Supplementary Material

TCTP protein degradation by targeting mTORC1 and signaling through S6K, Akt and Plk1 sensitizes lung cancer cells to DNA-damaging drugs

Mini Jeong, Mi Hyeon Jeong, Jung Eun Kim, Serin Cho, Kyoung Jin Lee, Serkin Park, Jeongwon Sohn, Yun Gyu Park



Supplementary Fig. S1. TCTP protein levels were determined in various lung cancer cells including human lung carcinogenesis model cell lines (BEAS-2B, 1799, 1198, and 1170-I cells). Total proteins were analyzed by immunoblotting. β -actin was used as an internal control. Duplicate experiments were performed.







Supplementary Figures S2. mTORC2 inhibition had no effect on TCTP degradation and the activation of Akt and PLK1 by rapamycin. (A) Transfection with Rictor siRNA had no effect on TCTP degradation and the activation of Akt and PLK1 by rapamycin. (B) Rictor siRNA used in Supplementary Fig. S2A effectively suppresses mTORC2 activity. A549 cells were transfected with 100 pM siRNA for 24 hours and then treated with 100 pM rapamycin (A) or 4 μ g/ml of insulin (B) for 24 hours. Cell lysates were analyzed by immunoblotting. The band intensities were quantified and normalized to those of internal controls or the total forms, and fold changes compared with controls are presented as numbers below the bands. GAPDH protein was used as an internal control. Triplicate experiments were performed.



Supplementary Figure S3. TCTP is involved in cell survival against DNA-damaging agents. (A) TCTP knockdown by siRNA transfection enhanced p53 induction by cisplatin. A549 cells were transfected with TCTP siRNA or negative control siRNA at a final concentration of 100 pM for 24 hours and treated with 5 µM cisplatin for 3 days. (B) Irrespective of the functional status of p53, co-treatment with cisplatin and rapamycin increased the cleaved form of PARP and decreased Mcl-1. A549 cells were treated with 200 pM rapamycin and/or 10 µM cisplatin. H1299 cells were treated with 200 pM rapamycin and/or 30 µM cisplatin. After 24 hours, the cells were harvested and analyzed. (C) Overexpression of wild-type TCTP abolished the synergistic effect of rapamycin and doxorubicin on the change in protein level of cleaved PARP. A549 cells were transfected with 5.5 μ g of a plasmid expressing wild-type TCTP or an empty vector (pcDNA4, as the negative control) for 24 hours and treated with 100 pM rapamycin and/or 4 µM doxorubicin for 3 days. (D) The overexpression and knockdown of TCTP were confirmed using qRT-PCR. A549 cells were transfected with 5.5 µg of plasmid expressing wild-type TCTP or empty vector (pcDNA4) or 100 pM TCTP siRNA for 24 hours. Total protein was analyzed by immunoblotting and GAPDH protein was used as an internal control (A, B, and C). The band intensities were quantified and normalized to those of internal controls and fold changes compared with controls are presented as numbers below the bands. Total cellular RNA was extracted and analyzed by qRT-PCR and the TCTP mRNA levels were normalized to GAPDH mRNA levels (D). Triplicate experiments were performed.













Supplementary Figure S4. Rapamycin augments the efficacy of cisplatin and induces S6K inhibition and Akt activation in an A549 lung cancer xenograft model. (A) The body weight of the mice was measured as an indicator of general health conditions. (B) The within-group of tumor volumes (mm3) at each measurement point (shown in Fig. 7B) are cumulatively presented as means \pm SD. (C) Tumor tissue homogenates were analyzed by immunoblotting and triplicate experiments were performed. (D-F) The expression levels of p-S6K1, p-S6, and p-Akt obtained in Supplementary Fig. S4C were quantified, normalized to the total forms or GAPDH, and are presented as the mean \pm SD. (G) Down-regulation of Mcl-1 expression by rapamycin was abolished by pretreatment with PLK1 inhibitor. A549 cells were preincubated for 2 hours with 5 μ M cyclapolin 9 and then treated for 24 hours with 100 pM rapamycin.



Supplementary Figure S5. Treatment with 100 pM rapamycin, a concentration adequate to reduce the TCTP protein level, has little effect on cell cycle progression. Non-synchronized A549 cells were treated for 24 hours with various concentrations of rapamycin. For analysis of DNA content, cells were stained for 30 minutes at 37°C using Dulbecco's phosphate buffered saline containing 100 μ g/ml of propidium iodide and 20 μ g/ml of RNase, and the percentages of cells in the G1, S, and G2/M phases of the cell cycle were determined by flow cytometry. DNA content histograms and cell cycle distributions are shown in the upper and lower panels, respectively. Duplicate experiments were performed.

