-up, whichever occurred first). In the multivariable Cox proportional hazards model, the prognostic value of SF mutations was evaluated in the context of demographic and clinical variables (including age, gender, type of AML, white blood cell count, type of stem cell transplantation administered) as well as the ELN 2017 risk classification or modified ELN 2017 classification (all of which were also significantly associated with OS and EFS in univariable Cox model, Supplemental Table S8). In order to assess the individual and combined prognostic value of *RUNX1* and *ASXL1* with SF mutations, a modified ELN 2017 classification was generated by excluding *RUNX1* and *ASXL1* from the list of genetic criteria due to their frequent co-occurrence with SF mutations (hence patients carrying these mutations were re-classified based on the remaining criteria). The proportional hazard assumption was evaluated for each variable using the Schoenfeld test and upon examination of the plots of Schoenfeld residuals. Type of stem cell transplantation violated the proportional hazard assumption and therefore it was used as strata variable in all the multivariable Cox regression models (for variables used in the model as strata the statistics are not calculated and therefore, they do not appear in the results). In addition, the interaction between SF mutations and *RUNX1* variables violated the proportional hazard assumption and was additionally evaluated as time-dependent co-variate. The Akaike Information Criterion (AIC) was used to compare the fit of the baseline models with the fit of models containing variables of interest (SF mutations or their interaction with other genetic mutations). This AIC criterion informs not only about the goodness of fit of the model but also penalizes on the number of variables in the model which makes it an attractive method to compare models (the lower the AIC value the better, regardless of the magnitude of difference). To further substantiate the differences in the fit of the models, we have additionally performed ANOVA analyses to compare the fit of the baseline and new model for each tested variable. In all analyses p-value < 0.05 was considered statistically significant (for Fisher's test the 0.05 cut-off was applied to BH-corrected p-values). All statistical analyses were carried out in R version 3.6.3/R studio version 1.2.5, including the following packages: survminer (version 0.4.6)⁹, survival (version 3.1-11, <https://CRAN.R-project.org/package=survival>)¹⁰, ggplot2 (version 3.2.1)¹¹, ComplexHeatmap (version 2.2.0)¹². The patient numbers in particular analyses vary depending on the amount of available data for specific variables.

125 **References**

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152 **SUPPLEMENTAL RESULTS**
153

154 **Study Design**

155 In this study, two independent cohorts of AML patients were included (the HOVON and MLL cohort, see
156 Materials and Methods in the main paper). Acknowledging the heterogenous origin of our cohort, the
157 genetic landscape as well as clinical features and treatment outcome of patients of both cohorts were
158 carefully compared to assure that the inter-cohort heterogeneity will not bias the analysis.

159 The mutational profiles of patients included in both cohorts were primarily defined based on routine
160 molecular diagnostics. In case of MLL patients, molecular diagnostics were additionally complemented
161 by whole genome sequencing (WGS). The routine diagnostics data for 430 patients of the HOVON
162 dataset were complemented by targeted sequencing using Illumina TruSight Myeloid Panel. The
163 complete genetic landscape of both HOVON as well as MLL patients was found to be consistent and
164 typical of AML with frequencies of cytogenetic and molecular aberrations being consistent with those
165 reported in previous studies (data not shown).^{1,2} In agreement, no significant differences in overall
166 survival were found between HOVON (with all patients treated with intensive chemotherapy) and
167 intensively treated MLL patients (Figure S9A). The experimental drugs included in HOVON studies
168 (Clofarabine, Laromustine and G-SCF) did not improve treatment outcome, corresponding previously
169 reported results (more details regarding treatment protocols can be found in the methods section).³⁻⁵
170 Altogether, this cohort of 1447 patients, of which 1223 were treated with intense chemotherapy, allowed
171 us to address our research questions, including the assessment of effects of relatively low frequency
172 events such as SF mutations in subgroups of patients, and maximized the benefits to be derived from
173 existing cohort studies.

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175 **Co-occurring SF mutations**

176 Nine patients in the analyzed cohort had coinciding mutations in two different SF genes (Supplemental
177 Table S5). In almost all cases a common (recurrent) allele for a particular SF was paired with a less
178 common allele of the second SF. Furthermore, these patients were almost exclusively over 65 years of
179 age and in majority presented with mutations in *ASXL1*. Interestingly, a recent study uncovered that less
180 common alleles with reduced effects on alternative splicing were enriched in patients with double SF
181 mutations as compared to single mutants.²

182

183 **The Influence of Gene Interactions on Survival in Younger AML Patients**

184 It was previously shown that in AML patients younger than 60 years harboring *RUNX1* mutations had
185 an independent negative prognostic value.⁶ Therefore, we assessed the association of the interaction
186 between SFmut4 (as well as *SRSF2* mutations) and *RUNX1* mutations within this subgroup of patients
187 (Figure S10). Again, the co-occurrence of *SRSF2* or SFmut4 with *RUNX1* mutations was associated
188 with adverse outcome, while the presence of *RUNX1* mutations without SFmut4 or *SRSF2* mutations
189 did not indicate inferior survival. In this subset of patients (with age below 60 years) the observed
190 associations were even stronger than in the total cohort.

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193 **References**

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207 mutational epistasis in myeloid neoplasms. *Blood*. 2020;doi:10.118.
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| 209 | SUPPLEMENTAL TABLES |
| 210 | |
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| Supplemental Table 1. Additional Patient Characteristic | |
|---|-------------|
| Prior Disease, n (%) | |
| None | 1079 (88.2) |
| Hematological Disorder | 68 (5.6) |
| Other Cancer | 53 (4.3) |
| Other Disease | 1 (0.1) |
| Missing | 22 (1.8) |
| Prior Treatment, n (%) | |
| None | 1139 (93.1) |
| Chemo- or Radiotherapy | 60 (4.9) |
| Missing | 24 (2.0) |

213

214 **Table S1. Additional characteristics of patients treated with intensive chemotherapy.**

215 FAB, French American British Classification; RAEB, Refractory Anemia with Excess Blasts.

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| Gene | Evaluated cases | Positive cases | Negative Cases | Percentage Positive (among evaluated) | Percentage Positive (among total cases) |
|--------------|-----------------|----------------|----------------|---------------------------------------|---|
| <i>SF1</i> | 558 | 2 | 556 | 0.36 | 0.14 |
| <i>SF3A1</i> | 558 | 1 | 557 | 0.18 | 0.07 |
| <i>SF3B1</i> | 1447 | 41 | 1406 | 2.83 | 2.83 |
| <i>SRSF2</i> | 1447 | 148 | 1299 | 10.23 | 10.23 |
| <i>U2AF1</i> | 988 | 31 | 957 | 3.14 | 2.14 |
| <i>U2AF2</i> | 558 | 2 | 556 | 0.36 | 0.14 |
| <i>ZRSR2</i> | 988 | 15 | 973 | 1.52 | 1.04 |
| SFmut7 | 661 | 231 | 430 | 34.90 | 15.96 |
| SFmut4 | 1039 | 229 | 810 | 22.00 | 15.80 |

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218 **Table S2. The frequencies of SF mutations.**

