

SUPPLEMENTAL MATERIAL

Mr-AbaA regulates conidiation by interacting with the promoter region of *Mr-veA* and *Mr-wetA* in *Metarhizium robertsii*

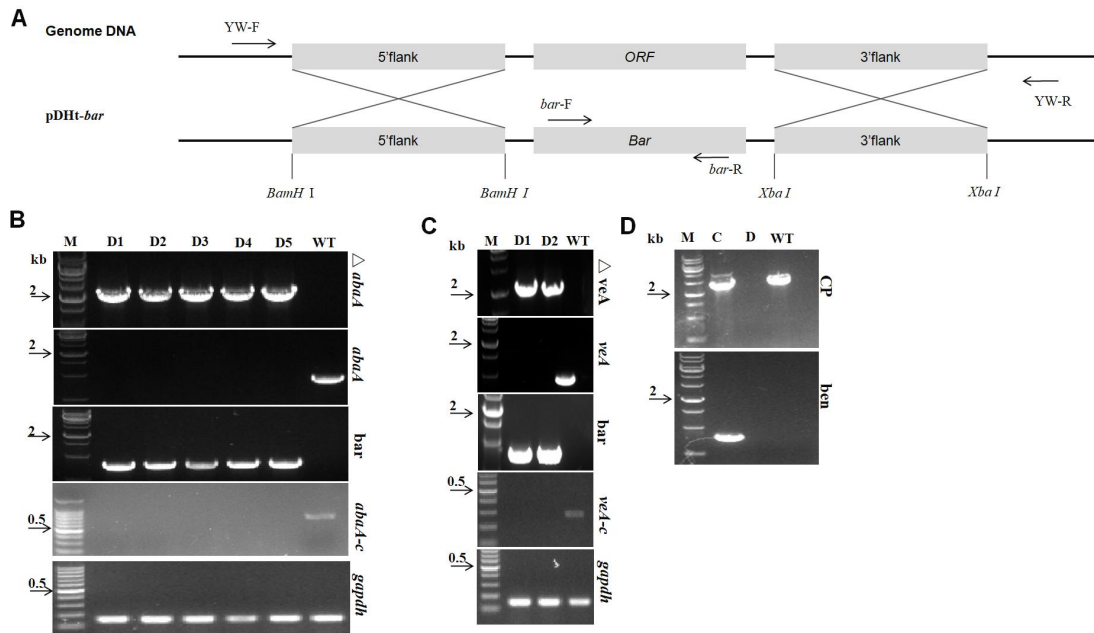


Figure S1. Gene deletions of *Mr-abaA*, and *Mr-veA* in *M. robertsii* ARSEF 23. (A) Schematic diagram of gene deletions by the homologous recombination approach. (B, C, D) Confirmation of the *Mr-abaA*, *Mr-veA* and the complemented strain of *Mr-veA* via PCR and RT-PCR. The targeted genes and *gapdh* were amplified using DNA from a different strain or cDNA as templates, respectively. M: marker. D: gene deletion mutant; WT: the wild-type strain. C: complemented strains

