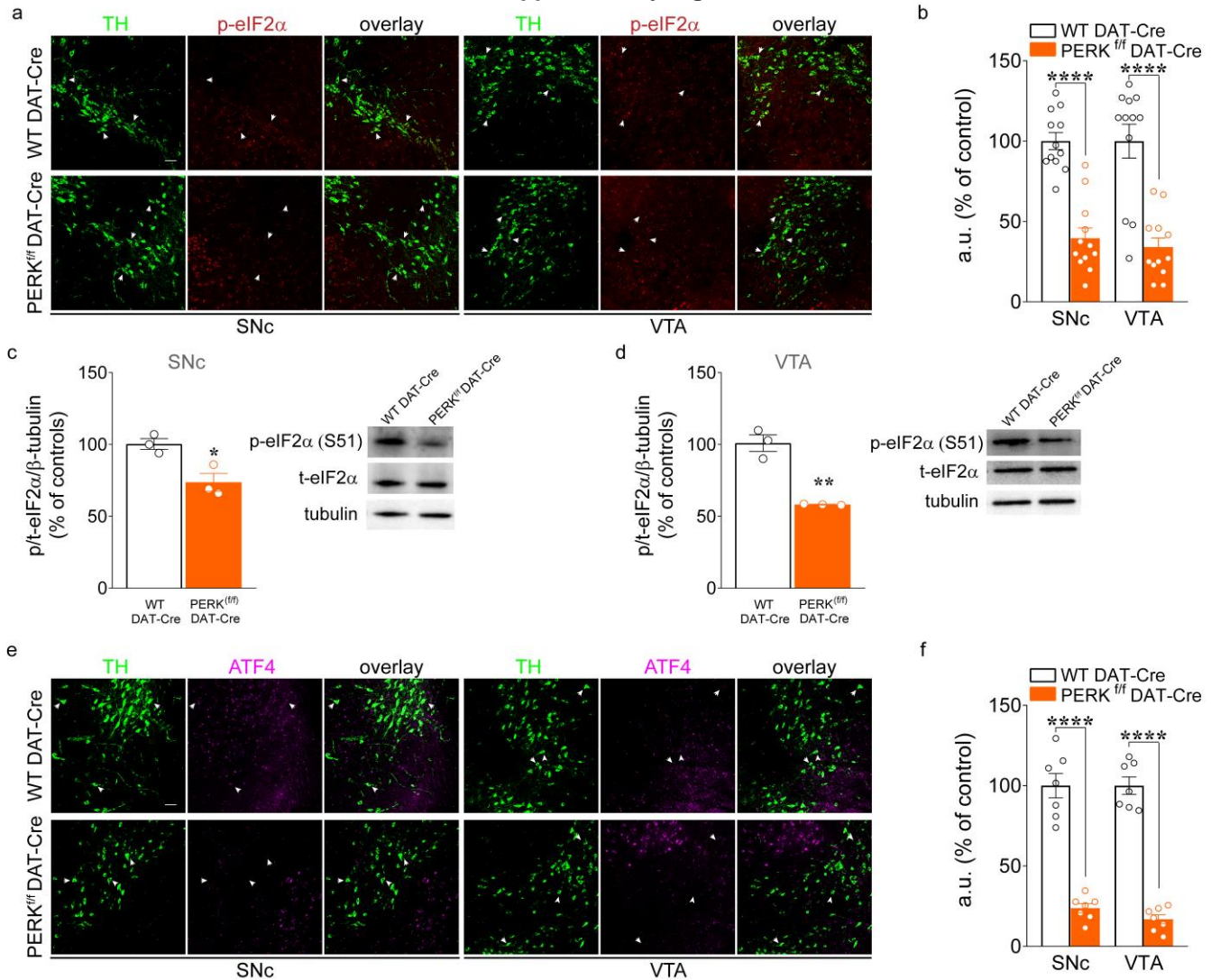


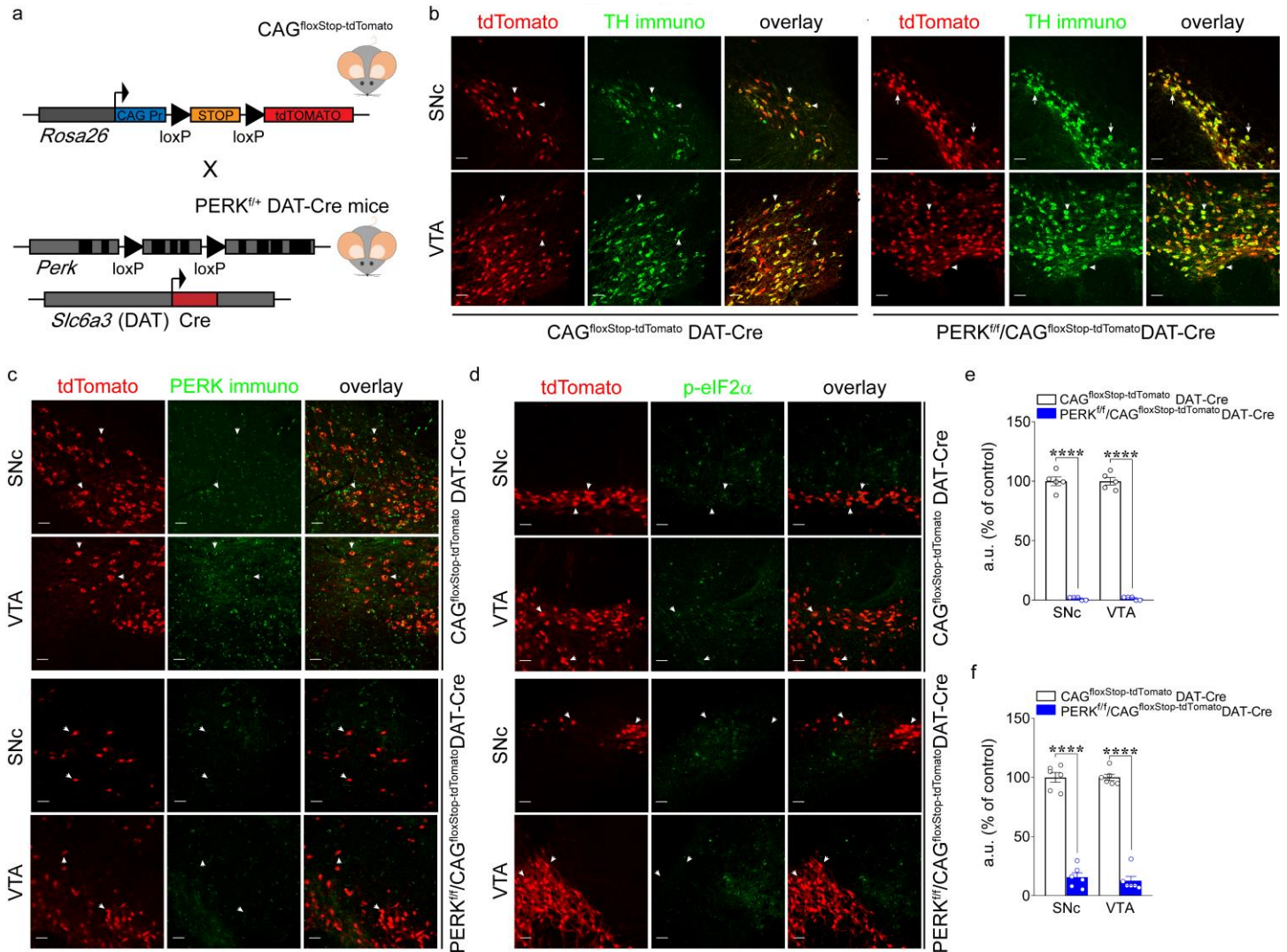
# Supplementary Figures

## Supplementary Figure 1



**Supplementary Figure 1. Cell-specific deletion of PERK in VTA and SNc DA neurons results in reduced UPR after thapsigargin-induced ER-stress.** (a) Immunofluorescent detection of TH<sup>+</sup> neuron (green) and phosphorylated eIF2α (red) in SNc and VTA DA neurons of PERK<sup>f/f</sup> DAT-Cre and WT DAT-Cre mice, in thapsigargin-treated brain slices (scale bars represent 50 μm). Arrows indicate dopaminergic neurons (green) and p-eIF2α (red) co-stain. (b) Summary plot showing reduced p-eIF2α signal after ER-stress induction, expressed as fluorescent arbitrary units (a.u.; % of control) in TH<sup>+</sup> neurons from SNc and VTA of PERK<sup>f/f</sup> DAT-Cre vs. WT DAT-Cre mice. Cell soma intensity was measured in ImageJ. Statistical significance was determined by using Student's *t* test (PERK<sup>f/f</sup> DAT-Cre vs. WT DAT-Cre mice; unpaired *t* test; SNc,  $t_{(22)} = 7.277$ ,  $P < 0.0001$ ; VTA,  $t_{(22)} = 5.496$ ,  $P < 0.0001$ ). Data are shown as mean ± s.e.m. of  $n = 12$  slices per group (average of  $n = 40$  somas per slice,  $n = 2$  slices per mouse, from three independent experiments) \*\*\*\* $P < 0.0001$ . (c,d) Representative western blots (right panel) and quantification of phosphorylation of eIF2α in SNc (c) and VTA (d) of PERK<sup>f/f</sup> DAT-Cre