SUPPLEMENTAL INFORMATION

Fig S1. Genes with reduced expression in the SCN after light stimulation. *Related to Figure 1.*

(A) Heatmap of the light suppressed genes.

(B-E) Representative light suppressed genes that were significantly changed after light stimulation at 30 min-, 1-, 3-, or 6- hours of

(F-H) Pathway analysis of light-reduced genes at different timepoints.



Fig S2. Genomic annotation of binding sites for H3K27ac. *Related to Figure 2.* Binding peaks were annotated in Chipseeker, promoter (\leq 1kb), promoter (1-2kb), promoter (2-3kb), 5'UTR, 3'UTR, 1st Exon, other Exon, 1st Intron, other Intron, downstream (\leq 300) and distal intergenic.

(A) Genomic annotation of the binding sites for H3K27ac from CT17 dark SCN.(B) Genomic annotation of the binding sites for H3K27ac from SCN after 1-hour light exposure.

(C) Comparison of H3K27ac sites distribution between dark and light exposed SCN. (D-E) UCSC genome browser views of *Nr4a1* and *Per2*. NPAS4 ChIP-seq (Kim et al., 2010) is shown in dark blue. H3K4me3, H3K27ac ChIP-seq, and light pulse bulk RNA-seq are shown as indicated. The bed file of the promoter (-1kb - +100 bp) is shown in green while the bed file of H3K27ac sites is shown in red. The H3K27ac sites for *Fos* and *Nr4a1* are shaded blue. The promoter region is highlighted in shaded grey.



1st Exon Other Exon 1st Intron Other Intron

Downstream (<=300) Distal Intergenic

Light

0

25

50

Percentage (%)

75

100

Fig S3. 10x sample sequencing metrics and quality control plots. *Related to Figure 3.*

X axis: a total of ten 10X Genomics single-nucleus libraries were involved in the study. Each library was generated from 7 pooled male SCN pairs.

- (A) Number of UMIs detected in each library. Nuclei containing more than 10,000 UMI were filtered out. Dashed line: 10,000 UMI. Number of genes detected in each library. Cells containing more than 4,000 genes were filtered out. Dashed line: 4000 UMI.
- (B) Quality control screening for mitochondria contamination and potential doublets. Left: Scatter plot showing the number of UMI (X-axis) vs Percentage of mitochondria (Y-axis). Right: Scatter plot showing the number of UMI (X-axis) vs Number of genes (Y-axis). Nuclei containing more than 10% mitochondrial transcripts and more than 10,000 UMI were filtered out.
- (C)Distribution of UMIs and detected genes per cells in 45 clusters. An average of 1901 UMI counts and 1100 genes were obtained per cell across all clusters.
- (D)Cell distribution for each cell type. Open bar, cells from light sample libraries; filled bar, cells from dark sample libraries.
- (E) Total extracted reads per library, unique mapped reads per library, and assigned reads per library.





D1

D3

D2

L0

D4

L1

L2

L3

L4

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Fig S4. Comparison with cell types described in analogous studies. *Related to Figure 4.*

- (A) Heatmap shows the correlation between cell types discovered in a dataset from mouse light-pulsed visual cortex (Hrvatin et al., 2018) and this study. Fisher's exact test and color reflects the adjusted *p* value.
- (B) Heatmap shows the correlation between cell types discovered in a single cell dataset from mouse arcuate hypothalamus and median eminence (Campbell et al., 2017) and this study. Fisher's exact test and color reflects the adjusted *p* value.
- (C) Heatmap shows the correlation between cell types discovered in a dataset from mouse SCN (Wen et al., 2020) and this study. Fisher's exact test and color reflects the adjusted *p* value.





Fig S5. Genes used to identify SCN cells and FISH confirmation of IEG expression in SCN peptidergic neurons. *Related to Figure 5.*

- (A) Three groups of marker genes were used to identify potential SCN cell clusters: canonical SCN marker genes, non-SCN marker genes, and unspecific marker genes. The clusters marked with Y are potential SCN clusters and they were used for reclustering. The clusters marked with X do not express canonical SCN markers and therefore were not used for the reclustering. Size of the dot reflects the percentage of cells that expressed genes of interest and the color intensity reflects the average expression.
- (B) Clustree analysis depicting the stability of the cluster across different resolutions. Nodes are colored by stability score. Size corresponds with the number of nuclei. Arrow width corresponds with the proportion of nuclei shared between clusters at different resolutions.
- (C) Expression of Fos, Egr1 and Per1 (green) in Vip (red) cells.
- (D) Expression of Fos, Egr1 and Per1 (green) in Avp (red) cells.
- (E) Expression of Fos, Egr1 and Per1 (green) in Prok2 (red) cells.
- (F) Expression of *Fos, Egr1 and Per1* (green) in *Gfap* (red) cells. Dapi (blue) was used to stain nuclei. *Map2* is a neuronal marker. SCNs were dissected after a 1-hour light pulse at CT17. Scale bar 25 μm.



Vip+

Avp+

Prok2/Cck+

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Fos C Vip Fos	D Avp Fos 25 μm	E Prok2 Fos
Egr1	Avp Egr1	Prok2 Egrif
Per1	Avp Per1	Prok2 Per1

 Gfap, Map2, Dapi

 Fos
 Map2
 Dapi

 Gfap
 Egr1
 Map2
 Dapi

 Fos
 Gfap
 Egr1
 Map2
 Dapi

 Per1
 Gfap
 Per1
 Map2
 Dapi

Fig S6. Phase of entrainment in *Npas4* mutants to 12-h light:12-h dark (LD 12:12). *Related to Figure 6.*

- (A) Representative actogram of *Npas4^{fx/fx}* and *Npas4* cKO per sex. Records are double-plotted, and each horizontal line represents 48 hr of activity.
- (B) Average period in constant darkness and constant light (*Npas4*^{fx/fx}, n = 12, *Npas4* cKO, n = 13). Mean ± s.e.m. are shown. (**** p < 0.0001, ** p < 0.01, unpaired Student's *t* test)

