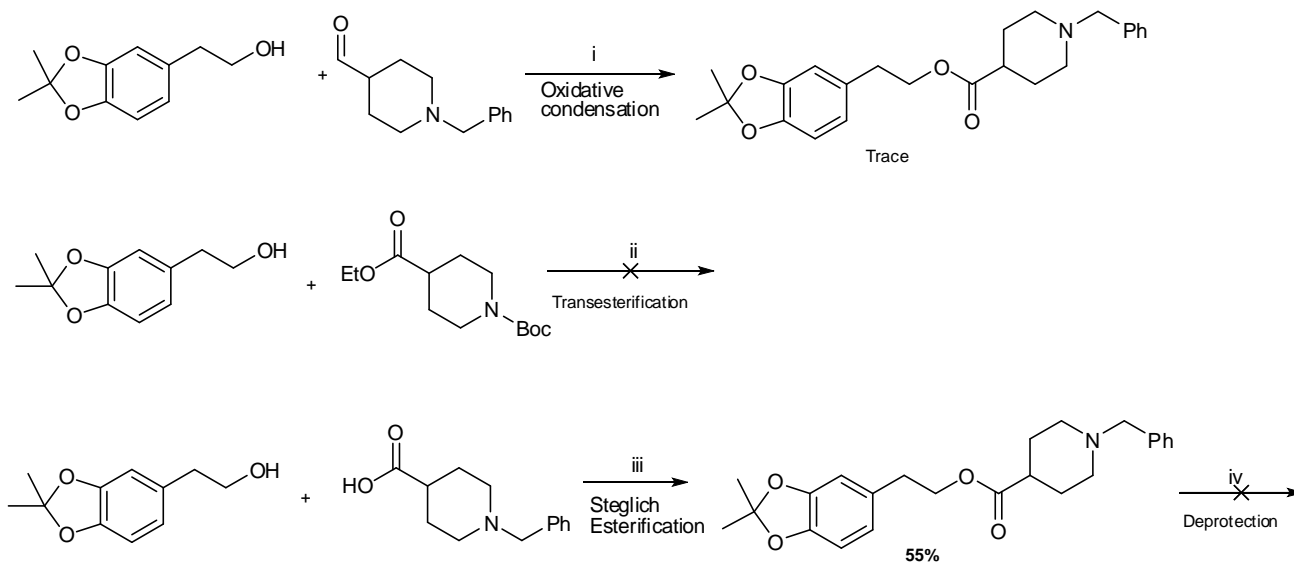


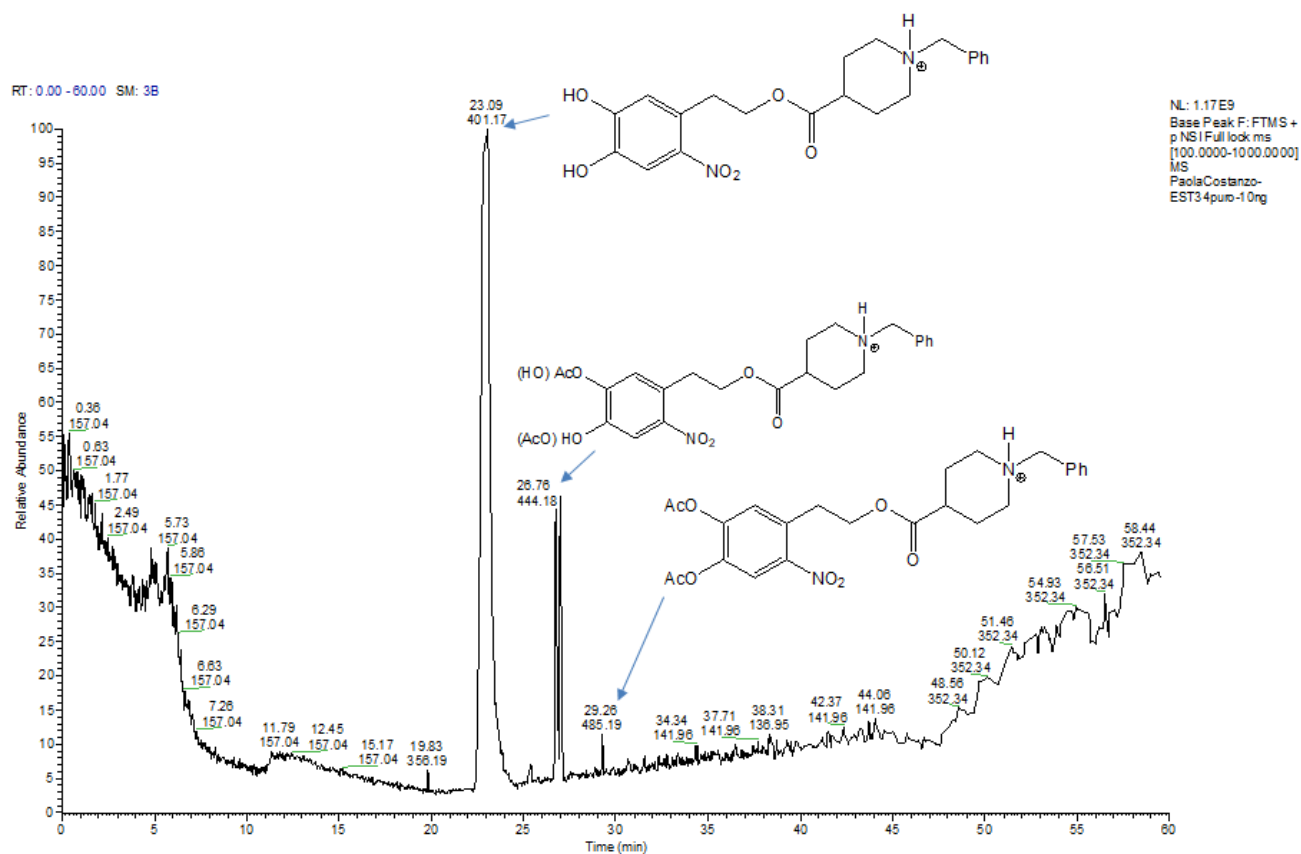
## Supplementary Material

### 1 Synthetic pathways tested for the synthesis of the hydroxytyrosol-donepezil hybrid



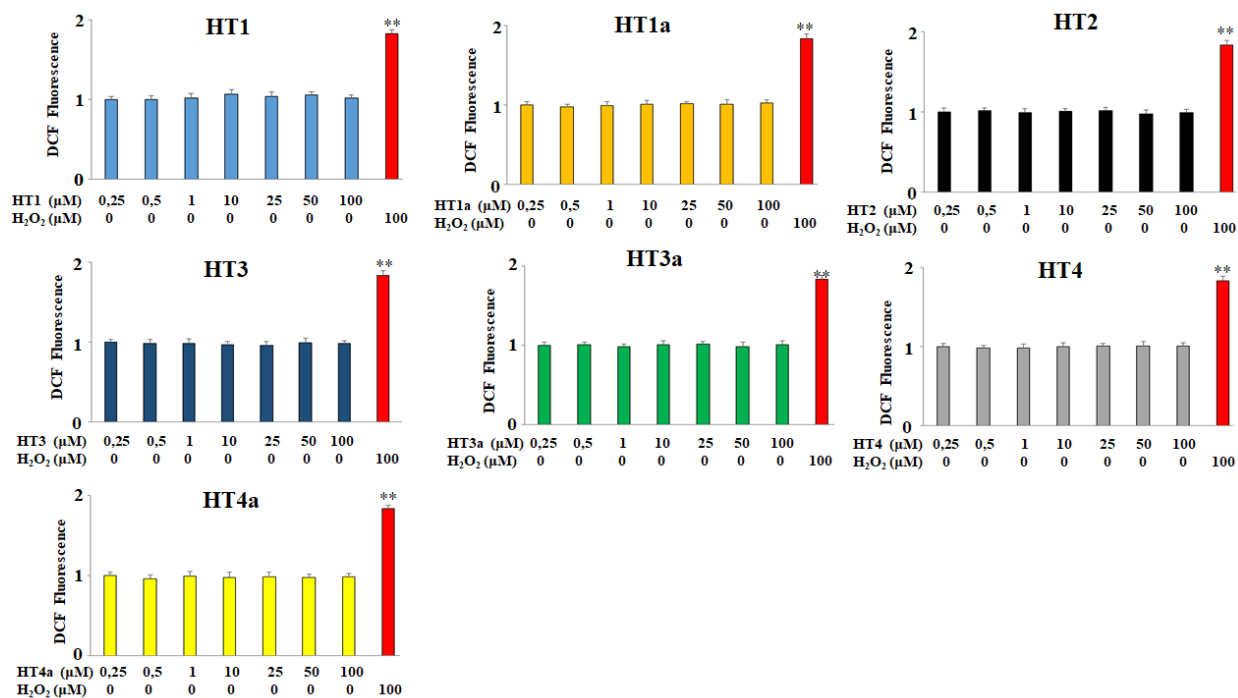
**Supplementary Figure 1.** Reaction conditions: (i)  $I_2$ ,  $K_2CO_3$ ,  $t$ -BuOH, rt,  $N_2$ , 4 days; (ii)  $Sc(OTf)_3$ , MW, 160 °C, 40 min; or  $MgO$ , 70 °C,  $N_2$ , o.n.; (iii) DIC, DMAP,  $CH_2Cl_2$  dry / DMF dry (1:1), 0 °C to rt, o.n.,  $N_2$ ; (iv) Amberlist, MeOH, reflux.