and WT DAT-Cre mice. Summary plot showed a significant decrease in the phosphorylation of eIF2α in both SNc (c; unpaired *t* test,  $t_{(4)} = 3.667$ ,  $P = 0.023$ ;  $n = 3$  independent lysates from 3 mice per group) and VTA (d; unpaired *t* test,  $t_{(4)} = 7.414$ ,  $P = 0.002$ ;  $n = 3$  independent lysates from 3 mice per group) of PERK<sup>f/f</sup> DAT-Cre mice compared with controls after treatment with thapsigargin. (e) Immunofluorescent detection of TH<sup>+</sup> neuron (green) and ATF4 (magenta) in SNc and VTA DA neurons of PERK<sup>f/f</sup> DAT-Cre and WT DAT-Cre mice, in thapsigargin-treated brain slices (scale bars represent 50 μm). Arrows indicate dopaminergic neurons (green) and ATF4 (magenta) co-stain. (f) Summary plot showing reduced ATF4 signal after ER-stress induction, expressed as fluorescent arbitrary units (a.u.; % of control) in TH<sup>+</sup> neurons from SNc and VTA of PERK<sup>f/f</sup> DAT-Cre vs. WT DAT-Cre mice. Cell soma intensity was measured in ImageJ. Statistical significance was determined by using Student's *t* test (PERK<sup>f/f</sup> DAT-Cre vs. WT DAT-Cre mice; unpaired *t* test; SNc,  $t_{(12)} = 9.407$ ,  $P < 0.0001$ ; VTA,  $t_{(12)} = 13.66$ ,  $P < 0.0001$ ). Data are shown as mean ± s.e.m. of  $n = 7$  slices per group (average of  $n = 40$  somas per slice,  $n = 2-3$  slices per mouse, from three independent experiments) \*\*\*\* $P < 0.0001$ .

