

Supporting Information

Rh(III)-Catalyzed Three-Component *syn*-Carboamination of Alkenes Using Arylboronic Acids and Dioxazolones

Sumin Lee and Tomislav Rovis*

Email: tr2504@columbia.edu

Department of Chemistry, Columbia University, New York, New York 10027

Contents

1. General information	2
2. Preparation of Starting Materials	2
3. General procedure for the carboamination of alkenes.....	5
4. Detailed optimization table	6
5. Product characterization.....	11
6. Mechanistic studies	32
7. Limitation	45
8. Reference	46
9. NMR Spectra.....	48

1. General information

All reactions were carried out using oven-dried glassware with magnetic stirring unless otherwise noted. Anhydrous solvents and reagents were purchased from commercial sources and used without further purification. Flash chromatography was conducted either manually on SiliCycle® SilicaFlash® P60 (230-400 mesh) silica gel, SiliCycle® prep TLC (TLG-R10011B-341, thickness 1000um) or automatically *via* a Teledyne Isco Lumen CombiFlash with RediSep Rf Disposable Flash columns. Thin layer chromatography (TLC) was performed on Silicycle 250µm silica gel 60 Å plates. Visualization was accomplished with UV light (254 nm) and KMnO₄. ¹H, ¹⁹F NMR, and ¹³C NMR spectra were collected at ambient temperature on Bruker 400 MHz or Bruker Avance III 500 MHz spectrometers unless otherwise noted. Chemical shifts (δ) are reported in parts per million (ppm), coupling constants (*J*) are reported in Hz, and multiplicity is described using the following abbreviations: singlet (s), broad (b), multiplet (m), doublet (d), triplet (t), quartet (q), or combinations thereof. ¹H NMR spectra were referenced to 7.26 ppm (CHCl₃) or 0.0 ppm (TMS). ¹³C NMR spectra were referenced to 77.16 ppm (CDCl₃) and all peaks given are singlet unless otherwise noted. Diastereomeric and regioisomeric ratios were measured by integration of ¹H NMR spectra of product mixtures prior to purification. High resolution mass spectra (HRMS) were obtained from Columbia University Mass Spectrometry Facility on a JOEL JMSHX110HF mass spectrometer using the ASAP ionization model. Infrared spectra were collected on a Perkin Elmer Spectrum Two FT-IR Spectrometer.

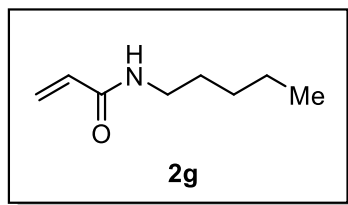
2. Preparation of Starting Materials

All boronic acids (**1a-1r**), alkenes **2b**, **2c**, **2d**, **2e**, and **2i** were purchased from commercial sources and used without purification. Alkene **2f**,¹ **2j**,² **2k**,³ **2l**,⁴ **2m**⁵ and dioxazolone **3c**,⁶ **3d**,⁷ **3e**,⁸ **3f**,⁷ **3g**,⁶ and **3h**⁷ were synthesized following literatures. Alkenes **2g**, **2h**, and dioxazolone **3b** were synthesized following the procedures below.

General procedure A: the synthesis of acrylamides (**2g-2h**)

To a solution of primary amine (or corresponding HCl salt, 1 equiv) and triethylamine (1.1 equiv) in dichloromethane (0.5 M) was added acryloyl chloride (1.1 equiv) in dichloromethane at 0 °C. The mixture was stirred at 0 °C for 15 minutes and warmed to room temperature. The reaction was stirred at room temperature for 3 hours. The volatiles was removed by a rotary evaporator then ethyl acetate was added and filtered through celite. The filtrate was washed with 1M HCl, saturated NaHCO₃ solution, and brine.

The organic layer was dried over anhydrous MgSO_4 and concentrated. The residue was purified by chromatography on silica gel (hexane/ethyl acetate) to afford the corresponding acrylamides.



Yield: Prepared according to general procedure **A** from 1-aminopentane (10 mmol). 78%, 1.11g, colorless oil.

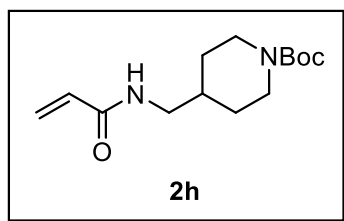
$R_f = 0.39$ (EA/Hex 2:3).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.23 (d, $J = 17.1$ Hz, 1H), 6.11 (dd, $J = 17.0, 10.2$ Hz, 1H), 6.05 (s, 1H), 5.58 (dt, $J = 10.0$ Hz, 1H), 3.28 (q, $J = 7.3, 6.7$ Hz, 2H), 1.50 (q, $J = 7.2$ Hz, 2H), 1.36 – 1.21 (m, 4H), 0.87 (t, $J = 6.8$, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 165.75, 131.22, 125.99, 39.70, 29.33, 29.18, 22.43, 14.04.

IR (CDCl_3 , cm^{-1}) ν 3276, 3079, 2957, 2929, 2861, 1656, 1623, 1548, 1407, 1245, 985, 954, 727.

HRMS (ASAP) m/z [$\text{C}_8\text{H}_{16}\text{NO}$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 142.1232, found 142.1236.



Yield: Prepared according to general procedure **A** from 1-Boc-4-(aminomethyl)piperidine (10 mmol). 65%, 1.74g, colorless oil.

$R_f = 0.18$ (EA/Hex 2:3).

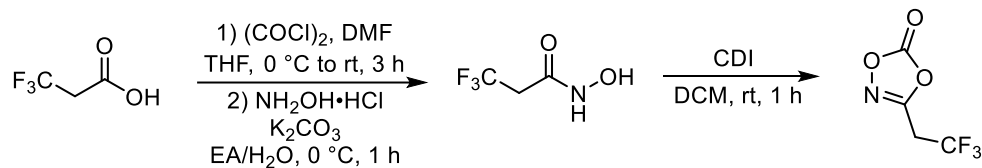
$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.26 (dd, $J = 17.0, 1.6$ Hz, 1H), 6.10 (dd, $J = 17.0, 10.2$ Hz, 1H), 6.06 (s, 1H), 5.61 (dd, $J = 10.2, 1.6$ Hz, 1H), 4.16 – 3.97 (m, 2H), 3.30 – 3.12 (m, 2H), 2.65 (t, $J = 12.7$ Hz, 2H), 1.73 – 1.61 (m, 3H), 1.42 (s, 9H), 1.19 – 1.04 (m, 2H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 165.82, 154.95, 130.86, 126.74, 79.57, 45.10, 43.76, 36.57, 29.92, 28.59.

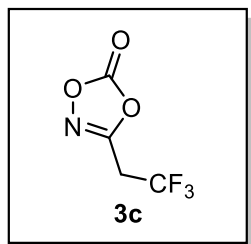
IR (CDCl₃, cm⁻¹) ν 3302, 3080, 2976, 2926, 2854, 2244, 1658, 1546, 1424, 1365, 1226, 1167, 1143, 906, 727, 647.

HRMS (ESI) m/z [C₁₄H₂₄N₂NaO₃]⁺ ([M+Na]⁺) calculated 291.1685, found 291.1699.

Preparation of 3c



Oxalyl chloride (9.8 mmol, 0.98 equiv) and catalytic amount (5 – 10 drops) of DMF were added dropwise to a solution of 3,3,3-trifluoropropanoic acid (10 mmol, 1.0 equiv) in THF (0.3 M) at 0 °C. The reaction mixture was warmed to room temperature and stirred for 3 hours. After the full conversion of acid, the crude material was taken without concentration. In a separate flask, hydroxylamine hydrochloride (11 mmol, 1.1 equiv) was dissolved in 0.1 M of ethyl acetate/H₂O 2:1 biphasic mixture. K₂CO₃ (20 mmol, 2 equiv) was added to the solution and cooled to 0 °C. The crude THF solution of acid chloride was added dropwise to the mixture and stirred for 1 hour at 0 °C. After completion of the reaction, the reaction mixture was extracted with ethyl acetate. The combined organic layers were washed with brine, dried over anhydrous MgSO₄, filtered, and concentrated. The residue was used without further purification. To a solution of 3,3,3-trifluoro-*N*-hydroxypropanamide (3.22 mmol, 1.0 equiv) in DCM (0.2 M), CDI (3.53 mmol, 1.2 equiv) was added as one portion. After stirring for 1 hour at room temperature, the reaction was quenched with 1 M HCl and extracted with DCM three times. The combined organic layer was washed with brine and dried over sodium sulfate. The solvent was removed under reduced pressure to afford 3-(2,2,2-trifluoroethyl)-1,4,2-dioxazol-5-one (**3c**). No further purification was required.



Yield: 44% (over 2 steps), 2.05 mmol, 346 mg, colorless oil.

¹H NMR (400 MHz, CDCl₃) δ 3.56 (q, J = 9.1 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 158.61 (q, J = 3.9 Hz), 153.03, 122.37 (q, J = 278.2 Hz), 31.48 (q, J = 34.9 Hz).

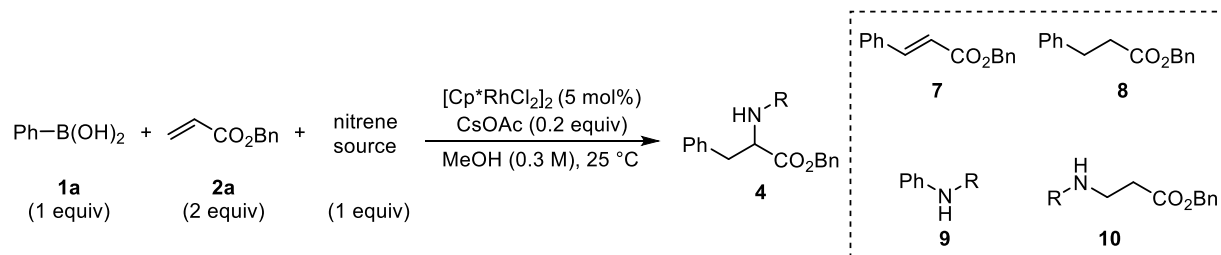
¹⁹F NMR (376 MHz, CDCl₃) δ -63.40 (t, J = 9.1 Hz).

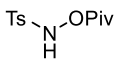
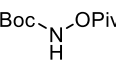
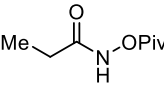
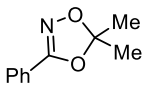
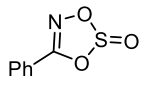
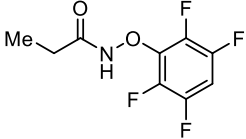
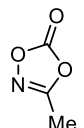
3. General procedure for the carboamination of alkenes

[Cp*RhCl₂]₂ (0.0050 mmol, 5 mol%), sodium bicarbonate (0.03 mmol, 0.3 equiv), and boronic acid (0.25 mmol, 2.5 equiv) were measured in a 0.5-dram vial with a magnetic stir bar. In a separate vial, dioxazolone (0.1 mmol, 1 equiv) and alkene (0.3 mmol, 3 equiv) were dissolved in 0.10 mL of MeOH and transferred to the first vial. It was rinsed with an additional 0.07 mL of MeOH and transferred again to the first vial. The reaction mixture was then stirred at 25 °C for 16 hours. After the completion of the reaction, the solvent was removed by a rotary evaporator. Ethyl acetate was added to the residue and filtered through a short pad of celite and washed with ethyl acetate three times. The filtrate was concentrated by rotary evaporator and a crude ¹H NMR spectrum was collected with 1,3,5-trimethoxybenzene as the internal standard. Purification was performed by flash chromatography or preparational TLC using ethyl acetate and hexane or DCM and MeOH as the eluent.

4. Detailed optimization table

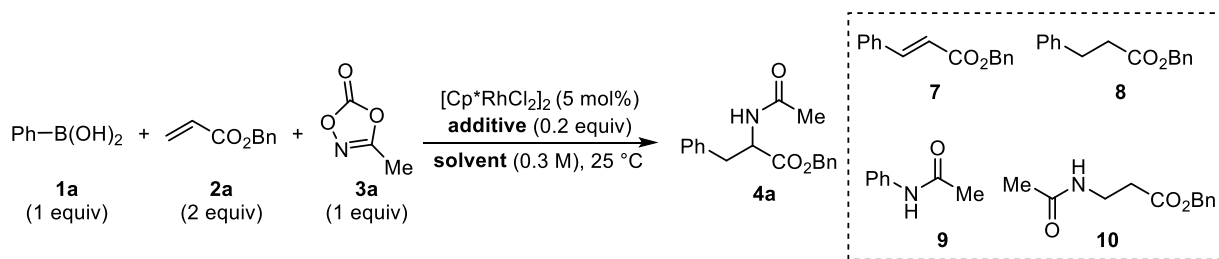
Table S1. Nitrene precursor screening.^a



entry	nitrene source (0.2 equiv)	4 (%)	7 (%)	8 (%)	9 (%)	10 (%)
1		-	-	-	-	-
2	Ts-N_3	-	-	-	-	-
3		-	-	-	>95	-
4		-	-	-	>95	-
5		-	-	-	-	-
6		-	-	-	-	-
7		-	80	6	-	-
8		8	24	< 5	10	< 5

^aReaction were conducted on 0.1 mmol scale according to the general procedure. Yield was determined by ¹H NMR spectroscopy of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. The yield of side products was calculated based on 0.1 mmol.

Table S2. Solvent and additive screening.^a



entry	solvent (0.3 M)	additive (0.2 equiv)	4a (%)	7 (%)	8 (%)	9 (%)	10 (%)
1	MeOH	CsOAc	8	24	<5	10	<5
2	MeOH	CsOPiv	<5	13	<5	<5	<5
3	MeOH	Cs ₂ CO ₃	48	28	<5	9	6
4	THF	Cs ₂ CO ₃	29	20	<5	<5	6
5	TFE	Cs ₂ CO ₃	16	20	<5	46	9
6	DCE	Cs ₂ CO ₃	17	46	<5	21	11
7	EtOH	Cs ₂ CO ₃	28	30	<5	24	<5
8	HFIP	Cs ₂ CO ₃	<5	10	<5	35	6
9	MeCN	Cs ₂ CO ₃	<5	24	<5	10	8
10	Acetone	Cs ₂ CO ₃	15	22	<5	7	<5
11	<i>i</i> -PrOH	Cs ₂ CO ₃	16	14	<5	11	<5
12	<i>t</i> -BuOH	Cs ₂ CO ₃	6	10	<5	8	<5
13	MeOH	Li ₂ CO ₃	44	30	6	9	<5
14	MeOH	Na ₂ CO ₃	45	29	6	8	5
15	MeOH	K ₂ CO ₃	42	30	7	8	5
16	MeOH	NaHCO ₃	44	28	8	8	<5
17	MeOH	NaF	48	33	<5	7	8
18	MeOH	NaOAc	<5	20	<5	6	<5

^aReaction were conducted on 0.1 mmol scale according to the general procedure. Yield was determined by ¹H NMR spectroscopy of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. The yield of side products was calculated based on 0.1 mmol.

Table S3. Concentration, additive, and temperature screening^a

Reaction scheme: Ph-B(OH)_2 (**1a**, 1 equiv) + $\text{CH}_2=\text{CHCO}_2\text{Bn}$ (**2a**, 2 equiv) + N-methylimidazole (**3a**, 1 equiv) $\xrightarrow[\text{temp. (}^\circ\text{C)}]{[\text{Cp}^*\text{RhCl}_2]_2 \text{ (5 mol\%)} \text{ NaHCO}_3 \text{ (equiv), MeOH (conc.)}$ $\text{Ph-CH}_2\text{-CH(Me)-NH-CO-Me-CO}_2\text{Bn}$ (**4a**)

Side products shown in a dashed box: **7** ($\text{Ph-CH=CH-CO}_2\text{Bn}$), **8** ($\text{Ph-CH}_2\text{-CH}_2\text{-CO}_2\text{Bn}$), **9** (Ph-NH-CO-Me), **10** ($\text{Me-CO-NH-CH}_2\text{-CH}_2\text{-CO}_2\text{Bn}$).

entry	concentration (M)	additive (equiv)	temperature ($^\circ\text{C}$)	4a (%)	7 (%)	8 (%)	9 (%)	10 (%)
1	0.1	NaHCO ₃ (0.2)	25	32	32	7	8	9
2	0.3	NaHCO ₃ (0.2)	25	45	24	5	8	6
3	0.6	NaHCO ₃ (0.2)	25	59	22	<5	8	6
4	0.6	NaHCO ₃ (0.1)	25	56	20	<5	8	6
5	0.6	NaHCO ₃ (0.3)	25	59	20	<5	9	6
6	0.6	NaHCO ₃ (0.6)	25	60	20	<5	8	6
7	0.6	NaHCO ₃ (1.0)	25	59	20	<5	8	6
8	0.6	NaHCO ₃ (0.3)	40	56	34	<5	11	13
9	0.6	NaHCO ₃ (0.3)	60	42	40	<5	13	20

^aReaction were conducted on 0.1 mmol scale according to the general procedure. Yield was determined by ¹H NMR spectroscopy of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. The yield of side products was calculated based on 0.1 mmol.

Table S4. Effect of ratio of each component.^a

entry	1a (equiv) / 2a (equiv) / 3a (equiv)	4a (%)	7 (%)	8 (%)	9 (%)	10 (%)
1	1 / 1 / 1	55	20	<5	16	5
2	1 / 1 / 2	54	10	<5	35	<5
3	1 / 1 / 3	44	6	<5	51	<5
4	1 / 2 / 1	59	22	<5	8	6
5	1 / 3 / 1	57	22	<5	5	5
6	2 / 1 / 1	64	20	5	22	8
7	3 / 1 / 1	58	16	<5	24	7
8	2 / 2 / 1	71	36	<5	8	13
9	3 / 2 / 1	73	32	<5	9	14
10	4 / 2 / 1	75	34	6	9	15
11	2 / 3 / 1	76	30	<5	6	15
12	2 / 4 / 1	76	32	<5	<5	15
13	3 / 3 / 1	77	34	<5	<5	15
14	2.5 / 3 / 1	77	35	<5	<5	15

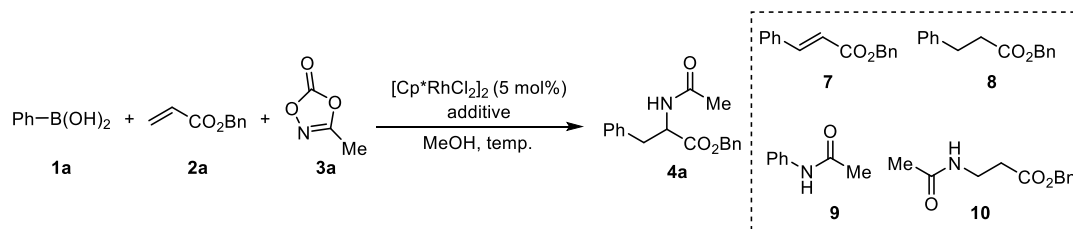
^aReaction were conducted on 0.1 mmol scale according to the general procedure. Yield was determined by ¹H NMR spectroscopy of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard. The yield of side products was calculated based on 0.1 mmol.

Table S5. Carbon source screening.^a

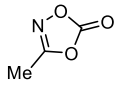
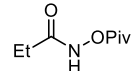
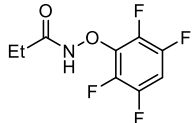
entry	carbon source (2.5 equiv)	yield (%)
1	Ph-B(OH) ₂	77
2		39
3	Ph-BF ₃ K	trace

^aReaction were conducted on 0.1 mmol scale according to the general procedure. Yield was determined by ¹H NMR spectroscopy of the crude reaction mixture using 1,3,5-trimethoxybenzene as an internal standard.

Table S6. Summary of the detailed optimization table.



- Different nitrene sources give different side products.

nitrene source	4a (%)	7 (%)	8 (%)	9 (%)	10 (%)
	8	24	< 5	10	< 5
	-	-	-	>95	-
	-	80	6	-	-

1a (1 equiv), **2a** (2 equiv), **3a** (1 equiv), CsOAc (0.2 equiv), MeOH (0.3 M)

- Acetates inhibit product formation, **7** becomes major product.

additive	4a (%)	7 (%)	8 (%)	9 (%)	10 (%)
NaHCO ₃	44	28	8	8	5
NaOAc	<5	20	<5	6	<5

1a (1 equiv), **2a** (2 equiv), **3a** (1 equiv), additive (0.2 equiv), MeOH (0.3 M)

- Acidic solvent leads to more direct C-N coupling product (**9**).

solvent	4a (%)	7 (%)	8 (%)	9 (%)	10 (%)
MeOH	48	28	<5	9	6
TFE	16	20	<5	46	9

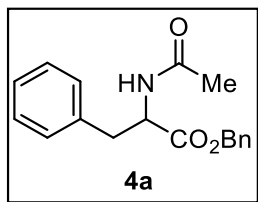
1a (1 equiv), **2a** (2 equiv), **3a** (1 equiv), Cs₂CO₃ (0.3 equiv), MeOH (0.3 M)

- Higher temperature give low chemoselectivity

temperature	4a (%)	7 (%)	8 (%)	9 (%)	10 (%)
25 °C	59	20	<5	9	6
60 °C	42	40	<5	13	20

1a (1 equiv), **2a** (2 equiv), **3a** (1 equiv), NaHCO₃ (0.3 equiv), MeOH (0.6 M)

5. Product characterization



Yield: 68%, 20 mg, white solid.

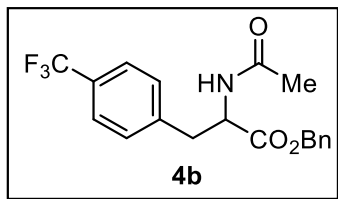
R_f = 0.26 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.29 (m, 3H), 7.31 – 7.26 (m, 2H), 7.26 – 7.17 (m, 3H), 7.04 – 6.95 (m, 2H), 5.93 (d, *J* = 7.9 Hz, 1H), 5.17 (d, *J* = 12.1 Hz, 1H), 5.12 (d, *J* = 12.1 Hz, 1H), 4.92 (dt, *J* = 7.9, 5.7 Hz, 1H), 3.18 – 3.03 (m, 2H), 1.98 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.62, 169.68, 135.82, 135.17, 129.43, 128.76, 128.72, 128.69, 128.67, 127.20, 67.40, 53.26, 37.94, 23.27.

IR (CDCl₃, cm⁻¹) ν 3277, 3063, 3031, 2932, 1738, 1652, 1540, 1372, 1209, 1173, 734, 696.

HRMS (ASAP) *m/z* [C₁₈H₂₀NO₃]⁺ ([M+H]⁺) calculated 298.1443, found 298.1437.



Yield: 62%, 23 mg, white solid.

R_f = 0.38 (EA/Hex 2:3).

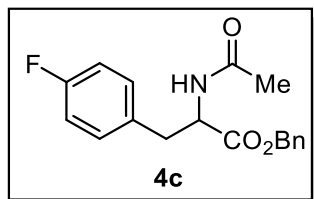
¹H NMR (400 MHz, CDCl₃) δ 7.44 (d, *J* = 8.0 Hz, 2H), 7.40 – 7.34 (m, 3H), 7.31 – 7.24 (m, 2H), 7.07 (d, *J* = 7.9 Hz, 2H), 5.98 (d, *J* = 7.3 Hz, 1H), 5.19 (d, *J* = 12.0 Hz, 1H), 5.09 (d, *J* = 12.0 Hz, 1H), 4.97 – 4.90 (m, 1H), 3.23 – 3.11 (m, 2H), 1.99 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.28, 169.74, 140.03 (d, *J* = 1.6 Hz), 134.89, 129.80, 129.45 (q, *J* = 32.7 Hz), 128.92, 128.84, 125.51 (q, *J* = 3.7 Hz), 124.17 (q, *J* = 270.9 Hz), 67.66, 53.10, 37.80, 23.28.

¹⁹F NMR (376 MHz, CDCl₃) δ -62.51.

IR (CDCl₃, cm⁻¹) ν 3276, 3066, 3036, 2939, 1740, 1654, 1541, 1324, 1164, 1122, 1067, 904, 729, 699.

HRMS (ASAP) m/z [C₁₉H₁₉F₃NO₃]⁺ ([M+H]⁺) calculated 366.1317, found 366.1310.



Yield: 60%, 20 mg, white solid

R_f = 0.25 (EA/Hex 2:3).