## 2 LC-HRMS identification of HT2 acetylation products

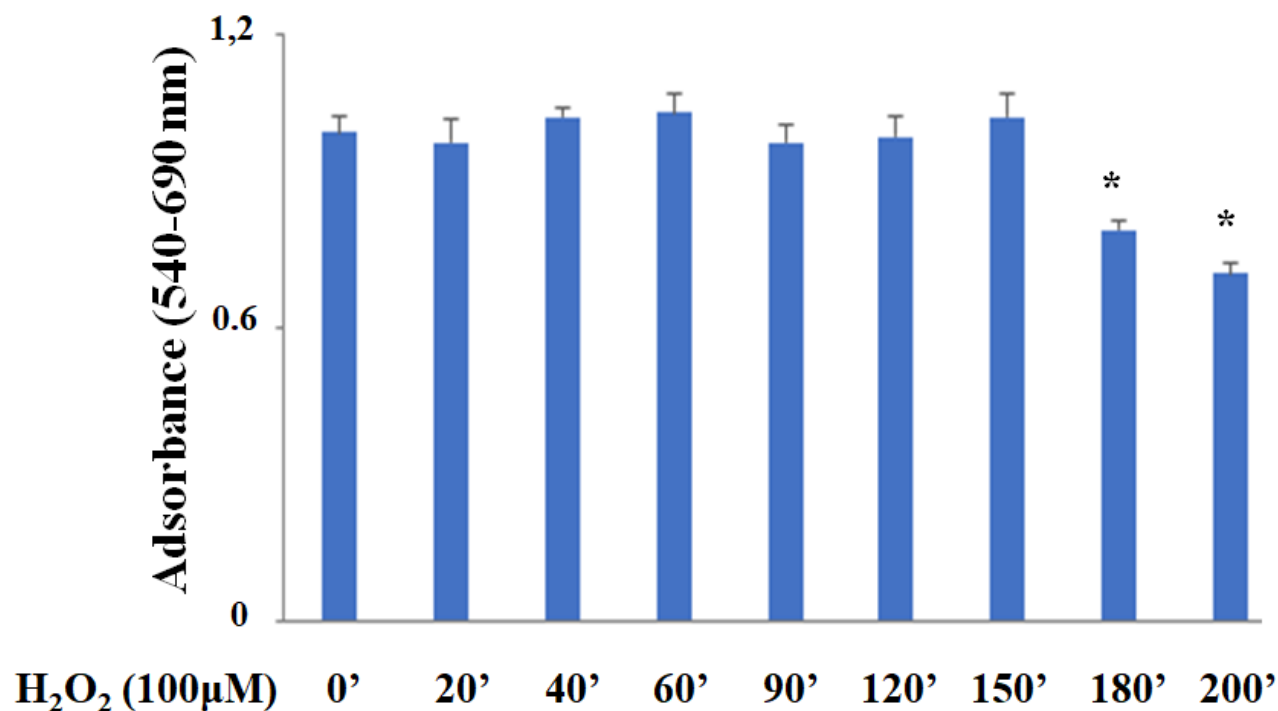


**Supplementary Figure 2.** Detection of the acetylated and di-acetylated form of **HT2** in the reaction crude. **[HT2-monoacetylated + H]<sup>+</sup>**: 443.1811 (theoretical **[M+H]<sup>+</sup>** m/z 443.1813); **[HT2-diacetylated + H]<sup>+</sup>**: 485.1917 (theoretical **[M+H]<sup>+</sup>** m/z 485.1918).

### 3 Evaluation of pro-oxidant effect for donepezil hybrids



**Supplementary Figure 3.** Donepezil hybrids have no pro-oxidant effects. Human neuroblastoma (SH-SY5Y) cell line was treated with donepezil hybrids for 24 h at the concentrations shown in the figure. No concentration showed accumulation of ROS. H<sub>2</sub>O<sub>2</sub> was used as positive control. Three independent experiments were carried out, with values expressed as the mean  $\pm$  standard deviation (sd). \*\* denotes  $p < 0.01$  vs. control. Analysis of Variance (ANOVA) was followed by a Tukey–Kramer comparison test.

4 SH-SY5Y Cell viability after treatment with H<sub>2</sub>O<sub>2</sub>

**Supplementary Figure 4.** Measurement of cell viability following treatment with H<sub>2</sub>O<sub>2</sub>. Cell viability following treatment with H<sub>2</sub>O<sub>2</sub> 100 μM was determined for several times (0-200'). The values of three independent experiments are expressed as the means ± standard deviations. \* denotes  $p < 0.05$  vs. control. Analysis of Variance (ANOVA) was followed by the Tukey–Kramer comparison test.

## 5 CUPRAC test

The CUPRAC test was performed as reported by Ozyürek et al. (Ozyürek et al., 2008) on the new compounds in the unacetylated form.

**Supplementary Table 1.** Concentration linearity range for all the unprotected hybrids.

Antioxidant	Linear equation and correlation coefficient	$\epsilon$ (L mol <sup>-1</sup> cm <sup>-1</sup> )	TEAC
Trolox	A= 0,0133, c - 0,0414, r = 0,9979	0,0133	1
HT	A= 0,0375, c - 0,0201, r = 0,9991	0,0375	2,82
HT1	A= 0,0447, c - 0,0336, r = 0,9943	0,0447	3,36
HT2	A= 0,0369, c+ 0,0178, r = 0,998	0,0369	2,77
HT3	A= 0,0239, c - 0,0192, r = 0,9945	0,0239	1,80
HT4	A= 0,0132, c - 0,0164, r = 0,9948	0,0132	0,99

## 6 Characterization of compounds

**3,4-dihydroxyphenethyl 1-benzylpiperidine-4-carboxylate (HT1):** White power, Yield 65%,  $R_f$  (DCM:MeOH:TEA – 84:13:2) 0.6. HRMS:  $[M+H]^+$  m/z 356.1852 (theoretical  $[M+H]^+$  m/z 356.1856).  $^1H$  NMR  $\delta$  (ppm) (500 MHz,  $d_6$ -DMSO): 8.64-8.71 (m br, 2H, OH), 7.21-7.33 (m, 5H), 6.63 (d,  $J=8$  Hz, 1H), 6.59 (d, 1H,  $J=2$  Hz, 1H), 6.45 (dd,  $J=8$  Hz and 2 Hz, 1H), 4.13 (t,  $J=7.0$  Hz, 2H), 3.43 (s, 2H), 2.66-2.72 (m, 4H), 2.22-2.36 (m, 1H), 1.96-2.01 (t,  $J=11.2$  Hz, 2H), 1.72-1.76 (m, 2H), 1.49-1.57 (m, 2H).  $^{13}C$  NMR (500 MHz,  $d_6$ -DMSO):  $\delta$  174.16, 144.96, 143.60, 128.46, 128.48, 128.02, 126.73, 119.36, 116.11, 115.34, 64.51, 62.13, 52.03, 45.63, 33.70, 27.81.

**4,5-dihydroxy-2-nitrophenethyl 1-benzylpiperidine-4-carboxylate (HT2):** Orange powder, Yield=85%,  $R_f$  (CHCl<sub>3</sub>:AcOEt:TEA-60:38:2) 0.56. HRMS:  $[M+H]^+$  m/z 440.2058 (theoretical  $[M+H]^+$  m/z 440.2068).  $^1H$  NMR  $\delta$  (ppm) (300MHz,  $d_6$ -DMSO): 7.47 (s, 1H), 7.21-7.33 (m, 6H), 4.23 (t,  $J=6.3$  Hz, 2H), 3.42 (s, 2H), 3.08 (t,  $J=6.3$  Hz, 2H), 2.69 (br d,  $J=10.7$  Hz, 2H), 2.18-2.25 (m, 1H), 1.96 (br t,  $J=9.3$  Hz, 2H), 1.70-1.73 (m, 2H), 1.46-1.53 (m, 2H);  $^{13}C$  NMR (500 MHz,  $d_6$ -DMSO). 174.16, 152.53, 144.35, 138.71, 138.34, 128.74, 128.12, 126.83, 126.75, 118.54, 111.85, 63.29, 62.22, 52.15, 32.15, 30.40, 29.22, 27.85.

**4-hydroxy-3-methoxyphenethyl 1-benzylpiperidine-4-carboxylate (HT3):** White power, Yield=45%,  $R_f$  (CH<sub>2</sub>Cl<sub>2</sub>:MeOH:TEA – 90:8:2) 0.70. HRMS:  $[M+H]^+$  m/z 370.2007 (theoretical  $[M+H]^+$  m/z 370.2013).  $^1H$  NMR  $\delta$  (ppm) (500 MHz, CDCl<sub>3</sub>): 7.20-7.32 (m, 5H), 6.83 (d,  $J=7.8$  Hz, 1H), 6.68-6.71 (m, 2H), 4.25 (t,  $J=6.9$  Hz, 2H), 3.86 (s, 3H), 3.48 (s, 2H), 2.80-2.87 (m, 2H), 2.24-2.31 (m, 1H), 2.00-2.03 (m, 2H), 1.83-1.86 (m, 2H), 1.70-1.79 (m, 2H).  $^{13}C$  NMR (500 MHz, CDCl<sub>3</sub>): 175.31, 146.68, 144.53, 129.8, 129.34, 128.5, 127.25, 121.84, 114.58, 111.60, 65.21, 63.41, 56.09, 41.36, 35.04, 32.12, 28.42.