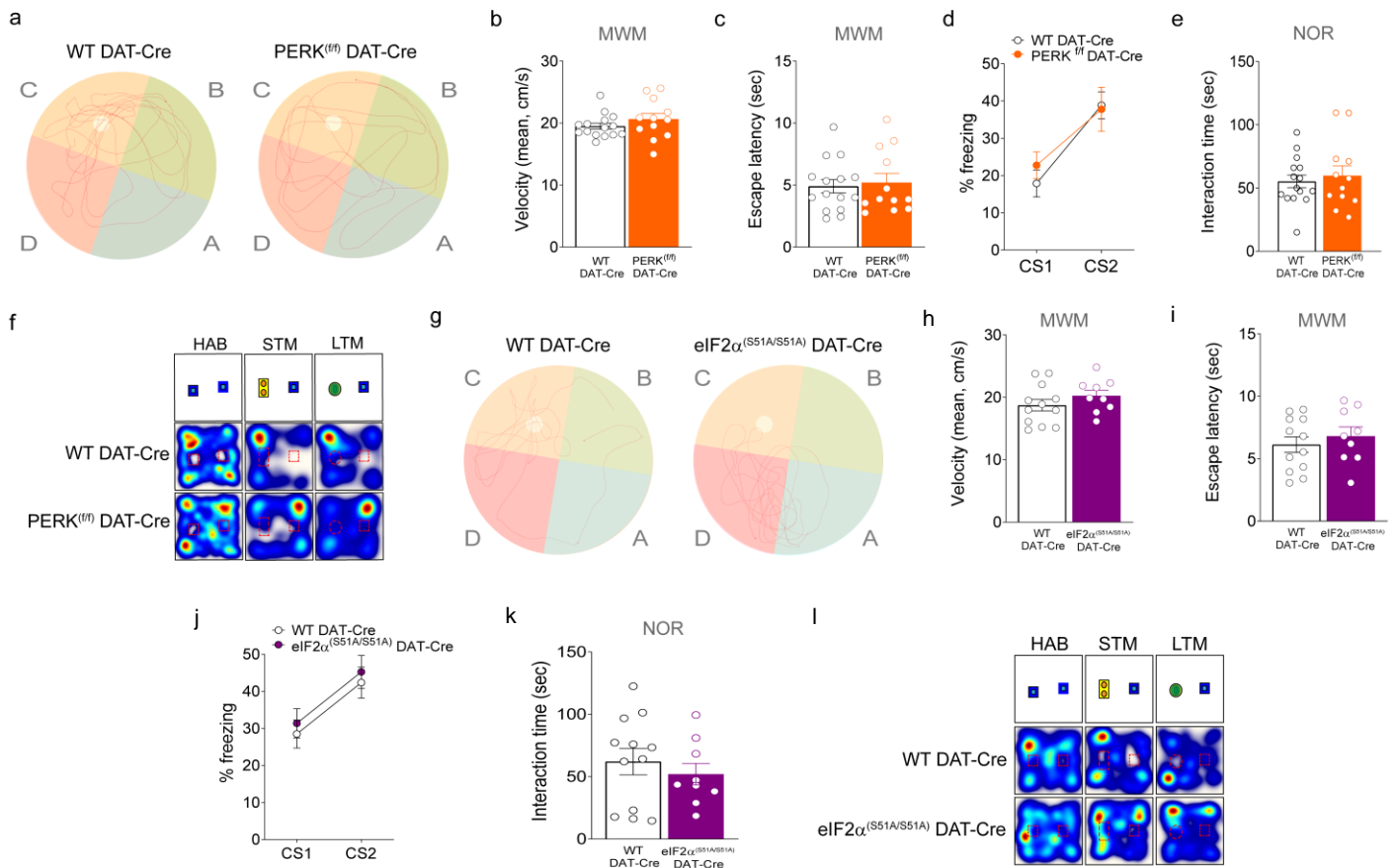
## Supplementary Figure 2



### Supplementary Figure 2. Gt(ROSA)26Sor<sup>tm14(CAG-tdTomato)Hze</sup> Cre-dependent reporter mice crossed with PERK<sup>fl/fl</sup> DAT-Cre mice confirm positive targeting of DAT<sup>+</sup> neurons for deletion of PERK and reduction of phosphorylated eIF2α.

(a) Schematic representation of DAT<sup>+</sup> neuron-specific expression of tdTomato in DAT-Cre mice crossed with Gt(ROSA)26Sor<sup>tm14(CAG-tdTomato)Hze</sup> Cre-dependent reporter mice. (b) Immunofluorescent detection of tdTomato (red) DAT<sup>+</sup> neurons and TH-IR (green) in SNc and VTA DA neurons of PERK<sup>fl/fl</sup>/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre and WT/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre (control) mice, confirming specificity of the Cre-system (scale bars represent 50 μm). Arrows indicate dopaminergic neurons (green) and tdTomato (red) co-stain. (c) Immunofluorescent detection of tdTomato (red) DAT<sup>+</sup> neurons and PERK (green) expression in SNc and VTA DA neurons of PERK<sup>fl/fl</sup>/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre and WT/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre mice, confirming positive targeting of DAT neurons for the deletion of PERK (scale bars represent 50 μm). Arrows indicate dopaminergic neurons (tdTomato; red) and PERK (green) co-stain. (d) Immunofluorescent detection of tdTomato (red) DAT<sup>+</sup> neurons and phosphorylated eIF2α (green) expression in SNc and VTA DA neurons of PERK<sup>fl/fl</sup>/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre and WT/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre mice, confirming reduction of phosphorylated eIF2α following deletion of PERK in DA neurons (scale bars represent 50 μm). Arrows indicate dopaminergic neurons (tdTomato; red) and p-eIF2α (green) co-stain. (e,f) Summary plot showing reduced PERK (e) and p-eIF2α (f) signal expressed as fluorescent arbitrary units (a.u.; % of control) in tdTomato<sup>+</sup> (dopaminergic) neurons from SNc and VTA of PERK<sup>fl/fl</sup>/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre vs. WT/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre mice. Cell soma intensity was measured in ImageJ. Statistical significance was determined by using Student's *t* test (PERK<sup>fl/fl</sup>/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre vs. WT/CAG<sup>floxedStop-tdTomato</sup>DAT-Cre mice; e, unpaired *t* test, SNc,  $t_{(8)} = 27.04$ ,  $P < 0.0001$ ; VTA,  $t_{(8)} = 31.14$ ,  $P < 0.0001$ ; f, unpaired *t* test, SNc,  $t_{(10)} = 15.31$ ,  $P < 0.0001$ ; VTA,  $t_{(10)} = 19.00$ ,  $P < 0.0001$ ) Data are shown as mean ± s.e.m. of  $n = 5-6$  slices per group (average of  $n = 40$  somas per slice,  $n = 2-3$  slices per mouse, from three independent experiments) \*\*\*\* $P < 0.0001$ .