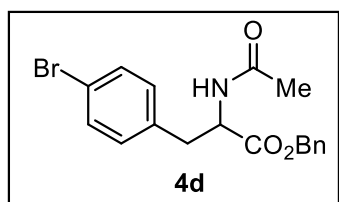
¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.34 (m, 3H), 7.33 – 7.19 (m, 2H), 6.96 – 6.84 (m, 4H), 5.97 (d, J = 7.7 Hz, 1H), 5.19 (d, J = 12.0 Hz, 1H), 5.10 (d, J = 12.1 Hz, 1H), 4.89 (dt, J = 7.8, 5.7 Hz, 1H), 3.15 – 3.00 (m, 2H), 1.98 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.49, 169.69, 162.08 (d, J = 245.4 Hz), 135.05, 131.51 (d, J = 3.3 Hz), 130.91 (d, J = 8.0 Hz), 128.85, 128.81, 115.48 (d, J = 21.3 Hz), 67.50, 53.30, 37.17, 23.28. (Two aromatic peaks overlapped).

¹⁹F NMR (376 MHz, CDCl₃) δ -115.54 – -115.76 (m).

IR (CDCl₃, cm⁻¹) ν 3278, 3066, 2935, 1738, 1653, 1541, 1508, 1373, 1220, 1176, 905, 828, 729, 698.

HRMS (ASAP) m/z [C₁₈H₁₉FNO₃]⁺ ([M+H]⁺) calculated 316.1349, found 316.1348.



Yield: 65%, 25 mg, white solid.

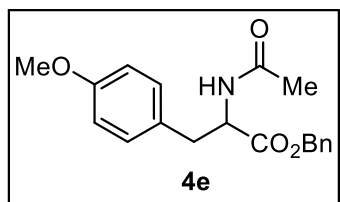
R_f = 0.31 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.34 (m, 3H), 7.33 – 7.25 (m, 4H), 6.87 – 6.79 (m, 2H), 5.96 (d, J = 7.7 Hz, 1H), 5.19 (d, J = 12.0 Hz, 1H), 5.09 (d, J = 12.1 Hz, 1H), 4.89 (dt, J = 7.8, 5.7 Hz, 1H), 3.13 – 2.99 (m, 2H), 1.98 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.38, 169.70, 134.99, 134.84, 131.73, 131.14, 128.87, 128.85, 128.83, 121.21, 67.54, 53.11, 37.40, 23.26.

IR (CDCl_3 , cm^{-1}) ν 3277, 3063, 3034, 2930, 1739, 1652, 1539, 1488, 1209, 1128, 1071, 813, 749.

HRMS (ASAP) m/z [$\text{C}_{18}\text{H}_{19}\text{BrNO}_3$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 376.0548, found 376.0555.



Yield: 60%, 20 mg, white solid.

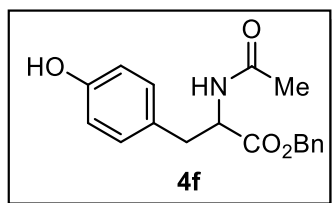
R_f = 0.33 (EA/Hex 2:3).

^1H NMR (400 MHz, CDCl_3) δ 7.41 – 7.33 (m, 3H), 7.33 – 7.27 (m, 2H), 6.94 – 6.86 (m, 2H), 6.78 – 6.70 (m, 2H), 5.91 (d, J = 7.9 Hz, 1H), 5.17 (d, J = 12.1 Hz, 1H), 5.11 (d, J = 12.1 Hz, 1H), 4.88 (dt, J = 7.9, 5.7 Hz, 1H), 3.76 (s, 3H), 3.14 – 3.00 (m, 2H), 1.97 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.70, 169.69, 158.76, 135.17, 130.40, 128.72, 128.66, 127.68, 114.05, 67.33, 55.29, 53.38, 37.05, 23.26. (Two aromatic peaks overlapped)

IR (CDCl_3 , cm^{-1}) ν 3278, 3063, 3034, 2953, 2934, 2836, 1739, 1653, 1511, 1373, 1246, 1177, 1033, 698.

HRMS (ASAP) m/z [$\text{C}_{19}\text{H}_{22}\text{NO}_4$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 328.1549, found 328.1549.



Yield: 70%, 22 mg, white solid.

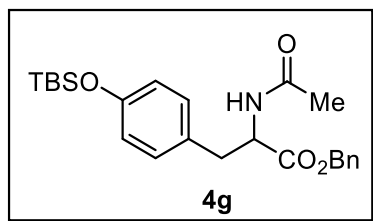
R_f = 0.17 (EA/Hex 2:3).

^1H NMR (400 MHz, CDCl_3) δ 7.41 – 7.23 (m, 5H), 7.02 (s, 1H), 6.87 – 6.79 (m, 2H), 6.77 – 6.62 (m, 2H), 6.10 (d, J = 8.1 Hz, 1H), 5.18 (d, J = 12.1 Hz, 1H), 5.12 (d, J = 12.1 Hz, 1H), 4.90 (dt, J = 8.0, 5.9 Hz, 1H), 3.06 (dd, J = 14.1, 5.8 Hz, 1H), 2.97 (dd, J = 14.1, 6.0 Hz, 1H), 1.97 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.84, 170.51, 155.65, 135.10, 130.43, 128.78, 128.73, 128.71, 126.90, 115.70, 67.52, 53.54, 37.22, 23.16.

IR (CDCl_3 , cm^{-1}) ν 3298, 3062, 3033, 2930, 1737, 1652, 1514, 1451, 1374, 1213, 905, 730, 699.

HRMS (ASAP) m/z [$\text{C}_{18}\text{H}_{20}\text{NO}_4$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 314.1392, found 314.1390.



Yield: 66%, 28 mg, colorless oil.

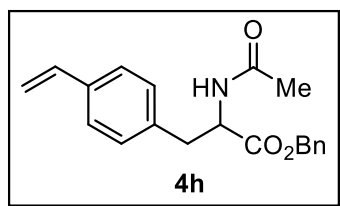
R_f = 0.38 (EA/Hex 2:3).

^1H NMR (400 MHz, CDCl_3) δ 7.41 – 7.29 (m, 5H), 6.89 – 6.79 (m, 2H), 6.73 – 6.63 (m, 2H), 5.89 (d, J = 7.9 Hz, 1H), 5.16 (d, J = 12.1 Hz, 1H), 5.11 (d, J = 12.2 Hz, 1H), 4.88 (dt, J = 7.9, 5.7 Hz, 1H), 3.11 – 2.96 (m, 2H), 1.97 (s, 3H), 0.97 (s, 9H), 0.18 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.71, 169.65, 154.88, 135.24, 130.41, 128.77, 128.72, 128.67, 128.36, 120.21, 67.36, 53.35, 37.16, 25.79, 23.28, 18.30, -4.29.

IR (CDCl_3 , cm^{-1}) ν 3282, 3063, 3033, 2954, 2930, 2888, 2857, 1740, 1655, 1508, 1254, 906, 838, 728, 697.

HRMS (ASAP) m/z [$\text{C}_{24}\text{H}_{34}\text{NO}_4\text{Si}$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 428.2257, found 428.2254.



Yield: 55%, 18 mg, white solid.

R_f = 0.31 (EA/Hex 2:3).

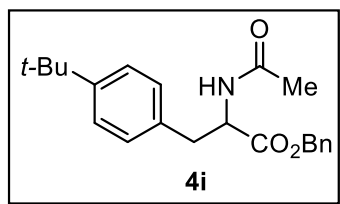
^1H NMR (400 MHz, CDCl_3) δ 7.39 – 7.33 (m, 3H), 7.34 – 7.22 (m, 4H), 6.99 – 6.91 (m, 2H), 6.67 (dd, J = 17.6, 10.9 Hz, 1H), 5.94 (d, J = 7.9 Hz, 1H), 5.70 (dd, J = 17.6, 1.0 Hz, 1H), 5.23 (dd, J = 10.9, 0.9 Hz,

1H), 5.17 (d, $J = 12.1$ Hz, 1H), 5.11 (d, $J = 12.1$ Hz, 1H), 4.92 (dt, $J = 7.8, 5.8$ Hz, 1H), 3.17 – 3.03 (m, 2H), 1.98 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.59, 169.72, 136.58, 136.51, 135.40, 135.12, 129.61, 128.77, 128.74, 128.70, 126.50, 113.90, 67.43, 53.25, 37.69, 23.27.

IR (CDCl_3 , cm^{-1}) ν 3278, 3063, 2930, 2252, 1739, 1654, 1539, 1511, 1373, 1209, 1178, 904, 727, 699, 649.

HRMS (ASAP) m/z [$\text{C}_{20}\text{H}_{22}\text{NO}_3$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 324.1600, found 324.1605.



Yield: 55%, 19 mg, white solid.

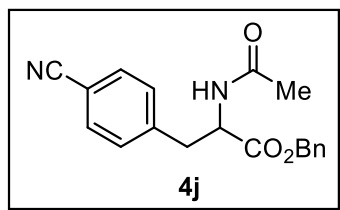
$R_f = 0.37$ (EA/Hex 2:3).

^1H NMR (400 MHz, CDCl_3) δ 7.38 – 7.33 (m, 3H), 7.32 – 7.27 (m, 2H), 7.26 – 7.22 (m, 2H), 6.97 – 6.91 (m, 2H), 5.93 (d, $J = 7.7$ Hz, 1H), 5.18 – 5.09 (m, 2H), 4.92 (dt, $J = 7.9, 5.7$ Hz, 1H), 3.08 (d, $J = 5.7$ Hz, 2H), 1.98 (s, 3H), 1.29 (s, 9H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.76, 169.74, 150.05, 135.21, 132.63, 129.10, 128.74, 128.70, 128.66, 125.60, 67.39, 53.26, 37.39, 34.55, 31.45, 23.29.

IR (CDCl_3 , cm^{-1}) ν 3280, 3062, 3033, 2960, 2868, 1740, 1654, 1537, 1515, 1368, 1190, 906, 730, 697.

HRMS (ASAP) m/z [$\text{C}_{22}\text{H}_{28}\text{NO}_3$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 354.2069, found 354.2072.



Yield: 51%, 16 mg, yellow oil.

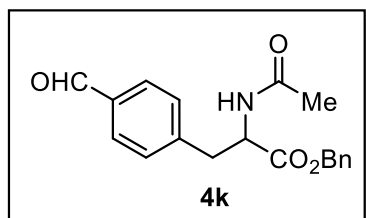
$R_f = 0.23$ (EA/Hex 1:1).

¹H NMR (400 MHz, CDCl₃) δ 7.48 – 7.38 (m, 2H), 7.42 – 7.31 (m, 3H), 7.31 – 7.28 (m, 2H), 7.08 – 7.02 (m, 2H), 6.03 (d, *J* = 7.5 Hz, 1H), 5.22 (d, *J* = 12.0 Hz, 1H), 5.08 (d, *J* = 11.9 Hz, 1H), 4.92 (dt, *J* = 7.5, 5.8 Hz, 1H), 3.16 (qd, *J* = 13.8, 5.9 Hz, 2H), 1.99 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.07, 169.75, 141.57, 134.82, 132.27, 130.22, 129.01, 128.99, 128.86, 118.76, 111.11, 67.70, 53.00, 38.13, 23.24.

IR (CDCl₃, cm⁻¹) ν 3283, 3064, 3035, 2936, 2228, 1736, 1656, 1534, 1505, 1374, 1268, 1211, 1175, 905, 731, 698.

HRMS (ASAP) *m/z* [C₁₉H₁₉N₂O₃]⁺ ([M+H]⁺) calculated 323.1396, found 323.1390.



Yield: 64%, 21 mg, white solid.

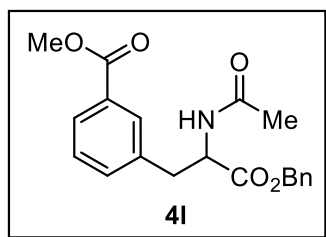
R_f = 0.20 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 9.94 (s, 1H), 7.73 – 7.66 (m, 2H), 7.41 – 7.31 (m, 3H), 7.32 – 7.27 (m, 2H), 7.17 – 7.10 (m, 2H), 6.05 (d, *J* = 7.7 Hz, 1H), 5.20 (d, *J* = 12.1 Hz, 1H), 5.10 (d, *J* = 12.0 Hz, 1H), 4.96 (dt, *J* = 7.7, 5.9 Hz, 1H), 3.19 (qd, *J* = 13.8, 5.9 Hz, 2H), 1.99 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 191.90, 171.23, 169.76, 143.15, 135.40, 134.90, 130.15, 129.97, 128.90, 128.89, 128.83, 67.63, 53.06, 38.20, 23.24.

IR (CDCl₃, cm⁻¹) ν 3284, 3062, 3034, 2932, 2831, 2738, 2251, 1738, 1695, 1656, 1606, 1534, 1374, 1306, 1211, 1170, 905, 730, 698.

HRMS (ASAP) *m/z* [C₁₉H₂₀NO₄]⁺ ([M+H]⁺) calculated 326.1392, found 326.1387.



Yield: 65%, 23 mg, yellow oil.

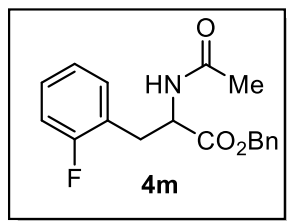
R_f = 0.19 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.90 (dt, *J* = 7.7, 1.5 Hz, 1H), 7.79 (t, *J* = 1.8 Hz, 1H), 7.41 – 7.32 (m, 3H), 7.36 – 7.24 (m, 3H), 7.19 (dt, *J* = 7.7, 1.5 Hz, 1H), 6.04 (d, *J* = 7.8 Hz, 1H), 5.28 – 5.09 (m, 2H), 4.94 (dt, *J* = 7.7, 5.9 Hz, 1H), 3.89 (s, 3H), 3.20 (dd, *J* = 13.8, 6.0 Hz, 1H), 3.13 (dd, *J* = 13.9, 5.7 Hz, 1H), 1.99 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.42, 169.79, 166.94, 136.35, 134.99, 133.92, 130.58, 130.49, 128.77, 128.73, 128.68, 128.46, 67.57, 53.28, 52.25, 37.79, 23.22. (Two aromatic peaks overlapped)

IR (CDCl₃, cm⁻¹) ν 3282, 3064, 3034, 2951, 1719, 1655, 1537, 1436, 1283, 1202, 1109, 908, 747, 733, 697.

HRMS (ASAP) *m/z* [C₂₀H₂₂NO₅]⁺ ([M+H]⁺) calculated 356.1498, found 356.1502.



Yield: 74%, 23 mg, yellow oil.

R_f = 0.45 (EA/Hex 2:3).

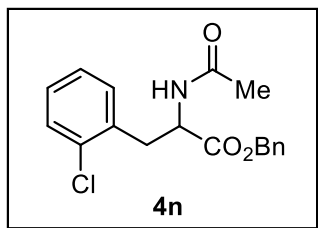
¹H NMR (400 MHz, CDCl₃) δ 7.01 – 7.33 (m, 3H), 7.33 – 7.28 (m, 2H), 7.24 – 7.17 (m, 1H), 7.08 – 6.95 (m, 3H), 6.04 (d, *J* = 8.0 Hz, 1H), 5.17 (d, *J* = 12.2, 1H), 5.12 (d, *J* = 12.2 Hz, 1H), 4.91 (dt, *J* = 8.0, 6.0 Hz, 1H), 3.22 (ddd, *J* = 13.9, 6.0, 1.2 Hz, 1H), 3.12 (ddd, *J* = 13.9, 6.1, 1.1 Hz, 1H), 1.96 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.52, 169.79, 161.48 (d, *J* = 245.2 Hz), 135.17, 131.80 (d, *J* = 5.1 Hz), 129.12 (d, *J* = 8.3 Hz), 128.73, 128.64, 128.60, 124.32 (d, *J* = 3.9 Hz), 123.02 (d, *J* = 16.0 Hz), 115.49 (d, *J* = 22.5 Hz), 67.56, 52.67, 31.52 (d, *J* = 1.8 Hz), 23.17.

¹⁹F NMR (376 MHz, CDCl₃) δ -117.50 (m).

IR (CDCl₃, cm⁻¹) ν 3277, 3064, 3036, 2936, 1739, 1654, 1539, 1493, 1454, 1373, 1271, 1209, 1230, 1176, 1132, 755, 697.

HRMS (ASAP) *m/z* [C₁₈H₁₉FNO₃]⁺ ([M+H]⁺) calculated 316.1349, found 316.1350.



Yield: 37%, 12 mg, colorless liquid.

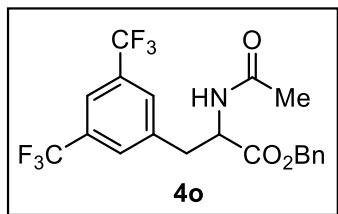
R_f = 0.26 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.24 (m, 6H), 7.21 – 7.05 (m, 3H), 6.01 (d, *J* = 8.2 Hz, 1H), 5.17 (d, *J* = 12.2 Hz, 1H), 5.12 (d, *J* = 12.2 Hz, 1H), 4.96 (ddd, *J* = 8.2, 7.1, 6.3 Hz, 1H), 3.32 (dd, *J* = 13.9, 6.3 Hz, 1H), 3.18 (dd, *J* = 13.9, 7.1 Hz, 1H), 1.95 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.65, 169.76, 135.18, 134.62, 134.17, 131.46, 129.78, 128.73, 128.69, 128.61, 128.50, 127.08, 67.52, 52.70, 35.55, 23.23.

IR (CDCl₃, cm⁻¹) ν 3275, 3064, 2939, 1742, 1653, 1543, 1444, 1373, 1269, 1177, 1134, 1052, 751, 697.

HRMS (ASAP) *m/z* [C₁₈H₁₉ClNO₃]⁺ ([M+H]⁺) calculated 332.1053, found 332.1052.



Yield: 52%, 23 mg, white solid.

R_f = 0.46 (EA/Hex 2:3).

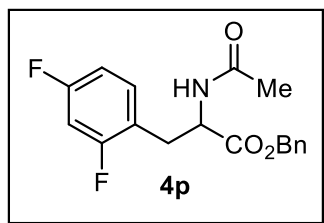
¹H NMR (400 MHz, CDCl₃) δ 7.74 (s, 1H), 7.54 – 7.49 (m, 2H), 7.42 – 7.33 (m, 3H), 7.33 – 7.25 (m, 2H), 6.07 (d, *J* = 7.2 Hz, 1H), 5.14 (s, 2H), 4.93 (ddd, *J* = 7.2, 6.4, 5.3 Hz, 1H), 3.32 (dd, *J* = 13.9, 6.3 Hz, 1H), 3.21 (dd, *J* = 13.9, 5.3 Hz, 1H), 2.00 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 170.93, 169.87, 138.76, 134.58, 131.84 (q, *J* = 33.3 Hz), 129.72 (d, *J* = 3.9 Hz), 129.03, 128.94, 128.63, 123.31 (q, *J* = 272.7 Hz), 121.24 (p, *J* = 3.9 Hz), 67.95, 53.19, 37.62, 23.13.

¹⁹F NMR (376 MHz, CDCl₃) δ -62.87 (s).

IR (CDCl₃, cm⁻¹) ν 3271, 3068, 2957, 1741, 1654, 1545, 1378, 1276, 1171, 1129, 900, 700, 682.

HRMS (ASAP) m/z $[C_{20}H_{18}F_6NO_3]^+$ ($[M+H]^+$) calculated 434.1191, found 434.1189.



Yield: 65%, 22 mg, colorless oil.

R_f = 0.28 (EA/Hex 2:3).

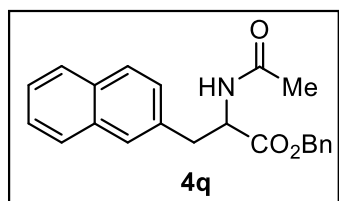
1H NMR (400 MHz, $CDCl_3$) δ 7.42 – 7.28 (m, 5H), 6.97 (td, J = 8.6, 6.3 Hz, 1H), 6.78 – 6.67 (m, 2H), 6.03 (d, J = 7.9 Hz, 1H), 5.14 (s, 2H), 4.89 (dt, J = 7.9, 6.0 Hz, 1H), 3.19 (ddd, J = 14.1, 6.1, 1.2 Hz, 1H), 3.07 (dd, J = 14.1, 5.9 Hz, 1H), 1.97 (s, 3H).

^{13}C NMR (101 MHz, $CDCl_3$) δ 171.41, 169.77, 162.26 (dd, J = 248.4, 12.1 Hz), 161.42 (dd, J = 247.8, 11.9 Hz), 135.05, 132.37 (dd, J = 9.6, 6.2 Hz), 128.77, 128.75, 128.70, 118.91 (dd, J = 16.3, 3.8 Hz), 111.46 (dd, J = 21.2, 3.7 Hz), 103.93 (dd, J = 25.7, 26.0 Hz), 67.66, 52.61, 31.10, 23.18.

^{19}F NMR (376 MHz, $CDCl_3$) δ -111.23 (p, J = 7.7 Hz), -113.16 (q, J = 8.7 Hz).

IR ($CDCl_3$, cm^{-1}) ν 3278, 3066, 3029, 2939, 1740, 1654, 1540, 1504, 1374, 1273, 1174, 1136, 1094, 965, 850, 747, 697.

HRMS (ASAP) m/z $[C_{18}H_{18}F_2NO_3]^+$ ($[M+H]^+$) calculated 334.1255, found 334.1255.



Yield: 52%, 18 mg, white solid.

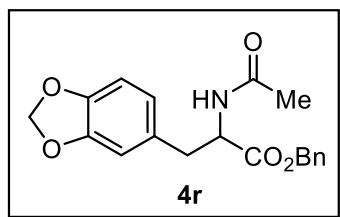
R_f = 0.26 (EA/Hex 2:3).

1H NMR (400 MHz, $CDCl_3$) δ 8.08 (d, J = 8.6 Hz, 1H), 7.89 – 7.80 (m, 1H), 7.73 (d, J = 8.3 Hz, 1H), 7.55 – 7.41 (m, 2H), 7.34 – 7.23 (m, 4H), 7.20 – 7.09 (m, 3H), 5.99 (d, J = 7.8 Hz, 1H), 5.10 – 4.97 (m, 3H), 3.65 – 3.49 (m, 2H), 1.91 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.88, 169.84, 135.03, 134.01, 132.46, 132.37, 128.93, 128.67, 128.58, 128.49, 128.06, 127.63, 126.42, 125.90, 125.41, 123.72, 67.36, 53.47, 35.22, 23.23.

IR (CDCl_3 , cm^{-1}) ν 3277, 3062, 2953, 2251, 1739, 1652, 1539, 1511, 1372, 1176, 905, 778, 728, 698.

HRMS (ASAP) m/z [$\text{C}_{22}\text{H}_{22}\text{NO}_3$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 348.1600, found 348.1603.



Yield: 65%, 22 mg, white solid.

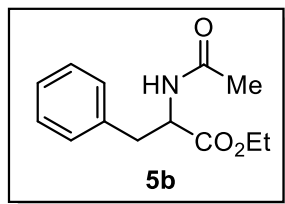
R_f = 0.37 (EA/Hex 1:1).

^1H NMR (400 MHz, CDCl_3) δ 7.40 – 7.34 (m, 3H), 7.34 – 7.29 (m, 2H), 6.64 (d, J = 7.9 Hz, 1H), 6.49 (d, J = 1.7 Hz, 1H), 6.42 (dd, J = 7.9, 1.7 Hz, 1H), 5.95 (d, J = 8.0 Hz, 1H), 5.91 (s, 2H), 5.18 (d, J = 12.1 Hz, 1H), 5.12 (d, J = 12.1 Hz, 1H), 4.87 (dt, J = 7.8, 5.7 Hz, 1H), 3.07 – 2.98 (m, 2H), 1.99 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.59, 169.70, 147.87, 146.81, 135.14, 129.40, 128.79, 128.74, 128.71, 122.53, 109.71, 108.41, 101.09, 67.43, 53.42, 37.68, 23.31.

IR (CDCl_3 , cm^{-1}) ν 3281, 3065, 3034, 2927, 2893, 1738, 1654, 1537, 1501, 1489, 1443, 1371, 1246, 1191, 1037, 904, 729, 699.

HRMS (ASAP) m/z [$\text{C}_{19}\text{H}_{20}\text{NO}_5$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 342.1341, found 342.1337.



Yield: 68%, 16 mg, white solid.