**4-hydroxyphenethyl 1-benzylpiperidine-4-carboxylate (HT4):** White power, Yield=55%;  $R_f$  (DCM:MeOH:TEA – 90:8:2) 0.37. HRMS:  $[M+H]^+$  m/z 340.1898 (theoretical  $[M+H]^+$  m/z 340.1907).  $^1H$  NMR  $\delta$  (ppm) (500 MHz, CDCl<sub>3</sub>): 7.23-7.32 (m, 5H), 7.03 (d,  $J=8.4$  Hz, 2H), 6.72 (d,  $J=8.4$  Hz, 2H), 4.23 (t,  $J=6.9$  Hz, 2H), 3.50 (s, 2H), 2.83 (t,  $J=6.9$  Hz, 4H), 2.23-2.33 (m, 1H), 2.00-2.08 (m, 2H), 1.82-1.86 (m, 2H), 1.70-1.79 (m, 2H).  $^{13}C$  NMR (500 MHz, CDCl<sub>3</sub>):  $\delta$  175.31, 154.89, 137.84, 130.22, 129.68, 129.57, 128.46, 127.39, 115.68, 65.31, 63.40, 52.95, 41.23, 34.46, 32.12, 28.16.

**4-(2-(1-benzylpiperidine-4-carboxyloxy)ethyl)-1,2-phenylene diacetate (HT1a):** Orange oil, Yield=55%,  $R_f$  (CH<sub>2</sub>Cl<sub>2</sub>:MeOH:TEA-90:8:2) 0.52. HRMS:  $[M+H]^+$  m/z 401.1706 (theoretical  $[M+H]^+$  m/z 401.1707).  $^1H$  NMR  $\delta$  (ppm) (CDCl<sub>3</sub>, 300 MHz): 7.22-7.37 (m, 5H), 7.08-7.18 (m, 2H), 7.00-7.08 (m, 1H), 4.29 (t,  $J=9.9$  Hz, 2H), 3.50 (s, 2H), 2.92 (t,  $J=9.9$  Hz, 2H), 2.84 (d,  $J=11$  Hz, 2H), 2.22-2.38 (m, 7H), 1.92-2.10 (m, 2H), 1.90-1.65 (m, 4H);  $^{13}C$  NMR (CDCl<sub>3</sub>, 300 MHz).  $\delta$ : 179.1, 175.2, 168.4, 146.3, 142.5, 137.8, 135.3, 129.9, 128.7, 127.5, 124.3, 65.4, 63.3, 52.4, 41.9, 34.6, 28.1, 20.7.

**4-acetoxy-3-methoxyphenethyl 1-benzylpiperidine-4-carboxylate (HT3a):** Orange oil, Yield=98%,  $R_f$  (CH<sub>2</sub>Cl<sub>2</sub>:MeOH:TEA – 90:8:2) 0.77; HRMS:  $[M+H]^+$  m/z 412.2111 (theoretical  $[M+H]^+$  m/z 412.2118).  $^1H$  NMR  $\delta$  (ppm) (500 MHz, CDCl<sub>3</sub>): 7.15-7.24 (m, 5H), 6.87 (d,  $J=8.0$  Hz,

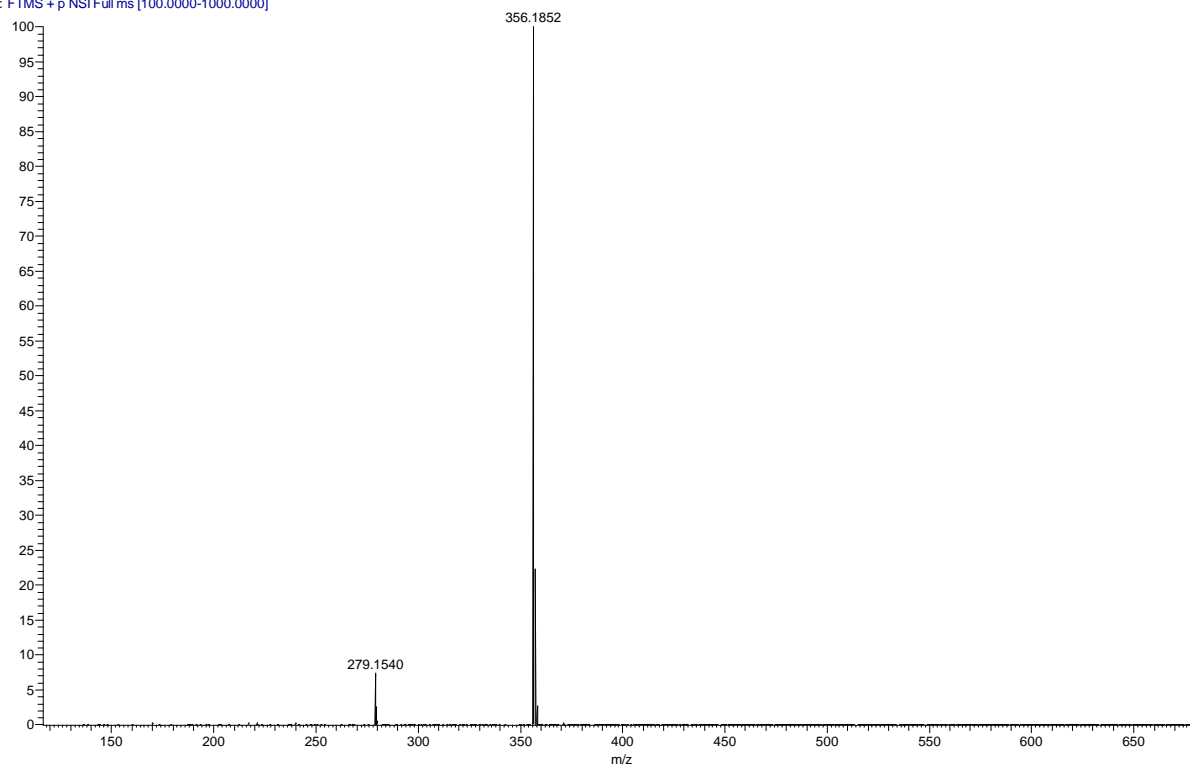
1H), 6.69-6.75 (td,  $J=8.0$  and  $J=1.8$  Hz, 2H), 4.22 (t,  $J=7.2$  Hz, 2H), 3.74 (s, 3H), 3.41 (s, 2H), 2.84 (t,  $J=7.0$  Hz, 2H), 2.75 (br d,  $J=11.0$  Hz, 2H), 2.24 (s, 3H), 2.16-2.22 (m, 1H), 1.94 (t br,  $J=11.0$  Hz, 2H), 1.77-1.79 (m, 2H), 1.63-1.73 (m, 2H).  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 500 MHz): 175.32, 169.34, 151.13, 138.6, 136.99, 129.28, 128.4, 127.19, 122.88, 121.27, 113.27, 64.76, 63.44, 56.07, 53.07, 41.42, 35.26, 28.49, 20.88.

**4-acetoxyphenethyl 1-benzylpiperidine-4-carboxylate (HT4a):** Incolor oil; Yield=85%;  $R_f$  ( $\text{CHCl}_3$ :AcOEt:TEA-60:38:2) 0.28. HRMS:  $[\text{M}+\text{H}]^+$   $m/z$  382.2005 (theoretical  $[\text{M}+\text{H}]^+$   $m/z$  382.2013);  $^1\text{H}$  NMR  $\delta$  (ppm) (500 MHz,  $\text{CDCl}_3$ ): 7.15-7.24 (m, 5H), 7.14 (d,  $J=8.4$  Hz, 2H), 6.94 (d,  $J=8.5$  Hz, 2H), 4.21 (t,  $J=6.8$  Hz, 2H), 3.41 (s, 2H), 2.85 (t,  $J=6.9$  Hz, 2H), 2.73-2.77 (br d,  $J=11.5$  Hz, 2H), 2.16-2.23 (m, 4H), 1.91-1.96 (br t,  $J=10.8$  Hz, 2H), 1.74-1.81 (m, 2H), 1.61-1.70 (m, 2H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  175.31, 169.77, 149.54, 138.58, 135.66, 130.08, 129.32, 128.40, 127.19, 121.75, 64.78, 63.45, 53.08, 41.37, 34.72, 28.1, 21.35.