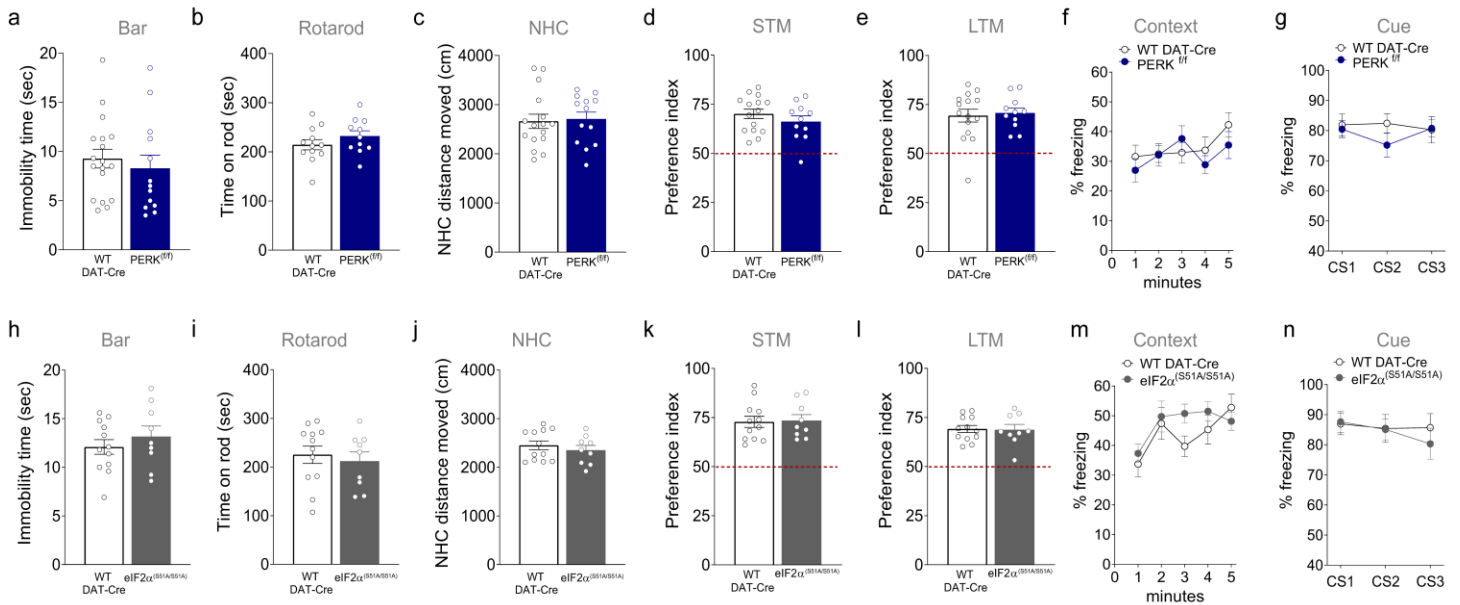
### Supplementary Figure 3



### Supplementary Figure 3. PERK<sup>fl/fl</sup> DAT-Cre and eIF2α<sup>(S51A/S51A)</sup> DAT-Cre mice exhibit normal vision during MWM test and, normal freezing behavior and object interaction during training in FC and NOR tests, respectively.