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220 Table S3 and S4 can be found in attached Excel File.
221
222 **Tables S3, S4 and S6-S13.xlsx**
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| Sample ID | Gender | Age | SF Mutations (VAF) | Other Mutations (VAF) | Cytogenetics and Karyotype |
|-----------|--------|------|--|--|----------------------------|
| #1 | M | 67,9 | <i>SRSF2</i> P95H (0.4693) <i>SF3B1</i> T871I(0.3964) | <i>IDH2</i> (0.3885), <i>RUNX1</i> (0.1513), <i>SMC3</i> (0.4646) | Trisomy 11 |
| #2 | M | 75,1 | <i>SRSF2</i> P95H (0.5407) <i>SF1</i> G497S (0.4834) | <i>ASXL1</i> (0.5192), <i>NRAS</i> (0.1724), <i>TET2</i> (0.9794) | Normal Karyotype |
| #3 | F | 84 | <i>SRSF2</i> P95H (0.5229) <i>SF1</i> K341N (0.4877) | <i>ASXL1</i> (0.2857), <i>TET2</i> (0.4154), <i>WT1</i> (0.3875) | Trisomy 8 |
| #4 | M | 78,3 | <i>SRSF2</i> P95H (0.1053) <i>ZRSR2</i> R169X (0.2449) | <i>CUX1</i> (0.092), <i>IDH1*</i> , <i>TET2</i> (0.381) | Normal Karyotype |
| #5 | F | 63,3 | <i>SF3B1</i> K666N (0.4567) <i>ZRSR2</i> G438R442dup (0.4713) | <i>NRAS</i> (VAF 0.0989) | Monosomy 7 |
| #6 | M | 80,3 | <i>SF3B1</i> D781Q (0.3364) <i>U2AF2</i> - (0.1132) | <i>ASXL1</i> (0.4302), <i>RUNX1</i> (0.2804) | Normal Karyotype |
| #7 | M | 80,1 | <i>U2AF1</i> Q157P (0.2595) <i>ZRSR2</i> Q120RfsX10 (0.7391) | <i>ASXL1</i> (0.4145), <i>KRAS</i> (0.0693) | Del7q |
| #8 | M | 64,6 | <i>U2AF1</i> Q157P (0.4142) <i>ZRSR2</i> S445R448dup (1) | <i>ASXL1</i> (0.4057), <i>BCOR</i> (0.0506), <i>EZH2</i> (0.4828), <i>FLT3^{TD}</i> (AR >0.5), <i>RUNX1</i> (0.4706), <i>SMC1A</i> (0.8116), <i>WT1</i> (0.5263) | Trisomy 8 |
| #9* | F | 45 | <i>SF3B1</i> - <i>U2AF1</i> - | <i>RUNX1</i> , <i>NRAS</i> , <i>STAG2</i> | Trisomy 8 |

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226 **Table S5. Characteristics of patients with co-occurring SF mutations at diagnosis.** Note: Copy
227 number variation data was not included in the analysis and therefore VAFs should be interpreted with
228 caution. * - no VAF data available; Male; F, Female; VAF, Variant Allele Frequency.

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231 Tables S6-S13 can be found in attached Excel File.

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233 **Tables S3, S4 and S6-S13.xlsx**

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235 **SUPPLEMENTAL FIGURE LEGENDS**
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237 **Figure S1. Overall survival of AML patients in the relation to the presence of individual SF**
238 **mutations.** Kaplan-Meier curves for overall survival in relation to the mutation status of *SRSF2*, *SF3B1*,
239 *U2AF1*, *ZRSR2*, *SF3A1* or *U2AF2* are depicted.

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241 **Figure S2. Event-free survival of AML patients in the relation to the presence of individual SF**
242 **mutations.** Kaplan-Meier curves for event-free survival in relation to the mutation status of *SRSF2*,
243 *SF3B1*, *U2AF1*, *ZRSR2*, *SF3A1* or *U2AF2* are depicted.

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245 **Figure S3. The influence of SF mutations on survival within ELN risk groups.** The figure depicts
246 Kaplan-Meier curves for event-free survival and overall survival in relation to SFmut4 status (A),
247 mutation status of *SRSF2* (B) or mutation status of *SF3B1* (C) within the favorable, intermediate or
248 adverse risk groups as defined by the ELN 2017 classification.

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250 **Figure S4. Multivariable analysis of overall survival of AML patients in relation to the presence**
251 **of SF mutations.** A,B. - Multivariable Cox regression analysis of overall survival in relation to SFmut4
252 with complete (A) and modified (B) ELN 2017 classification. In the modified ELN 2017 classification
253 *RUNX1* and *ASXL1* mutations were excluded, so that patients carrying *RUNX1* or *ASXL1* mutations
254 were re-classified based on the presence of the rest of aberrations in this classification system. Type of
255 stem cell transplantation violated the proportional hazard assumption and therefore it was used as strata
256 variable in all the multivariable Cox regression models (for variables used in the model as strata the
257 statistics are not calculated and therefore, they do not appear in the results). WBC – white blood cell
258 count; sAML – secondary AML, tAML - treatment-related AML.

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260 **Figure S5. Survival of AML patients in relation to the presence of a mutation in *RUNX1* or *ASXL1*.**
261 A - Multivariable Cox regression analysis of event-free (left) or overall survival (right) in relation to
262 mutations in *RUNX1* including modified ELN 2017 classification. B - Multivariable Cox regression
263 analysis of event-free (left) or overall survival (right) in relation to mutations in *ASXL1* including modified
264 ELN 2017 classification. In the modified ELN 2017 classification *RUNX1* and *ASXL1* mutations were
265 excluded, so that patients carrying *RUNX1* or *ASXL1* mutations were re-classified based on the
266 presence of the rest of aberrations in this classification system. In both models type of stem cell
267 transplantation was included as strata. WBC – white blood cell count; sAML – secondary AML, tAML -
268 treatment-related AML.

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270 **Figure S6. Influence of interactions between SF mutations and *RUNX1* as well as *ASXL1***
271 **mutations on overall survival.** A - Kaplan Meier curves for overall survival in relation to the mutation
272 status of *SRSF2* (left) or SFmut4 (right) in combination with *RUNX1* mutations. B - Kaplan Meier curves
273 for event-free and overall survival based on the mutation status of *SRSF2* (left) or SFmut4 (right) in
274 combination with *ASXL1* mutations.

275

276 **Figure S7. Analysis of survival in relation to interactions of SF mutations with *RUNX1* and *ASXL1***
277 **mutations in adverse risk category according to ELN 2017 classification.** A - Kaplan Meier curves
278 for event-free survival (left) and overall survival (right) in relation to the mutation status of *SRSF2* and
279 *RUNX1* within the adverse ELN 2017 risk group. B - Kaplan Meier curves for event-free survival (left)
280 and overall survival (right) in relation to the SFmut4 and *RUNX1* mutations.

281
282 **Figure S8. Multivariable analysis of survival of AML patients in relation to the interaction of SF**
283 **mutations with mutations in *RUNX1* or *ASXL1*.** A – Multivariable Cox regression analysis of overall
284 survival in relation to the mutation status of *SRSF2* or SFmut4 and *RUNX1* including modified ELN 2017
285 classification. B,C – Multivariable Cox regression analysis of event-free (B) and overall survival (C) in
286 relation to the mutation status of *SRSF2* or SFmut4 and *ASXL1* including modified ELN 2017
287 classification. In the modified ELN 2017 classification *RUNX1* and *ASXL1* mutations were excluded, so
288 that patients carrying *RUNX1* or *ASXL1* mutations were re-classified based on the presence of the rest
289 of aberrations in this classification system. Type of stem cell transplantation violated the proportional
290 hazard assumption and therefore it was used as strata variable in all the multivariable Cox regression
291 models (for variables used in the model as strata the statistics are not calculated and therefore, they do
292 not appear in the results). WBC – white blood cell count; sAML – secondary AML, tAML - treatment-
293 related AML.

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295 **Figure S9. Survival analysis of our combined cohort.** The figure depicts Kaplan-Meier curves for
296 overall and event free survival in all HOVON and MLL patients (A) and patients with complete remission
297 only (B).

298
299 **Figure S10. Survival of young and old AML patients in relation to the presence of SF mutations**
300 **and *RUNX1* mutations.** A - Kaplan Meier curves for overall survival in relation to the mutation status of
301 *SRSF2* in combination with *RUNX1* mutations in AML patients younger than 60 years (left), or 60 years
302 and older (right). B - Kaplan Meier curves for overall survival in relation to the presence of SFmut4 in
303 combination with *RUNX1* mutations in AML patients younger than 60 years (left), or 60 years and older
304 (right). C - Kaplan Meier curves for event-free survival in relation to the mutation status of *SRSF2* in
305 combination with *RUNX1* mutations in AML patients younger than 60 years (left), or 60 years and older
306 (right). D - Kaplan Meier curves for event-free survival in relation to the presence of SFmut4 in
307 combination with *RUNX1* mutations in AML patients younger than 60 years (left), or 60 years and older
308 (right).