Primers for plasmid construction					
TCTP(Ser46Ala)	Forward	5'-ATTGATGAC <u>GCG</u> CTCATTGG-3'			
	Reverse	5'-CCACCAATGAG <u>CGC</u> GTCA-3'			
TCTP (Ser64Ala)	Forward	5'-GCGAAGGTACCGAA <u>GCA</u> ACAG-3'			
	Reverse	5'- ACCAGTGATTACTGT <u>TGC</u> TTCGGT-3'			
TCTP (Thr65Val)	Forward	5'-GCGAAGGTACCGAAAGC <u>GTA</u> GT-3'			
	Reverse	5'-GACACCAGTGATTAC <u>TAC</u> GCTTTCG-3'			

В

The sequences of siRNAs				
Raptor (NM_020761.2)	5'-CCUCACUUUAUUUCCAUGU(dTdT)-3'			
Rictor (NM_001285439.2)	5'- GUCCUUCACGAGAGACAGU(dTdT)-3'			
PLK1 (NM_005030.3)	5'-CCGGGAAAAAGAAGAACCAG(dTdT)-3'			
TCTP (NM_003295.2)	5'- GCAUGGUUGCUCUAUUGGA(dTdT)-3'			
GFP	5'-GCAUCAAGGUGAACUUCAA(dTdT)-3'			
AccuTarget Negative control siRNA	5'-CCUACGCCACCAAUUUCGU(dTdT)-3'			

С

Primers for qRT-PCR						
	Forward	5'-ATCCAGATGGCATGGTTGCT-3'				
TCTP (INIM_003295.2)	Reverse	5'-CGCAGGGATTTCTTTCTTTGC-3'				
GAPDH (NM_001256799.3)	Forward	5'-CAATGACCCCTTCATTGACC-3'				
	Reverse	5'-GATCTCGCTCCTGGAAGATG-3'				

Name	Supplier	Cat no.
ТСТР	Abcam	ab37506
phospho-TCTP-Ser46	Cell Signaling Technology	5251
Raptor	Cell Signaling Technology	2280
Rictor	Cell Signaling Technology	2114
S6K	Cell Signaling Technology	2708
phospho-S6K-T389	Cell Signaling Technology	9234
Cdc25C	Cell Signaling Technology	4688
phospho-Cdc25C-Ser198	Cell Signaling Technology	9529
Akt	Cell Signaling Technology	9272
phospho-Akt-T308	Cell Signaling Technology	9275
phospho-Akt-S473	Cell Signaling Technology	9271
PLK1	Cell Signaling Technology	4513
phospho-PLK1-Thr210	Cell Signaling Technology	9062
phospho-S6	eBioscience	14-9007
Ubiquitin	Santa Cruz Biotech	sc-8017
НА	Santa Cruz Biotech	sc-7392
GAPDH	Santa Cruz Biotech	sc-32233
McI-1	Santa Cruz Biotech	sc-12756
PARP	Santa Cruz Biotech	sc-8007
p53	Santa Cruz Biotech	sc-126
β-actin	AB Frontier, Korea	LF-PA0209
α-tubulin	AB Frontier, Korea	LF-PA0146

Supplementary Information

(Original blot/gel images)

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Figure 1A



Figure 1C



Figure 1D





Figure 2A



Figure 2B



Figure 2C



Figure 2D







Figure 3B

A549



Figure 3C





Figure 3D



Figure 3E



Figure 4A











Figure 4C



Figure 4D









Figure 4E







Figure 4F

Figure 5A













Figure 5C



55kDa





Figure 5C







5kDa

Figure 5E













p-S6





Figure 6B













Figure 6D







Supplementary Figure. S1 – replicate blot















Supplementary Figure. S2A – replicate blot











Supplementary Figure. S2B – replicate blot



Insulin	-	+	-	+
si-rictor	-	-	+	+
Rictor	-	An or state	Process	Variation
p-AKT(S473)	-	-		~
AKT		-	-	-
α-tubulin	1	-	-	1









Supplementary Figure. S3C – replicate blot



Supplimentary Figure. S4C











Supplimentary Figure. S4C



AKT











Supplimentary Figure. S4G