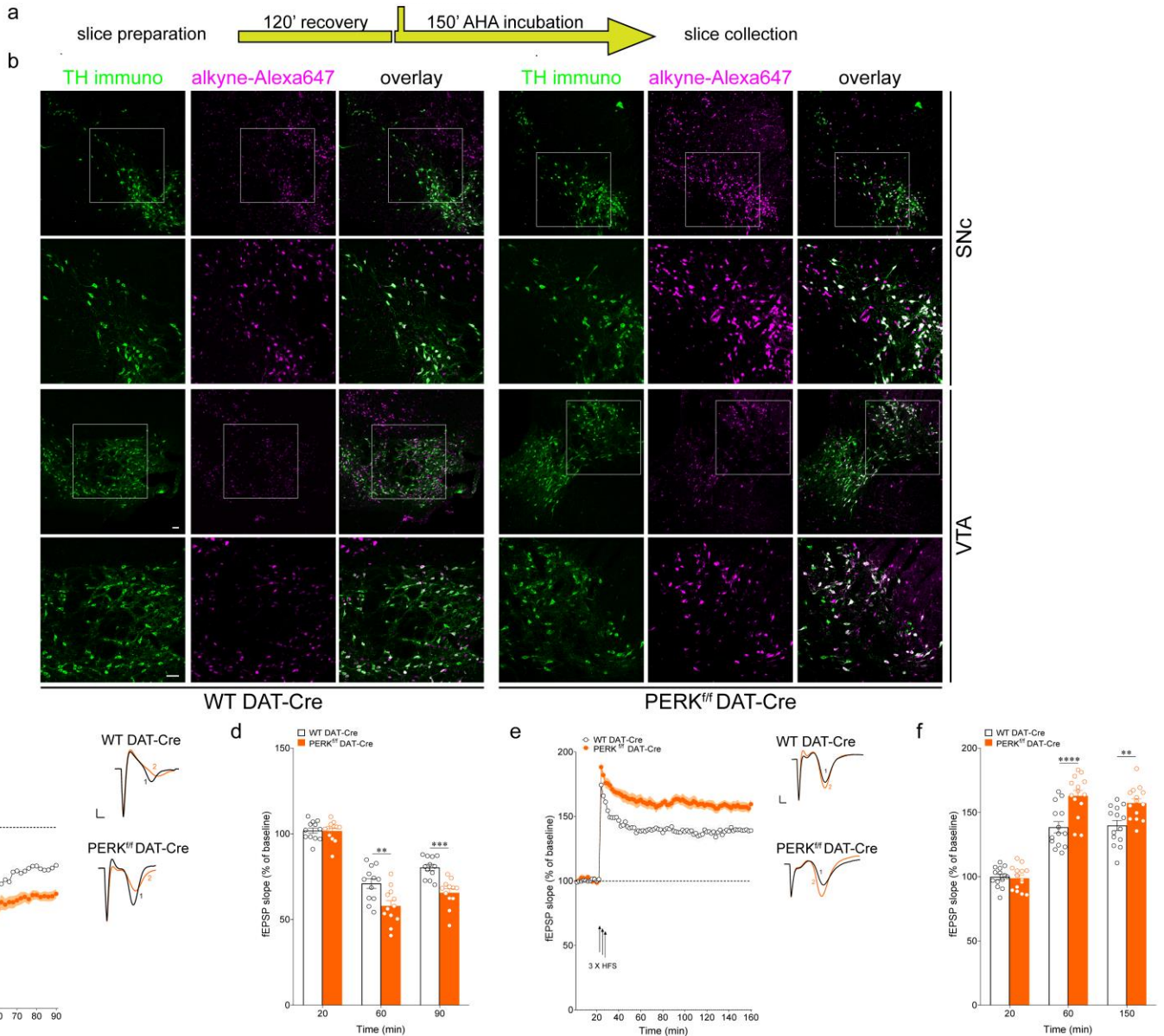
(a) Representative swim paths in the MWM test. (b-e) Summary plot of (b) average velocity during the MWM test (unpaired *t* test;  $t_{(25)} = 1.15$ ,  $P = 0.26$ ); (c) escape latency during visible platform test (unpaired *t* test;  $t_{(25)} = 0.33$ ,  $P = 0.74$ ); (d) average percentage of freezing during training in the associative threat memory test (two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, time x genotype,  $F_{(1,25)} = 0.76$ ,  $P = 0.39$ ), and (e) interaction time with familiar objects during habituation in the novel object recognition test (unpaired *t* test;  $t_{(25)} = 0.513$ ,  $P = 0.61$ ) in 3-month old PERK<sup>fl/fl</sup> DAT-Cre versus WT DAT-Cre mice. (f) Representative heat maps of the interaction with the objects for each genotype in the novel object recognition test. (g) Representative swim paths in the MWM test. (h-k) Summary plot of (h) average velocity during the MWM test (unpaired *t* test;  $t_{(19)} = 1.15$ ,  $P = 0.27$ ); (i) escape latency during visible platform test (unpaired *t* test;  $t_{(19)} = 0.71$ ,  $P = 0.49$ ); (j) average percentage of freezing during training in the associative threat memory test (two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, time,  $F_{(1,19)} = 78.34$ ,  $P < 0.001$ ), and (k) interaction time with familiar objects during habituation in the novel object recognition test (unpaired *t* test;  $t_{(19)} = 0.702$ ,  $P = 0.49$ ) in 3-month old eIF2α<sup>(S51A/S51A)</sup> DAT-Cre versus WT DAT-Cre mice. (l) Representative heat maps of the interaction with the objects for each genotype in the novel object recognition test.