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Figure S1

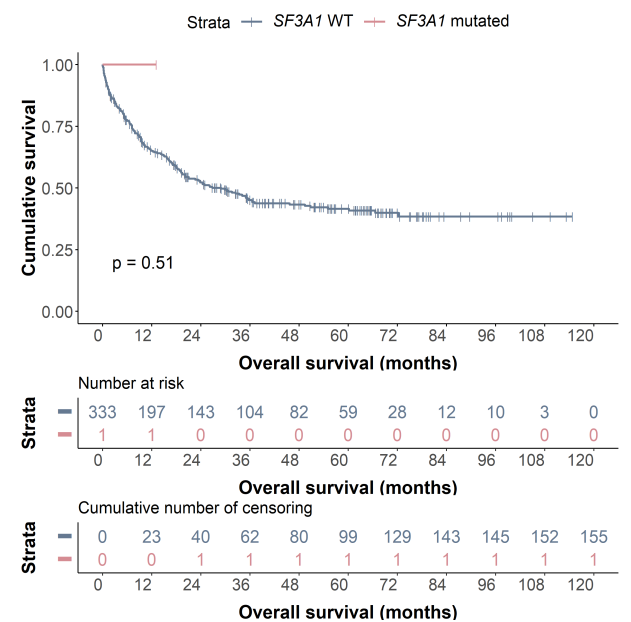
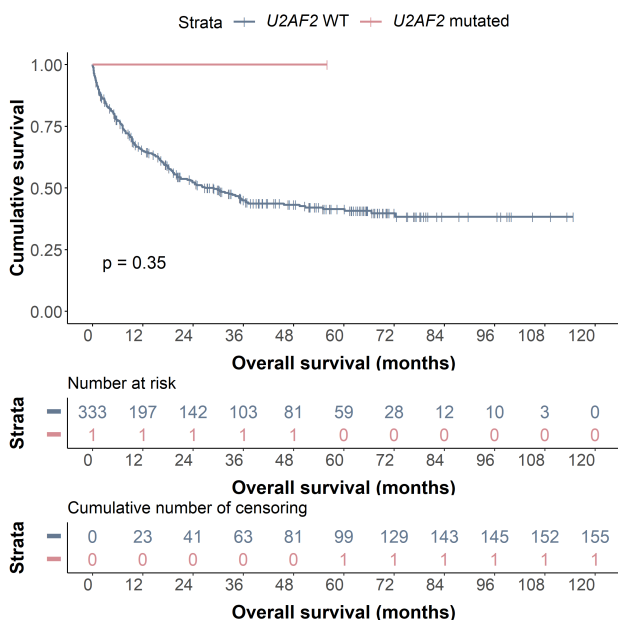
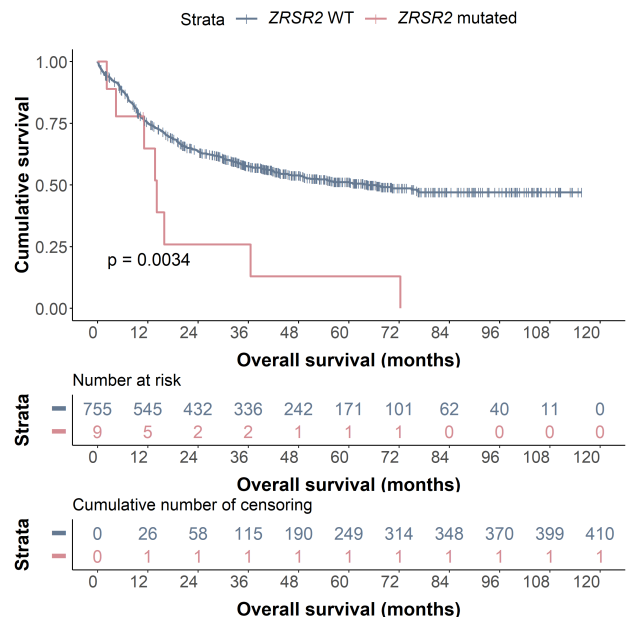
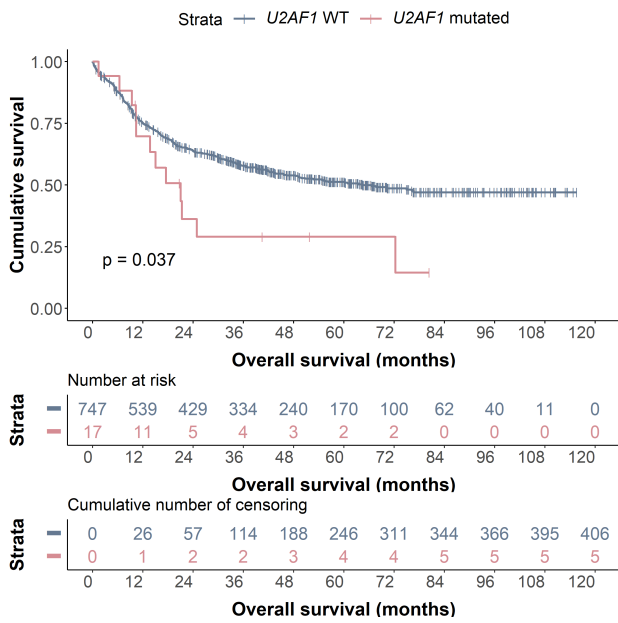
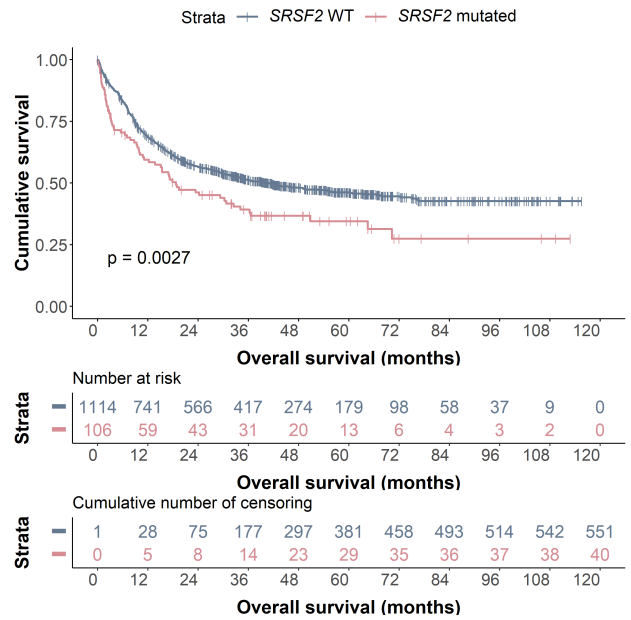
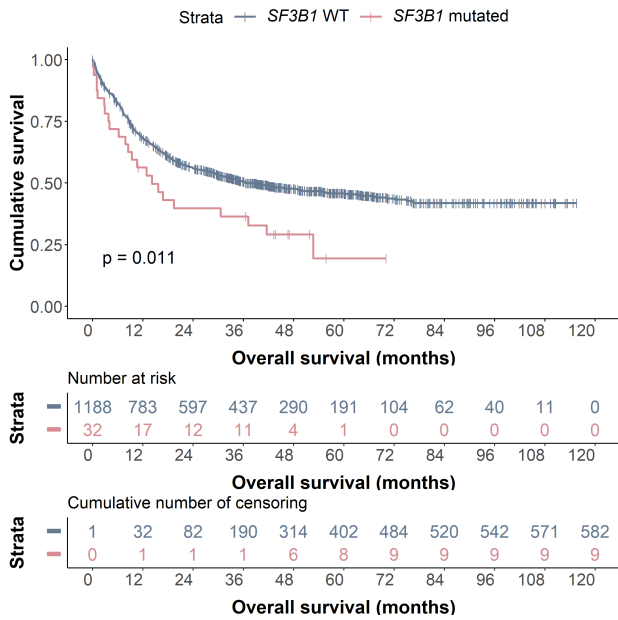