# 7 HRMS

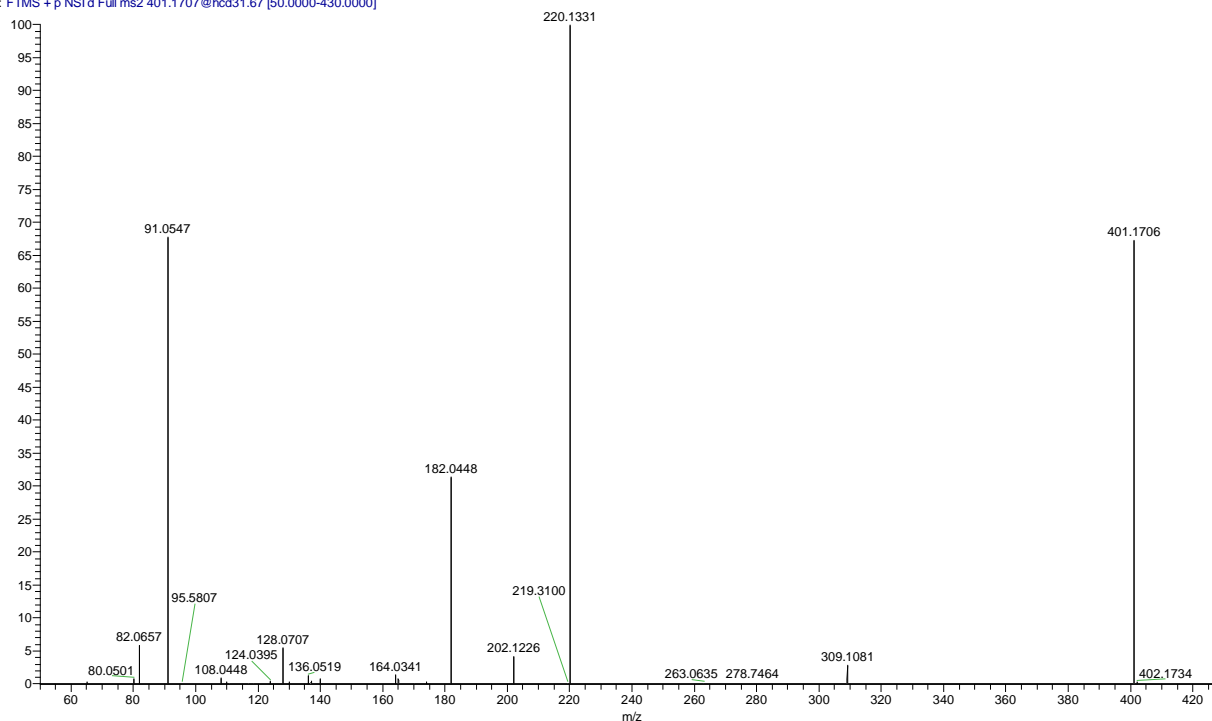
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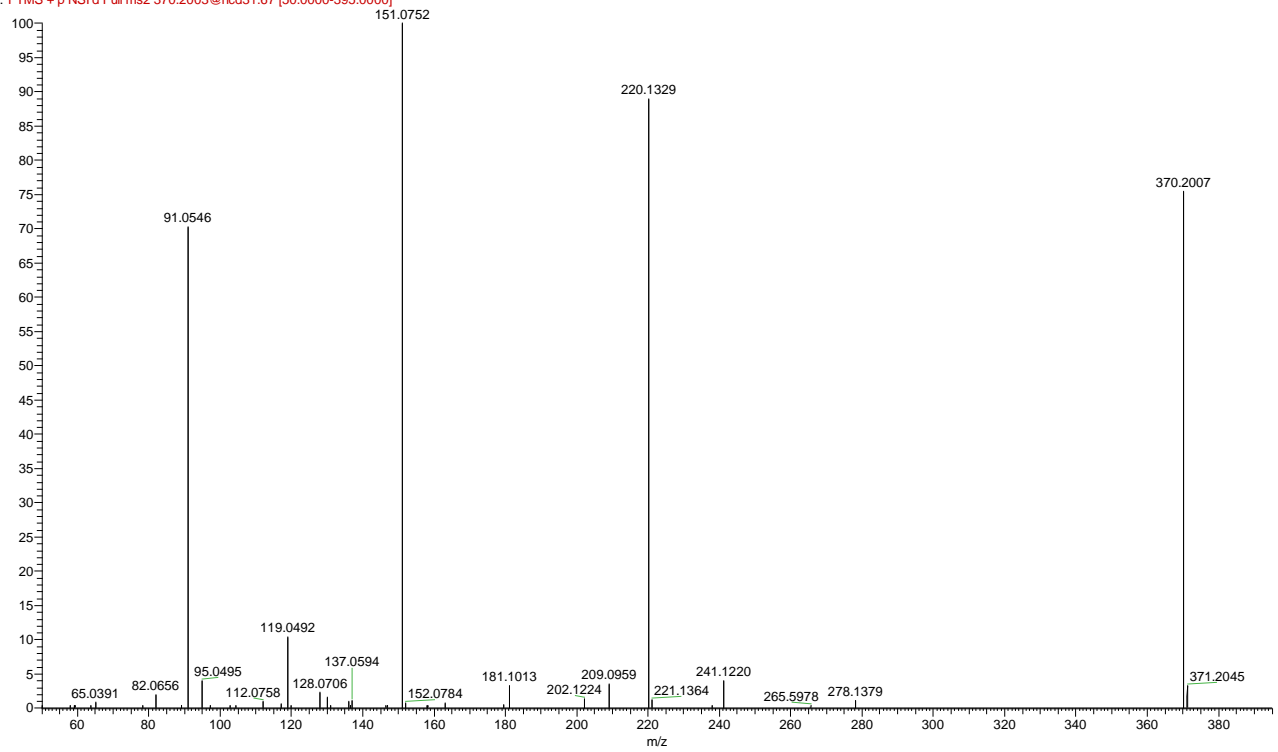
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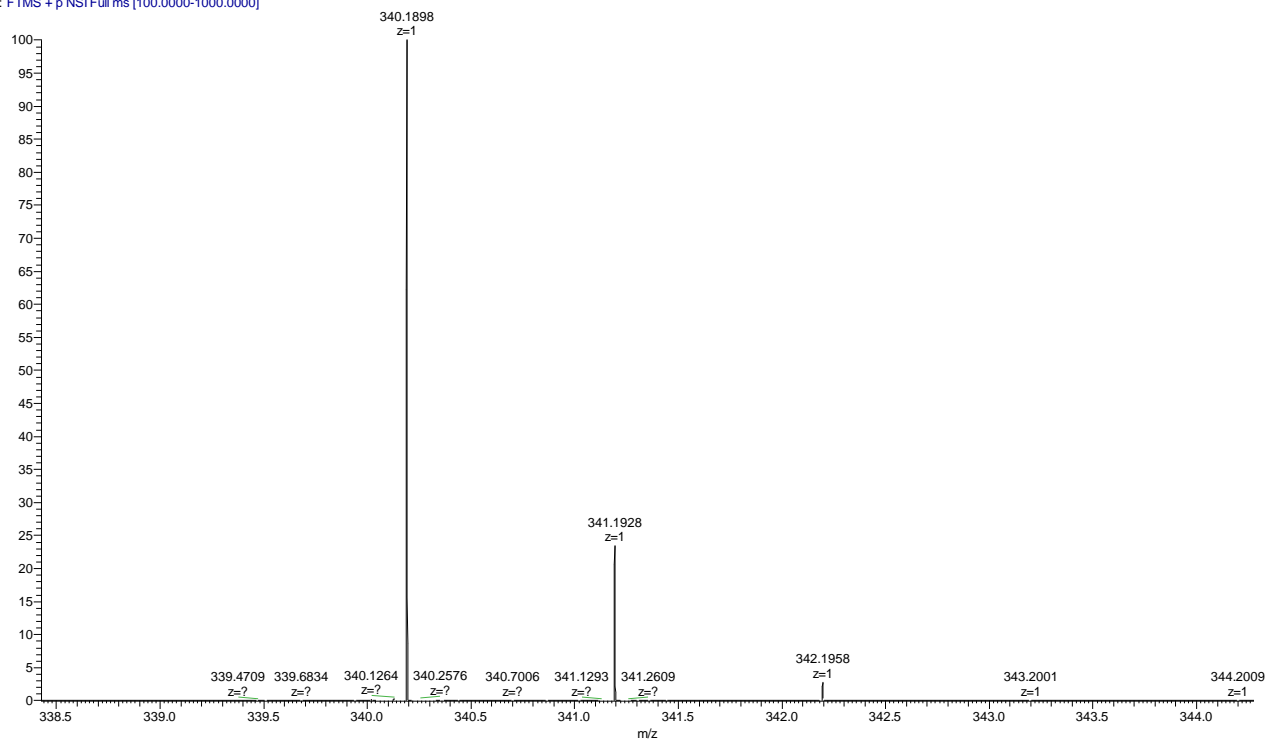
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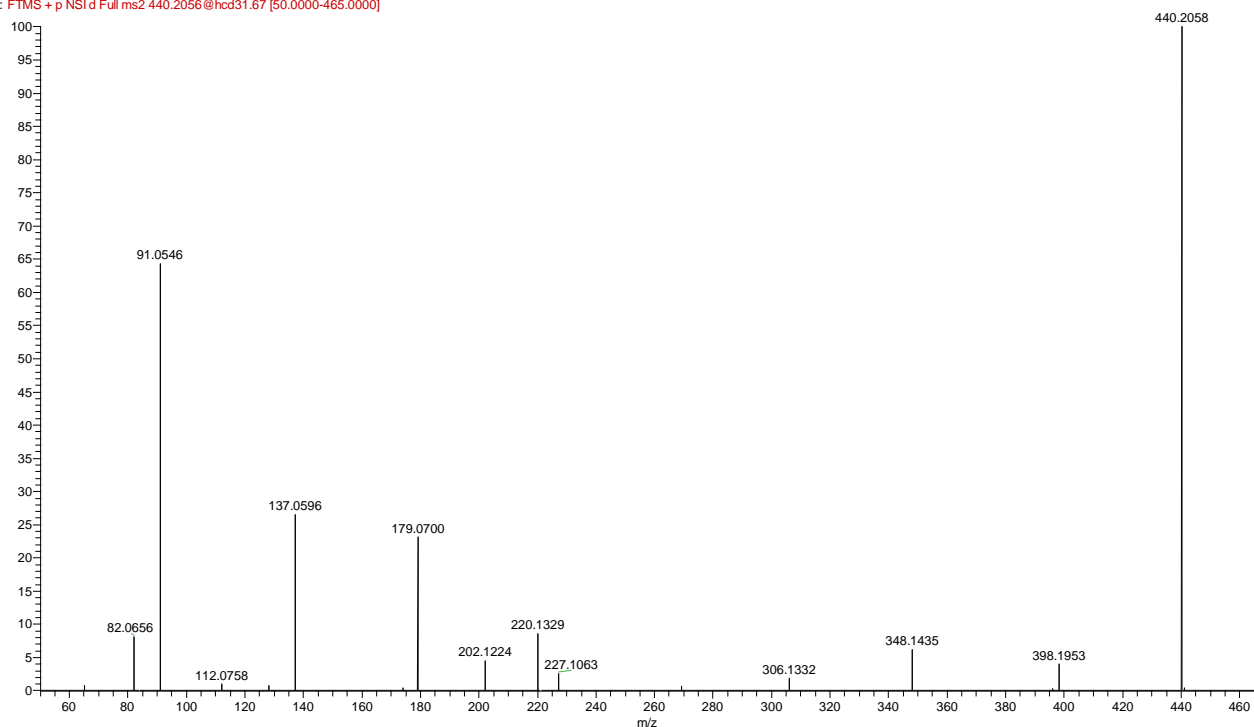