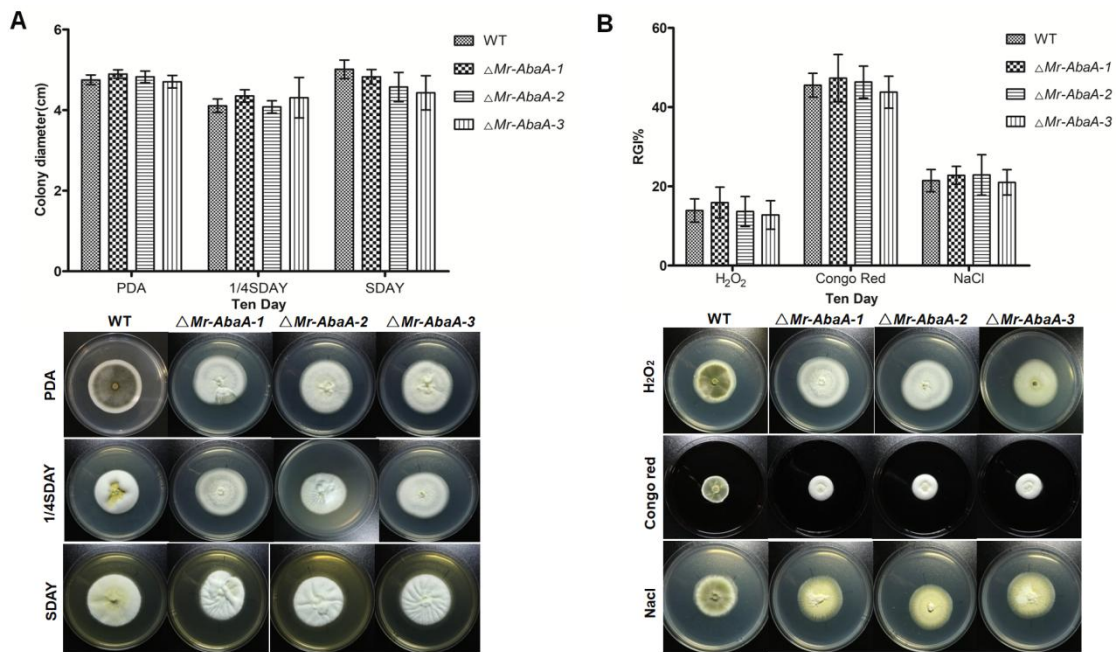


Figure S2. Effects of *Mr-abaA* deletion on hyphal growth of *M. robertsii* on different media. (A) The colony diameters of each strain were measured after 10 dpi on PDA, SDAY, and 1/4 SDAY plates. The colony phenotype of each strain cultured for 10 days on the above medium was evaluated. **(B)** Relative growth inhibition (RGI) of each strain fungal colony after 10 days of culture under H₂O₂, Congo red, and NaCl stress on PDA. The colony phenotype of each strain cultured under the above stress conditions for 10 days was evaluated.