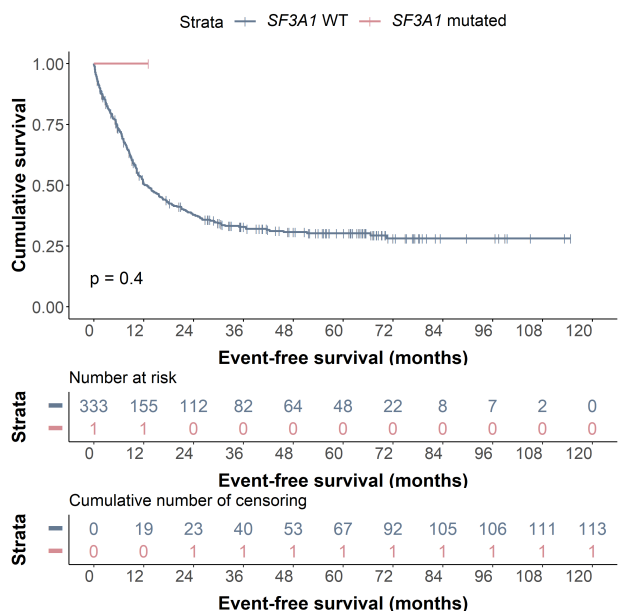
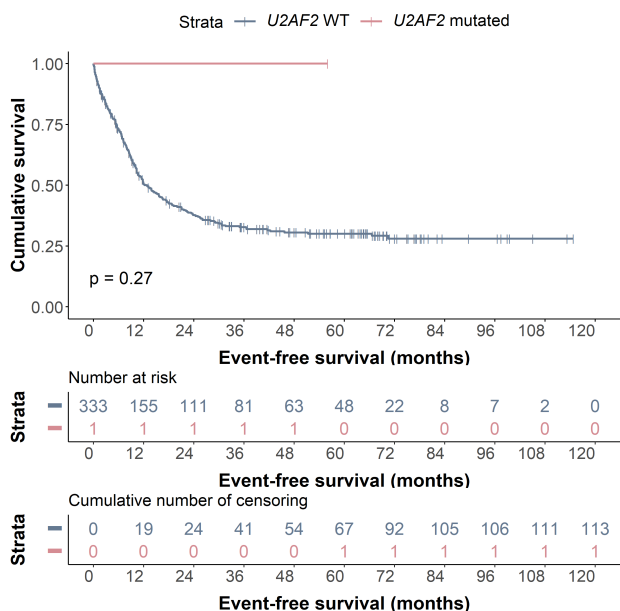
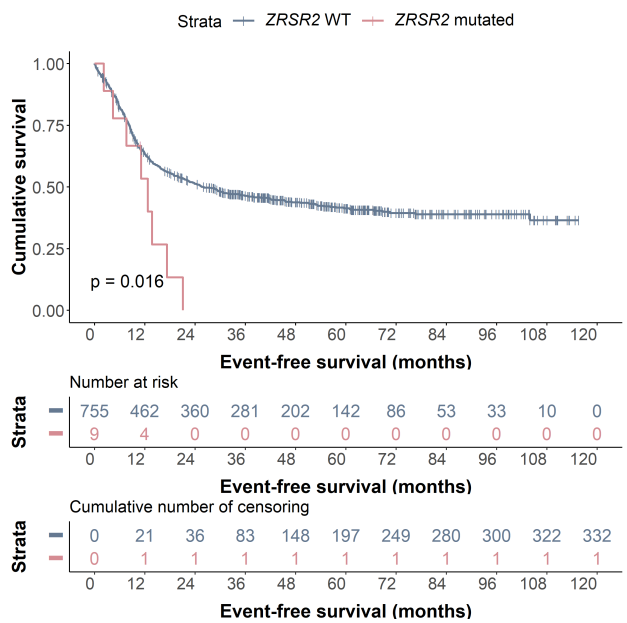
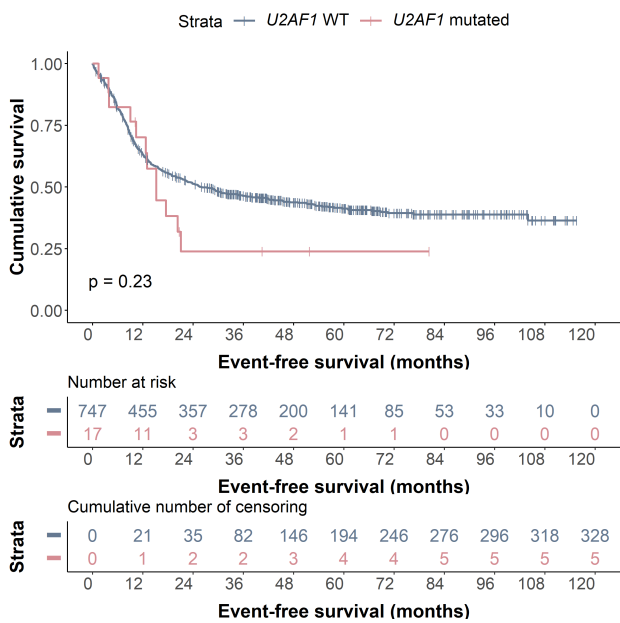
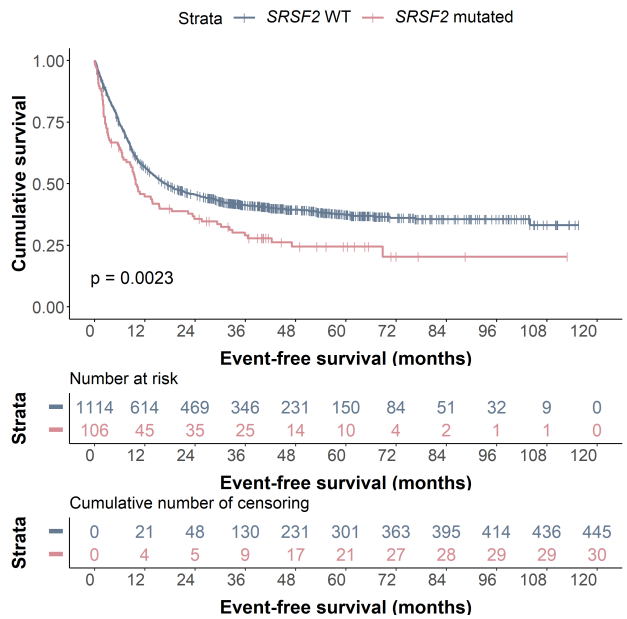
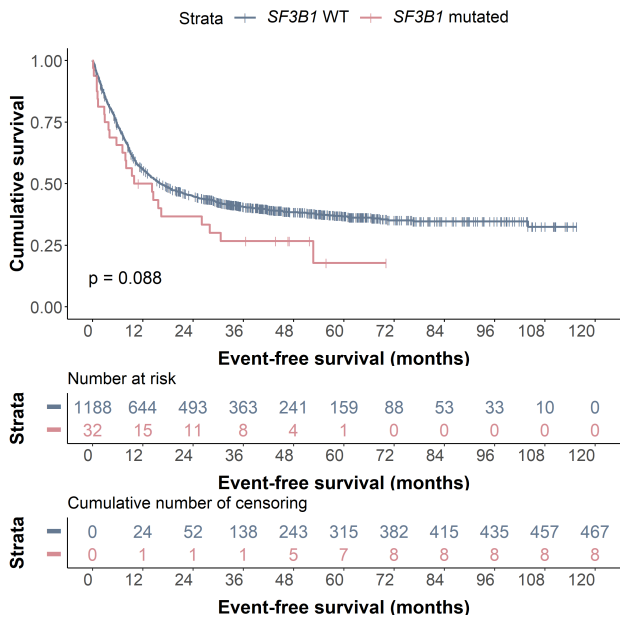
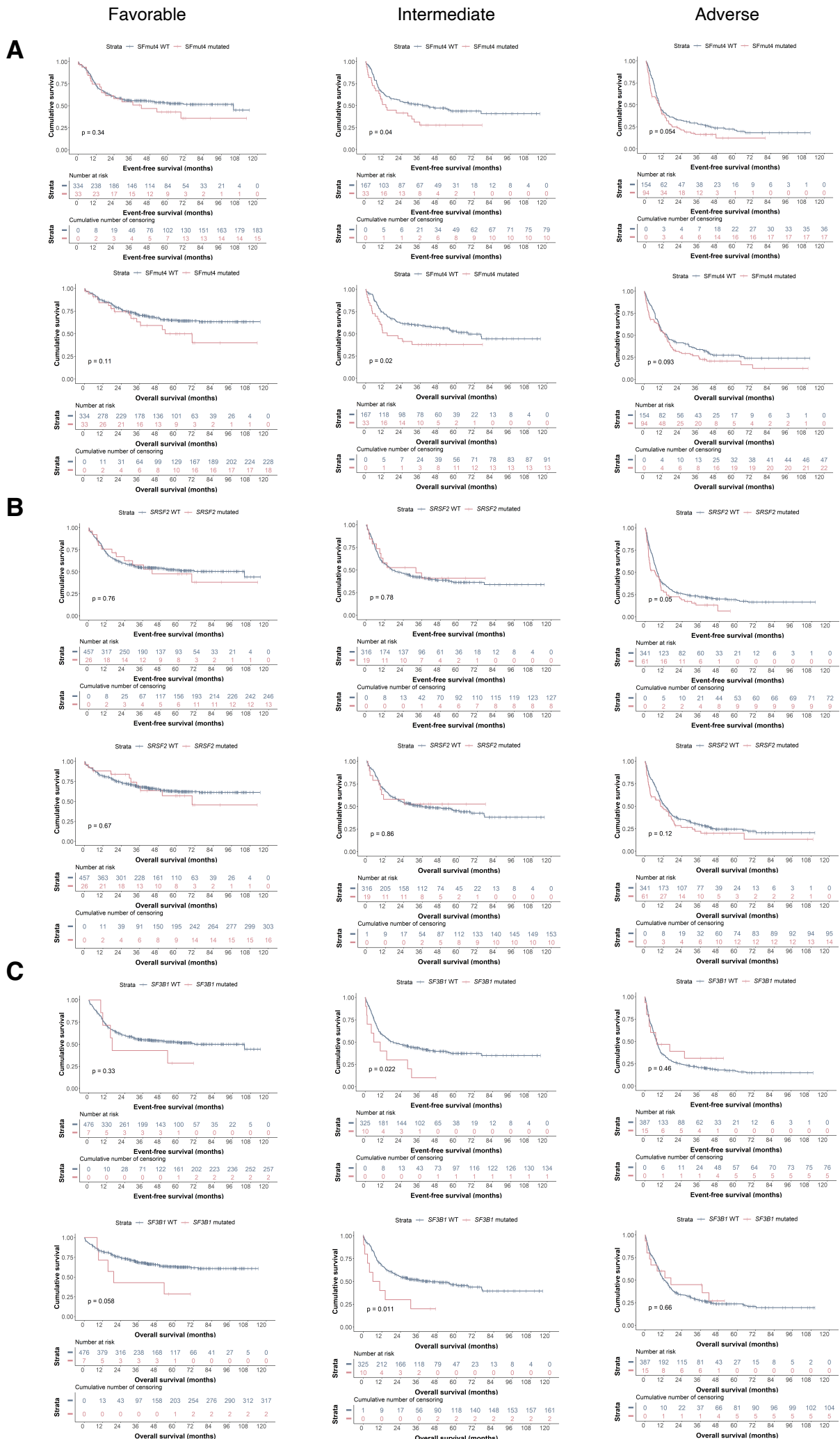
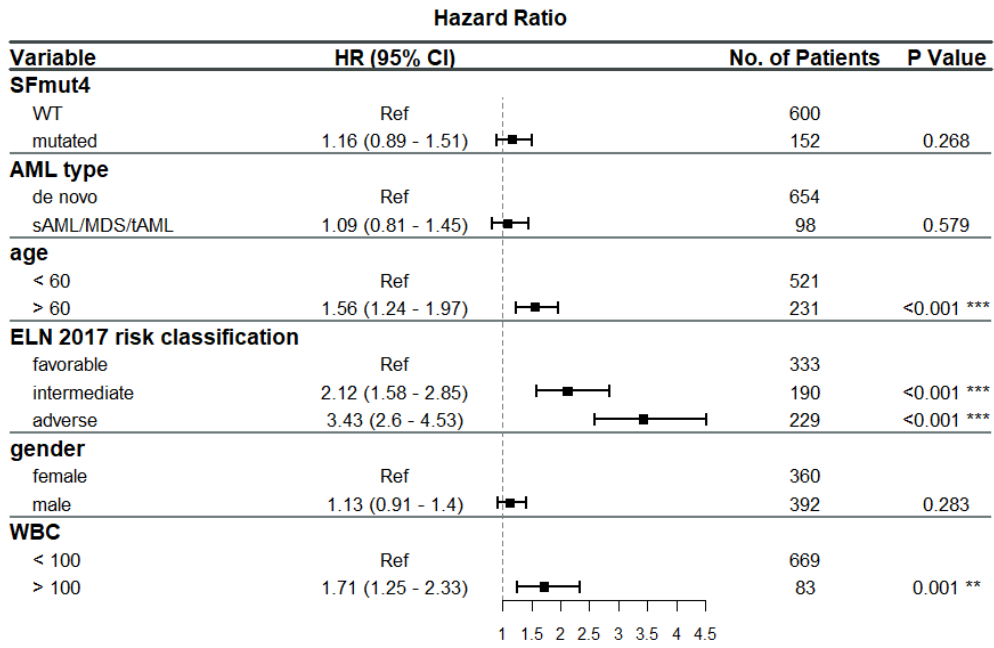