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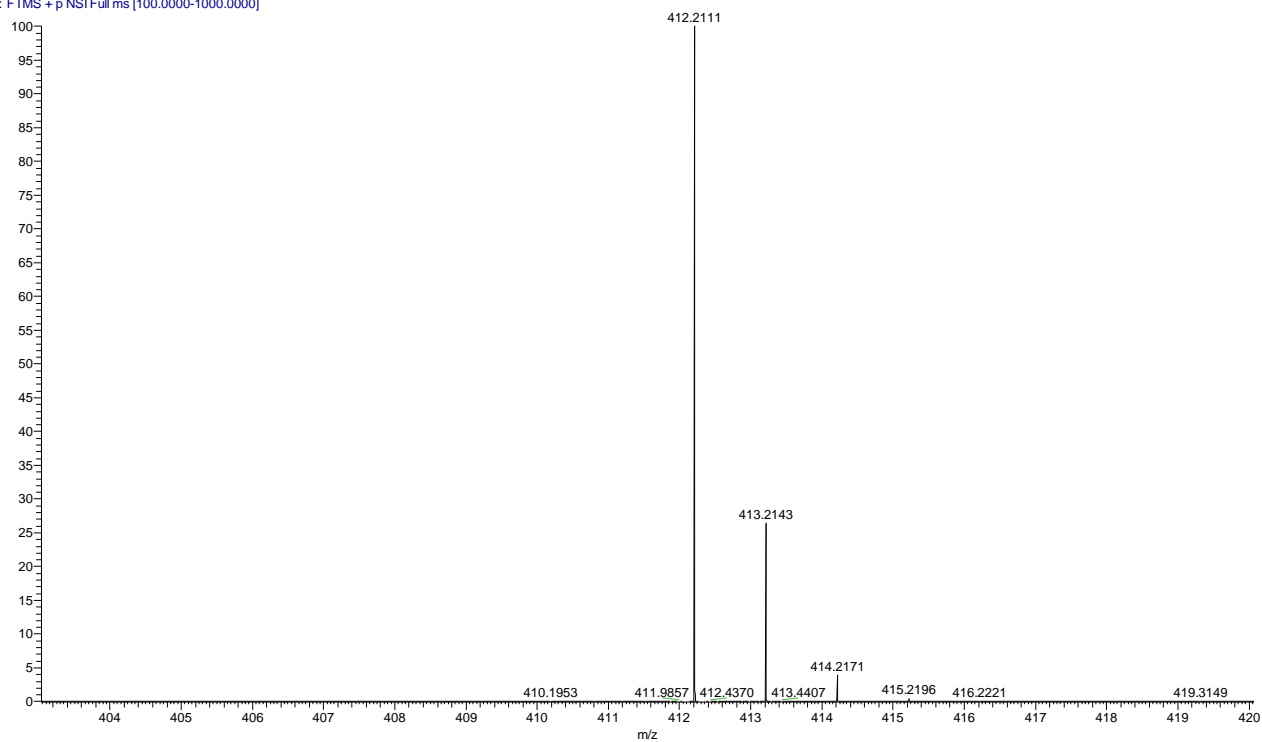
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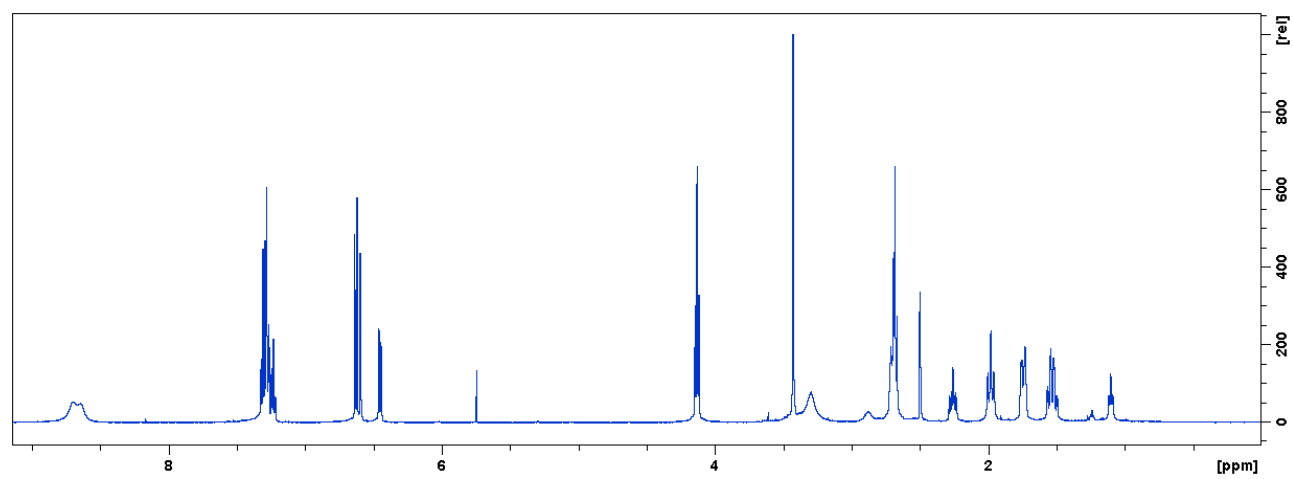
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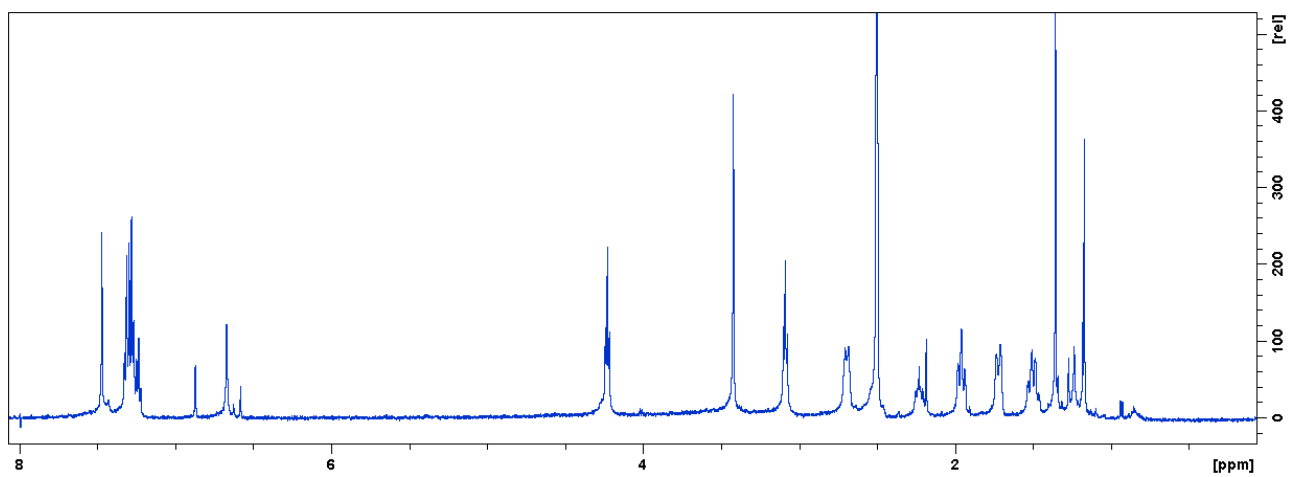