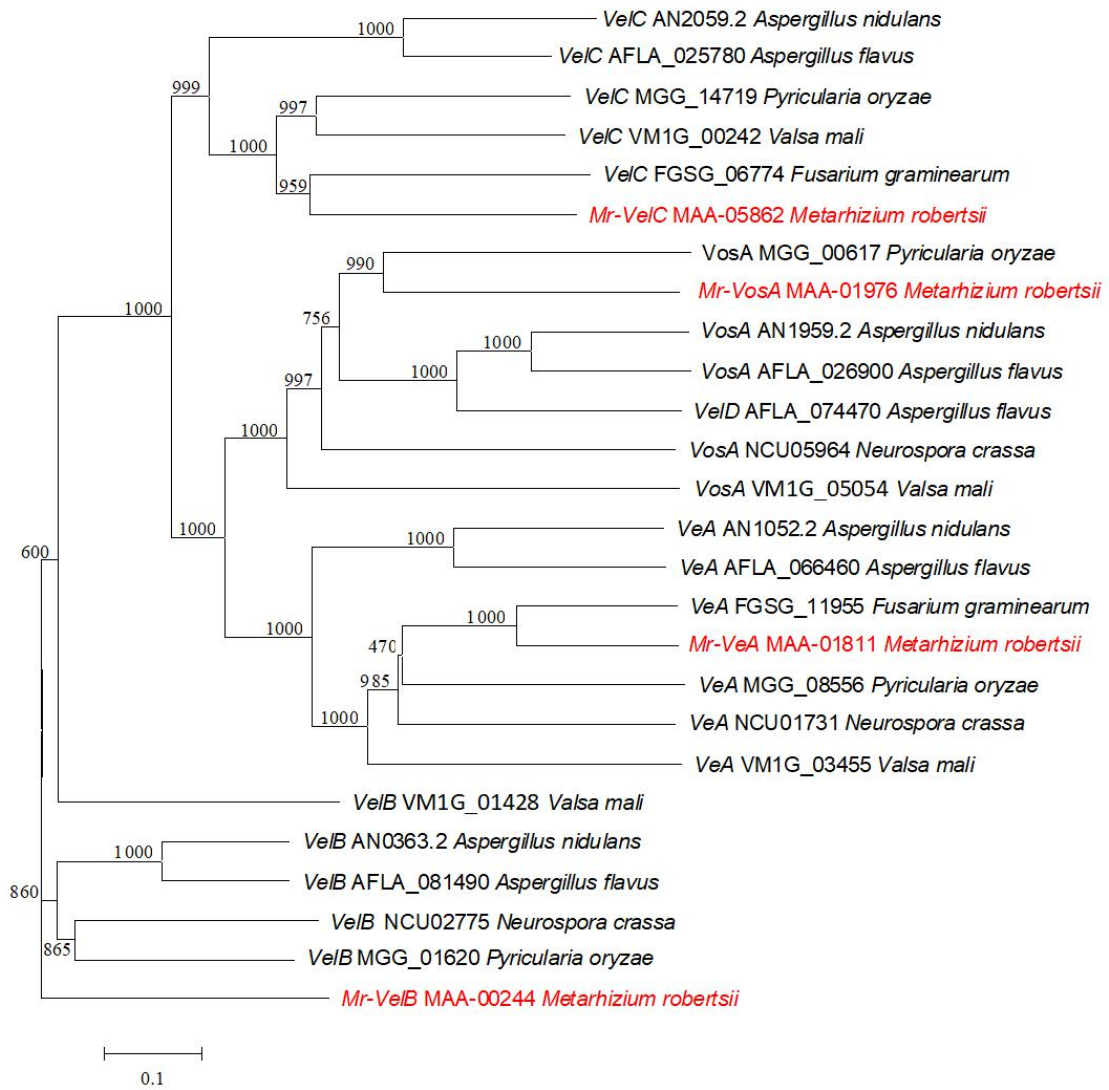


Figure S3. Phylogenetic tree analysis of the *M. robertsii* velvet family genes.

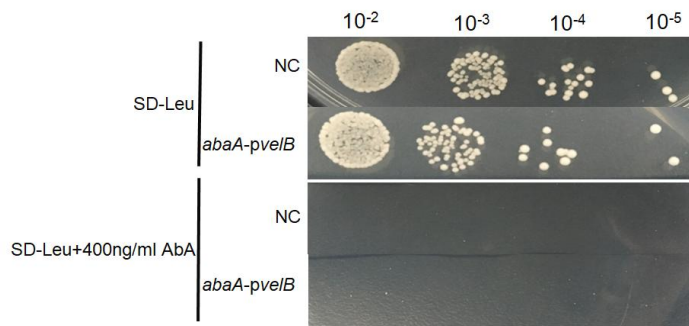


Figure S4. Analysis of the protein-DNA interaction. Yeast one-hybrid assay to test the interactions of Mr-AbaA with the *Mr-velB* promoter regions NC: Negative control. *abaA-pvelB*: interaction between the Mr-AbaA protein and the *Mr-velB* promoter region. AbA, aureobasidin A.

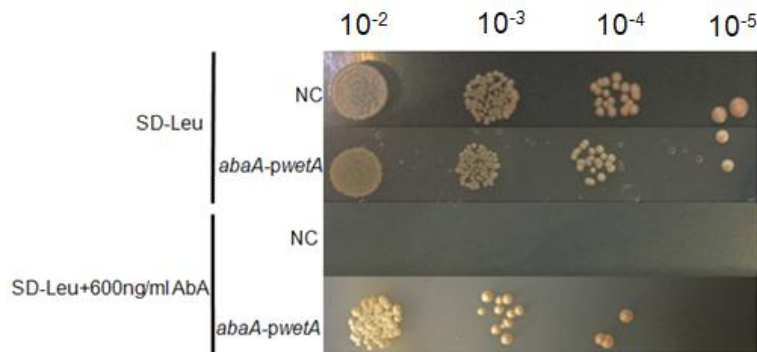


Figure S5. Analysis of the protein-DNA interaction. Yeast one-hybrid assay to test the interactions of Mr-AbaA with the *Mr-wetA* promoter regions NC: Negative control. *abaA-pwetA*: interaction between the Mr-AbaA protein and the *Mr-wetA* promoter region. AbA, aureobasidin A.