Figure S2

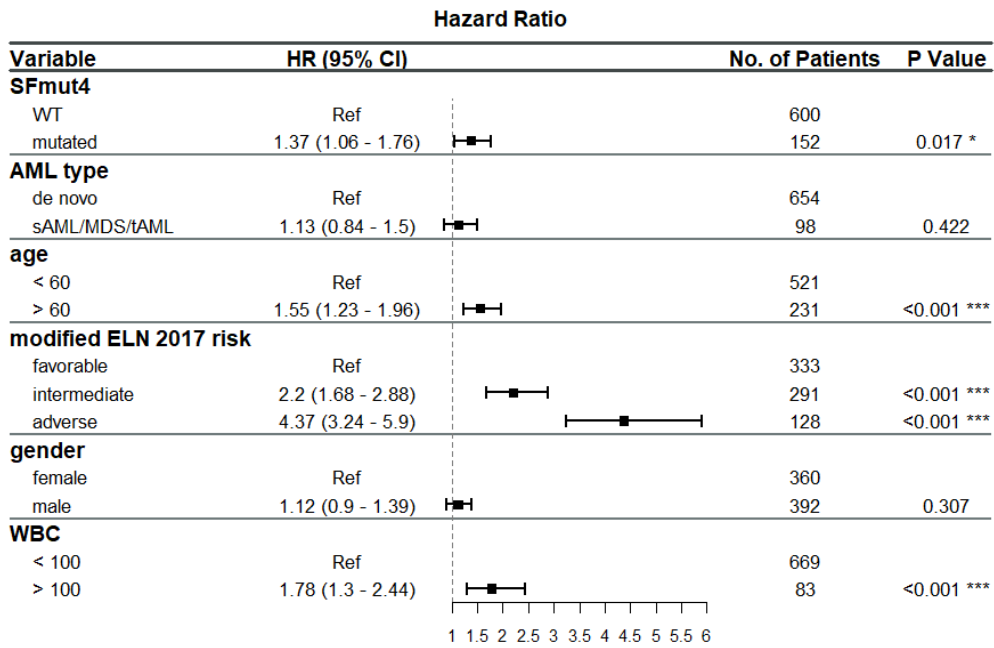




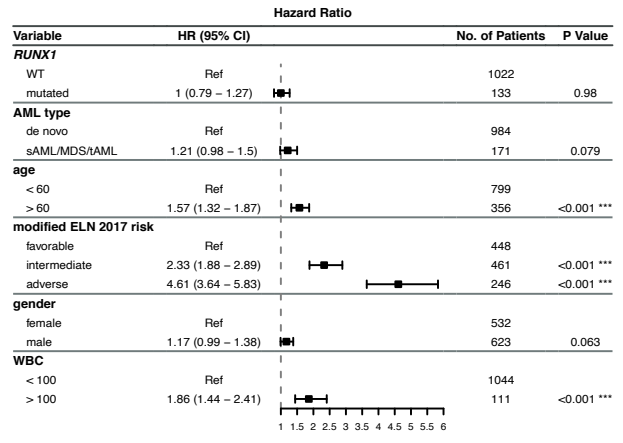
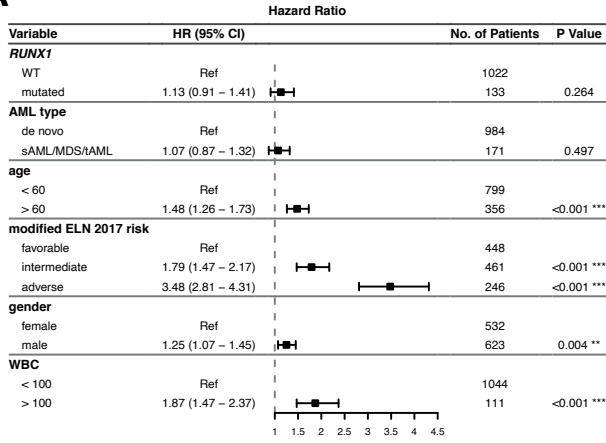
A



