



**Fig. S1: Experimental set-up of NanoLuc™ Luciferase assay and ligand binding data.**

(a) Schematic representation of NanoLuc™ Luciferase (nLuc) based MC4R ligand binding assay. nLuc protein was fused at the receptor N-terminus. Bioluminescence resonance energy transfer (BRET) is observed in dependence of the relative distance of the fluorescent (with the fluorophore 5-carboxytetramethylrhodamine (TAMRA)) labelled NDP- $\alpha$ -MSH (TAMRA-NDP) and nLuc-MC4R.

(b) Ligand binding assay workflow, HEK293T cells were infected 24 h prior, followed by media exchange against ligand titrations. Ligand binding equilibration is ensured by 2 h incubation time with subsequent addition of the nLuc substrate luciferin and measurement of the short-pass filter (460 nm) and long-pass filter (610 nm) using a fluorescent plate reader. The BRET ratio is the quotient of long-pass by short-pass.

(c) Titration of TAMRA-NDP from 10  $\mu$ M to 0.1 pM is plotted as dose-response measurement with an  $EC_{50}$  of 1.44 nM.

(d) Setmelanotide binding was determined by competing the agonist setmelanotide against 10 nM TAMRA-NDP with a resulting  $K_i$  of 1.41 nM. The addition of 20  $\mu$ M non-fluorescent labeled NDP- $\alpha$ -MSH enhanced the BRET effect induced by non-specific binding (binding control).

(a, b) was created with *Biorender.com*.