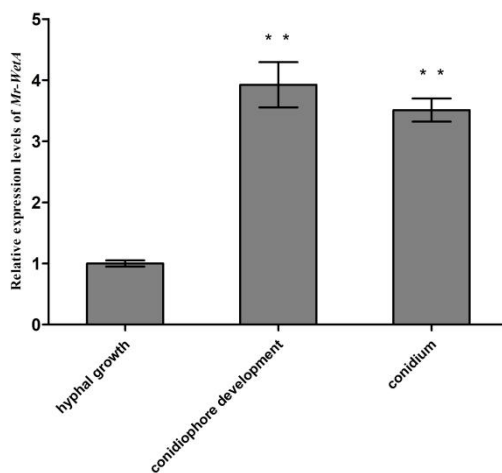


Figure S6. Transcriptional profiles of *Mr-wetA*.

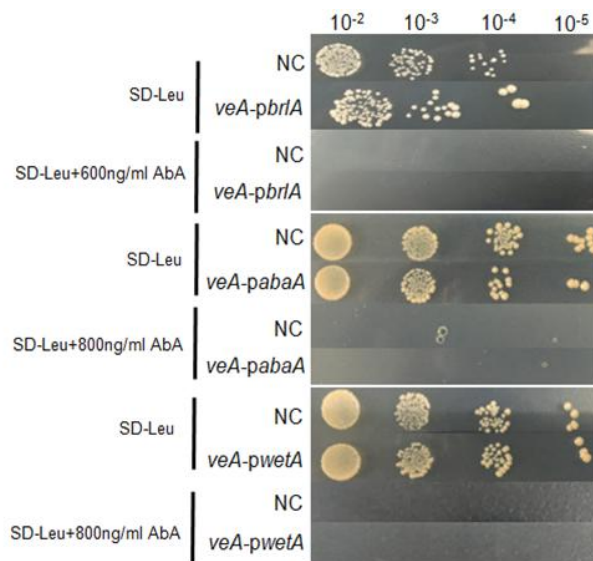


Figure S7. Analysis of the protein-DNA interaction. Yeast one-hybrid assay to test the interactions of Mr-VeA with the *Mr-brlA*, *Mr-abaA*, and *Mr-wetA* promoter regions. NC: Negative control. *veA-pbr1A*: interaction between the Mr-VeA protein and the *Mr-brlA* promoter region. *veA-pabaA*: interaction between the Mr-VeA protein and the *Mr-abaA* promoter region. *veA-pwetA*: interaction between the Mr-VeA protein and the *Mr-wetA* promoter region. AbA, aureobasidin A. The results showed that the Mr-VeA could not directly bind to the promoter regions of *Mr-brlA*, *Mr-abaA*, and *Mr-wetA*.

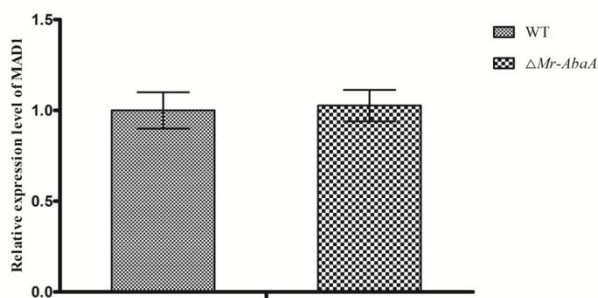


Figure S8. qRT-PCR analysis of the expression level of MAD1 between the WT and $\Delta Mr-abaA$.

Table S1 Primers used for functional analyses of two genes in *M. robertsii*.