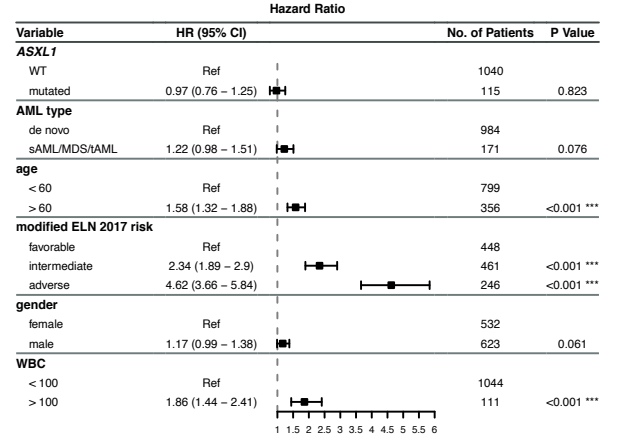
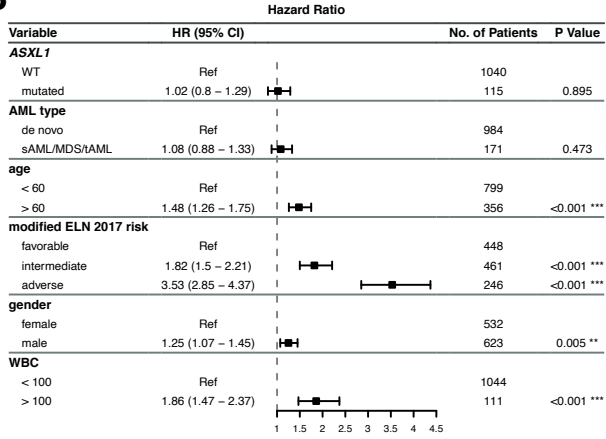
B



A



B



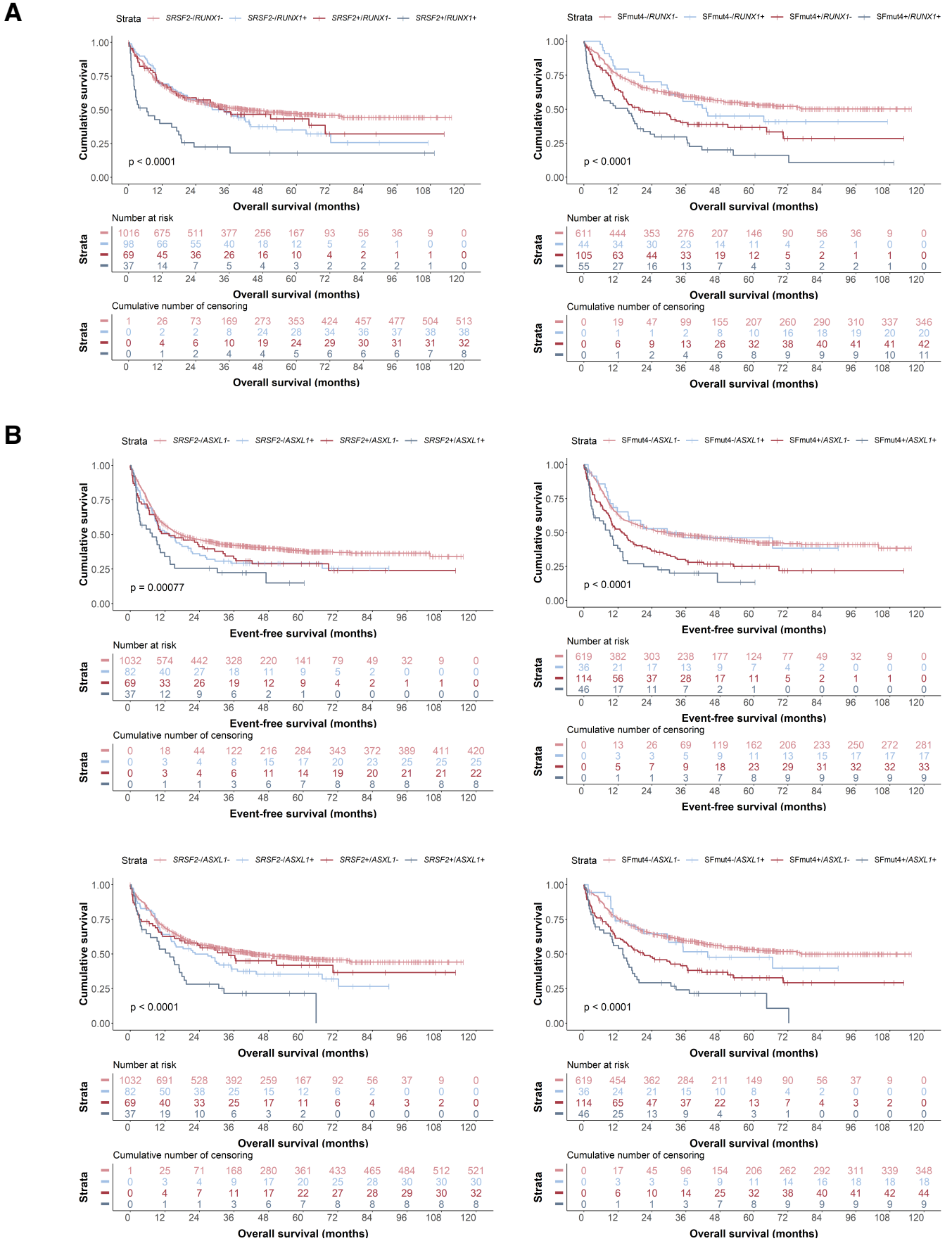
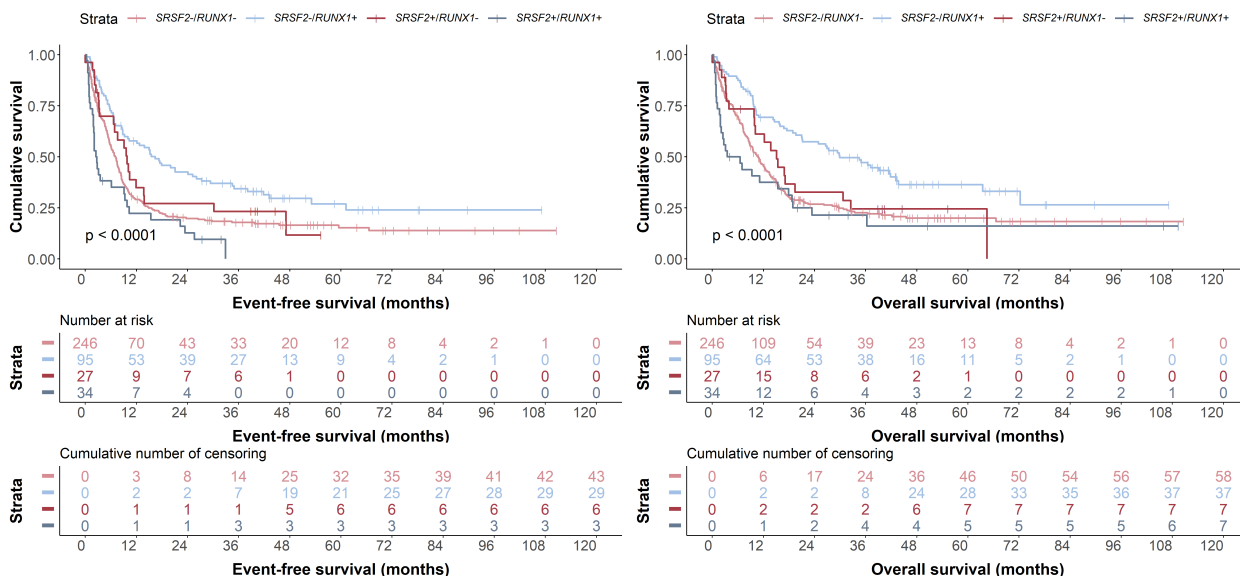
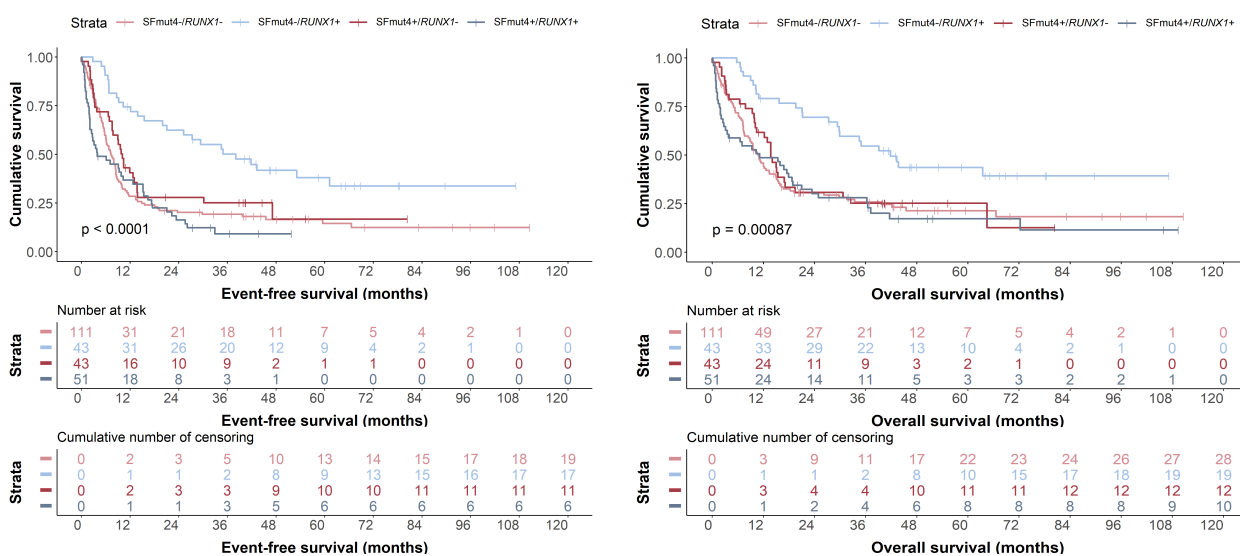