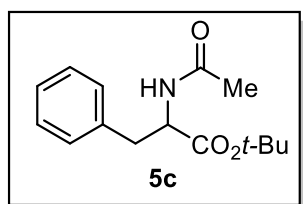
R_f = 0.30 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.33 – 7.19 (m, 3H), 7.14 – 7.07 (m, 2H), 5.93 (d, *J* = 7.8 Hz, 1H), 4.86 (dt, *J* = 7.9, 5.8 Hz, 1H), 4.17 (qd, *J* = 7.2, 1.2 Hz, 2H), 3.19 – 3.05 (m, 2H), 1.98 (s, 3H), 1.24 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 171.79, 169.72, 136.02, 129.42, 128.62, 127.18, 61.62, 53.26, 38.02, 23.25, 14.21.

IR (CDCl₃, cm⁻¹) ν 3283, 3063, 3030, 2982, 2932, 1737, 1656, 1542, 1374, 1212, 1028, 701.

HRMS (ASAP) *m/z* [C₁₃H₁₈NO₃]⁺ ([M+H]⁺) calculated 236.1287, found 236.1284.



Yield: 63%, 17 mg, yellowish oil.

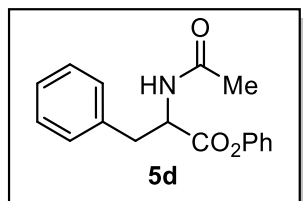
R_f = 0.31 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.18 (m, 3H), 7.18 – 7.05 (m, 2H), 5.97 (d, *J* = 7.7 Hz, 1H), 4.76 (dt, *J* = 7.8, 6.0 Hz, 1H), 3.09 (d, *J* = 6.0 Hz, 2H), 1.98 (s, 3H), 1.41 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 170.94, 169.64, 136.32, 129.62, 128.49, 127.08, 82.53, 53.65, 38.18, 28.07, 23.34.

IR (CDCl₃, cm⁻¹) ν 3285, 3064, 3030, 2978, 2931, 1732, 1655, 1544, 1498, 1369, 1257, 1226, 1154, 755, 738, 699.

HRMS (ESI) *m/z* [C₁₅H₂₁NO₃Na]⁺ ([M+H]⁺) calculated 286.1419, found 286.1433.



Yield: 65%, 18 mg, white solid.

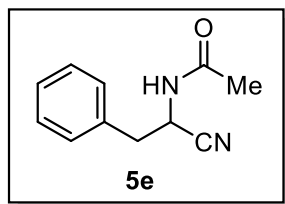
R_f = 0.28 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.19 (m, 8H), 7.03 – 6.95 (m, 2H), 6.04 (d, *J* = 7.8 Hz, 1H), 5.12 (dt, *J* = 7.7, 6.0 Hz, 1H), 3.34 – 3.20 (m, 2H), 2.01 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 170.50, 169.89, 150.38, 135.72, 129.63, 129.57, 128.86, 127.49, 126.33, 121.37, 53.44, 38.08, 23.23.

IR (CDCl₃, cm⁻¹) ν 3267, 3033, 2936, 1760, 1653, 1537, 1491, 1372, 1190, 1162, 1121, 908, 734, 699, 494.

HRMS (ESI) *m/z* [C₁₇H₁₈NO₃]⁺ ([M+H]⁺) calculated 284.1287, found 284.1281.



Yield: 70%, 13 mg, white solid.

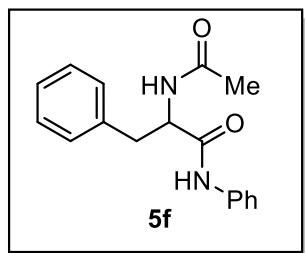
R_f = 0.27 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.28 (m, 3H), 7.31 – 7.24 (m, 2H), 6.17 (d, *J* = 8.6 Hz, 1H), 5.14 (ddd, *J* = 8.7, 7.4, 5.8 Hz, 1H), 3.08 (m, 2H), 1.98 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 169.56, 134.03, 129.57, 129.14, 128.09, 118.30, 41.66, 38.88, 22.89.

IR (CDCl₃, cm⁻¹) ν 3270, 3032, 2932, 2856, 2244, 1954, 1658, 1531, 1497, 1372, 1289, 749, 700.

HRMS (ASAP) *m/z* [C₁₁H₁₃N₂O]⁺ ([M+H]⁺) calculated 189.1028, found 189.1029.



Yield: 71%, 20 mg, white solid.

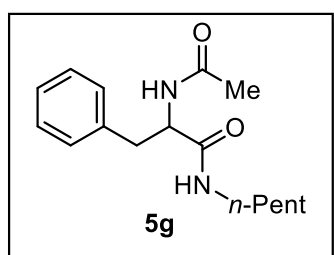
R_f = 0.42 (EA/Hex 2:3).

¹H NMR (400 MHz, DMSO-*d*₆) δ 10.07 (s, 1H), 8.27 (d, *J* = 8.2 Hz, 1H), 7.57 (d, *J* = 7.9 Hz, 2H), 7.34 – 7.24 (m, 6H), 7.21 – 7.16 (m, 1H), 7.05 (t, *J* = 7.3 Hz, 1H), 4.66 (td, *J* = 8.9, 5.3 Hz, 1H), 3.02 (dd, *J* = 13.7, 5.3 Hz, 1H), 2.84 (dd, *J* = 13.7, 9.4 Hz, 1H), 1.79 (s, 3H).

¹³C NMR (101 MHz, DMSO-*d*₆) δ 170.34, 169.26, 138.81, 137.72, 129.14, 128.68, 128.06, 126.33, 123.37, 119.34, 54.86, 37.76, 22.39.

IR (DMSO-*d*₆, cm⁻¹) ν 3446, 2249, 2123, 1767, 1665, 1548, 1222, 1052, 1024, 1006, 820, 757, 621.

HRMS (ASAP) *m/z* [C₁₇H₁₉N₂O₂]⁺ ([M+H]⁺) calculated 283.1447, found 283.1446.



Yield: 59%, 16 mg, yellow solid.

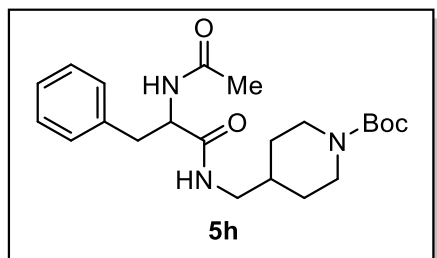
R_f = 0.29 (EA/Hex 1:1).

¹H NMR (400 MHz, CDCl₃) δ 7.30 – 7.24 (m, 2H), 7.27 – 7.16 (m, 3H), 6.70 (d, *J* = 8.1 Hz, 1H), 6.15 (t, *J* = 5.7 Hz, 1H), 4.64 (td, *J* = 8.3, 6.4 Hz, 1H), 3.24 – 3.11 (m, 1H), 3.10 – 2.94 (m, 3H), 1.96 (s, 3H), 1.40 – 1.18 (m, 4H), 1.23 – 1.06 (m, 2H), 0.85 (t, *J* = 7.3 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 170.96, 170.11, 137.01, 129.39, 128.67, 127.00, 54.93, 39.59, 39.03, 29.01, 28.99, 23.25, 22.38, 14.02.

IR (CDCl₃, cm⁻¹) ν 3277, 3085, 3030, 2956, 2929, 2859, 1638, 1552, 1497, 1372, 743, 698.

HRMS (ASAP) *m/z* [C₁₆H₂₅N₂O₂]⁺ ([M+H]⁺) calculated 277.1916, found 277.1913.



Yield: 74%, 30 mg, yellow oil.

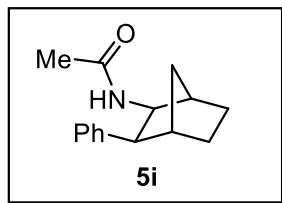
R_f = 0.22 (EA 100%).

¹H NMR (400 MHz, CDCl₃) δ 7.31 – 7.12 (m, 5H), 6.63 (d, *J* = 8.1 Hz, 1H), 6.43 (t, *J* = 5.6 Hz, 1H), 4.68 (td, *J* = 8.5, 6.3 Hz, 1H), 4.00 (s, 2H), 3.22 – 3.08 (m, 1H), 3.03 (dd, *J* = 13.4, 6.3 Hz, 1H), 2.96 (dd, *J* = 13.4, 8.7 Hz, 1H), 2.86 (s, 1H), 2.66 – 2.45 (m, 2H), 1.96 (s, 3H), 1.43 (s, 9H), 1.41 – 1.21 (m, 3H), 1.01 – 0.84 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 171.19, 170.17, 154.86, 136.81, 129.37, 128.72, 127.06, 79.52, 54.95, 44.91, 43.75, 38.98, 36.21, 29.64, 28.56, 23.26.

IR (CDCl₃, cm⁻¹) ν 3281, 2972, 2923, 2849, 1686, 1640, 1547, 1421, 1365, 1277, 1226, 1168, 1141, 734, 698.

HRMS (ESI) *m/z* [C₂₂H₃₃N₃NaO₄]⁺ ([M+Na]⁺) calculated 426.2369, found 426.2367.



Yield: 61%, 14 mg, white solid.

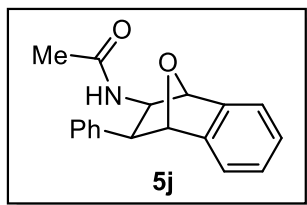
R_f = 0.42 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.45 – 7.35 (m, 2H), 7.32 – 7.22 (m, 3H), 4.79 (d, *J* = 8.7 Hz, 1H), 4.29 (t, *J* = 8.6 Hz, 1H), 3.19 (d, *J* = 8.6 Hz, 1H), 2.70 (s, 1H), 2.41 – 2.35 (m, 1H), 1.84 – 1.64 (m, 3H), 1.62 (s, 3H), 1.57 – 1.38 (m, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 169.17, 140.13, 128.54, 128.40, 126.47, 56.52, 52.12, 42.61, 40.28, 35.54, 30.09, 26.39, 23.24.

IR (CDCl₃, cm⁻¹) ν 3293, 3061, 2955, 2872, 1646, 1549, 1496, 1451, 1373, 1290, 1154, 905, 726, 700, 494.

HRMS (ASAP) *m/z* [C₁₅H₂₀NO]⁺ ([M+H]⁺) calculated 230.1545, found 230.1547.



Yield: 89%, 25 mg, white solid.

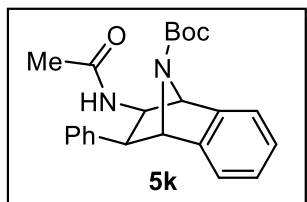
R_f = 0.42 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.47 – 7.26 (m, 7H), 7.30 – 7.20 (m, 2H), 5.56 (s, 1H), 5.31 – 5.23 (m, 2H), 4.49 (t, *J* = 8.5 Hz, 1H), 3.34 (d, *J* = 7.9 Hz, 1H), 1.58 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 169.86, 146.12, 143.36, 138.74, 128.93, 128.84, 127.64, 127.30, 127.27, 120.84, 119.13, 84.45, 84.30, 54.24, 50.12, 22.98.

IR (CDCl₃, cm⁻¹) ν 3428, 3308, 3040, 3006, 1656, 1495, 1371, 904, 727, 703, 648.

HRMS (ASAP) *m/z* [C₁₈H₁₈NO₂]⁺ ([M+H]⁺) calculated 280.1338, found 280.1337.



Yield: 55%, 21 mg, white solid.

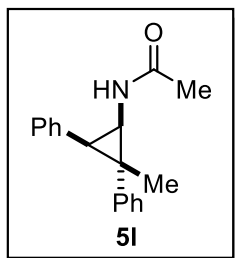
R_f = 0.26 (EA/Hex 2:3).

¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.35 (m, 1H), 7.35 – 7.28 (m, 2H), 7.28 – 7.20 (m, 4H), 7.19 – 7.13 (m, 2H), 5.26 (s, 1H), 5.18 (d, *J* = 9.4 Hz, 1H), 4.91 (s, 1H), 4.46 – 4.29 (m, 1H), 3.20 (d, *J* = 8.1 Hz, 1H), 1.51 (s, 3H), 1.34 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 169.62, 156.59, 146.57, 143.71, 138.95, 128.83, 128.68, 127.54, 127.29, 127.16, 121.79, 120.04, 99.74, 81.11, 67.68, 66.46, 53.64, 50.12, 28.44, 22.96. (Measured at 60 °C.)

IR (CDCl₃, cm⁻¹) ν 3315, 3028, 2979, 2927, 1698, 1667, 1367, 1336, 1278, 1255, 1158, 909, 729.

HRMS (ESI) *m/z* [C₂₃H₂₆N₂NaO₃]⁺ ([M+Na]⁺) calculated 401.1841, found 401.1851.



Yield: 43%, 11 mg, colorless oil.

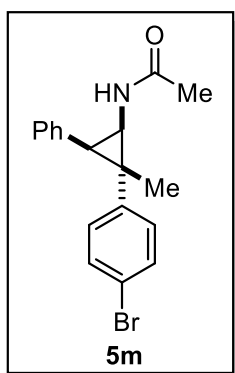
R_f = 0.37 (EA/Hex 2:3).

(major diastereomer) **¹H NMR** (400 MHz, CDCl₃) δ 7.64 – 7.58 (m, 2H), 7.48 – 7.42 (m, 2H), 7.42 – 7.32 (m, 5H), 7.32 – 7.25 (m, 1H), 5.37 – 5.32 (m, 1H), 3.50 (dd, *J* = 8.6, 5.0 Hz, 1H), 2.69 (dd, *J* = 8.6, 1.2 Hz, 1H), 2.02 (s, 3H), 1.27 (s, 3H).

(major diastereomer) **¹³C NMR** (101 MHz, CDCl₃) δ 171.48, 146.83, 134.92, 130.58, 129.25, 128.74, 128.57, 127.14, 126.65, 37.19, 30.92, 30.24, 23.36, 18.07.

IR (CDCl₃, cm⁻¹) ν 3303, 3056, 3024, 2927, 1651, 1601, 1495, 1444, 1370, 1268, 907, 763, 732, 700.

HRMS (ASAP) *m/z* [C₁₈H₂₀NO]⁺ ([M+H]⁺) calculated 266.1545, found 266.1544.



Yield: 38%, 13 mg, white solid.

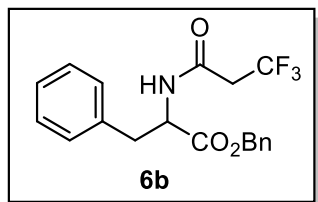
R_f = 0.32 (EA/Hex 2:3).

(major diastereomer) **¹H NMR** (400 MHz, CDCl₃) δ 7.54 – 7.42 (m, 6H), 7.39 – 7.30 (m, 3H), 5.34 (s, 1H), 3.41 (dd, *J* = 8.7, 4.7 Hz, 1H), 2.65 (dd, *J* = 8.7, 1.1 Hz, 1H), 2.01 (s, 3H), 1.23 (s, 3H).

(major diastereomer) **¹³C NMR** (101 MHz, CDCl₃) δ 171.57, 145.93, 134.53, 131.83, 130.53, 130.46, 129.37, 127.32, 120.50, 37.15, 30.69, 30.17, 23.32, 17.94.

IR (CDCl₃, cm⁻¹) ν 3302, 3054, 3026, 2931, 2979, 1653, 1492, 1444, 1369, 1090, 1009, 825, 734, 702, 532.

HRMS (ASAP) m/z [C₁₈H₁₉BrNO]⁺ ([M+H]⁺) calculated 344.0650, found 344.0644.



Yield: 73%, 27 mg, white solid.

R_f = 0.30 (EA/Hex 1:4).

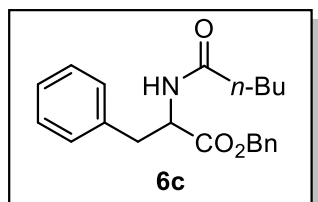
¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.35 (m, 3H), 7.34 – 7.28 (m, 2H), 7.24 – 7.17 (m, 3H), 7.01 – 6.92 (m, 2H), 6.19 (d, J = 7.6 Hz, 1H), 5.20 (d, J = 12.1 Hz, 1H), 5.14 (d, J = 12.1 Hz, 1H), 4.94 (dt, J = 7.7, 5.6 Hz, 1H), 3.24 – 3.09 (m, 2H), 3.04 (q, J = 10.5 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 170.94, 162.13 (q, J = 3.6 Hz), 135.23, 134.94, 129.44, 128.85, 128.84, 128.77, 127.42, 123.97 (q, J = 276.9 Hz), 67.74, 53.59, 41.69 (q, J = 29.7 Hz), 37.68. (Two aromatic peaks overlapped)

¹⁹F NMR (376 MHz, CDCl₃) δ -62.86 (t, J = 10.5 Hz)

IR (CDCl₃, cm⁻¹) ν 3313, 3066, 3032, 2957, 1740, 1664, 1542, 1388, 1261, 1240, 1213, 1191, 1136, 905, 733, 699.

HRMS (ASAP) m/z [C₁₉H₁₉F₃NO₃]⁺ ([M+H]⁺) calculated 366.1317, found 366.1319.



Yield: 53%, 18 mg, white solid.

R_f = 0.23 (EA/Hex 1:4).

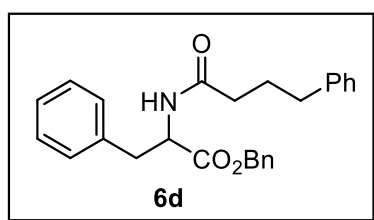
¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.30 (m, 3H), 7.33 – 7.27 (m, 2H), 7.24 – 7.19 (m, 3H), 7.03 – 6.96 (m, 2H), 5.87 (d, J = 7.8 Hz, 1H), 5.17 (d, J = 12.2 Hz, 1H), 5.11 (d, J = 12.1 Hz, 1H), 4.95 (dt, J = 7.9, 5.8

Hz, 1H), 3.19 – 3.04 (m, 2H), 2.20 – 2.12 (m, 2H), 1.62 – 1.50 (m, 2H), 1.35 – 1.24 (m, 2H), 0.88 (t, $J = 7.3$ Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 172.76, 171.72, 135.89, 135.20, 129.45, 128.75, 128.72, 128.68, 128.65, 127.17, 67.39, 53.04, 38.01, 36.40, 27.73, 22.42, 13.88.

IR (CDCl_3 , cm^{-1}) ν 3288, 3063, 3031, 2956, 2930, 2869, 1740, 1648, 1537, 1497, 1454, 1210, 1174, 742, 697.

HRMS (ASAP) m/z [$\text{C}_{21}\text{H}_{26}\text{NO}_3$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 340.1913, found 340.1908.



Yield: 54%, 22 mg, white solid.

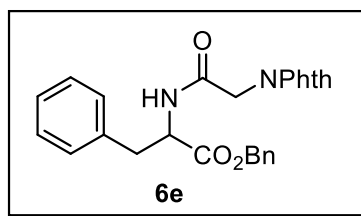
$R_f = 0.25$ (EA/Hex 1:4).

^1H NMR (400 MHz, CDCl_3) δ 7.42 – 7.08 (m, 13H), 7.04 – 6.95 (m, 2H), 5.86 (d, $J = 7.9$ Hz, 1H), 5.17 (d, $J = 12.1$ Hz, 1H), 5.11 (d, $J = 12.1$ Hz, 1H), 4.94 (dt, $J = 7.9, 5.9$ Hz, 1H), 3.19 – 3.03 (m, 2H), 2.59 (t, $J = 7.5$ Hz, 2H), 2.16 (dd, $J = 7.9, 6.6$ Hz, 2H), 1.92 (p, $J = 7.6$ Hz, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 172.37, 171.67, 141.50, 135.84, 135.17, 129.41, 128.74, 128.71, 128.67, 128.59, 128.49, 127.19, 126.06, 67.40, 53.07, 37.97, 35.70, 35.14, 27.01. (Two aromatic peaks overlapped)

IR (CDCl_3 , cm^{-1}) ν 3290, 3062, 3028, 2930, 2859, 1738, 1648, 1534, 1496, 1453, 1175, 907, 734, 696.

HRMS (ASAP) m/z [$\text{C}_{26}\text{H}_{28}\text{NO}_3$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 402.2069, found 402.2067.



Yield: 56%, 25 mg, white solid.

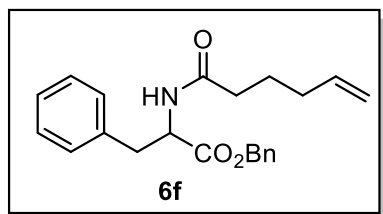
$R_f = 0.44$ (EA/Hex 2:3).

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.95 – 7.85 (m, 2H), 7.79 – 7.73 (m, 2H), 7.41 – 7.32 (m, 3H), 7.31 – 7.27 (m, 2H), 7.19 – 7.13 (m, 3H), 7.03 – 6.90 (m, 2H), 6.24 (d, $J = 7.6$ Hz, 1H), 5.18 (d, $J = 12.1$ Hz, 1H), 5.12 (d, $J = 12.0$ Hz, 1H), 4.91 (dt, $J = 7.8, 5.5$ Hz, 1H), 4.38 (d, $J = 16.1$ Hz, 1H), 4.30 (d, $J = 16.1$ Hz, 1H), 3.18 – 3.07 (m, 2H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 171.00, 167.76, 165.66, 135.39, 134.99, 134.38, 132.13, 129.58, 128.84, 128.80, 128.79, 128.68, 127.24, 123.80, 67.62, 53.53, 40.80, 37.69.

IR (CDCl_3 , cm^{-1}) ν 3346, 3066, 3033, 2947, 1774, 1716, 1419, 1393, 1192, 954, 904, 730, 716, 701.

HRMS (ASAP) m/z [$\text{C}_{26}\text{H}_{23}\text{N}_2\text{O}_5$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 443.1607, found 443.1608.



Yield: 61%, 21 mg, colorless oil.

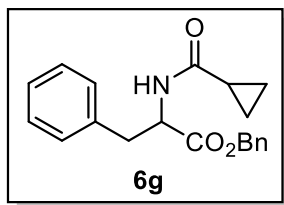
$R_f = 0.22$ (EA/Hex 1:4).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.42 – 7.33 (m, 3H), 7.33 – 7.28 (m, 2H), 7.24 – 7.19 (m, 3H), 7.03 – 6.96 (m, 2H), 5.86 (d, $J = 7.9$ Hz, 1H), 5.75 (ddt, $J = 17.0, 10.2, 6.7$ Hz, 1H), 5.18 (d, $J = 12.1$ Hz, 1H), 5.12 (d, $J = 12.1$ Hz, 1H), 5.02 – 4.92 (m, 3H), 3.20 – 3.05 (m, 2H), 2.19 – 2.15 (m, 2H), 2.07 – 2.02 (m, 2H), 1.76 – 1.66 (m, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 172.45, 171.68, 137.90, 135.86, 135.19, 129.44, 128.76, 128.73, 128.69, 128.67, 127.20, 115.48, 67.41, 53.06, 38.00, 35.76, 33.11, 24.64.

IR (CDCl_3 , cm^{-1}) ν 3293, 3064, 3031, 2931, 1742, 1648, 1539, 1498, 1454, 1175, 744, 698.

HRMS (ASAP) m/z [$\text{C}_{22}\text{H}_{26}\text{NO}_3$] $^+$ ($[\text{M}+\text{H}]^+$) calculated 352.1913, found 352.1911.



Yield: 53%, 17 mg, white solid.

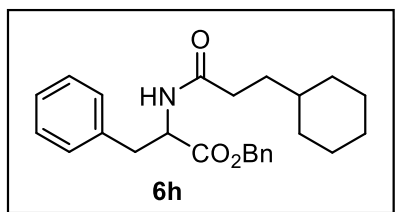
R_f = 0.27 (EA/Hex 1:4).

¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.34 (m, 3H), 7.32 – 7.28 (m, 2H), 7.25 – 7.19 (m, 3H), 7.07 – 6.98 (m, 2H), 6.10 (d, *J* = 7.8 Hz, 1H), 5.18 (d, *J* = 12.2 Hz, 1H), 5.12 (d, *J* = 12.1 Hz, 1H), 4.95 (dt, *J* = 7.9, 5.8 Hz, 1H), 3.18 – 3.07 (m, 2H), 1.36 (tt, *J* = 7.9, 4.6 Hz, 1H), 1.02 – 0.91 (m, 2H), 0.79 – 0.68 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 173.24, 171.74, 135.92, 135.21, 129.51, 128.74, 128.71, 128.66, 128.64, 127.15, 67.35, 53.34, 38.10, 14.75, 7.52, 7.45.

IR (CDCl₃, cm⁻¹) ν 3302, 3063, 3029, 3006, 2941, 2838, 1739, 1650, 1599, 1498, 1455, 1203, 1150, 1066, 732, 699.

HRMS (ASAP) *m/z* [C₂₀H₂₂NO₃]⁺ ([M+H]⁺) calculated 324.1600, found 324.1601.