8  $^1\text{H-NMR}$ 

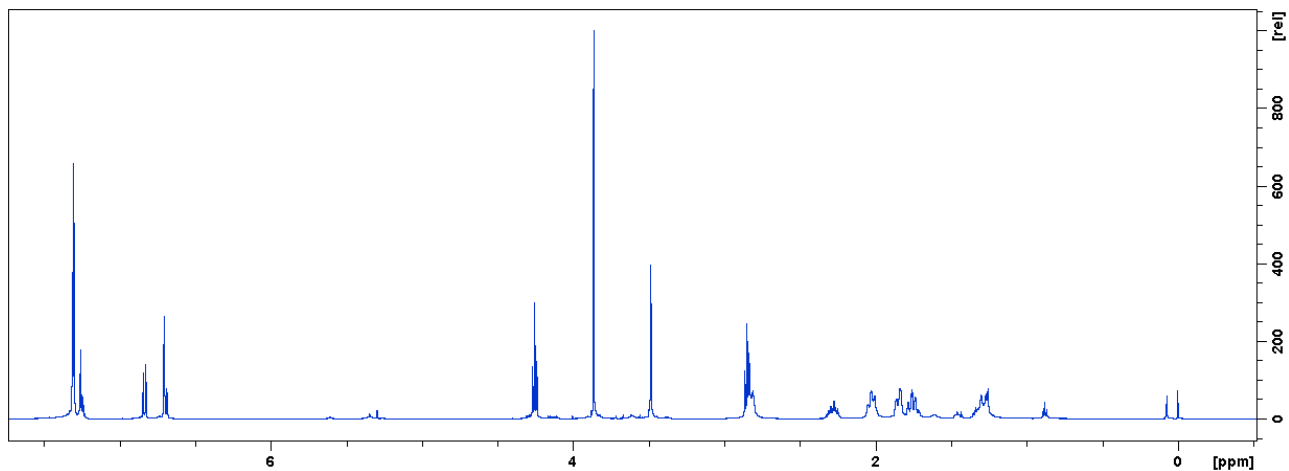
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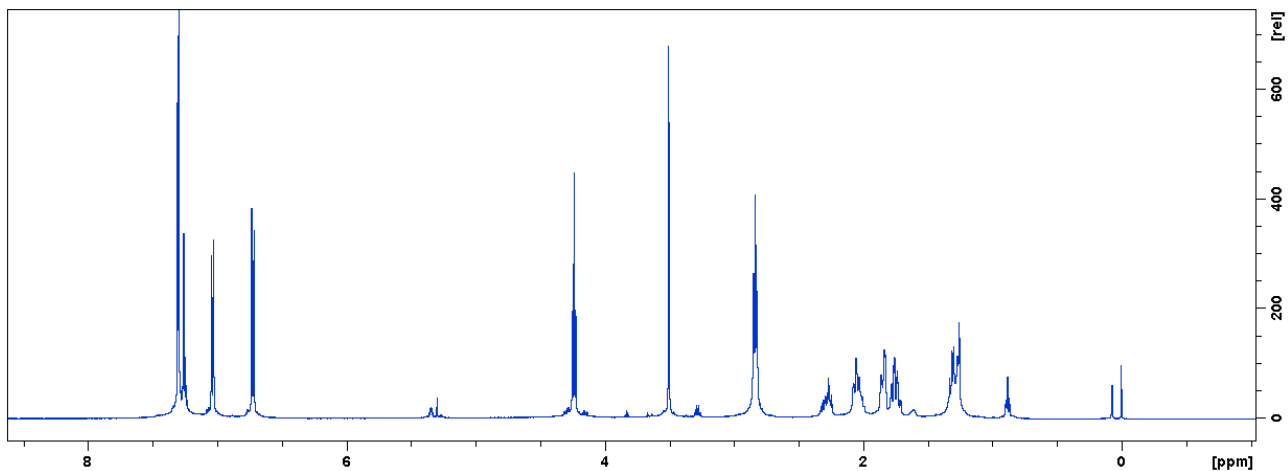
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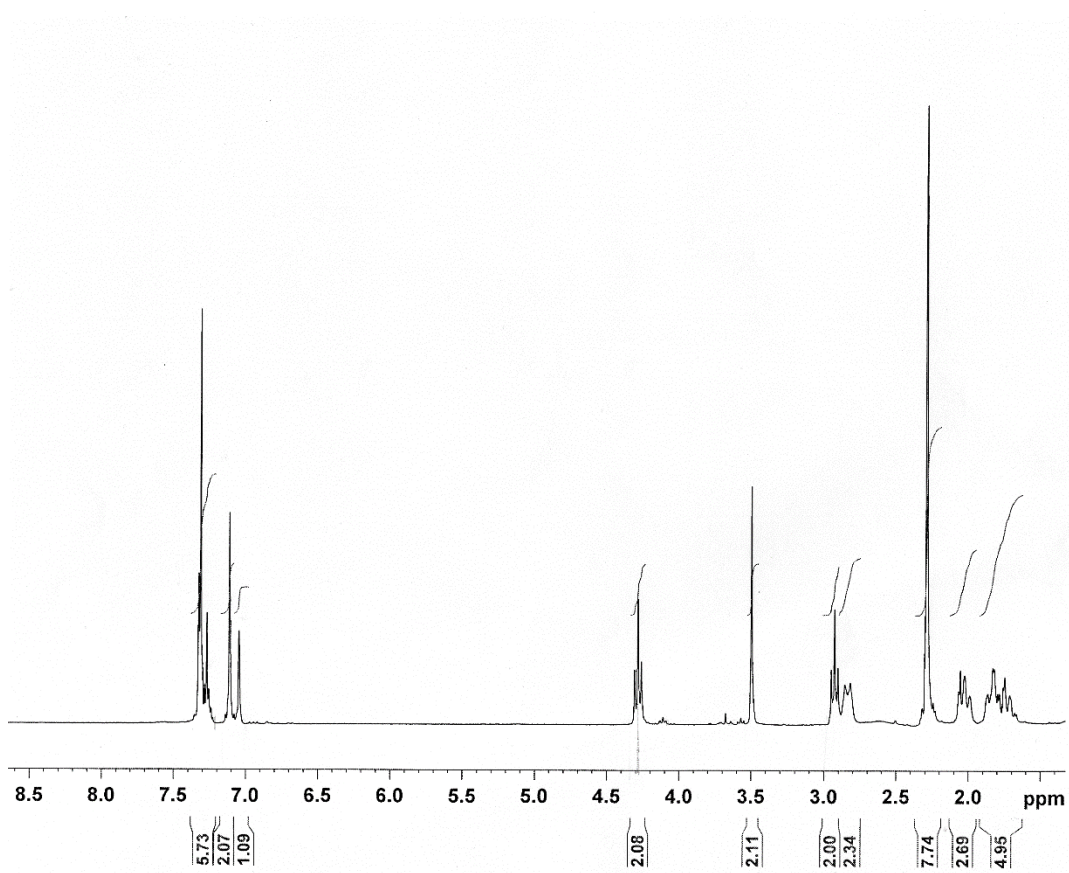
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# HT4