Figure S7

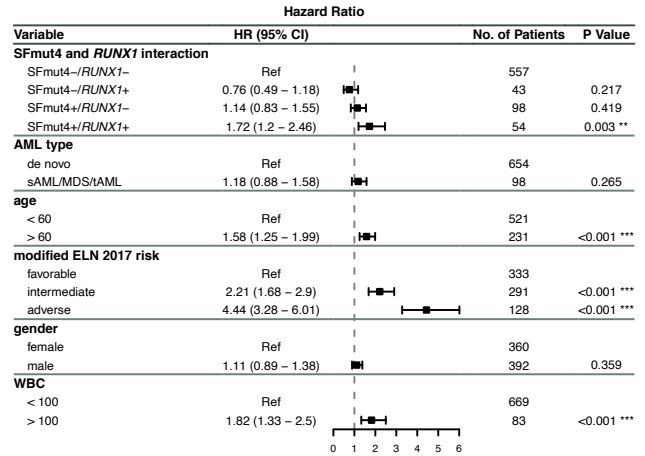
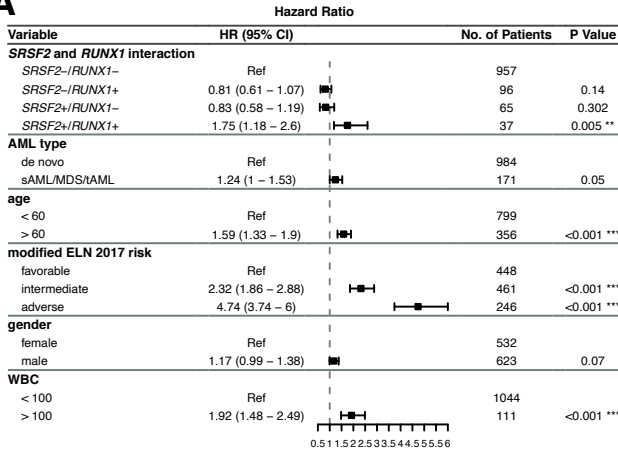
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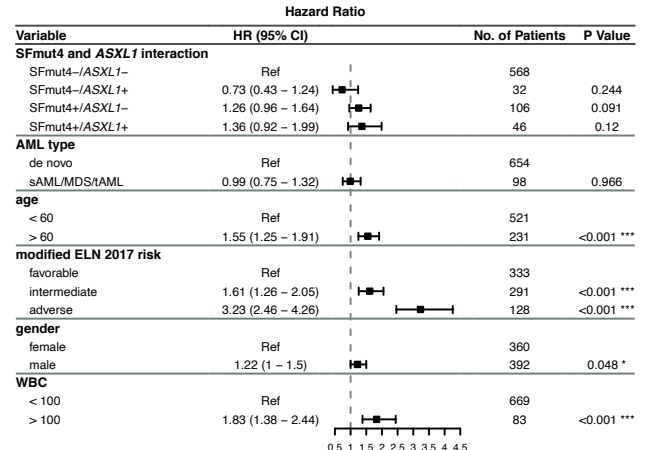
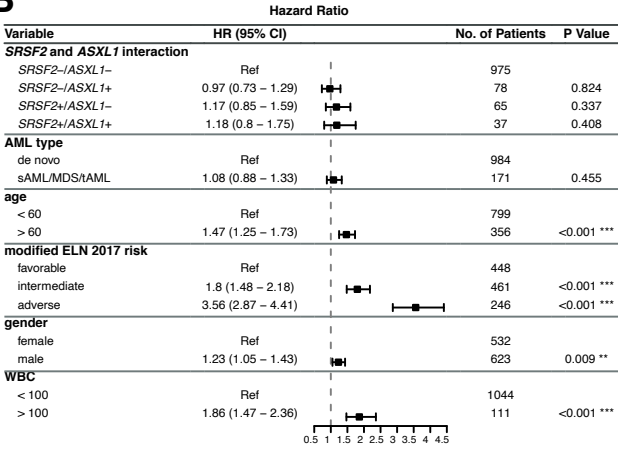
B



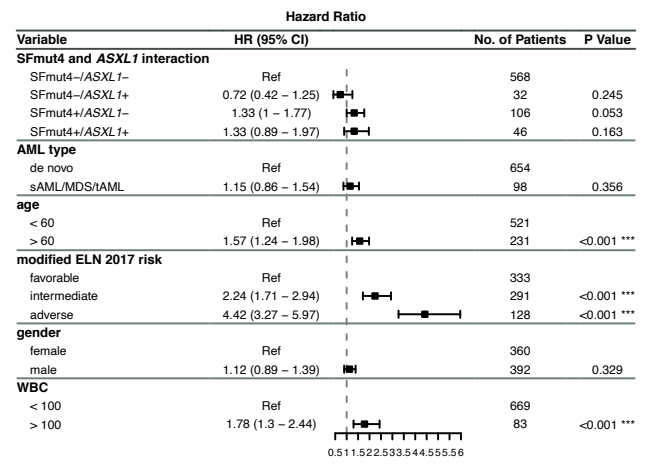
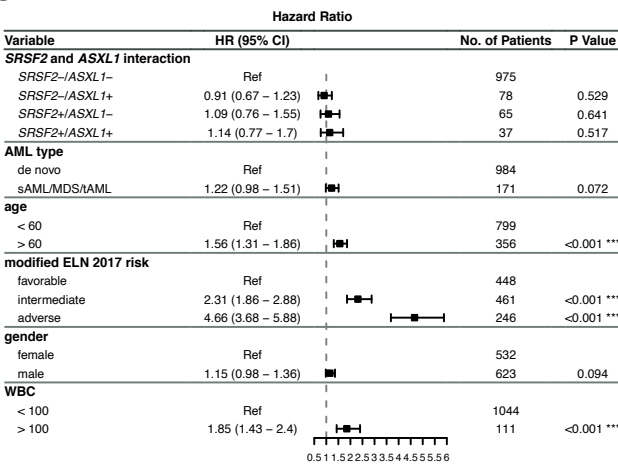
A