Yield: 57%, 22 mg, colorless oil.

R_f = 0.33 (EA/Hex 1:4).

¹H NMR (400 MHz, CDCl₃) δ 7.44 – 7.31 (m, 3H), 7.34 – 7.28 (m, 2H), 7.28 – 7.18 (m, 3H), 7.04 – 6.95 (m, 2H), 5.86 (d, *J* = 7.8 Hz, 1H), 5.18 (d, *J* = 12.1 Hz, 1H), 5.12 (d, *J* = 12.1 Hz, 1H), 4.95 (dt, *J* = 7.9, 5.8 Hz, 1H), 3.19 – 3.05 (m, 2H), 2.24 – 2.12 (m, 2H), 1.74 – 1.59 (m, 5H), 1.54 – 1.42 (m, 2H), 1.28 – 1.05 (m, 4H), 0.94 – 0.79 (m, 2H).

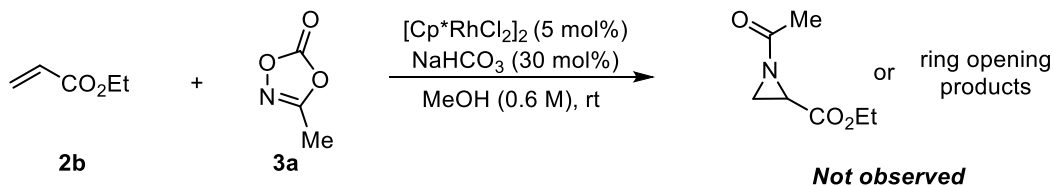
¹³C NMR (101 MHz, CDCl₃) δ 173.03, 171.73, 135.89, 135.21, 129.46, 128.76, 128.73, 128.68, 128.64, 127.18, 67.39, 53.05, 38.00, 37.34, 34.16, 33.17, 33.11, 33.03, 26.65, 26.33.

IR (CDCl₃, cm⁻¹) ν 3298, 3070, 3029, 2921, 2850, 1740, 1648, 1536, 1498, 1451, 1174, 905, 730, 698.

HRMS (ASAP) m/z [C₂₅H₃₂NO₃]⁺ ([M+H]⁺) calculated 394.2387, found 394.2384.

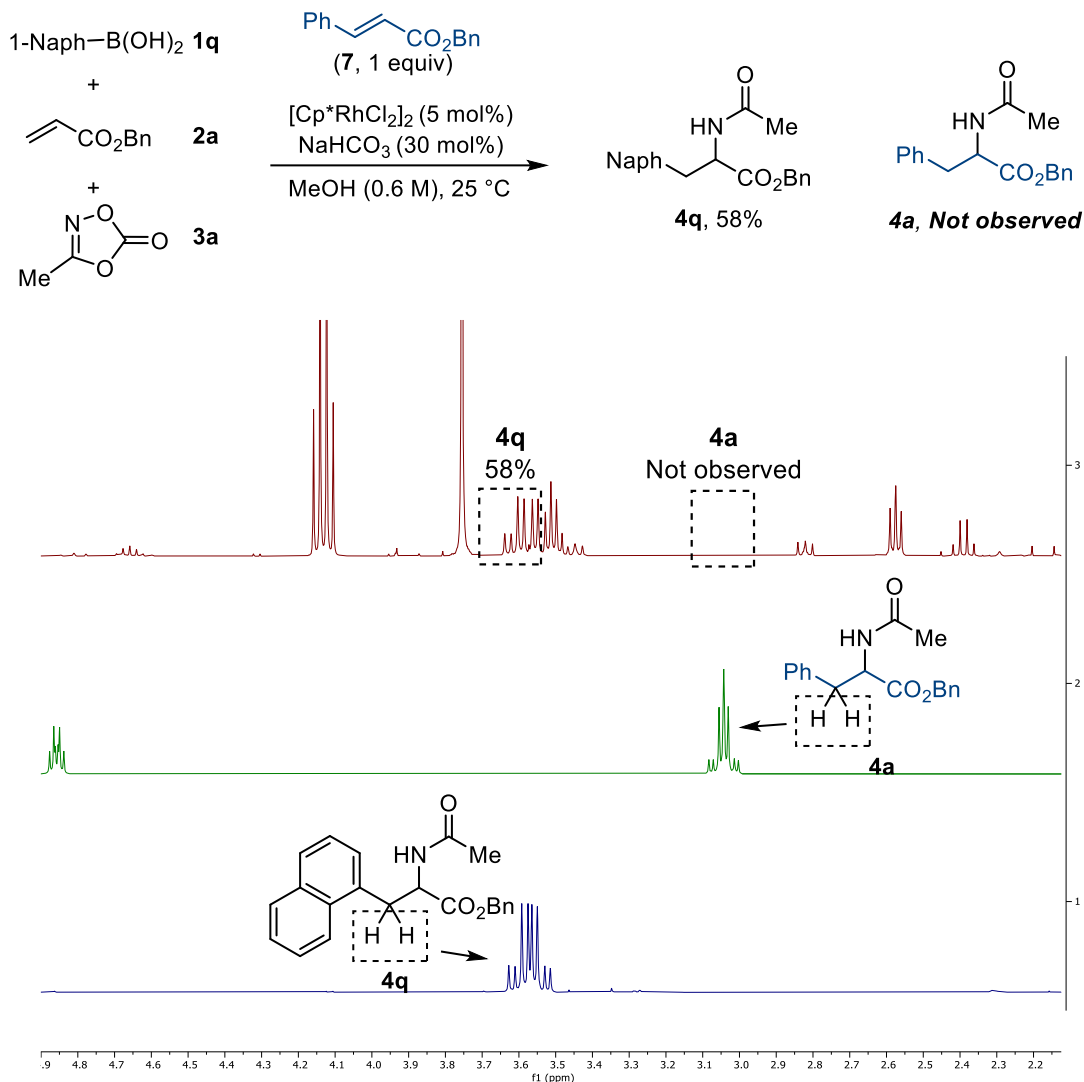
6. Mechanistic studies

Possible aziridine intermediate



$[\text{Cp}^*\text{RhCl}_2]_2$ (0.0050 mmol, 5 mol%) and sodium bicarbonate (0.03 mmol, 30 mol%) were measured in a 0.5-dram vial with a magnetic stir bar. In a separate vial, dioxazolone **3a** (0.1 mmol, 1 equiv) and ethyl acrylate **2b** (0.3 mmol, 3 equiv) were dissolved in 0.10 mL of MeOH and transferred to the first vial. It was rinsed with an additional 0.07 mL of MeOH and transferred again to the first vial. The reaction mixture was then stirred at room temperature for 16 hours. After the completion of the reaction, the solvent was removed by rotary evaporator. Ethyl acetate was added to the residue and filtered through a short pad of celite and washed with ethyl acetate three times. The filtrate was concentrated by rotary evaporator and a crude ^1H NMR spectrum was collected with 1,3,5-trimethoxybenzene as the internal standard. As a result, neither aziridine nor ring-opening products by methanol were observed. This result suggests Rh(III) -catalyzed aziridination and subsequent ring-opening by phenylboronic acid is unlikely for the mechanism of the carboamination.

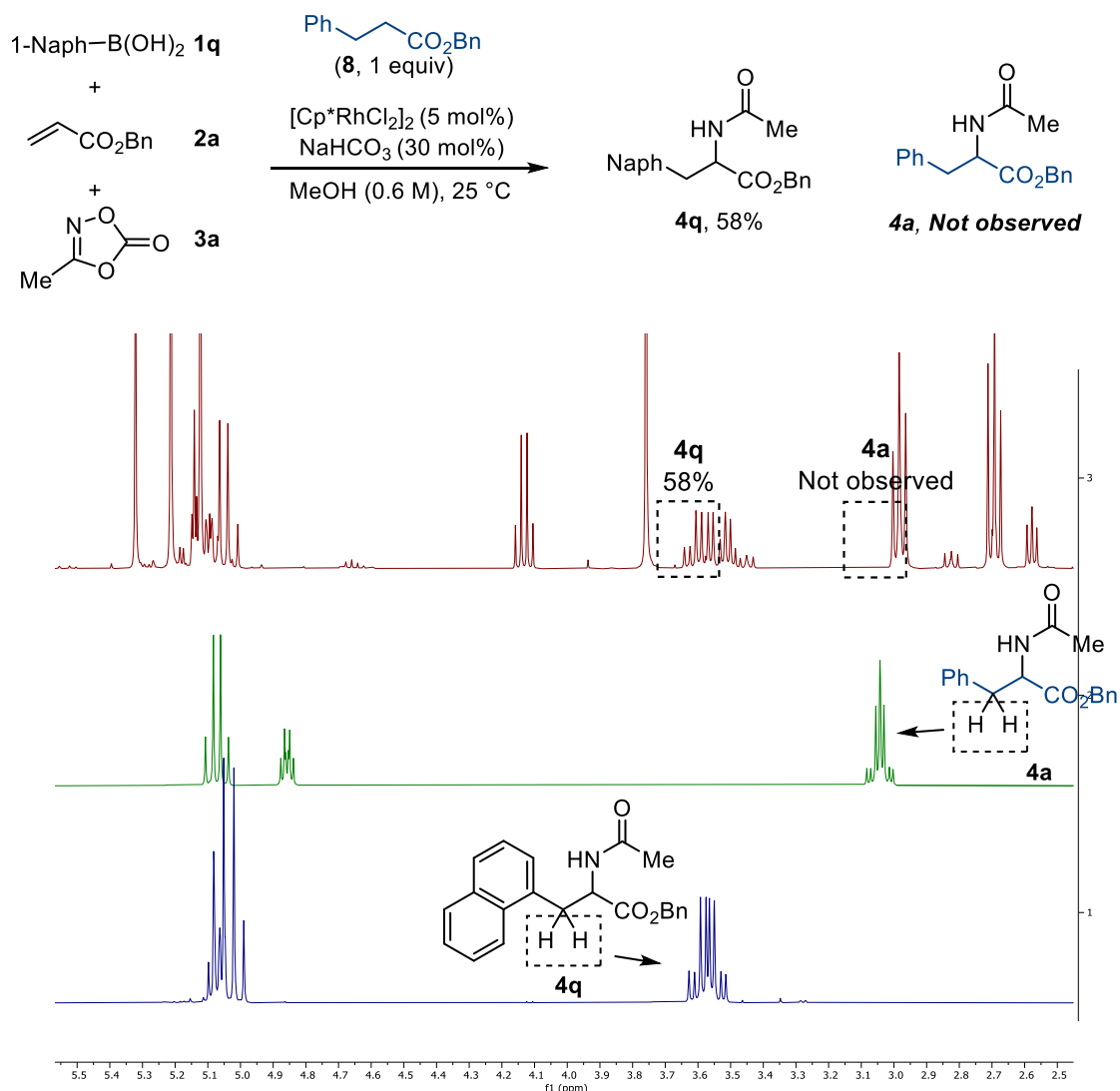
Reversibility of β -hydride elimination



Benzyl cinnamate **7** was prepared according to literature procedures.⁹ $[\text{Cp}^*\text{RhCl}_2]_2$ (0.0050 mmol, 5 mol%) and sodium bicarbonate (0.03 mmol, 30 mol%), and naphthalene-1-boronic acid **1q** (0.25 mmol, 2.5 equiv) were measured in a 0.5-dram vial with a magnetic stir bar. In a separate vial, benzyl cinnamate **7** (0.1 mmol, 1 equiv) was dissolved 0.10 mL of MeOH and transferred to the first vial. It was rinsed with an additional 0.07 mL of MeOH and transferred again to the first vial. Sequentially benzyl acrylate **2a** (0.3 mmol, 3 equiv) and dioxazolone **3a** (0.1 mmol, 1 equiv) were added to the reaction mixture using a micropipette. The reaction mixture was then stirred at room temperature for 16 hours. After the completion of the reaction, the solvent was removed by a rotary evaporator. Ethyl acetate was added to the residue and filtered through a short pad of celite and washed with ethyl acetate three times. The filtrate was concentrated by rotary

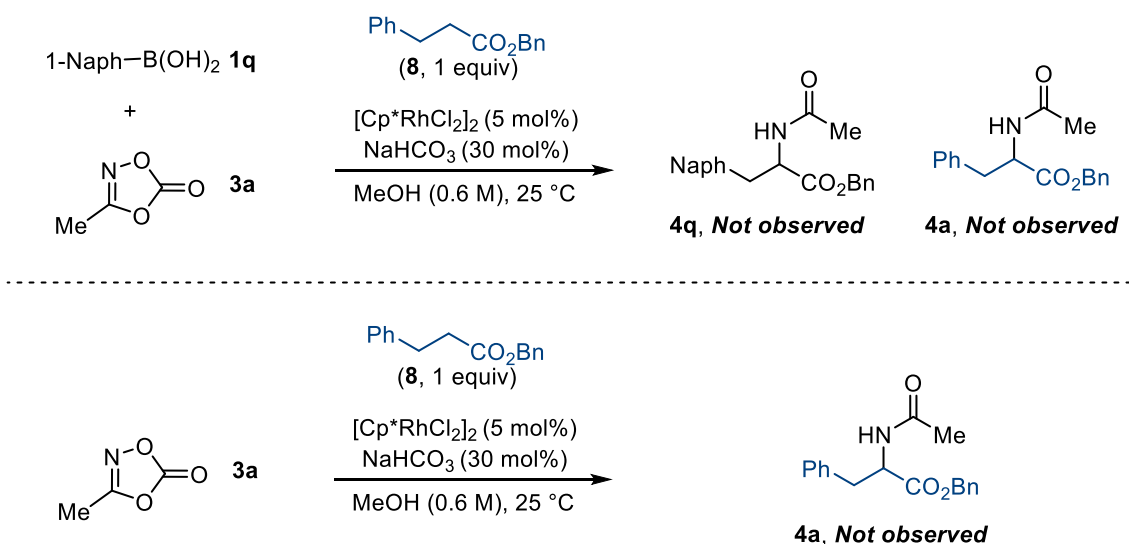
evaporator and a crude ^1H NMR spectrum was collected with 1,3,5-trimethoxybenzene as the internal standard).

Reversibility of proto-demetalation step



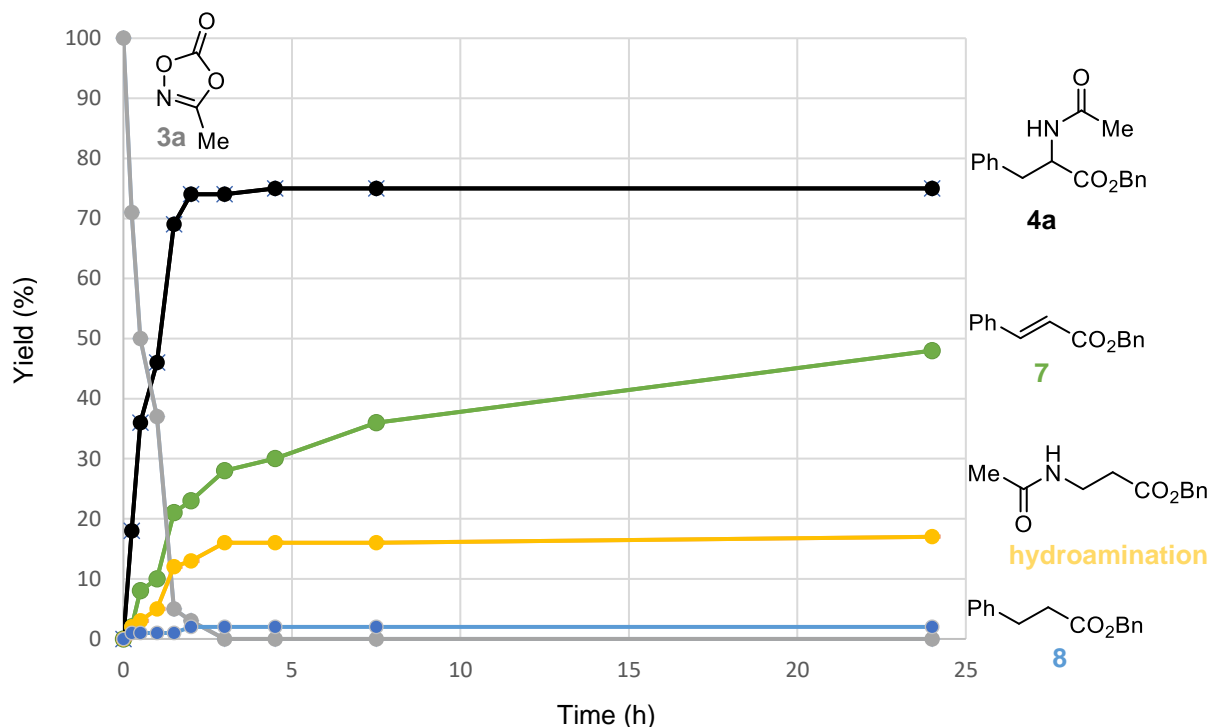
Benzyl 3-phenylpropanoate **8** was prepared according to literature procedures.¹⁰ [Cp*RhCl₂]₂ (0.0050 mmol, 5 mol%) and sodium bicarbonate (0.03 mmol, 30 mol%), and naphthalene-1-boronic acid **1q** (0.25 mmol, 2.5 equiv) were measured in a 0.5-dram vial with a magnetic stir bar. In a separate vial, Benzyl 3-phenylpropanoate **8** (0.1 mmol, 1 equiv) were dissolved 0.10 mL of MeOH and transferred to the first vial. It was rinsed with an additional 0.7 mL of MeOH and transferred again to the first vial. Sequentially benzyl acrylate **2a** (0.3 mmol, 3 equiv) and dioxazolone **3a** (0.1 mmol, 1 equiv) were added to the reaction mixture

using a micropipette. The reaction mixture was then stirred at room temperature for 16 hours. After the completion of the reaction, the solvent was removed by a rotary evaporator. Ethyl acetate was added to the residue and filtered through a short pad of celite and washed with ethyl acetate three times. The filtrate was concentrated by rotary evaporator and a crude ^1H NMR spectrum was collected with 1,3,5-trimethoxybenzene as the internal standard. While both reactions give the desired product (**4q**) from 1-naphthyl boronic acid (**2q**) in good yield, carboamination products (**4a**) from these side products (**7** or **8**) are not observed. This experiment suggests **7** and **8** are off-cycle side products that do not re-enter the catalytic cycle.



The same experiments were conducted without benzyl acrylate (**2a**) and without both naphthalene-1-boronic acid (**1q**) and benzyl acrylate (**2a**). As a result, carboamination products are not observed after the reaction, which again supports benzyl 3-phenylpropanoate (**8**) as an off-cycle side product.

Reaction progress monitoring



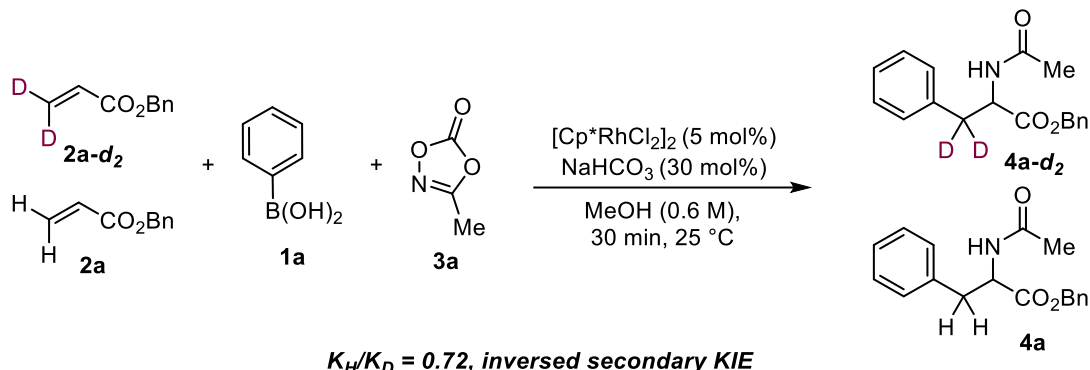
[Cp* RhCl_2] $_2$ (0.0050 mmol, 5 mol%) and sodium bicarbonate (0.03 mmol, 30 mol%), and phenylboronic acid **1a** (0.25 mmol, 2.5 equiv) were measured in a 0.5-dram vial with a magnetic stir bar. In a separate vial, benzyl acrylate **2a** (0.3 mmol, 3 equiv) and dioxazolone **3a** (0.1 mmol, 1 equiv) were dissolved in 0.10 mL of MeOH. It was rinsed with an additional 0.07 mL of MeOH and transferred again to the first vial. 10 separate reactions were set up and each time the solvent was removed by rotary evaporator and ethyl acetate was added to the residue and filtered through a short pad of celite and washed with ethyl acetate three times. The filtrate was concentrated by rotary evaporator and a crude ^1H NMR spectrum was collected with 1,3,5-trimethoxybenzene as the internal standard.

Synthesis of benzyl acrylate-3,3- d_2 (**2a-d₂**)

Benzyl acrylate-3,3- d_2 (**2a-d₂**) was synthesized following literature¹¹ using (benzyloxycarbonylmethyl)triphenylphosphonium bromide and paraformaldehyde- d_2 . To a solution of (benzyloxycarbonylmethyl)triphenylphosphonium bromide (10 mmol, 1 equiv) in a 2:1 mixture of Et $_2$ O/H $_2$ O (40 mL/20 mL), K $_2$ CO $_3$ was added portionwise over 15 minutes. After stirring 24 hours at 35 $^\circ\text{C}$, the layers were separated, and the water layer was extracted with Et $_2$ O (10 mL) three times. The combined

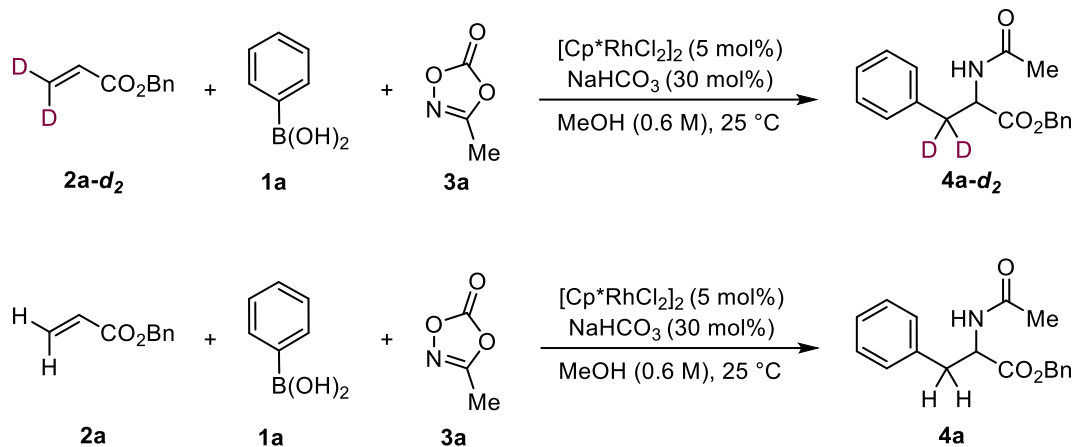
organic layer was dried with MgSO₄ and filtered. To this solution, paraformaldehyde-*d*₂ (10.5 mmol, 1.05 equiv) was added and refluxed for 24 hours. After careful removal of the solvent, the crude material was purified by column chromatography using pentane and Et₂O. Yield: 48%, colorless oil. ¹H NMR (500 MHz, CDCl₃) δ 7.44 – 7.30 (m, 5H), 6.18 (bs, 1H), 5.22 (s, 2H).