### Supplementary Figure 4



**Supplementary Figure 4. Either mice harboring floxed PERK gene (PERK<sup>ff</sup>) or eIF2α<sup>(S51A/S51A)</sup> mice show no differences in motor and cognitive phenotype compared to their WT-Cre littermates.** PERK<sup>ff</sup>, eIF2α<sup>(S51A/S51A)</sup> and their respective WT DAT-Cre littermates mice were subjected to a set of tests, including the bar (a,h), rotarod (b,i) and novel home (c,j) tests to investigate locomotor activity, and to NOR (d,e; k,l) and FC (f,g; m,n) tests to investigate memory. (a,h) Summary plot of immobility time (sec) during bar test in (a) PERK<sup>ff</sup> versus WT DAT-Cre mice (unpaired *t* test,  $t_{(29)} = 0.62$ ,  $P = 0.54$ ) and (h) eIF2α<sup>(S51A/S51A)</sup> versus WT DAT-Cre mice (unpaired *t* test,  $t_{(19)} = 0.83$ ,  $P = 0.42$ ). (b,i) Summary plot of latency to fall from the rotating rod measured as average of two days (4 trials/day) test in (b) PERK<sup>ff</sup> versus WT DAT-Cre mice (unpaired *t* test,  $t_{(21)} = 1.19$ ,  $P = 0.25$ ) and (i) eIF2α<sup>(S51A/S51A)</sup> versus WT DAT-Cre mice (unpaired *t* test,  $t_{(19)} = 0.50$ ,  $P = 0.62$ ). (c,j) Summary plot of the novelty-induced locomotor activity expressed as a novel home cage (NHC) distance moved (cm) in the first 10 minutes interval of a 60 minutes test during novel home cage test in (c) PERK<sup>ff</sup> versus WT DAT-Cre mice (unpaired *t* test,  $t_{(27)} = 0.23$ ,  $P = 0.82$ ) and (j) eIF2α<sup>(S51A/S51A)</sup> versus WT DAT-Cre mice (unpaired *t* test,  $t_{(19)} = 0.74$ ,  $P = 0.47$ ). (d,e) Summary plots of preference indices of mice towards a novel object introduced in the novel object recognition test in PERK<sup>ff</sup> versus WT DAT-Cre mice (unpaired *t* test; d,  $t_{(24)} = 1.06$ ,  $P = 0.30$ ; e,  $t_{(24)} = 0.30$ ,  $P = 0.76$ ). (k,l) Summary plots of preference indices of mice towards a novel object introduced in the novel object recognition test in eIF2α<sup>(S51A/S51A)</sup> versus WT DAT-Cre mice (unpaired *t* test; k,  $t_{(19)} = 0.16$ ,  $P = 0.88$ ; l,  $t_{(19)} = 0.12$ ,  $P = 0.90$ ). (f,g) Summary plots of average percentage of freezing during (f) exposure to the context 24 hours after training, and (g) exposure to 3 CS presentations in a novel context in the associative threat memory test PERK<sup>ff</sup> versus WT DAT-Cre mice (f, two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, genotype,  $F_{(1, 28)} = 0.38$ ,  $P = 0.54$ ; g, two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, genotype,  $F_{(1, 28)} = 0.53$ ,  $P = 0.47$ ). (m,n) Summary plots of average percentage of freezing during (m) exposure to the context 24 hours after training, and (n) exposure to 3 CS presentations in a novel context in the associative threat memory test eIF2α<sup>(S51A/S51A)</sup> versus WT DAT-Cre mice (m, two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, genotype,  $F_{(1, 22)} = 0.97$ ,  $P = 0.34$ ; n, two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, genotype,  $F_{(1, 22)} = 0.10$ ,  $P = 0.75$ ). 3-month old mice were analyzed using Student's *t* test or two-way RM ANOVA, followed by Bonferroni's test for multiple comparisons. All data are shown as mean ± s.e.m. of  $n = 13-18$  (a),  $n = 11-12$  (b),  $n = 13-16$  (c),  $n = 9-12$  (h-l),  $n = 11-15$  (d,e),  $n = 15$  (f,g), and  $n = 12$  (m,n) mice/genotype.

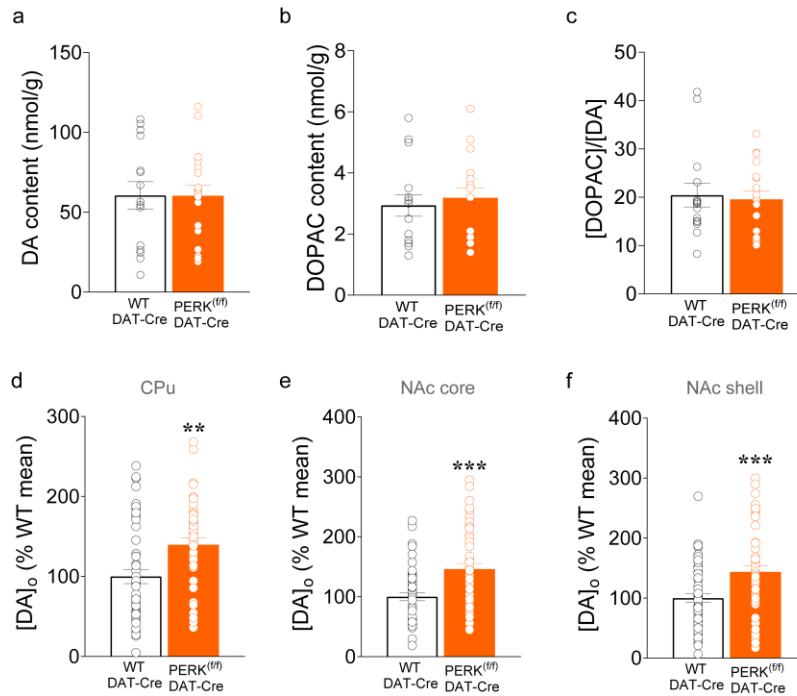
### Supplementary Figure 5



### Supplementary Figure 5. Deletion of PERK in DA neurons causes dysregulated *de novo* translation in mice and alters both striatal and hippocampal synaptic plasticity.

