Oh_Supplementary Fig.S1





Supplementary S1. B7-H3 does not affect cell viability in human HC PBMCs. (A) Monocytes
were cultured in M-CSF (40 ng/ml) for 3 days. B7-H3-specific siRNA or control siRNA (40 nM)
transfected OCPs were incubated in the presence or absence of RANKL (80 ng/ml). At day 0 and
3 of incubation, cell viability was measured using the Ez-cytox viability kit reagent. (B) OCPs
were cultured with M-CSF (20 ng/ml) and RANKL (40, 20, 5, 1 ng/ml) in the presence of B7-H3
Fc (5 µg/ml) or human IgG for 8 days. Cell viability was measured using the Ez-cytox viability
kit.



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Supplementary S2. Inhibition of B7-H3 suppresses RANKL- induced human osteocalstogensis. (A) OCPs were transiently transfected with B7-H3-specific siRNA or control siRNA (40 nM), then removed from serum for 6 hours. The cells were stimulated with RANKL (40 ng/ml) for the indicated times. Western blot was performed to detect expression of the

14	indicated proteins. The bar graphs show the densitometry analyses of the gels quantified relative
15	to the internal control. The mRNA levels were normalized relative to GAPDH expression. (B) In
16	the absence or presence of B7-H3-Fc (5 μ g/ml) for indicated time, monocytes were cultured with
17	M-CSF (20 ng/ml) for 2 days, after which, RANKL (20 ng/ml) was added and the cells cultured
18	for an additional 6 days. TRAP staining was performed and the number of TRAP-positive
19	multinucleated cells per well were counted as osteoclasts (scale bar, 200 μ m).
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Supplementary S3. B7-H3 affects osteoclast differentiation in mouse BMMs. (A, B) 24 25 Representative microCT images of the femur (upper panel) and trabecular (Tb) bone (lower panel) from WT or B7-H3^{-/-} male mice (n = 3 per group). (B) Tb bone volume per tissue volume (BV/TV), 26 27 Tb thickness (Th), number (No) and spacing (Sp) were determined by microCT analysis. (C) Mouse BMMs derived from WT or B7-H3^{-/-} male mice (n = 5 per group) were cultured in the 28 29 presence of M-CSF (50 ng/ml) and RANKL (100 ng/ml). After 4 days, TRAP staining was 30 performed, and the number of TRAP- positive multinucleated osteoclasts per well was counted. Actin ring in osteoclasts stained with FITC-phalloidin (Scale bar, 200 µm). (D) The expression of 31

32	mature osteoclast marker; mCtsk, mAcp5 and mDcstamp was analyzed by RT-qPCR at 4 days.
33	The mRNA levels were normalized relative to the expression of GAPDH. n.s., not significant.
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Supplementary S4. B7-H3 overexpression leads to decreased IFN signaling. (A, B) M-CSF
(40 ng/ml) was added on monocytes for 3 days and then the cells were transfected with human
pCMV3-B7-H3 ORF expression plasmid or pCMV3-negative control vector (2 µg) for 1 day. RTqPCR was used to evaluate the expression of B7-H3, IFNs, and IL-27. (C) Same as (A, B) except
the expression of IFN-inducible gene was analyzed. The mRNA levels were normalized relative
to GAPDH expression.

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Oh_Supplementary Fig.S5



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Supplementary S5. B7-H3 Fc induces IDO protein expression. (A) As described for Fig. 1A,
the expression of IDO mRNA during human osteoclast differentiation was detected by RT-qPCR.
(B) OCPs, cultured with M-CSF (20 ng/ml) for 2 days, were incubated with M-CSF (20 ng/ml)
and RANKL (40 or 20 ng/ml) in the presence of B7-H3-Fc (5 µg/ml) or human IgG for 6 days.
The expression of IDO, phospho-STAT1 (PY701), and STAT1 protein was analyzed by Western
blot.

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Oh_Supplementary Fig.S6





Supplementary S6. AhR siRNA is unable to inhibit osteoclastogenesis due to failure to inhibit
IDO. (A) In 1 out of 5 of the repeat experiments, AhR failed to reverse the inhibition of
osteoclastogenesis in B7-H3 deficient osteoclasts. The cells were stained for TRAP expression and
TRAP-positive multinucleated cells were counted as osteoclasts (Scale bar, 200 µm). (B) Wholecell lysates were immunoblotted with B7-H3, AhR and IDO Abs. The bar graphs show the
densitometry analyses of the gels referred to the relative internal control.