Intermolecular competition KIE



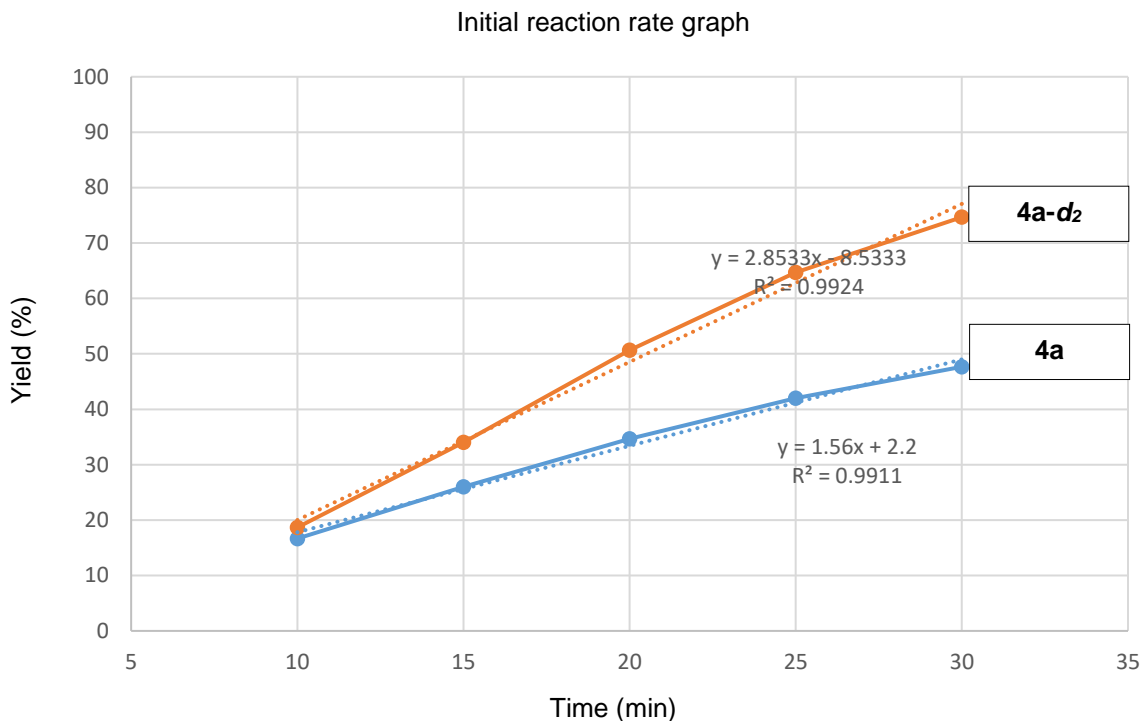
[Cp*RhCl₂]₂ (0.010 mmol, 5 mol%), sodium bicarbonate (0.06 mmol, 30 mol%), and phenylboronic acid **1a** (0.50 mmol, 2.5 equiv) were measured in a 1-dram vial with a magnetic stir bar. In a separate vial, **2a** (0.15 mmol, 1.5 equiv) and **2a-d**₂ (0.15 mmol, 1.5 equiv) were dissolved in 0.20 mL of MeOH containing 1,3,5-trimethoxybenzene as standard and transferred to the first vial. It was rinsed with an additional 0.13 mL of MeOH and transferred again to the first vial. After the addition of dioxazolone **3a** (0.20 mmol, 1 equiv), the reaction mixture was stirred at room temperature for 30 min. 20 uL aliquot was taken and diluted with 0.5 mL of MeCN-*d*₃ then analyzed by ¹H NMR. The KIE value was calculated based on the area of the 4.92 ppm (contribution from both molecules, **4a** and **4a-d**₂) and at 3.12 ppm (contribution solely from **4a**). The experiment was repeated three times and an average KIE value of 0.72 indicates sp² → sp³ hybridization change during the turnover limiting step, which suggests alkene migratory insertion as the turnover limiting step of the reaction.

Intermolecular parallel KIE



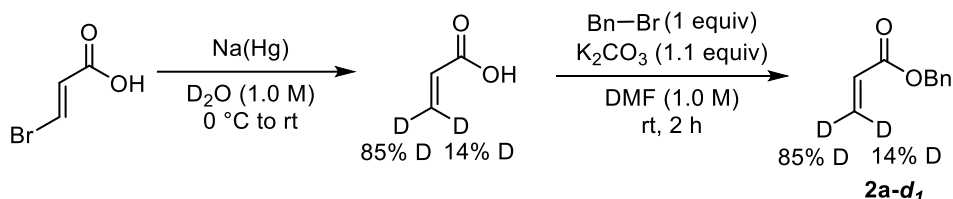
$$K_H/K_D = 0.55, \text{ inverted secondary KIE}$$

KIE was measured from two separated reactions, one with benzyl acrylate (**2a**) containing two C-H bonds and benzyl acrylate-3,3- d_2 (**2a-d₂**) containing two C-D bonds. $[\text{Cp}^*\text{RhCl}_2]_2$ (0.010 mmol, 5 mol%), sodium bicarbonate (0.06 mmol, 30 mol%), and phenylboronic acid **1a** (0.50 mmol, 2.5 equiv) were measured in a 1-dram vial with a magnetic stir bar. In a separate vial, **2a** (or **2a-d₂**) (0.60 mmol, 3 equiv) were dissolved in 0.20 mL of MeOH containing 1,3,5-trimethoxybenzene as standard and transferred to the first vial. It was rinsed with an additional 0.13 mL of MeOH and transferred again to the first vial. Dioxazolone **3a** (0.20 mmol, 1 equiv) was added to the reaction mixture using a micropipette and the reaction mixture was stirred at room temperature. To monitor the reaction progress, 20 μL aliquot was taken every 5 min and diluted with 0.5 mL of MeCN-d_3 . KIE value was calculated by comparing the initial reaction rate (slope) of each reaction (repeated 3 times each) by using ^1H NMR. Observed inverted secondary KIE (K_H/K_D) value of 0.55 suggests $\text{sp}^2 \rightarrow \text{sp}^3$ hybridization change occurs during the turnover limiting step.



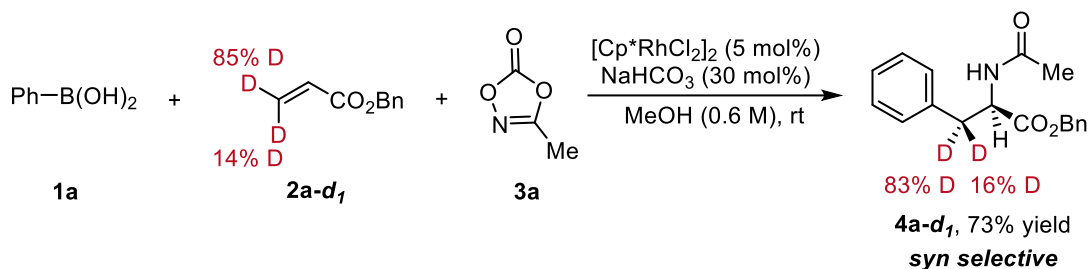
Syn addition of alkenes

Synthesis of benzyl acrylate-3-d₁ (2a-d₁)

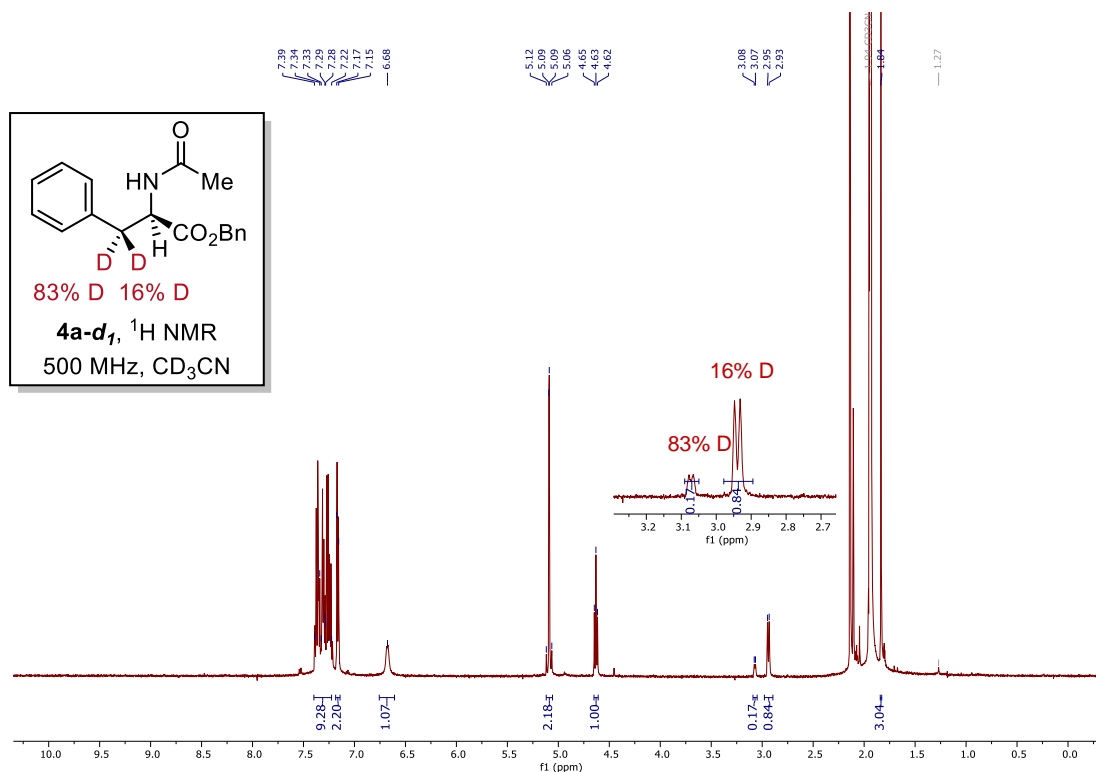


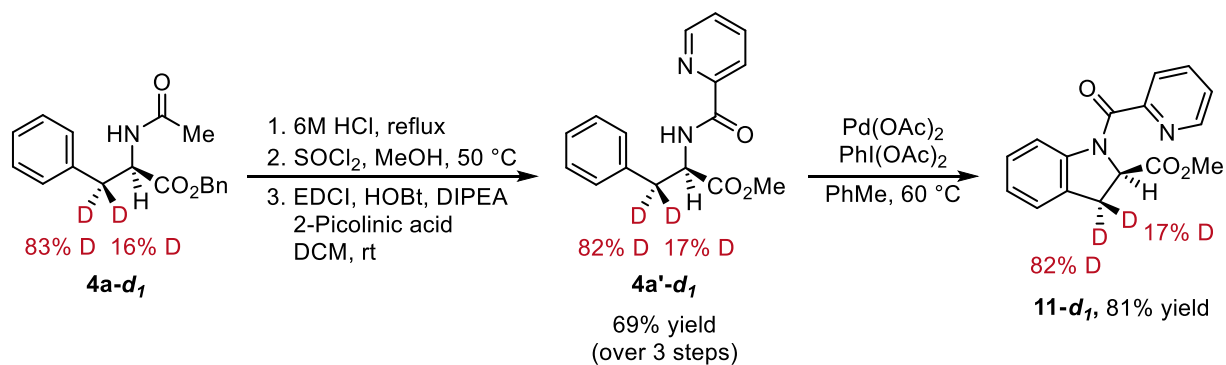
(*E*)-3-bromoacrylic acid was synthesized following literature.¹² To a solution of (*E*)-3-bromoacrylic acid (1.0 g, 6.62 mmol) in D₂O (1.0 M), 5.0 g of 20% sodium amalgam was added as one portion at 0 °C. After stirring 5 min at 0 °C for 5 min, the reaction mixture was warmed to room temperature and stirred additional 30 min. The reaction mixture was decanted and washed with water. The reaction mixture was acidified to pH ≈ 1 with 6 M HCl, then extracted with diethyl ether 3 times. The combined organic layer was washed with brine and dried over MgSO₄ and solvent was removed by a rotary evaporator. (*E*)-acrylic-3-d acid was used without further purification (around 20% of over reduction product was observed). To a mixture of K₂CO₃ (1.1 equiv) and benzyl bromide (1 equiv) in DMF (1.0 M), (*E*)-acrylic-3-d acid was added dropwise at room temperature. After stirring for two hours at room temperature, the reaction mixture was diluted

with diethyl ether and washed with water and brine. The organic layer was dried with MgSO_4 and concentrated. The crude material was purified by column chromatography using pentane and diethyl ether as eluent. **2a-d₁**. Yield: 20% (over two steps). Colorless oil. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.44 – 7.30 (m, 5H), 6.44 (d, $J = 17.3$ Hz, 0.86H), 6.17 (dt, $J = 17.2, 1.5$ Hz, 1H), 5.84 (d, $J = 10.5$ Hz, 0.15H).

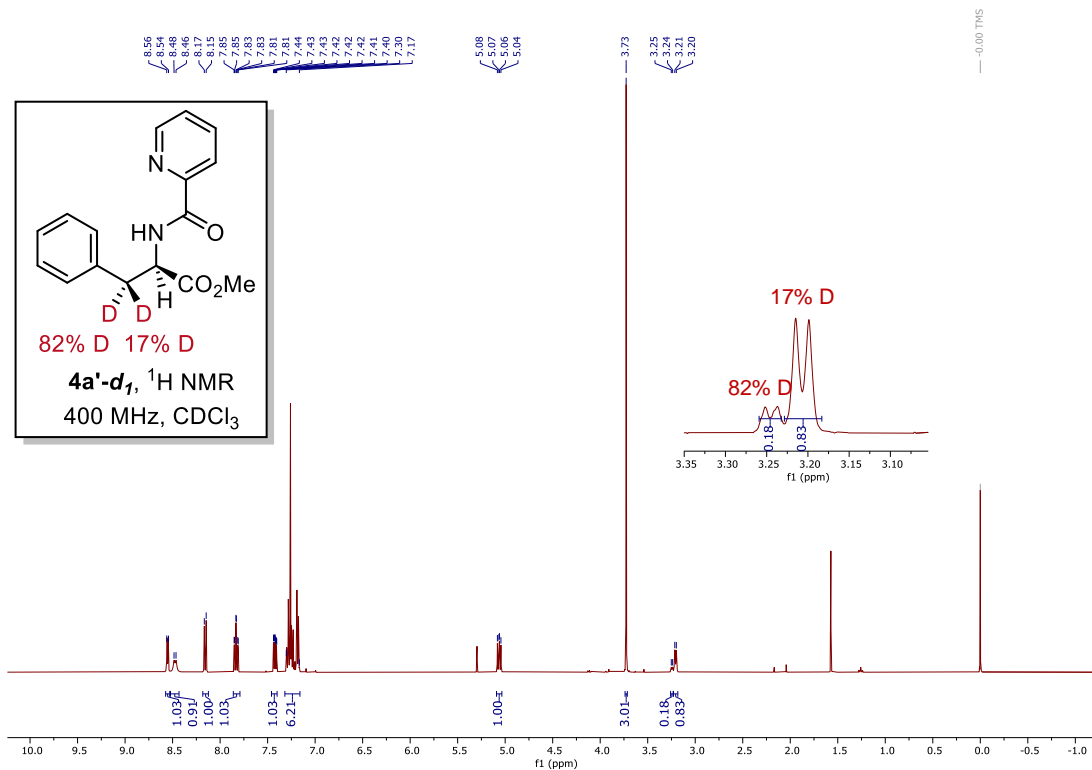


To a solution of $[\text{Cp}^*\text{RhCl}_2]_2$ (0.05 mmol, 5 mol%), sodium bicarbonate (0.30 mmol, 30 mol%), and phenylboronic acid **1a** (2.5 mmol, 2.5 equiv) in MeOH (1.7 mL, 0.6 M), **2a-d₁** (3.0 mmol, 3.0 equiv) and **3a** (1.0 mmol, 1 equiv) were added at room temperature and stirred overnight. The volatiles were removed by rotary evaporator and ethyl acetate was added and filtered through short pad of celite. After removing solvent using rotary evaporator, the crude material was purified by column chromatography using ethyl acetate and hexane as eluent to obtain **4a-d₁**. Yield: 73%, white solid. $^1\text{H NMR}$ (500 MHz, CD_3CN) δ 7.40 – 7.22 (m, 8H), 7.18 – 7.14 (m, 2H), 6.67 (s, 1H), 5.12 – 5.06 (m, 2H), 4.63 (t, $J = 7.8$ Hz, 1H), 3.07 (d, $J = 6.1$ Hz, 0.17H), 2.94 (d, $J = 8.0$ Hz, 0.84H), 1.84 (s, 3H).

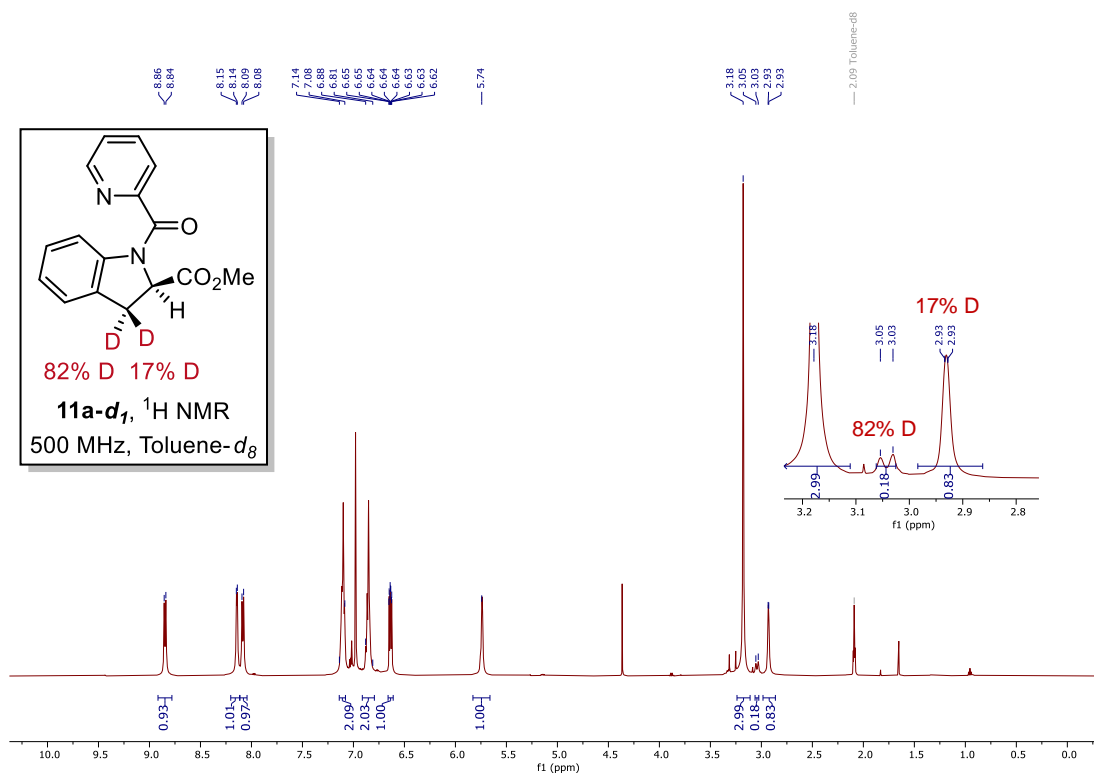




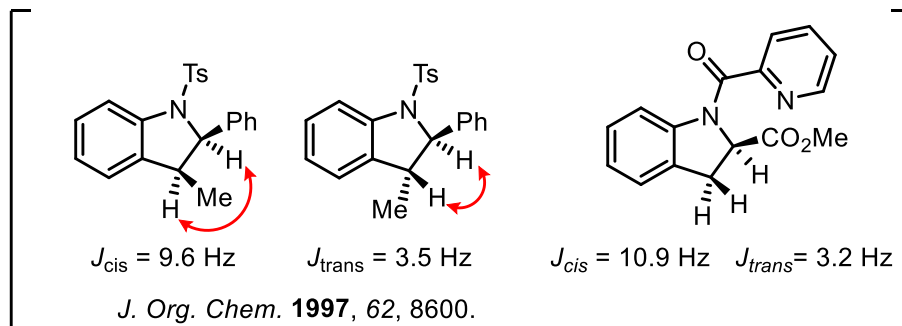
6 M HCl (0.1 M) was added to **4a-d₁** (1.38 mmol) and refluxed for 4 hours. After removing volatiles, diethyl ether was added, and the white solid was collected by filtration. The resulting solid was washed with diethyl ether and dried under high vacuum. The deprotected product was dissolved in MeOH (0.25 M) and SOCl₂ (1.5 equiv) was added dropwise at 0 °C. The reaction mixture was warmed to room temperature and heated at 50 °C for 3 hours. The volatiles were removed by a rotary evaporator and diethyl ether was added. After stirring for 10 min at 0 °C, the white solid was filtered and washed with diethyl ether to give crude deuterated phenylalanine methyl ester hydrochloride. The crude material (1 equiv) and 2-picolinic acid (1.1 equiv) were dissolved in DCM (0.2 M) and HOBt hydrate (1.1 equiv) and DIPEA (3.0 equiv) were added at room temperature. The reaction mixture was cooled to 0 °C and EDCI (1.1 equiv) was added. After stirring for 5 min, the reaction mixture was warmed to room temperature and stirred overnight. Water was added and extracted with DCM three times. The combined organic layer was washed with water and brine and dried with Na₂SO₄. After removing the solvent, the crude material was purified by column chromatography using ethyl acetate and hexane as eluent to obtain **4a'-d₁**. Yield: 69% (over three steps). White solid. ¹H NMR (400 MHz, CDCl₃) δ 8.55 (ddd, *J* = 4.8, 1.8, 0.9 Hz, 1H), 8.47 (d, *J* = 8.4 Hz, 1H), 8.16 (dt, *J* = 7.8, 1.1 Hz, 1H), 7.83 (td, *J* = 7.7, 1.7 Hz, 1H), 7.42 (ddd, *J* = 7.6, 4.8, 1.3 Hz, 1H), 7.33 – 7.14 (m, 5H), 5.06 (dd, *J* = 8.4, 6.3 Hz, 1H), 3.73 (s, 3H), 3.24 (d, *J* = 5.8 Hz, 0.18H), 3.21 (d, *J* = 6.5 Hz, 0.83H).

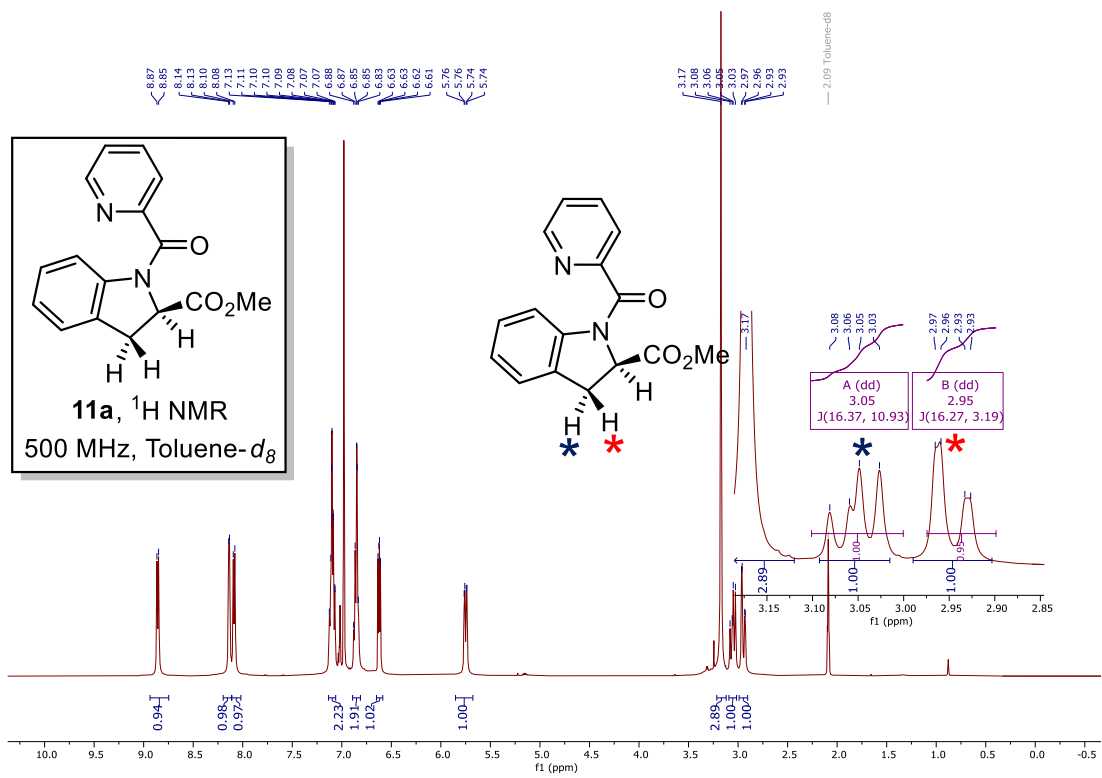


9a-d₁ was prepared following the literature.¹³ In a 20 mL vial, Pd(OAc)₂ (2 mol%, 0.018 mmol) and Phi(OAc)₂ (2 equiv. 1.78 mmol) were measured in the glovebox. In a separate flask, **4a'-d₁** (1 equiv, 0.89 mmol) was dissolved in toluene (0.125 M) and transferred to the first vial. After purging with Ar for 10 min, the vial was sealed and heated at 60 °C for 24 hours. The reaction mixture was concentrated and purified by column chromatography using ethyl acetate and hexane as eluent. **11a-d₁**. Yield: 81%, white solid. ¹H NMR (500 MHz, toluene-*d*₈) δ 8.85 (d, *J* = 8.2 Hz, 1H), 8.14 (d, *J* = 4.8 Hz, 1H), 8.09 (d, *J* = 7.9 Hz, 1H), 7.15 – 7.16 (m, 2H), 6.89 – 6.81 (m, 2H), 7.04 – 6.96 (m, 2H), 6.64 (ddd, *J* = 7.6, 4.7, 1.2 Hz, 1H), 5.77 – 5.71 (m, 1H), 3.18 (s, 3H), 3.04 (d, *J* = 11.8 Hz, 0.18H), 2.93 (s, 0.83H).