# HT1a



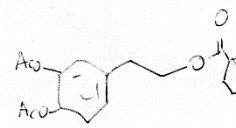
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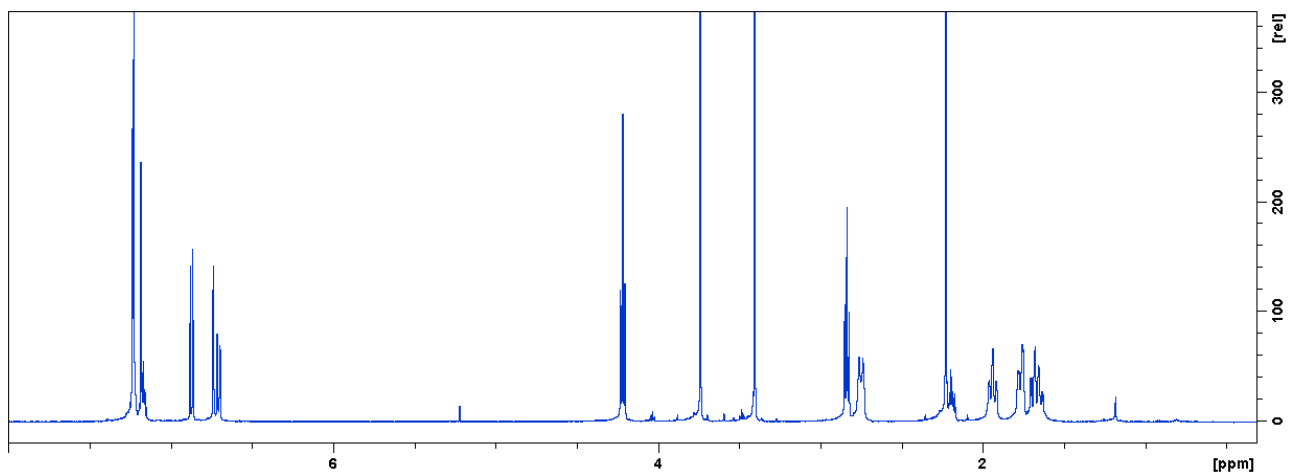
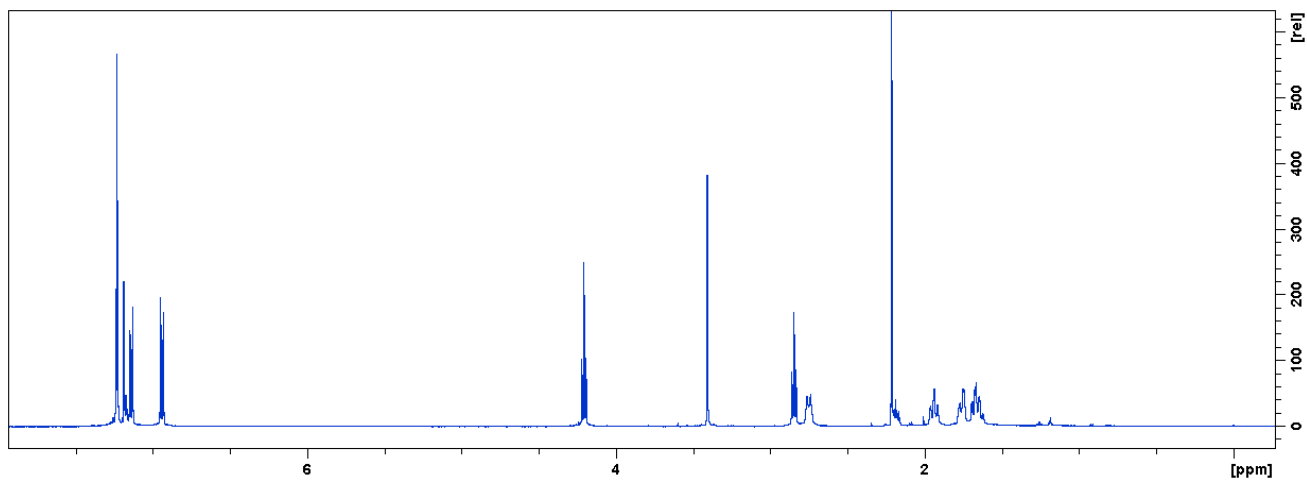
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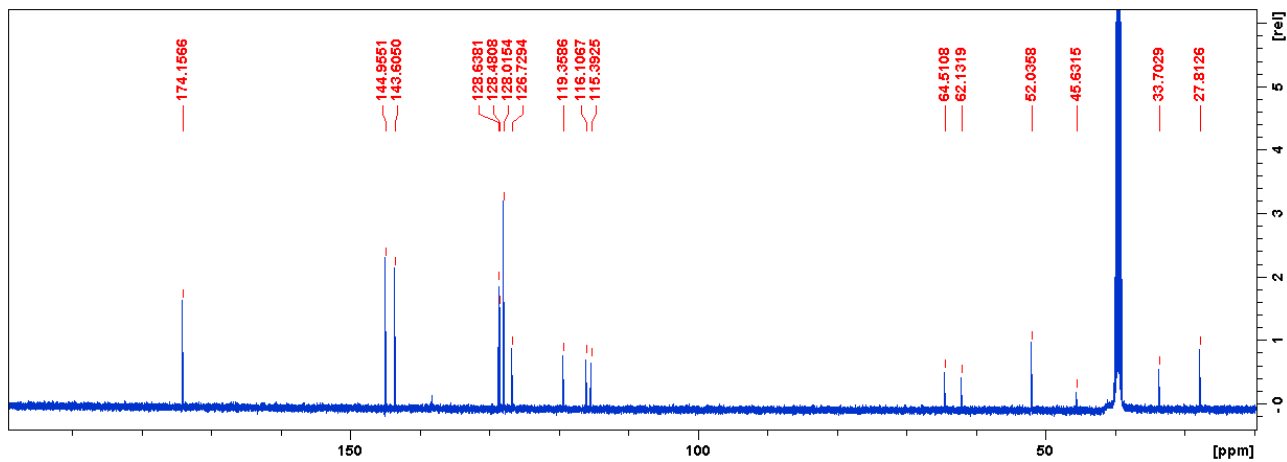
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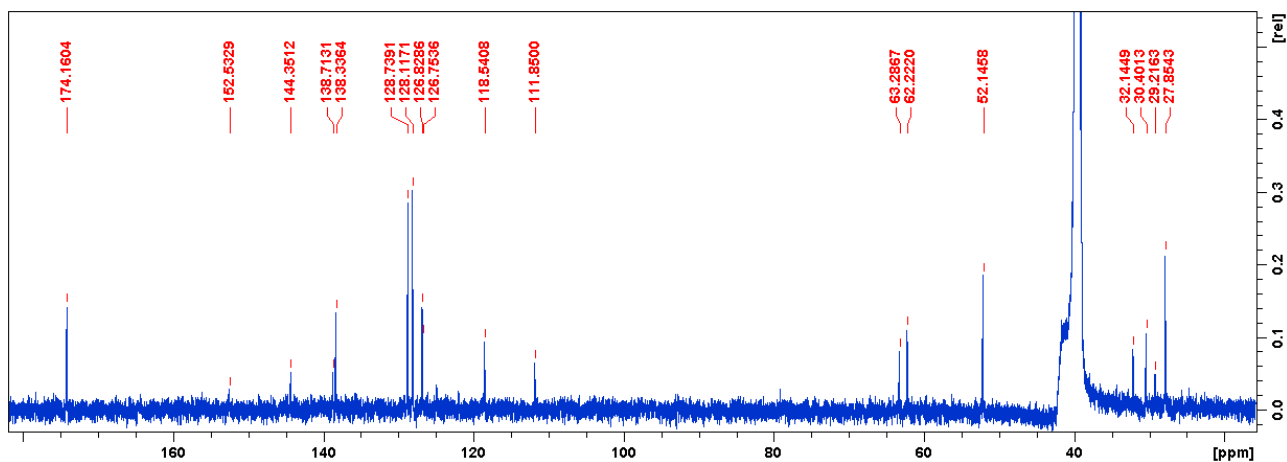
**HT3a****HT4a**

# 9 <sup>13</sup>C-NMR

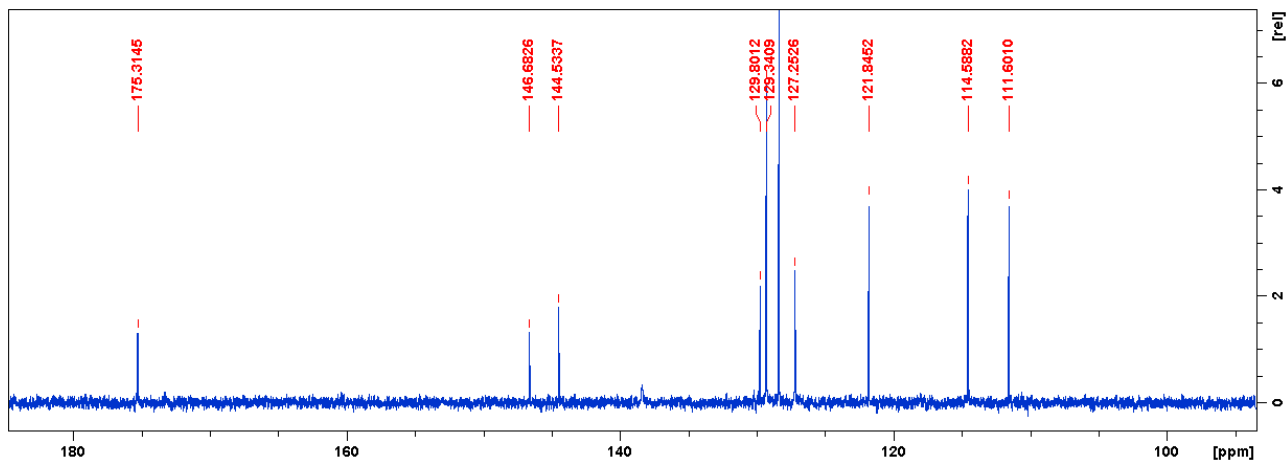
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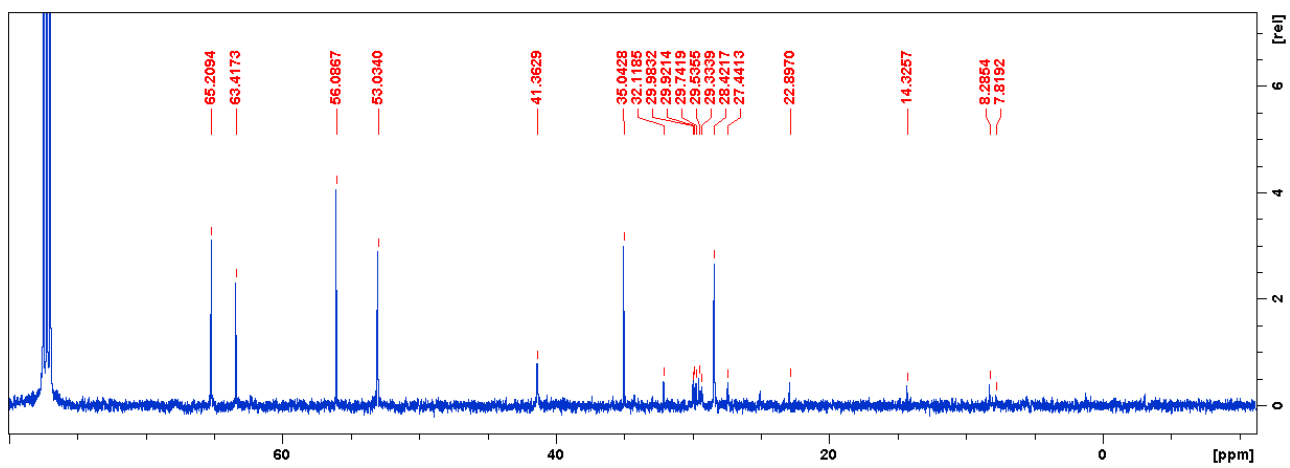


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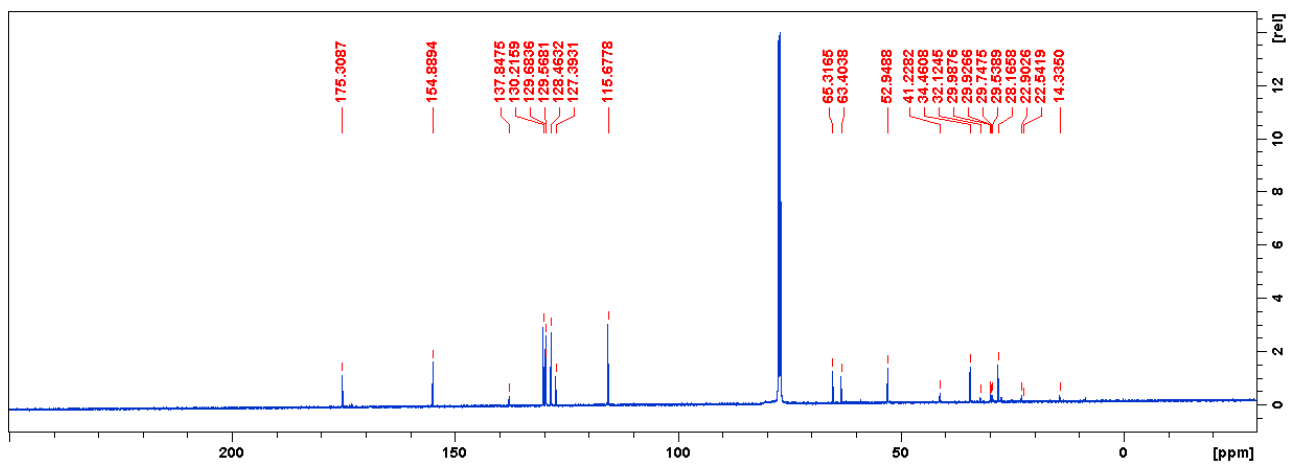