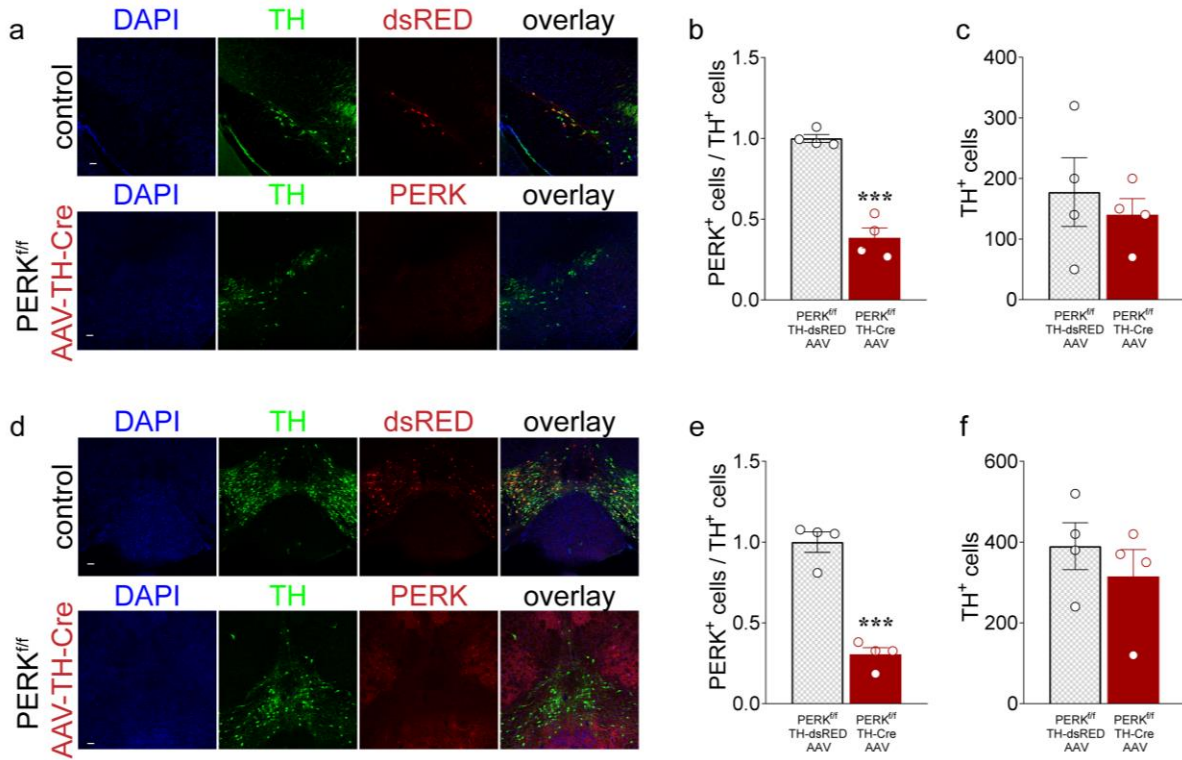
**(a)** Schematic for FUNCAT experiments. **(b)** Representative immunofluorescence images of TH-IR (green) and incorporation of AHA (magenta) detected by FUNCAT with alkyne-Alexa 647 in SNc and VTA DA neurons from midbrain coronal slices of 3-month old PERK<sup>f/f</sup> DAT-Cre mice and their WT DAT-Cre littermates. Insets in the first and third rows are magnified in the second and fourth rows, respectively (scale bars represent 50  $\mu$ m). **(c,d)** Striatal long-term depression (LTD) in 3-month old PERK<sup>f/f</sup> DAT-Cre vs. WT DAT-Cre mice. **(c)** Plot showing normalized fEPSP mean slope ( $\pm$ s.e.m. displayed every 2 min) recorded from striatal slices. **(d)** Mean fEPSPs at baseline (20 min), at 60 (40 min after tetanus) and at 90 min (70 min after tetanus). LTD evoked by 3 trains of high frequency stimulation (HFS) was significantly enhanced in PERK<sup>f/f</sup> DAT-Cre striatal slices at both 60 min and 90 min (two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, time  $\times$  genotype,  $F_{(2,44)} = 6.71$ ,  $P = 0.003$ ; time,  $F_{(2,44)} = 161.60$ ,  $P < 0.0001$ ; genotype  $F_{(1,22)} = 19.85$ ,  $P = 0.0002$ ;  $n = 12$  slices from 7 mice/genotype). Data are shown as mean  $\pm$  s.e.m. **\*\***  $P < 0.01$  and **\*\*\***  $P < 0.001$  PERK<sup>f/f</sup> DAT-Cre versus WT DAT-Cre mice. **(e,f)** Hippocampal late-phase long-term potentiation (L-LTP) in 3-month old PERK<sup>f/f</sup> DAT-Cre vs. WT DAT-Cre mice. **(e)** Plot showing normalized fEPSP mean slope ( $\pm$ s.e.m. displayed every 2 min) recorded from hippocampal slices. **(f)** Mean fEPSPs at baseline (20 min), at 60 (40 min after tetanus) and at 150 min (130 min after tetanus). L-LTP evoked by 3 trains of high frequency stimulation (HFS) was significantly increased in PERK<sup>f/f</sup> DAT-Cre hippocampal slices at both 60 min and 150 min (two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, time  $\times$  genotype,  $F_{(2,52)} = 8.36$ ,  $P = 0.0007$ ; time,  $F_{(2,52)} = 162$ ,  $P < 0.0001$ ; genotype  $F_{(1,26)} = 162$ ,  $P = 0.0004$ ;  $n = 14$  slices from 8 mice/genotype). Representative traces (**c,e** left panels) are superimposed fEPSPs (scale bars represent 1 mv/msec) recorded during baseline (1) and 60 min after HFS train (2). Arrows indicate delivery of HFS.