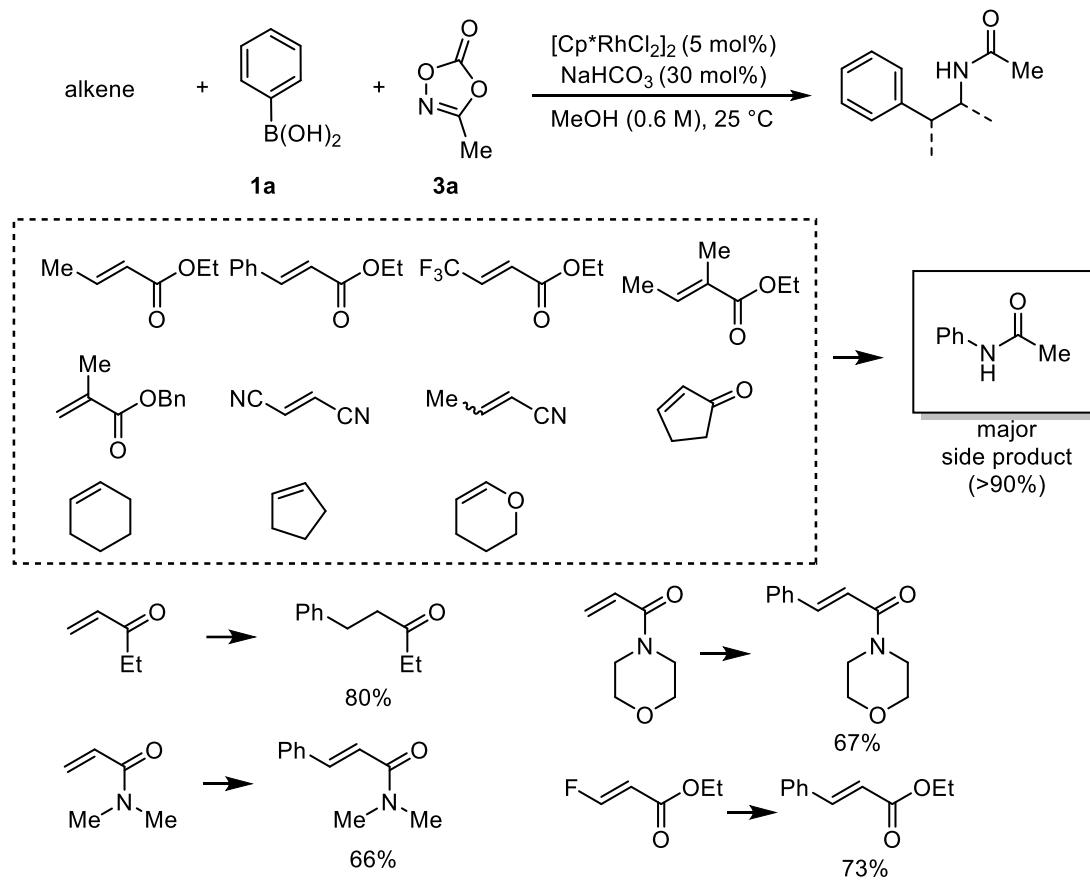
11a was synthesized from phenylalanine using same procedure described above. **11a**. White solid. ¹H NMR (500 MHz, toluene-*d*₈) δ 8.86 (d, $J = 8.2$ Hz, 1H), 8.14 (d, $J = 4.5$ Hz, 1H), 8.09 (d, $J = 7.9$ Hz, 1H), 7.09 (td, $J = 9.6, 7.8, 6.0$ Hz, 2H), 6.89 – 6.81 (m, 2H), 6.62 (dd, $J = 7.7, 4.8$ Hz, 1H), 5.75 (dd, $J = 10.9, 3.2$ Hz, 1H), 3.17 (s, 3H), 3.05 (dd, $J = 16.4, 10.9$ Hz, 1H), 2.95 (dd, $J = 16.3, 3.2$ Hz, 1H). The relative stereochemistry was assigned based on the literature.¹⁴ Benzylic proton with 10.9 Hz coupling constant was assigned as *cis* proton and proton with 3.2 Hz was assigned as *trans* proton which confirms the syn-carboamination of the reaction.



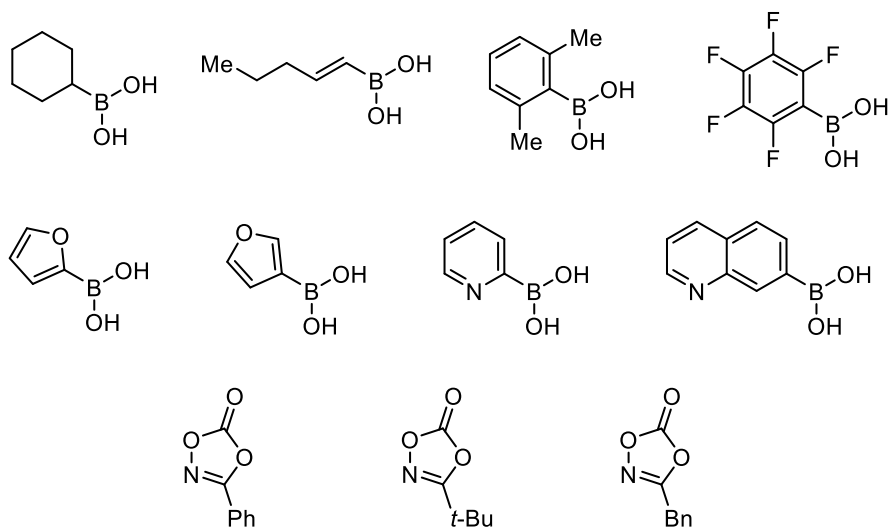


7. Limitation

<Unsuccessful alkene coupling partners>



<Unsuccessful boronic acids and dioxazolones>

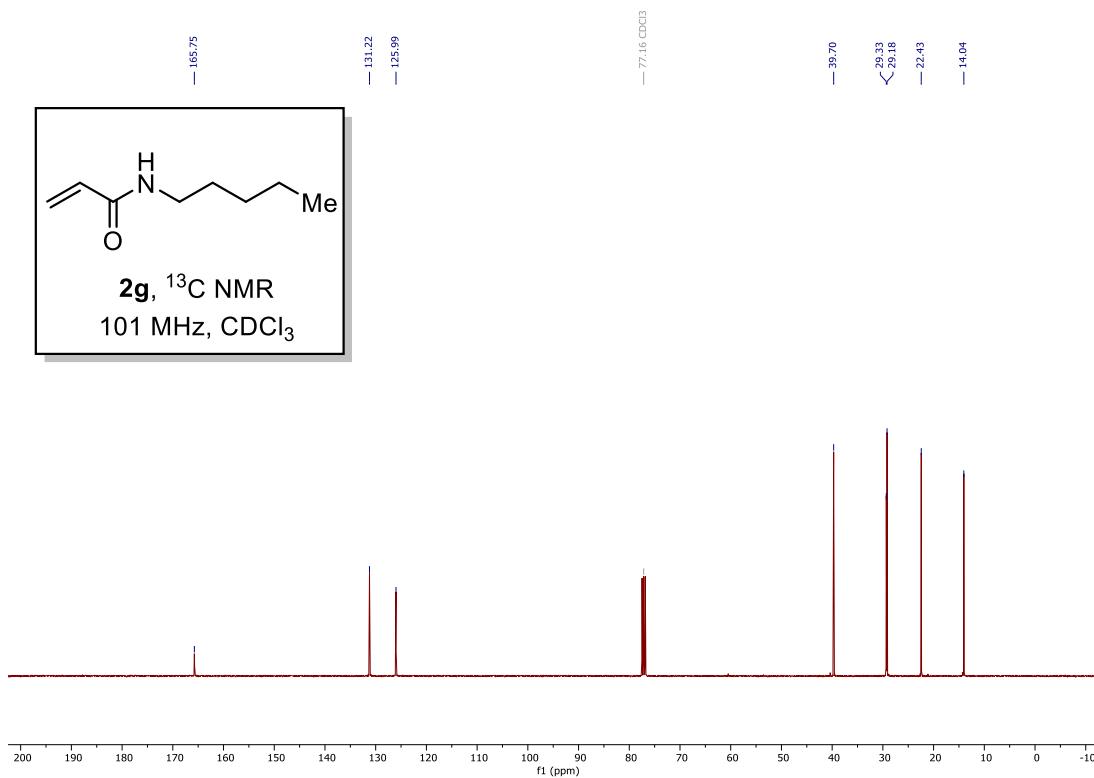
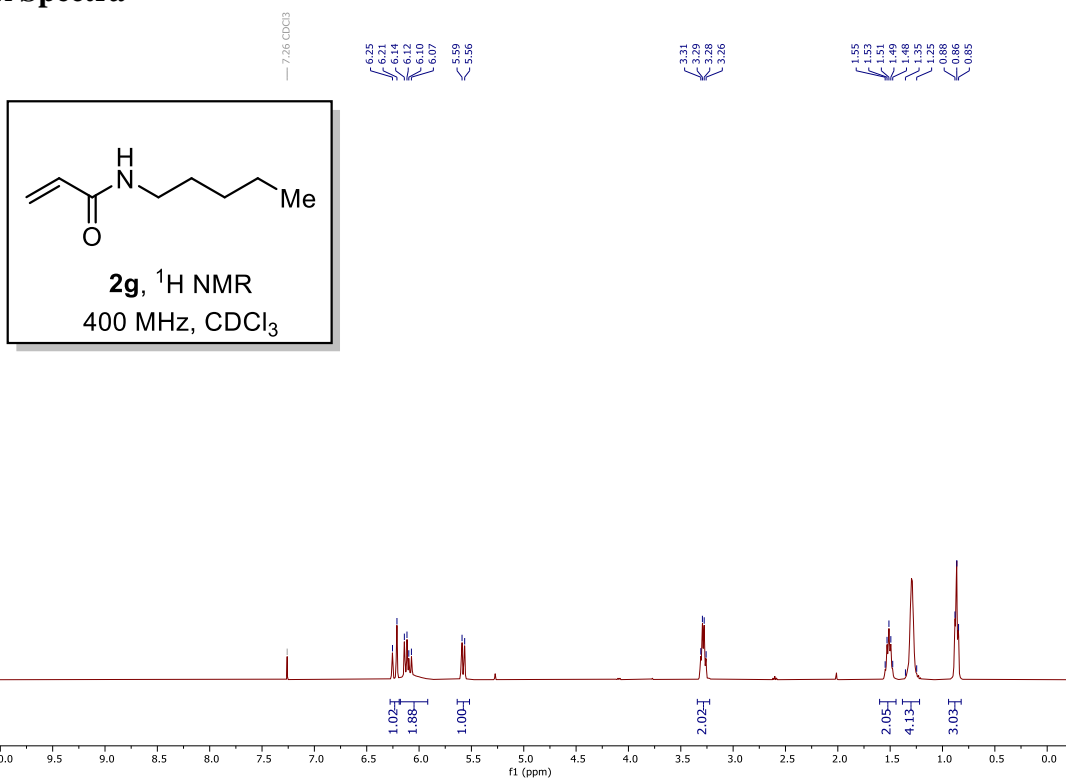


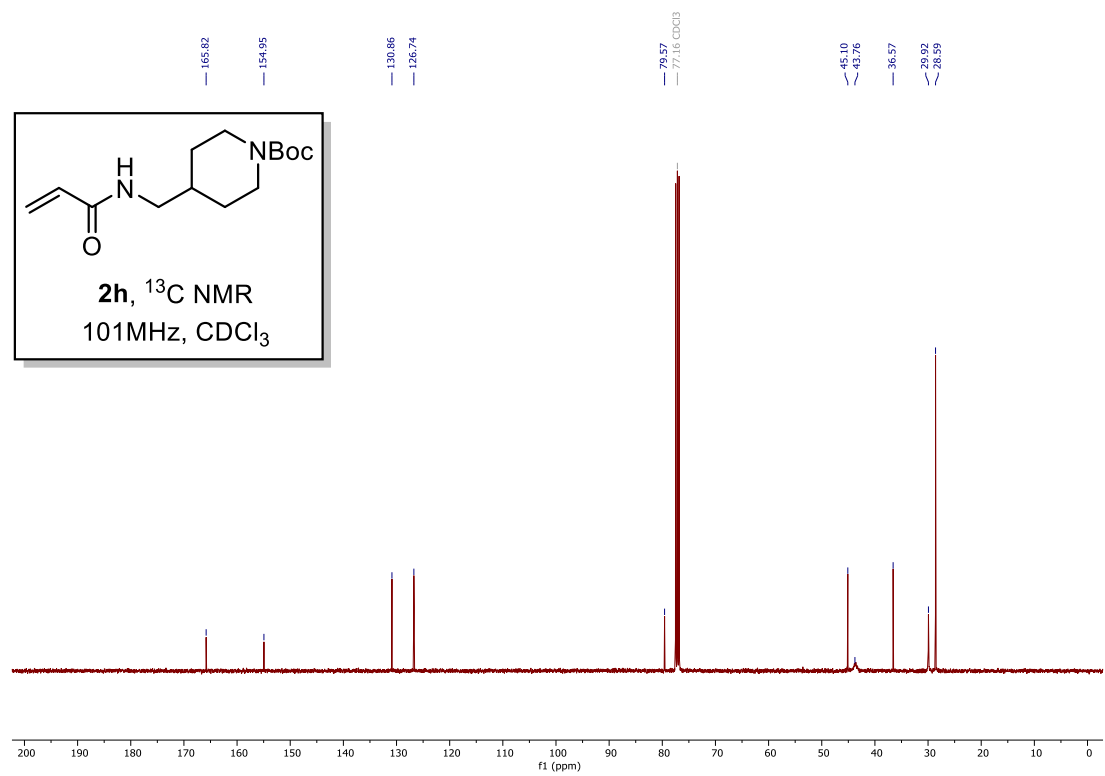
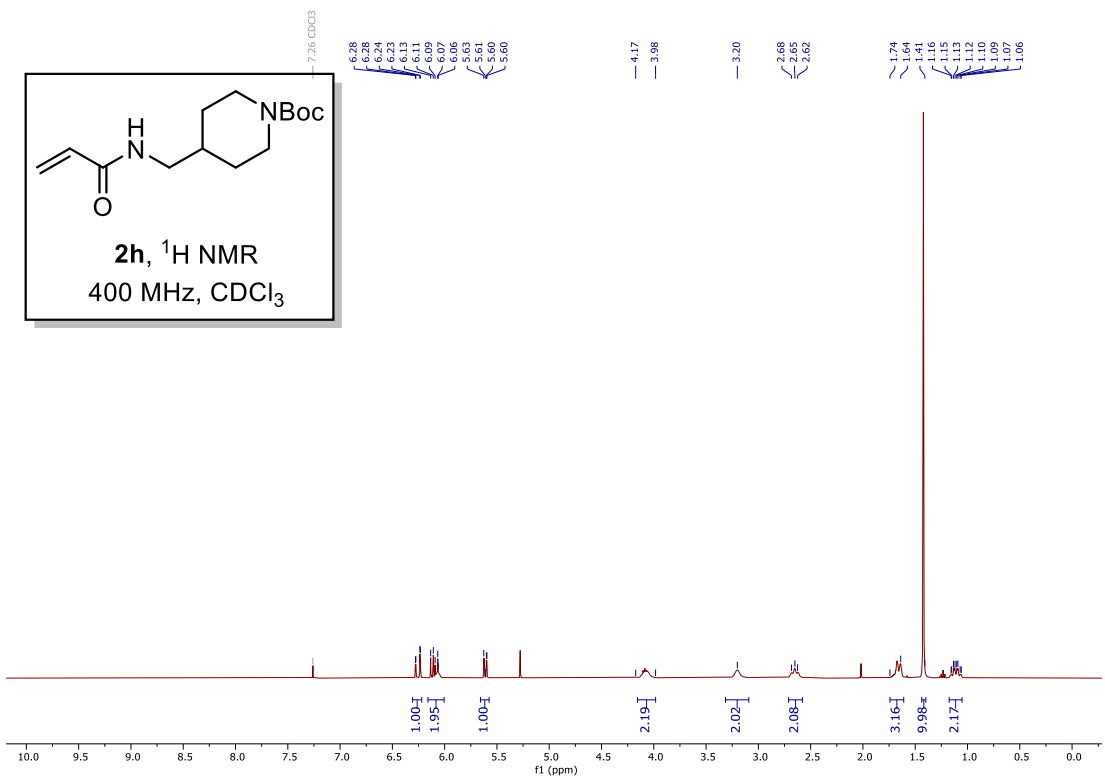
8. Reference

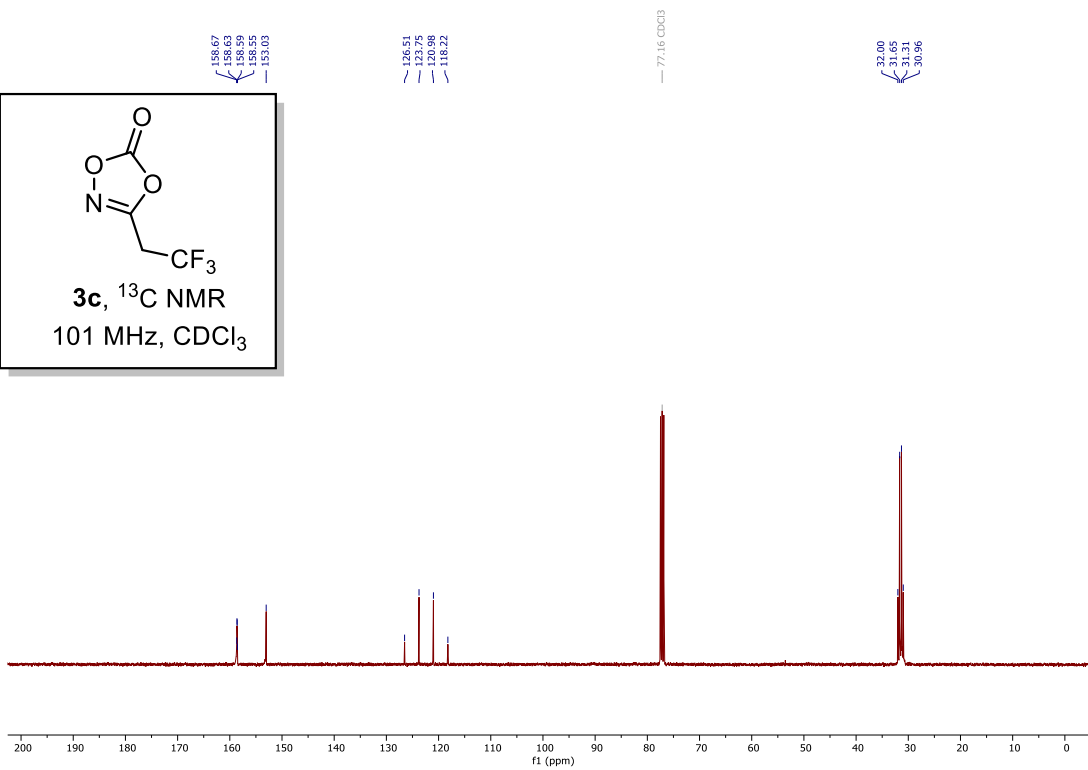
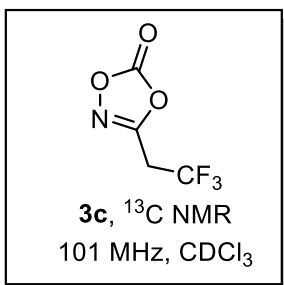
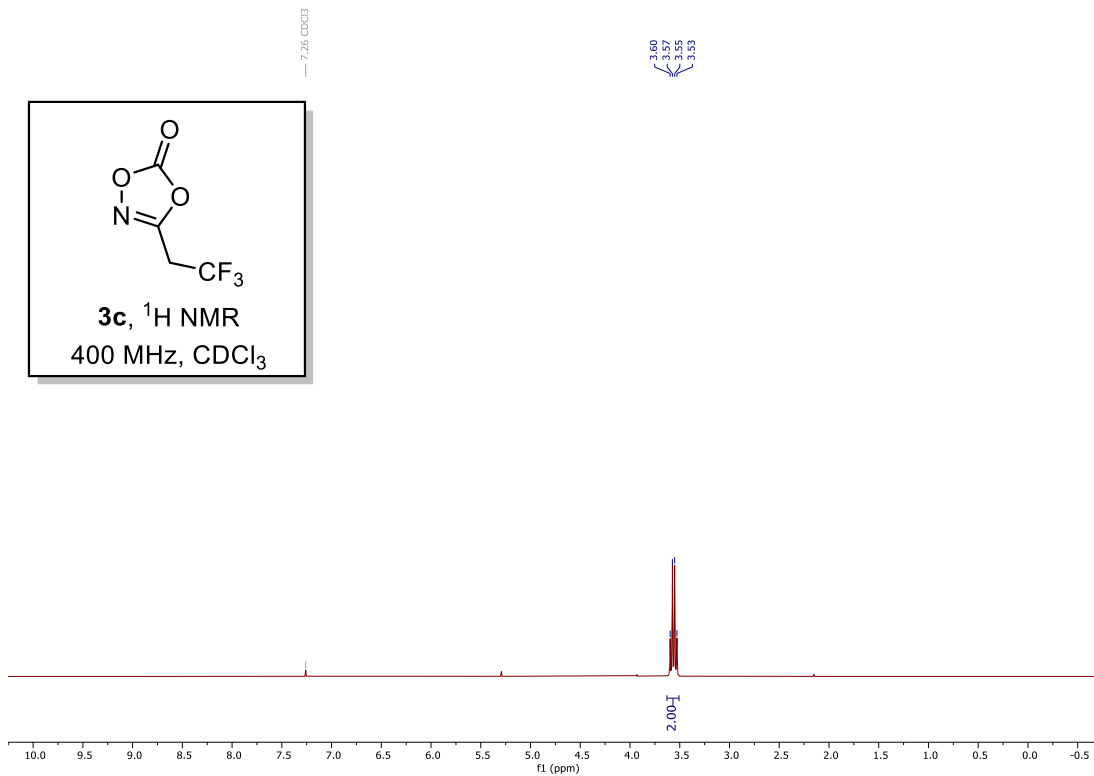
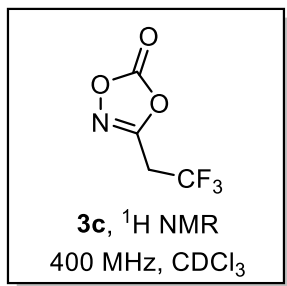
- (1) Chanthamath, S.; Takaki, S.; Shibatomi, K.; Iwasa, S. Highly Stereoselective Cyclopropanation of α,β -Unsaturated Carbonyl Compounds with Methyl (Diazoacetoxy)Acetate Catalyzed by a Chiral Ruthenium(II) Complex. *Angew. Chemie Int. Ed.* **2013**, *52*, 5818–5821.
- (2) Yang, W.; Luo, R.; Yang, D. Iridium-Catalyzed Asymmetric Ring-Opening of Oxabenzonorbornadienes with N-Substituted Piperazine Nucleophiles. *Molecules* **2015**, *20*, 21103.
- (3) Lautens, M.; Fagnou, K.; Zunic, V. An Expedient Enantioselective Route to Diaminotetralins: Application in the Preparation of Analgesic Compounds. *Org. Lett.* **2002**, *4*, 3465–3468.
- (4) Phan, D. H. T.; Kou, K. G. M.; Dong, V. M. Enantioselective Desymmetrization of Cyclopropenes by Hydroacylation. *J. Am. Chem. Soc.* **2010**, *132*, 16354–16355.
- (5) Cheng, X.; Zhu, L.; Lin, M.; Chen, J.; Huang, X. Rapid Access to Cyclopentadiene Derivatives through Gold-Catalyzed Cycloisomerization of Ynamides with Cyclopropenes by Preferential Activation of Alkenes over Alkynes. *Chem. Commun.* **2017**, *53*, 3745–3748.
- (6) Lei, H.; Rovis, T. Ir-Catalyzed Intermolecular Branch-Selective Allylic C–H Amidation of Unactivated Terminal Olefins. *J. Am. Chem. Soc.* **2019**, *141*, 2268–2273.
- (7) Hong, S. Y.; Park, Y.; Hwang, Y.; Kim, Y. B.; Baik, M.-H.; Chang, S. Selective Formation of γ -Lactams via C–H Amidation Enabled by Tailored Iridium Catalysts. *Science (80-.)*. **2018**, *359*, 1016 LP – 1021.
- (8) Farr, C. M. B.; Kazerouni, A. M.; Park, B.; Poff, C. D.; Won, J.; Sharp, K. R.; Baik, M.-H.; Blakey, S. B. Designing a Planar Chiral Rhodium Indenyl Catalyst for Regio- and Enantioselective Allylic C–H Amidation. *J. Am. Chem. Soc.* **2020**, *142*, 13996–14004.
- (9) Lutjen, A. B.; Quirk, M. A.; Barbera, A. M.; Kolonko, E. M. Synthesis of (E)-Cinnamyl Ester Derivatives via a Greener Steglich Esterification. *Bioorg. Med. Chem.* **2018**, *26*, 5291–5298.
- (10) Chen, H.; Xu, X.; Liu, L.; Tang, G.; Zhao, Y. Phosphorus Oxychloride as an Efficient Coupling Reagent for the Synthesis of Esters, Amides and Peptides under Mild Conditions. *RSC Adv.* **2013**, *3*, 16247–16250.
- (11) Wenzel, S. C.; Williamson, R. M.; Grünanger, C.; Xu, J.; Gerth, K.; Martinez, R. A.; Moss, S. J.; Carroll, B. J.; Grond, S.; Unkefer, C. J.; Müller, R.; Floss, H. G. On the Biosynthetic Origin of Methoxymalonyl-Acyl Carrier Protein, the Substrate for Incorporation of “Glycolate” Units into Ansamitocin and Soraphen A. *J. Am. Chem. Soc.* **2006**, *128*, 14325–14336.