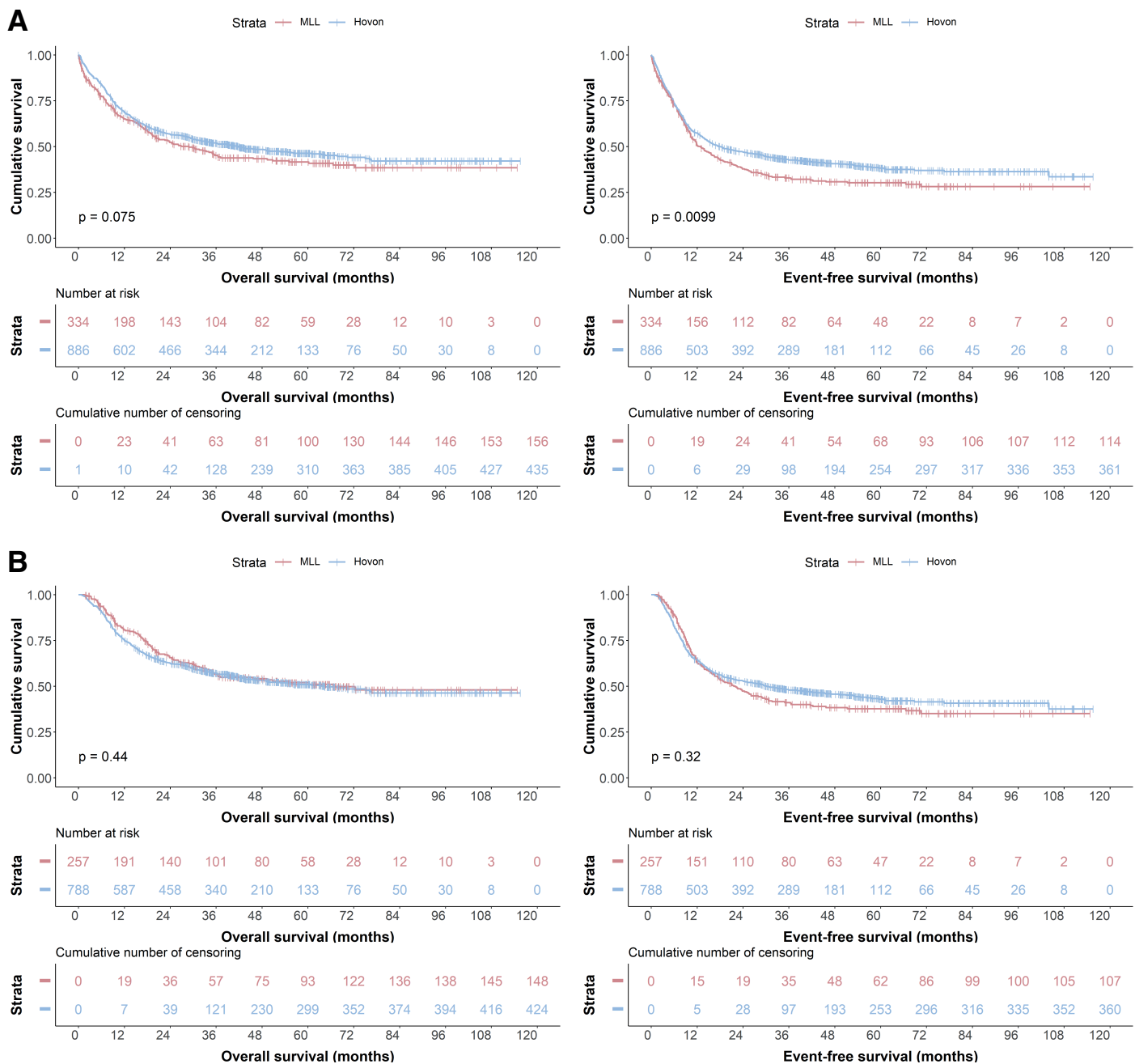


B



C

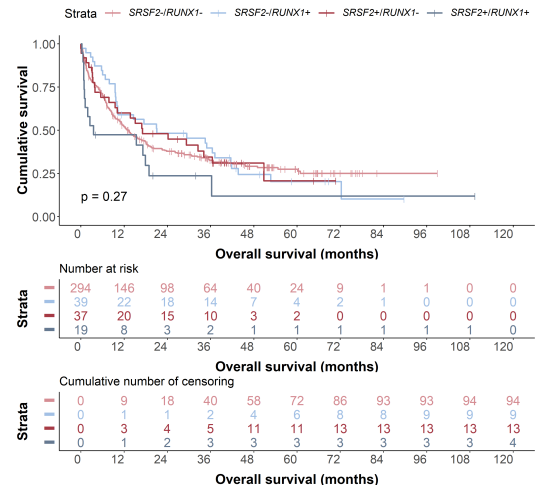
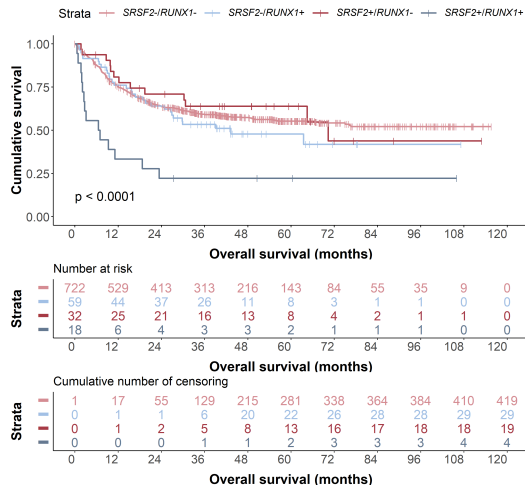




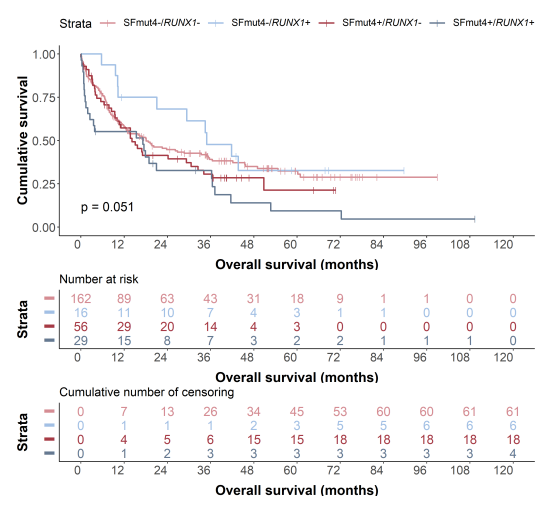
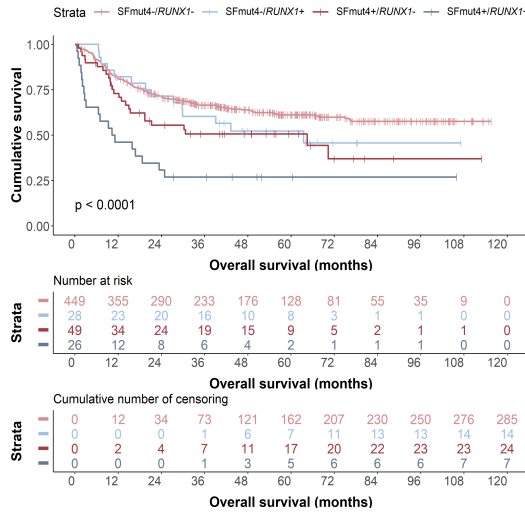
Young (<60 years)

Aged (>60 years)

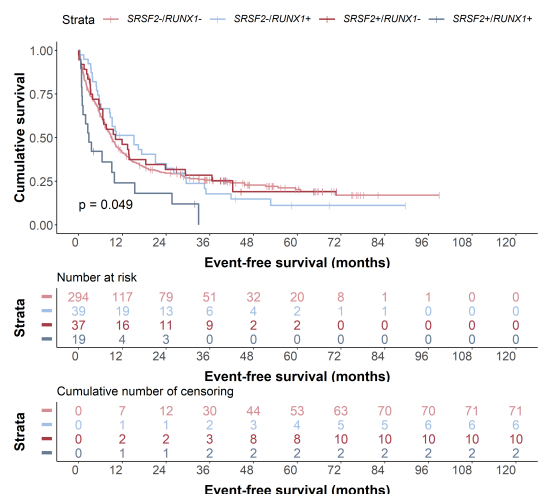
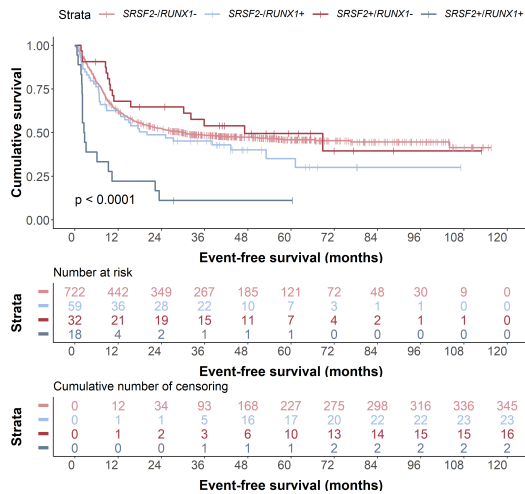
A



B



C



D