Primer name	Primer sequence(5' - 3') ^a	Purpose of use
Component of <i>Mr-abaA</i>		
<i>abaA</i> -SF	CGGGATCCTCTCAAGGCAATGGATGGGC	
<i>abaA</i> -SR	CGGGATCCGACGCAACGAACGAGAACCA	Amplifying upstream flanking sequence
<i>abaA</i> -XF	GCTCTAGATTATCTCCACGCCCTCCT	
<i>abaA</i> -XR	GCTCTAGATTTGAAATATCCGGTCCTTT	Amplifying downstream flanking sequence
<i>abaA</i> -MF	AGCACCATCTATGCAAACCA	
<i>abaA</i> -MR	GTAGGCAACATCATCCCGAA	PCR for <i>abaA</i> to screen the transformants
<i>abaA</i> -YWR	TCTTGTGCGGTGCGGATAG	PCR for Δ <i>abaA</i> to confirm the mutant
<i>abaA</i> -GFP-b1F	ttctcagcccggggatccAAGAAATCGAGAAGATGCAGCG	Amplifying the promoter region and
<i>abaA</i> -GFP-b1R	tgctaccatCCATCCAGCAGTGACGGGC	<i>Mr-abaA</i> sequence to construct the EGFP
<i>abaA</i> -GFP-b2F	tgctgatggATGGTGAGCAAGGGCGAGG	fusion protein
<i>abaA</i> -GFP-b2R	tcattctctcagcgatccTTACTTGTACAGCTCGTCCATGCC	Amplifying EGFP sequence
<i>abaA</i> -cdfsF	gccatggagccagtgaaattcATGTCTTCACTCTTTCAGCCAAGA	Amplifying the <i>Mr-abaA</i> ORF for yeast
<i>abaA</i> -cdfsR	atgccaccggggtgaaattcTCACCATCCAGCAGTGACGG	one-hybrid test
<i>p-wetA</i> -F	aattcagagctcggtaccgggATGTTTGAGCGGTAATGGC	
<i>p-wetA</i> -R	cgaggtcgacagatccccgggGGCGTGTGATGTTGTGTGT	Amplifying <i>Mr-wetA</i> promoter region
<i>p-veA</i> -F	aattcagagctcggtaccgggTGAGTTGATCGCCACCCTGC	
<i>p-veA</i> -R	cgaggtcgacagatccccgggTTTGTCTAGCAGTCTCAAGGTTG	Amplifying <i>Mr-veA</i> promoter region
<i>p-velB</i> -F	aattcagagctcggtaccgggTTGTGATTGACTCTTTGCGA	
<i>p-velB</i> -R	cgaggtcgacagatccccgggGTTCAAATGGTGGTTTCTGG	Amplifying <i>Mr-velB</i> promoter region
<i>abaA</i> -F	AGCACCATCTATGCAAACCA	
<i>abaA</i> -R	GTAGGCAACATCATCCCGAA	RT-PCR for <i>abaA-c</i> to confirm the mutant
Component of <i>Mr-VeA</i>		
<i>veA</i> -SF	CGGAATCCCCCTACTTGCATACGA	
<i>veA</i> -SR	CGGGATCCGCGAATGAACGGGATGGTG	Amplifying upstream flanking sequence
<i>veA</i> -XF	GGACTAGTTCGCTCTGCTACTTTGCCTA	
<i>veA</i> -XR	GGACTAGTCGCTCCACTTTTCTCTCTC	Amplifying downstream flanking sequence
<i>veA</i> -MF	GACTACCAAGGAGGAAAAAGAC	
<i>veA</i> -MR	ATAATAGGAGACACAGGCACAG	PCR for <i>Mr-veA</i> to screen the transformants
<i>veA</i> -YWR	GCGGGAAAGGAGAAAAATAGTT	PCR for Δ <i>Mr-veA</i> to confirm the mutant
<i>veA</i> -F	TGAAAGCCAATAGCGACAGAC	
<i>veA</i> -R	CGGGAAAGATGAAGTAACCAG	RT-PCR for <i>Mr-veA-c</i> to confirm the mutant
<i>veA</i> -cdfsF	gccatggagccagtgaaattcATGCCGCCCTCATCTGTACA	Amplifying the <i>Mr-veA</i> ORF for yeast
<i>veA</i> -cdfsR	atgccaccggggtgaaattcCTATTGGTACCGTTGAAGTGAATC	one-hybrid test
<i>p-brlA</i> -F	aattcagagctcggtaccgggGCCTCAACTTCACCATACATGGG	
<i>p-brlA</i> -R	cgaggtcgacagatccccgggGTGTTGTTATTGTTGCTGTTGCTG	Amplifying <i>Mr-brlA</i> promoter region
<i>p-abaA</i> -F	aattcagagctcggtaccgggCTGGGCGCTGTGTAGATAGATCT	
<i>p-abaA</i> -R	cgaggtcgacagatccccgggAACTGCAAAGCAAAAAAAGA	Amplifying <i>Mr-abaA</i> promoter region
<i>veA</i> -CPF	aagcttcgcaactgttctagaTTGATCGCCACCCTGCAT	
<i>veA</i> -CPR	cgcggtggcgccgctctagaGACGCCGACTCCTTCCATG	PCR for CP to screen the transformants
<i>ben</i> -F	CCTCCACATAGACCCGTGTTCTGT	