### Supplementary Figure 6



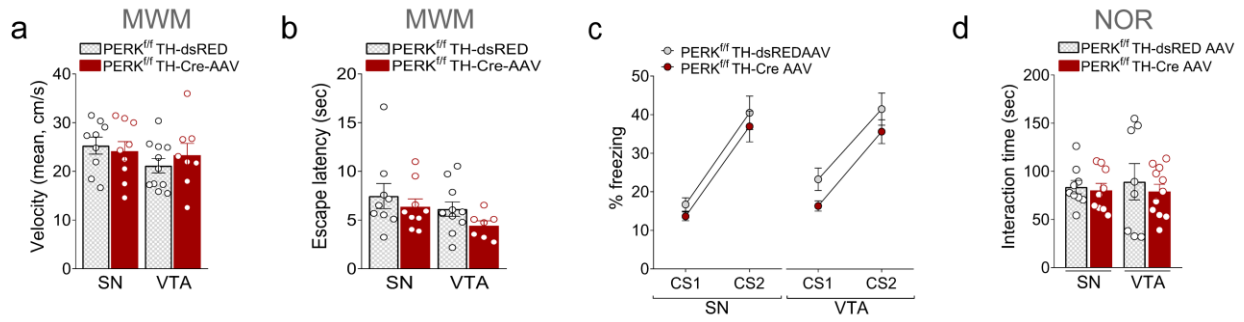
**Supplementary Figure 6. Deletion of PERK from DA neurons does not affect DA content and the effect on DA release does not involve ACh and nAChRs.** Tissue content (nmoles/g tissue wet weight) of DA (**a**), its metabolite DOPAC (**b**), was performed using HPLC with electrochemical detection in experimental striatal slices. The analysis of the metabolite/DA ratio ([DOPAC/DA]; **c**) is also reported. (**a-b**) Summary plot of average DA (**a**) and DOPAC (**b**) content in both dorsal (DS) and ventral (VS) striatum of WT DAT-Cre vs. PERK<sup>f/f</sup> DAT-Cre mice (unpaired *t*-test, n.s.). (**c**) Summary plot of average [DOPAC/DA] ratio (unpaired *t*-test, n.s.). All data are shown as mean ± s.e.m. of *n* = 15-18 mice per genotype (**a-c**). (**d-f**) Average single-pulse-evoked [DA]<sub>o</sub> in dStr (**d**, unpaired *t*-test:  $t_{(91)} = 3.160$ ,  $P < 0.01$ ), NAc core (**e**, unpaired *t*-test:  $t_{(96)} = 4.045$ ,  $P < 0.001$ ), and NAc shell (**f**, unpaired *t*-test:  $t_{(96)} = 4.491$ ,  $P < 0.001$ ) in the presence of dihydro- $\beta$ -erythroidine (Dh $\beta$ E; 1  $\mu$ M), a selective antagonist for  $\beta$ 2 subunit-containing ( $\beta$ 2\*) nAChRs that are enriched on DA axons. Data are are means ± s.e.m. normalized to 100% mean WT control for each region. \*\* $P < 0.01$ ; \*\*\* $P < 0.001$  for WT DAT-Cre vs. PERK<sup>f/f</sup> DAT-Cre mice, where *n* denotes the number of recording sites in each region sampled from 3 to 5 mice per genotype.

### Supplementary Figure 7



**Supplementary Figure 7. Selective deletion of PERK in DA neurons of the SNc and the VTA.** (a) Immunofluorescence detection of TH-IR (green) and either PERK or dsRED (red) in the SNc of PERK<sup>f/f</sup> TH-Cre-AAV and PERK<sup>f/f</sup> TH-dsRED AAV control mice (scale bars represent 50  $\mu$ m). (b) Summary data showing the ratio of TH<sup>+</sup> cells in the SNc that co-labeled for PERK in PERK<sup>f/f</sup> TH-Cre-AAV vs. PERK<sup>f/f</sup> TH-dsRED AAV control mice ( $n = 4$  mice each, unpaired t test,  $t_{(6)} = 9.373$ ,  $P < 0.001$ ). (c) Summary data showing the overall number of TH<sup>+</sup> cells in the SNc that co-labeled for PERK in PERK<sup>f/f</sup> TH-Cre-AAV vs. PERK<sup>f/f</sup> TH-dsRED AAV control mice ( $n = 4$  mice each, unpaired t test,  $t_{(6)} = 0.599$ ,  $P = 0.571$ ). (d) Immunofluorescence detection of TH-IR (green) and PERK or dsRED (red) in the VTA of PERK<sup>f/f</sup> TH-Cre-AAV and PERK<sup>f/f</sup> TH-dsRED AAV control mice (scale bars represent 50  $\mu$ m). (e) Summary data showing the ratio of TH<sup>+</sup> cells in the VTA that co-labeled for PERK in PERK<sup>f/f</sup> TH-Cre-AAV vs. PERK<sup>f/f</sup> TH-dsRED AAV control mice ( $n = 4$  mice each, unpaired t test,  $t_{(6)} = 9.120$ ,  $P < 0.001$ ). (f) Summary data showing the overall number of TH<sup>+</sup> cells in the VTA that co-labeled for PERK in PERK<sup>f/f</sup> TH-Cre-AAV vs. PERK<sup>f/f</sup> TH-dsRED AAV control mice ( $n = 4$  mice each, unpaired t test,  $t_{(6)} = 0.849$ ,  $P = 0.429$ ).

### Supplementary Figure 8



**Supplementary Figure 8. Selective PERK deletion in DA neurons of the SN or the VTA does not alter freezing behavior and object interaction during training.** (a-d) Summary plot of (a) average velocity during the MWM test (unpaired t test; d, n.s. in both SN and VTA); (b) escape latency during visible platform test (unpaired t test; d, n.s. in both SN and VTA); (c) average percentage of freezing during training in the associative threat memory test (two-way RM ANOVA, followed by Bonferroni's multiple comparisons test, n.s. in both SN and VTA), and (d) interaction time with familiar objects during habituation in the novel object recognition test (unpaired t test; d, n.s. in both SN and VTA) in 3-month old PERK<sup>fl/fl</sup> TH-Cre-AAV versus PERK<sup>fl/fl</sup> TH-dsRED AAV control mice.