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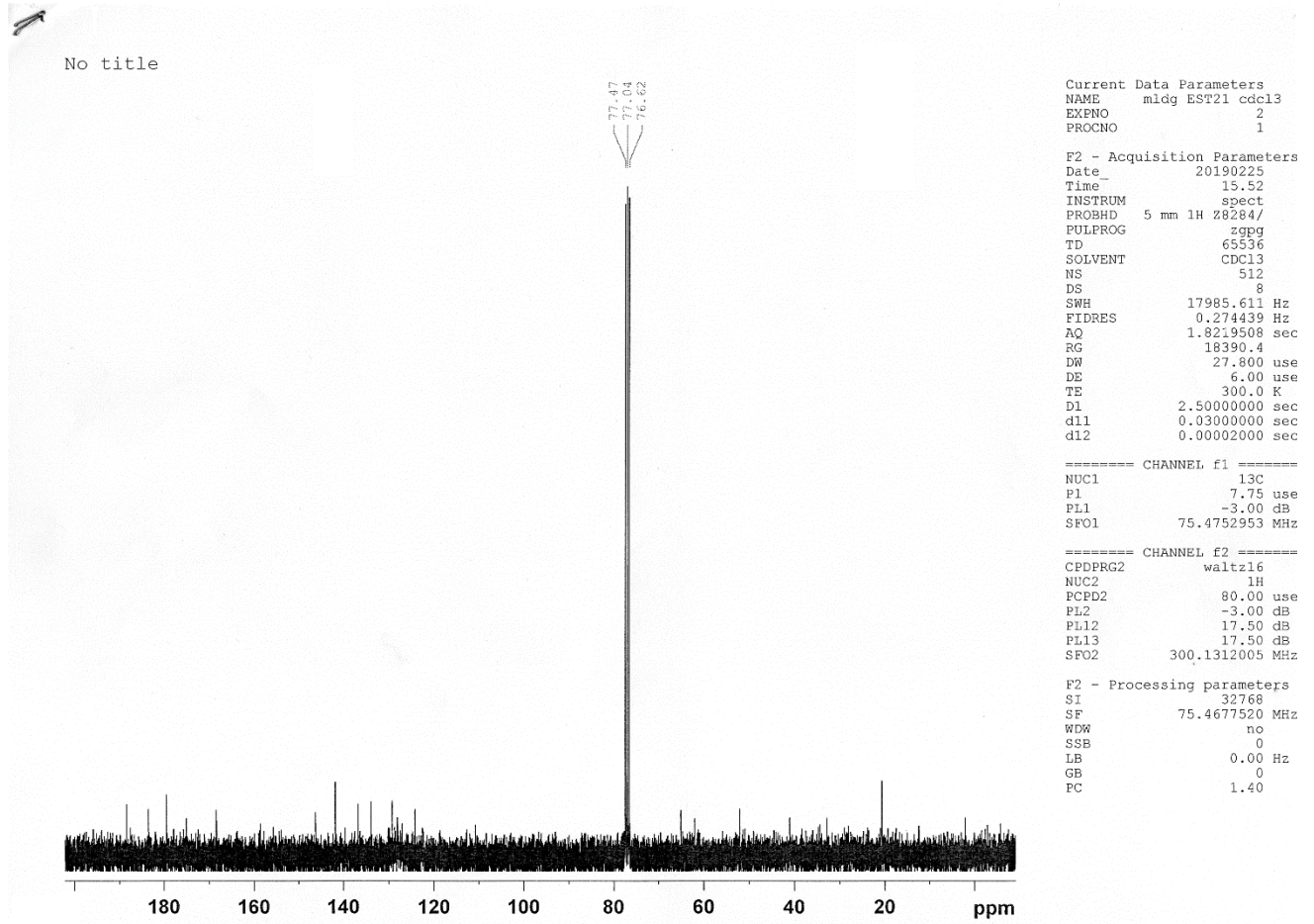




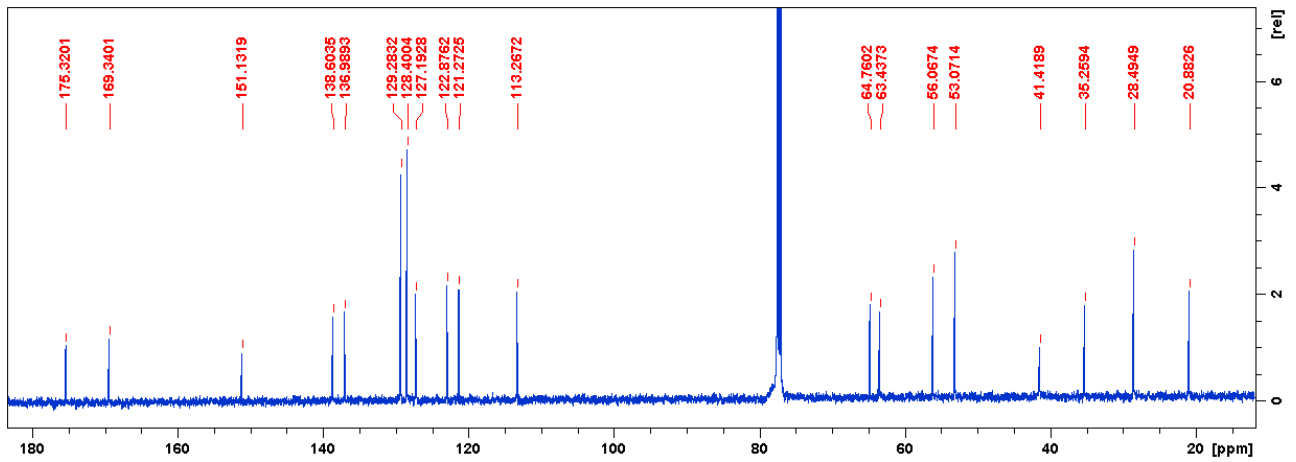
HT4



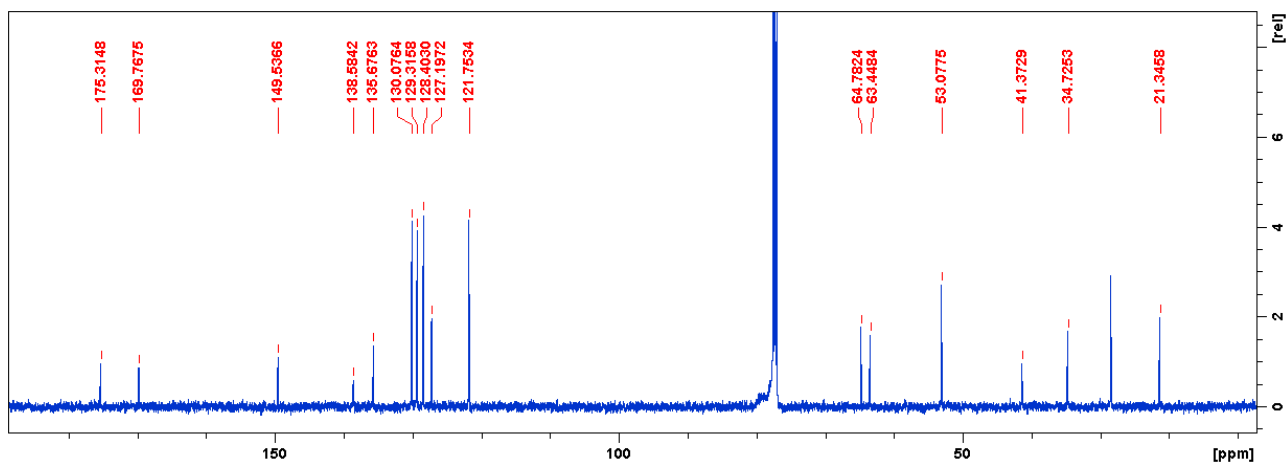
# HT1a



# HT3a



## HT4a



## 10 References

Ou, B., Hampsch-Woodill, M., and Prior, R. L. (2001). Development and validation of an improved oxygen radical absorbance capacity assay using fluorescein as the fluorescent probe, *J. Agric. Food Chem.* 49, 4619–4626. doi: 10.1021/jf010586o.

Ozyürek, M., Bektaşoğlu, B., Güçlü, K., Güngör, N., and Apak, R. (2008). Simultaneous total antioxidant capacity assay of lipophilic and hydrophilic antioxidants in the same acetone-water solution containing 2% methyl-beta-cyclodextrin using the cupric reducing antioxidant capacity (CUPRAC) method. *Anal. Chim. Acta.* 630(1), 28-39. doi: 10.1016/j.aca.2008.09.057.