<i>ben</i> -R	CAGAGGAGCCTGAATGTTGAGTG	PCR for CP to screen the transformants
<i>bar</i> -F	ATTTTGGTTTAGTCGTCCAGGCG	
<i>bar</i> -R	AGCTGCCAGAAACCCACGTCATG	PCR for mutants to screen the transformants

a: The lowercase letter in primers are required for cloning the PCR products into plasmids by homologous recombination and the red font showed the restriction enzyme.

Table S2 Primers used for qRT-PCR analysis

Gene	Tag locus	Annotation	primer (5'-3')
<i>gapdh</i>	MAA_07675	glyceraldehyde 3-phosphate dehydrogenase	GACTGCCCGCATTGAGAAG/AGATGGAGGAGTTGGTGTG
<i>fluG</i>	MAA_00122	protein fluG	TGCGGGTTGAATACGG/CTCCACCTCTTCTCTCTGA
<i>flbA</i>	MAA_06313	developmental regulator flbA	ACTCAAAGGGCATCACG/CAACAAAGCGGCGGAATA
<i>flbC</i>	MAA_04342	developmental regulator flbC	TTTCCAATCTACGACGACA/AGTCCGATTGATGGTCTTC
<i>flbD</i>	MAA_03655	developmental regulator flbD	AACGATGGGCTGAGATTG/GGTGATTGAGTTTCGGATG
<i>brlA</i>	MAA_10599	central regulator BrlA	ATCCTTTTGCTACTCACCA/ACCATTCCATCCAACCTCT
<i>abaA</i>	MAA_00694	central regulator AbaA	AAACCACTATTCTGTCTCC/AGCCTGCCTGTTACGATA
<i>wetA</i>	MAA_02845	central regulator WetA	CGACGAAATAGGAAAGCA/TGAAGTGGAGGAGATACGG
<i>stuA</i>	MAA_02988	APSES transcription factor StuA	GCAAGGCACCAACCCACT/TGCTGCTCCGTAGGCTGA
<i>veA</i>	MAA_01811	velvet family gene VeA	TGAAAGCCAATAGCGACAGAC/CGGAAAAGATGAAGTAACCAG
<i>velB</i>	MAA_00244	velvet family gene velB	CAGGAAAACCTCTTGGGA/TCGCCGAAACCGCACATT
<i>vosA</i>	MAA_01976	velvet family gene vosA	TTCACCATCACTCGGCACC/CTTTTCTTTCCAACGGC
<i>velC</i>	MAA_05862	velvet family gene velC	AGACGGGTTCAAATAGCC/TGTTTACTGAGGACGGAT
<i>hk1</i>	MAA_04209	Hexokinase	ATTCAAAGAACGTGGTGCC/ATTGCCAGCCTTGACTCTCT
<i>hk2</i>	MAA_05057	Hexokinase	GTCACCTCAAGCCGTACAG/CAACTCGGAATCTGTACG
<i>pfk</i>	MAA_09075	Phosphofructokinase	TGCGCAACATTATCCGCGAG/CTTCTGTACGCGGTTATGGG
<i>pk</i>	MAA_06851	Pyruvate kinase	TGTCTGGTGAACGGCAAAG/CAATGATACCACCAGCAGCC