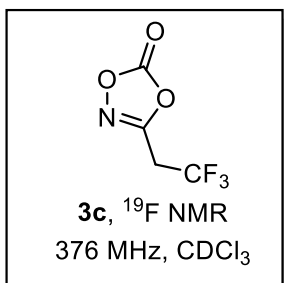
- (12) Luo, Y.; Roy, I. D.; Madec, A. G. E.; Lam, H. Enantioselective Synthesis of Allylboronates and Allylic Alcohols by Copper-Catalyzed 1,6-Boration. *Angew. Chem. Int. Ed.* **2014**, *53*, 4186–4190.
- (13) He, G.; Lu, C.; Zhao, Y.; Nack, W. A.; Chen, G. Improved Protocol for Indoline Synthesis via Palladium-Catalyzed Intramolecular C(Sp²)-H Amination. *Org. Lett.* **2012**, *14*, 2944–2947.
- (14) McComas, C. C.; Gilbert, E. J.; Van Vranken, D. L. Stereochemistry of 3-Alkylindole Dimerization: Acyclic $\delta 1, \delta 1'$ -Tryptophan Dimers. *J. Org. Chem.* **1997**, *62*, 8600–8603.

9. NMR Spectra

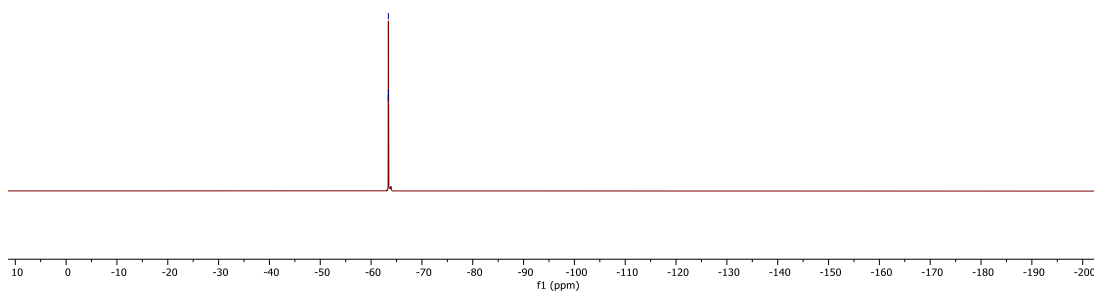
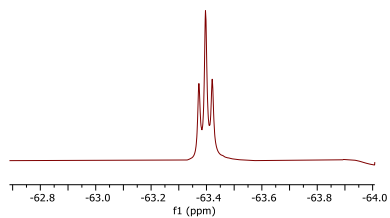


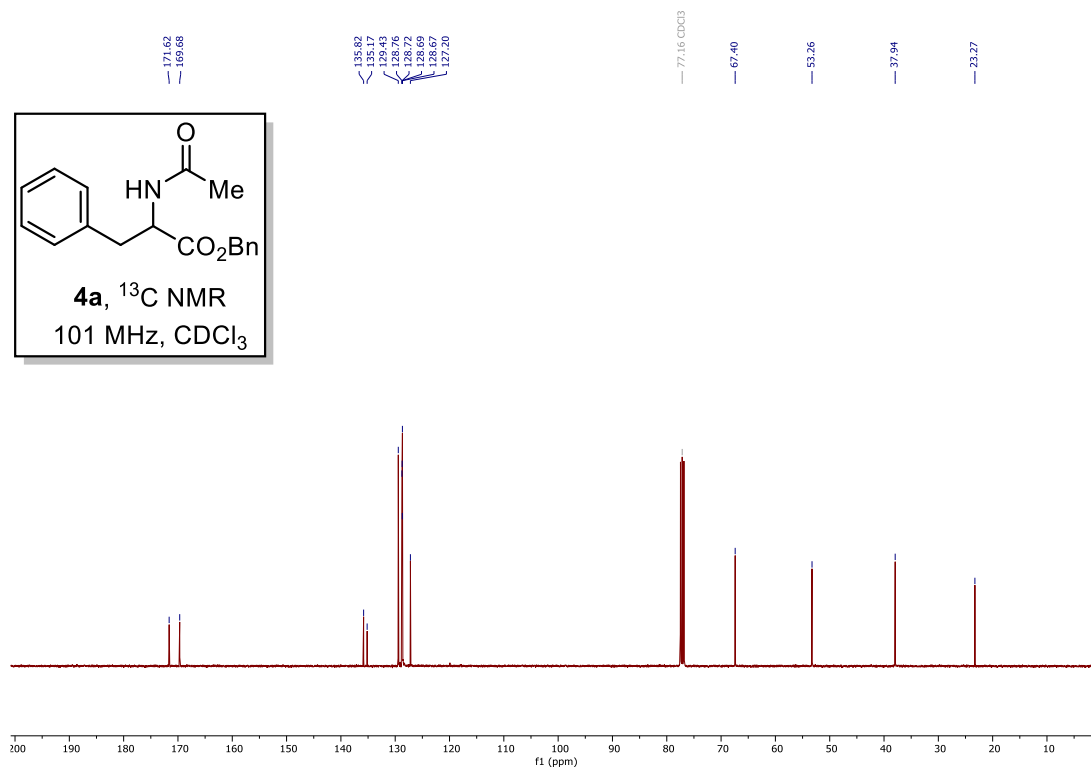
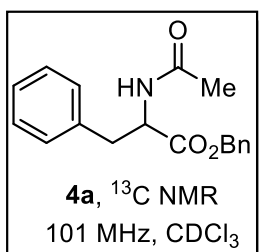
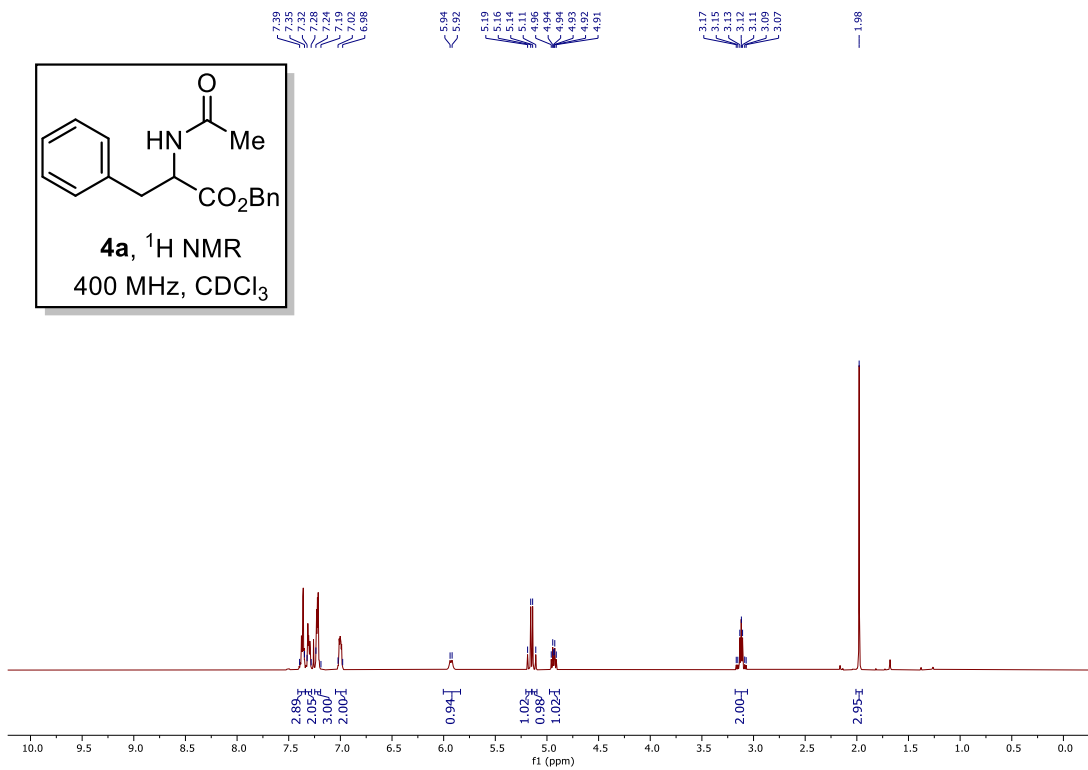
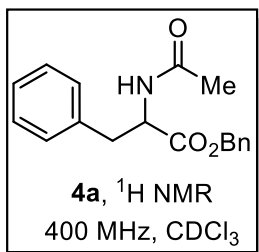


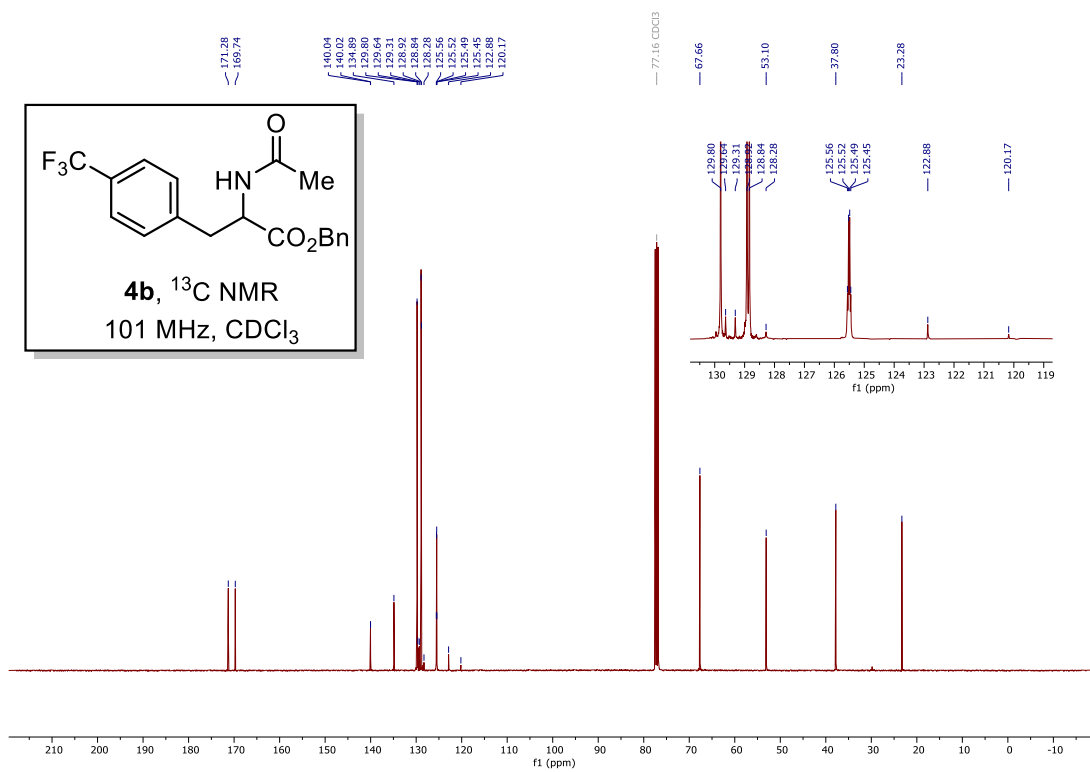
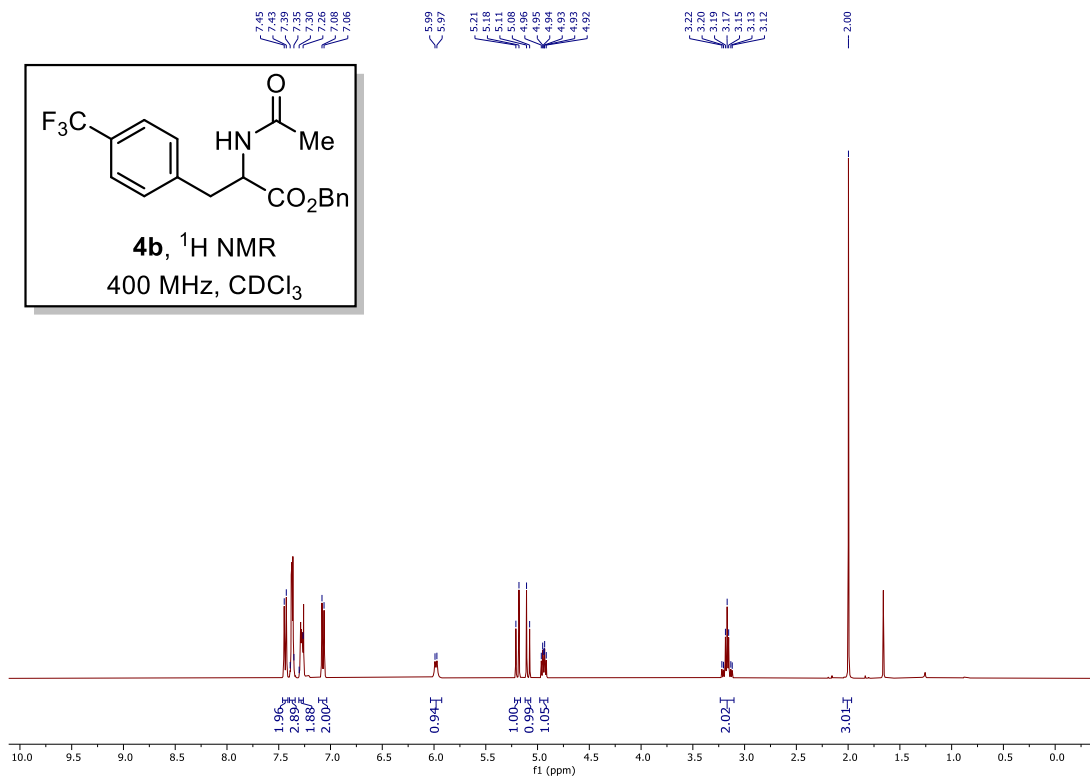


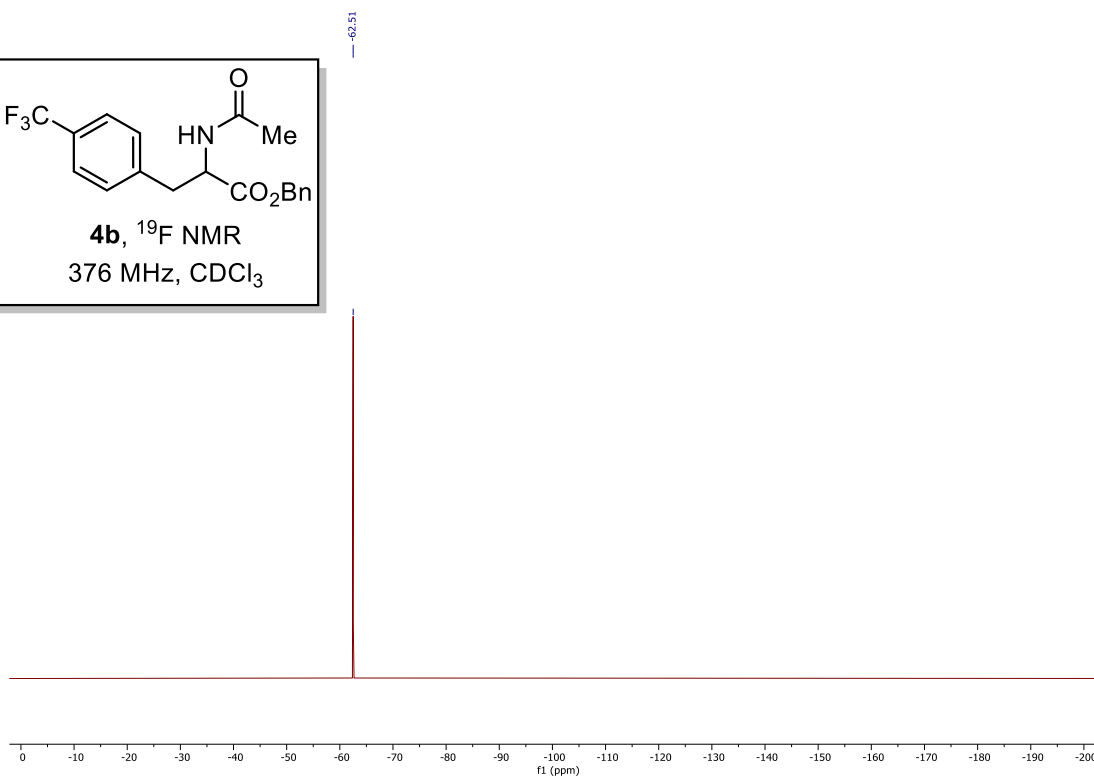
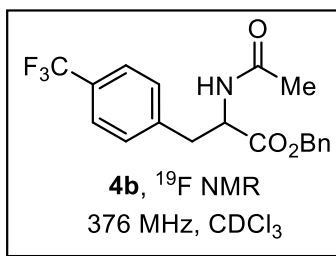


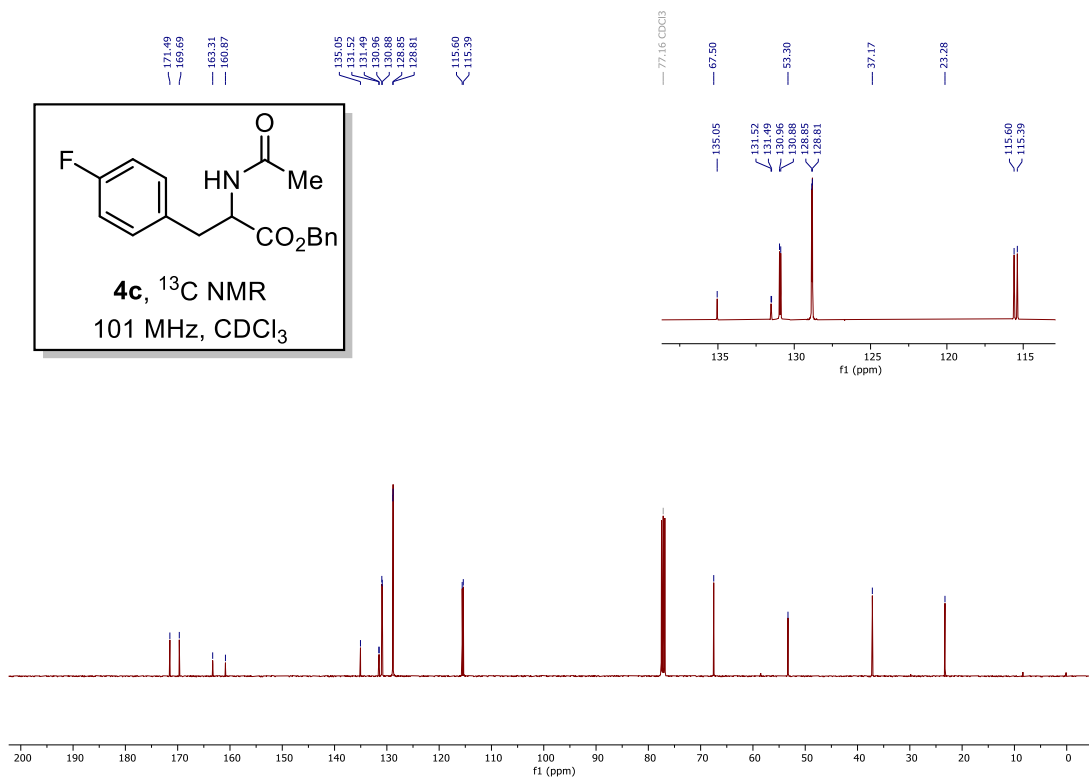
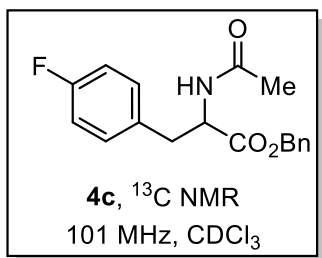
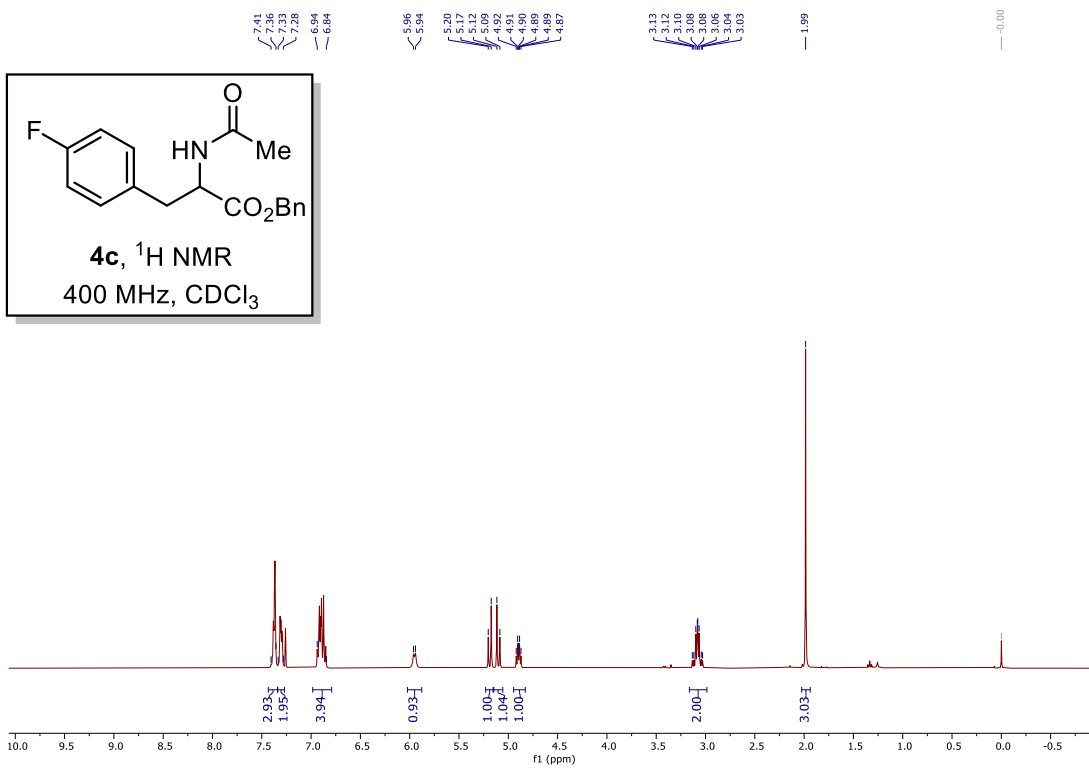
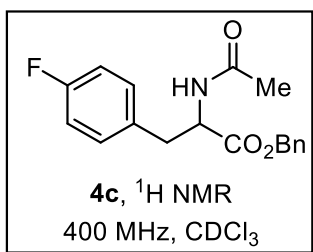
63.37
63.40
63.42

