Supplementary Figures

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PN16





Figure S1. IDH1 regulated expression levels of HIF-1 α and TGF- β 1. (A) Alpha-ketoglutaric acid blocks induction of HIF-1 α in cells expressing IDH1 R132H. PN12 (left) or PN16 (right) GSCs were transfected with wild-type IDH1 or IDH1 R132H, and different concentrations of octyl- α -KG ester were added to each type of transfected cell for 4 h. HIF-1 α protein levels were assayed by Western blotting. (B) IDH1 knockdown or mutant does not affect TGF- β 1 mRNA. After PN12 (left) or PN16 (right) GSCs were transfected with control or siRNA targeting IDH1 or with IDH1 R132H mutant, expression of TGF- β 1 mRNA was measured by quantitative polymerase chain reaction. (C) IDH1 knockdown or mutant reduces TGF- β 1 protein. After PN12 (left) or PN16 (right) GSCs was transfected with control or siRNA targeting IDH1 or with IDH1 R132H mutant, expression of TGF- β 1 protein was analyzed by Western blotting.



Figure S2. Competitive interaction of E2F4 and EP300 with Smad3. (A) Western blotting results showing the molecular subtype marker expression in proneural GSCs (PN12, PN16, and PN19), and mesenchymal GSCs (ME23, ME27, and ME29) with or without TGF-β1 treatment for 4 h or 8 h. Proneural markers: CD133 and SOX2. Mesenchymal markers: CD44 and POSTN. (B) Lysates from HEK293 cells expressing His-Smad3 and Myc-EP300 (or HA-E2F4) were subjected to IP followed by IB with anti-HA and anti-Myc antibodies. (C) GST pull down utilizing purified GST-Smad3 and HA-E2F4 (or Myc-EP300)-expressing HEK293 cell lysates, followed by IB with anti-HA and anti-Myc antibodies. IP, immunoprecipitation; IB, immunoblotting.



TGF- β /**Smad-signaling pathway.** (A) Domain structure of human DHHC proteins and Smad3 protein. (B) Palmitoylation and phosphorylation of Smad3 in ME27 cells transfected with Smad3 C421A mutant constructs or infected with ZDHHC19 siRNA for 48 h. (C) Palmitoylation of Smad3 in ME23 cells treated with 2-BP (a palmitoylation inhibitor) or PalmB (a de-palmitoylation inhibitor) for 24 h. (D) ME23 cells were cultured in medium containing 100 µM palmitic acid-azide for 6 h and the cell lysates were prepared for the Click-iT reaction, followed by immunoblotting with the Smad3 antibody. (E) Western blot analyses of Smad3 distribution in ME23 cells transfected with ZDHHC19 siRNA, Smad3 C421A mutant plasmid, or treated with the de-palmitoylation inhibitor, PalmB (1 µM) for 48 h. HAM, hydroxylamine; IP, immunoprecipitation.



Figure S4. Inhibitory effect of GN44028, 2-BP, C646, and (E)-SIS3 in proneural GSCs and mesenchymal GSCs. (A) Western blot analyses of E2F4 or HIF-1 α in proneural GSCs (PN12, PN16, and PN19) transfected with E2F4 shRNA or HIF-1 α shRNA, and ZDHHC19 in mesenchymal GSCs (ME23, ME27, and ME29) transfected with ZDHHC19 shRNA. (B) Viability of proneural PN12 GSCs treated with GN44028 at the indicated concentration for 24 h. (C) Viability of mesenchymal ME23 GSCs treated with C646, 2-BP, or (E)-SIS3 at the indicated concentration for 24 h.



Figure S5. Change in molecular subtype *in vivo*. (A) Results of real time polymerase chain reaction analysis showing the molecular subtype marker expression (proneural markers, CD133; mesenchymal markers, CD44) in mice bearing a single subtype of tumor and mice bearing a mixed orthotopic tumor with PN12 and ME23 cells treated with 10 mg/kg GN44028, 7.5 mg/kg 2-BP, and 15 mg/kg (E)-SIS3 alone or in combination (n=5 per group) after 6 weeks. (B) Results of acyl-biotin exchange and Western blotting showing palmitoylation or phosphorylation of Smad3 in mice

bearing a single subtype of tumor and mice bearing a mixed orthotopic tumor with PN12 and ME23 cells treated with 10 mg/kg GN44028, 7.5 mg/kg 2-BP, and 15 mg/kg (E)-SIS3 alone or in combination (n=5 per group) after 6 weeks.