<i>pc</i>	MAA_04366	Pyruvate carboxylase	TCGGGAGATATGCTGAACCC/ACACCAGTACCAGCAGAGTC
<i>icd</i>	MAA_03118	Isocitrate dehydrogenase	TAAGAATCCGGTCGTGGAAC/GTGACCTGGTCGTTGGTCTT
<i>pp</i>	MAA_02781	Pyruvate permease	TCATCCCCATCCACCTCATG/TAGACGCAGCCCATGAAGAT
<i>pdE1</i>	MAA_08787	Pyruvate dehydrogenase E1 component beta subunit	GTCAACTCTGCCGAAAGAC/CAGCCTTGAGAAGACCCTTG
<i>D-lcd</i>	MAA_07000	D-lactate dehydrogenase	TGCCTACGACCCATTTCCAT/TATTCACCAGCATCACCCCA
<i>icl</i>	MAA_04402	Isocitrate lyase	CCGTATGAAGCTACCAAG/AGGTCGTCGCCATAATGTC
<i>hsp70-L</i>	MAA_03832	Hsp70-like protein	AGGTCCTGCAGCTCAITTTCT/CCGCAAACCTCGTAGTCATG
<i>hsp20a</i>	MAA_10381	Heat shock protein Hsp20	CGCCCAAATAAAGGAGCCAG/CGCCCAAATAAAGGAGCCAG
<i>hsp20b</i>	MAA_07190	Heat shock protein Hsp20	ACATTGAATTCACCGAGCCG/CAGTACTGTACTTGGGCGC
<i>hsp30</i>	MAA_04014	Heat shock protein 30	CTGAGCCCGAGGAGAAG/GACACGGTTGGGAAAGT
<i>hsp40a</i>	MAA_02497	Heat shock protein 40	AGCGAGGCGAGGTGAA/GGTGGTGGCAGCATTT
<i>hsp40b</i>	MAA_03231	Heat shock protein 40	AGAGGAAGCCAACGACG/TCCGCAAGAGTAACAACGAG
<i>hsp40c</i>	MAA_06393	Heat shock protein 40	CGCCAACAGGAAAGACT/GCCGTGGTAAGAGGAAT
<i>hsp60</i>	MAA_07685	Heat shock protein 60	CGGCCAACTTTGACCAG/CAATGACAACGGAACCTT
<i>hsp70</i>	MAA_00810	Heat shock protein 70	TTCAAAGAGGGATACCACC/TTCAAAGAGGGATACCACC
<i>hsp90</i>	MAA_04726	Heat shock protein 90	TATGTCCCGCTGTCTT/TGTTCTGCTGGAGGGTC
<i>hsp104</i>	MAA_03534	Heat shock protein 104	TCTGCGGTCCCTCTGGT/CGGCTAAGTGCCTGTCTG
<i>catA</i>	MAA_05879	Catalase	ATCAACAGACCCGTTGTTC/GTGAAGAAGCCTTTGCTTGG
<i>catB</i>	MAA_06740	chanoclavine synthase catalase protein	TCCACTGGTTCCTGTATCC/GCATCTTCGGGGTCAATGAC
<i>p-cat</i>	MAA_10327	Peroxisomal catalase	CGATATTGGGGCACTGAAGC/ACGGGTTCCAGATCTCCTTG
<i>catC</i>	MAA_01261	Catalase	ATTGCCCATCTTCAACGG/CTGCCTGTCAAATCCGCT
<i>catD</i>	MAA_05785	Catalase	GAGATGGTGGCATGGTTGTC/CGTCACATCGCCAAAGAGAG