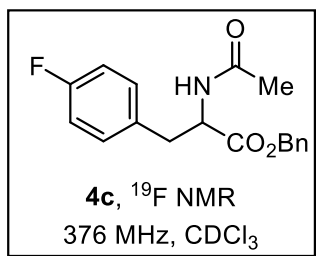




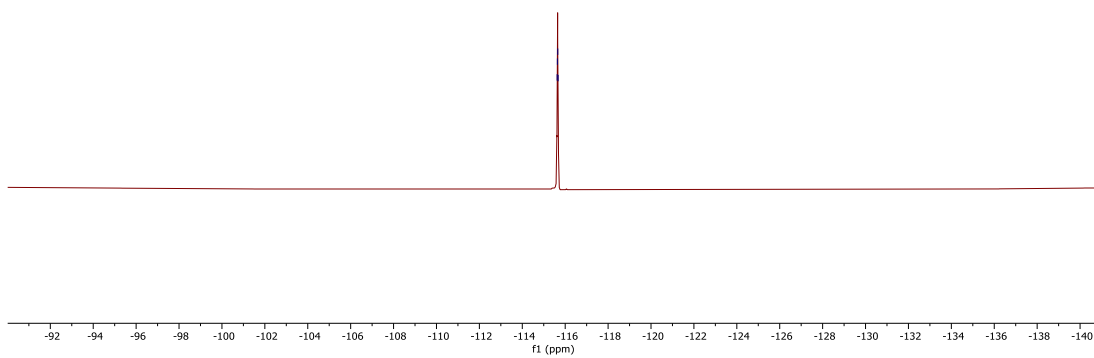


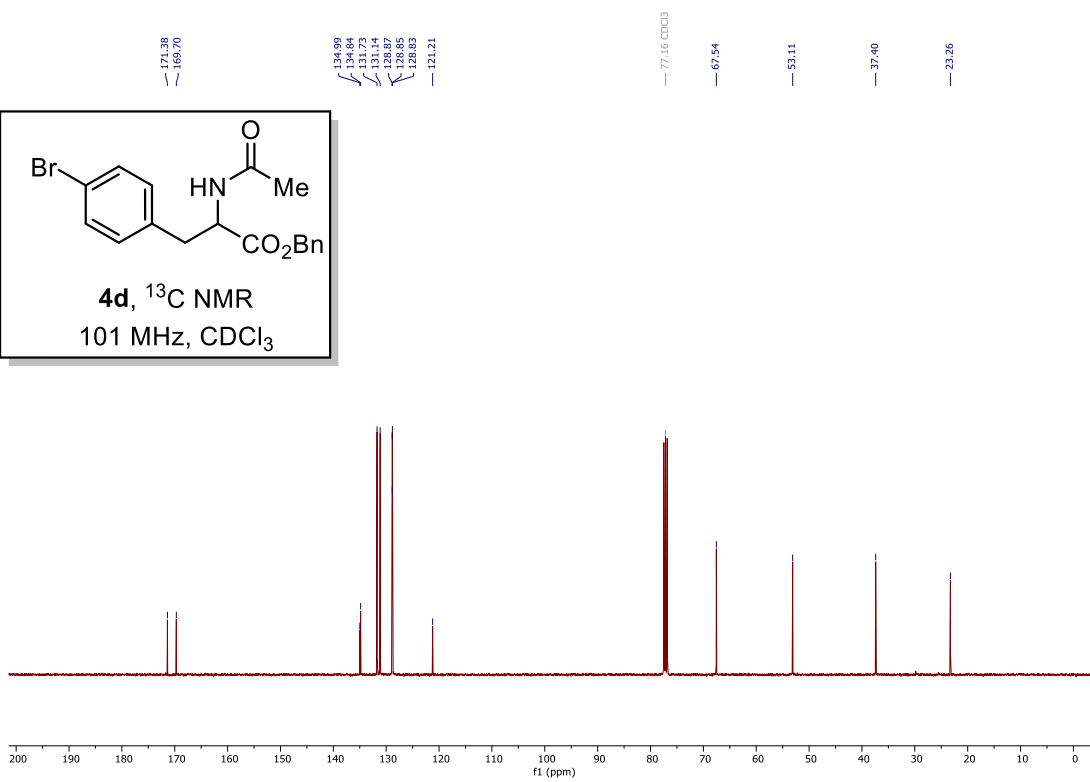
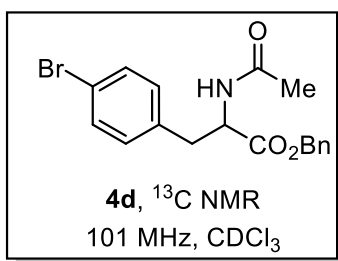
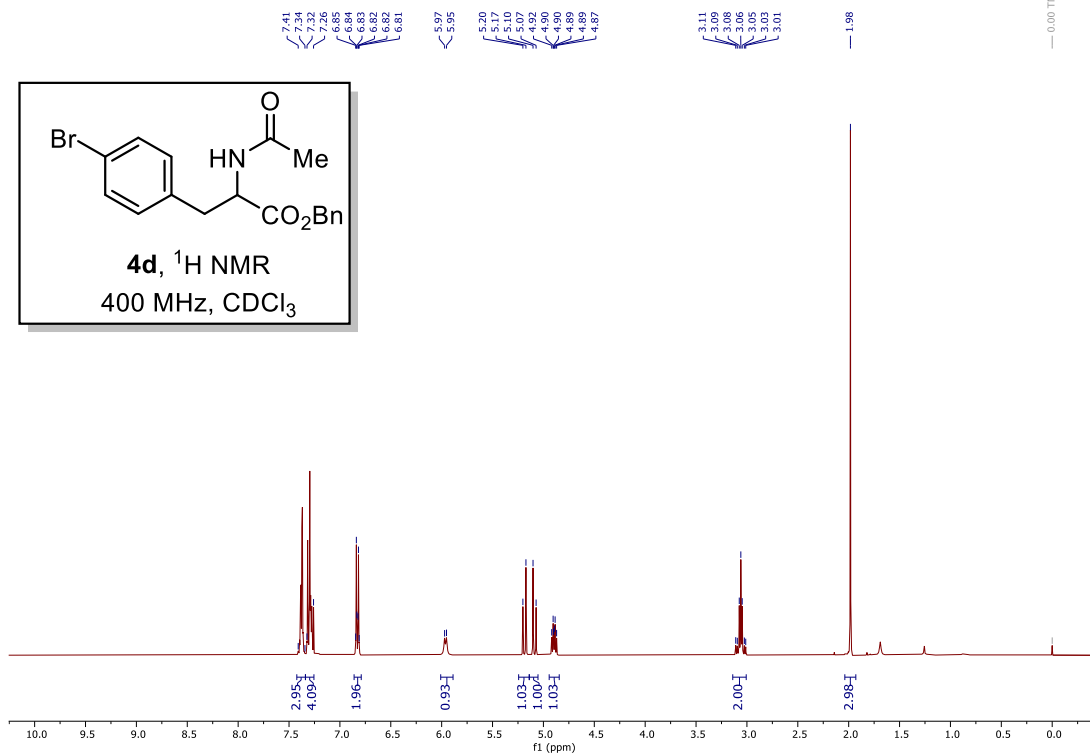
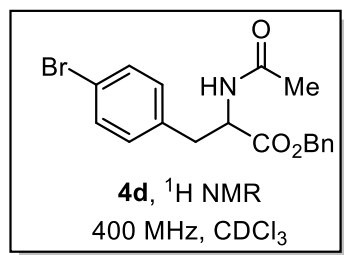


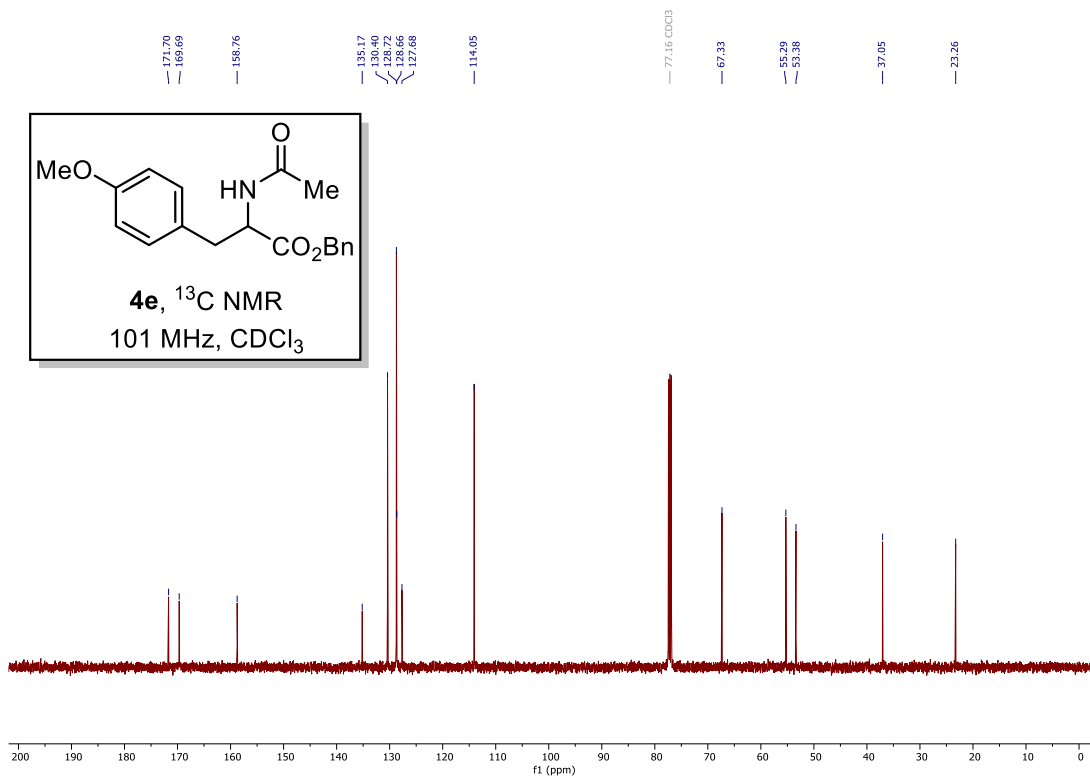
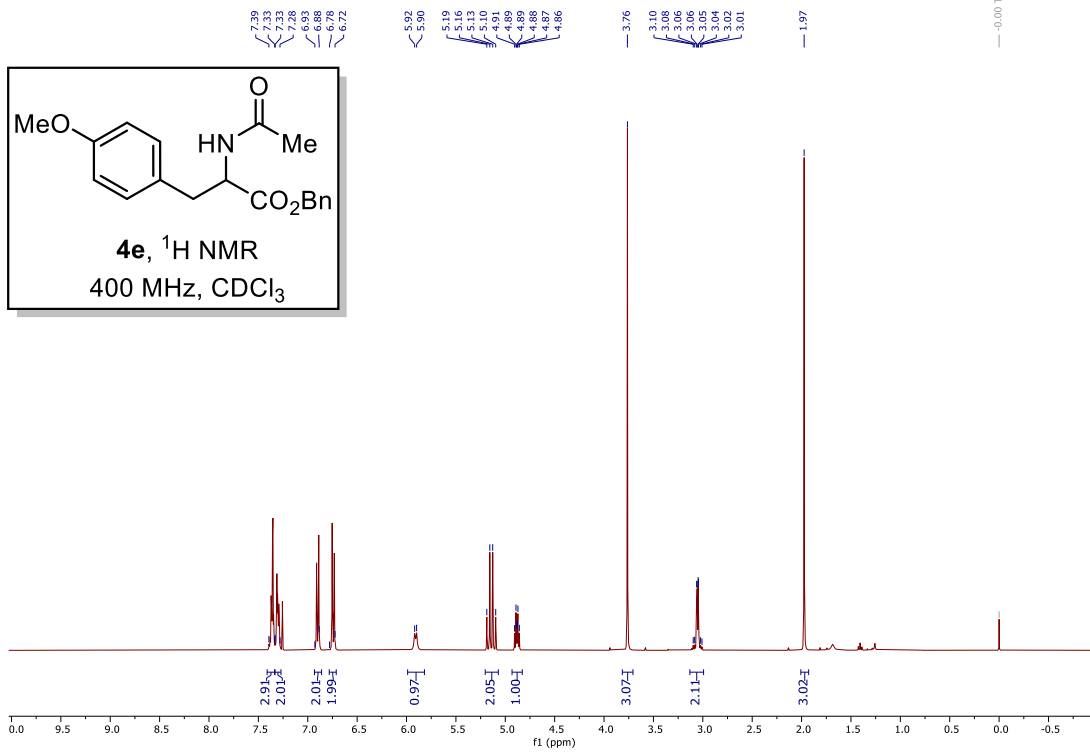


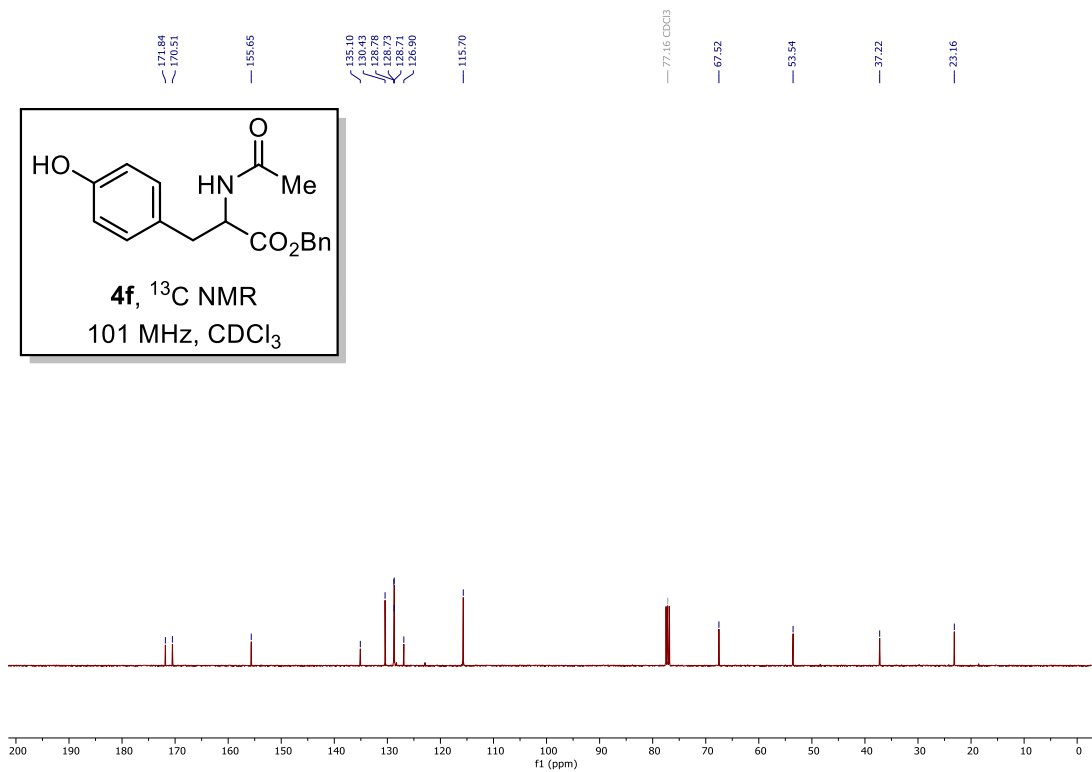
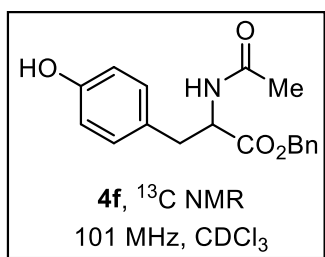
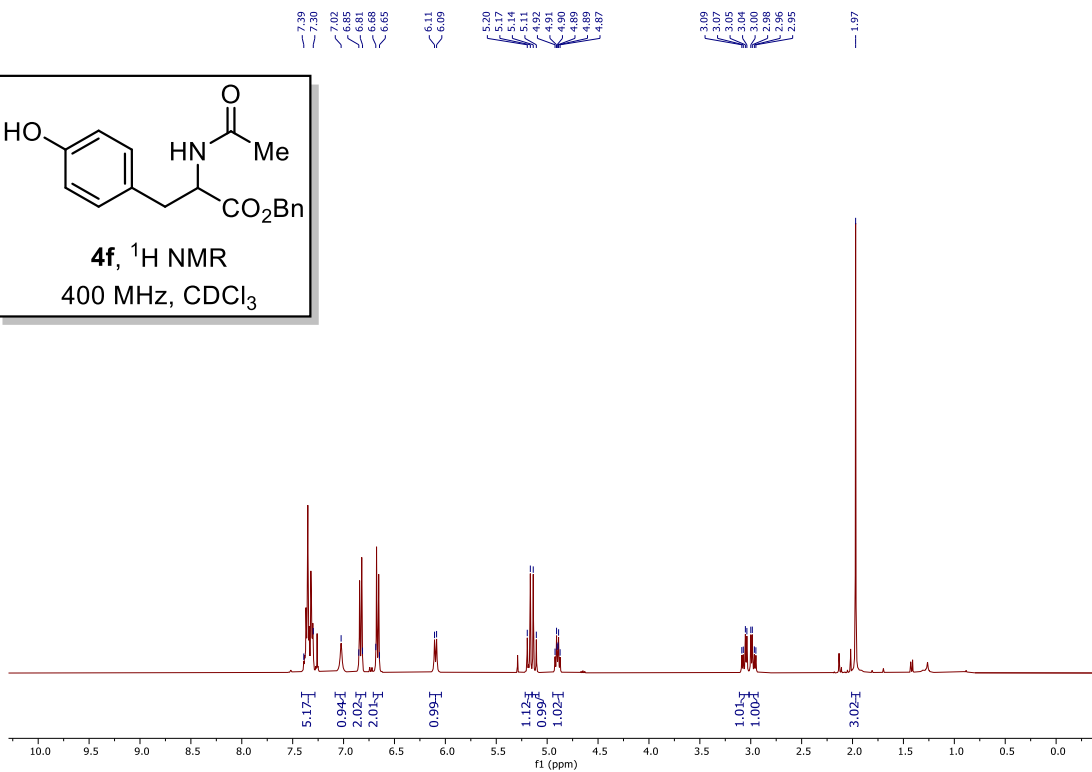
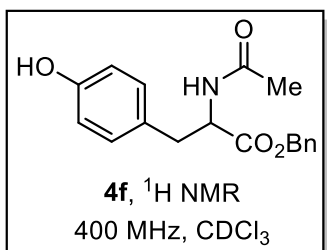


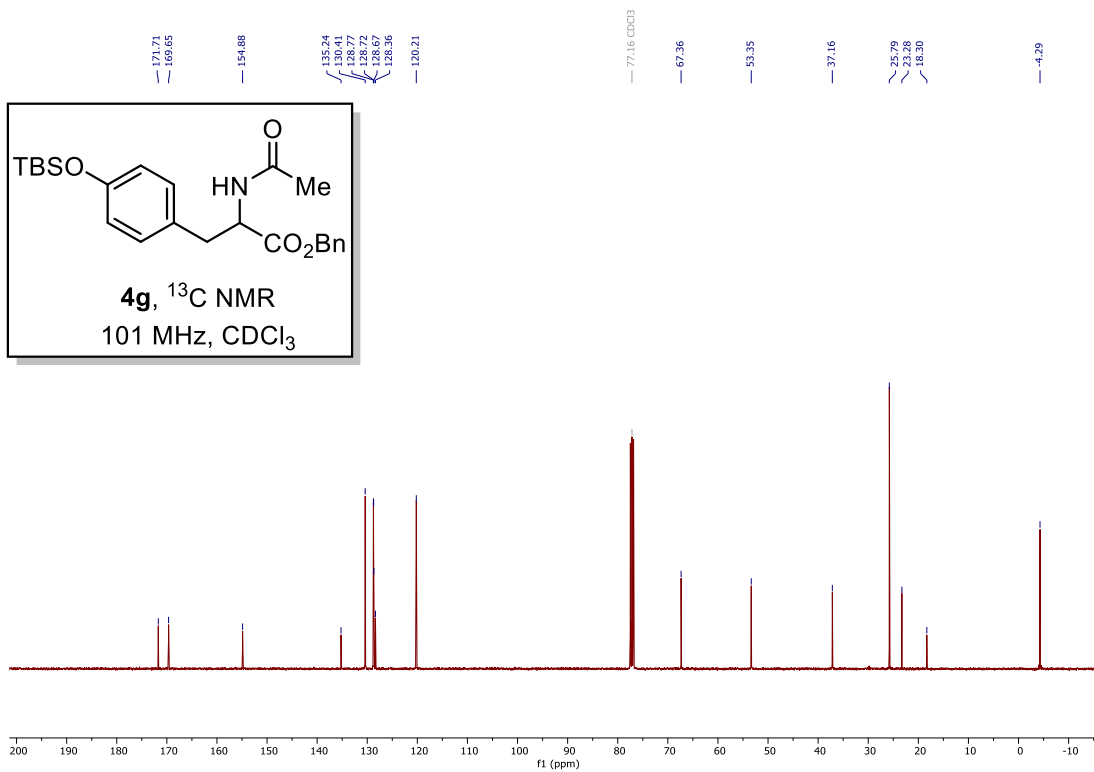
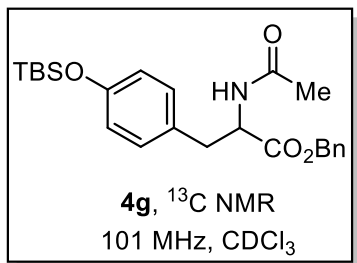
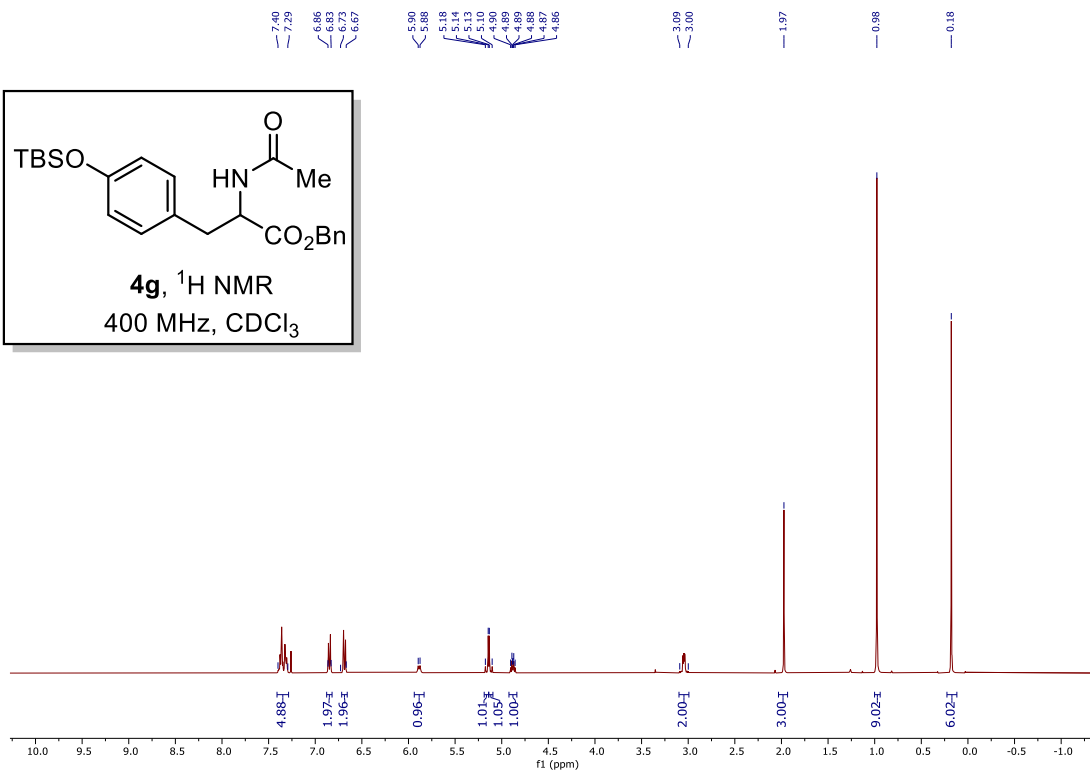
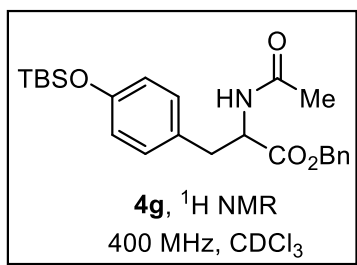
-115.62
-115.64
-115.65
-115.67

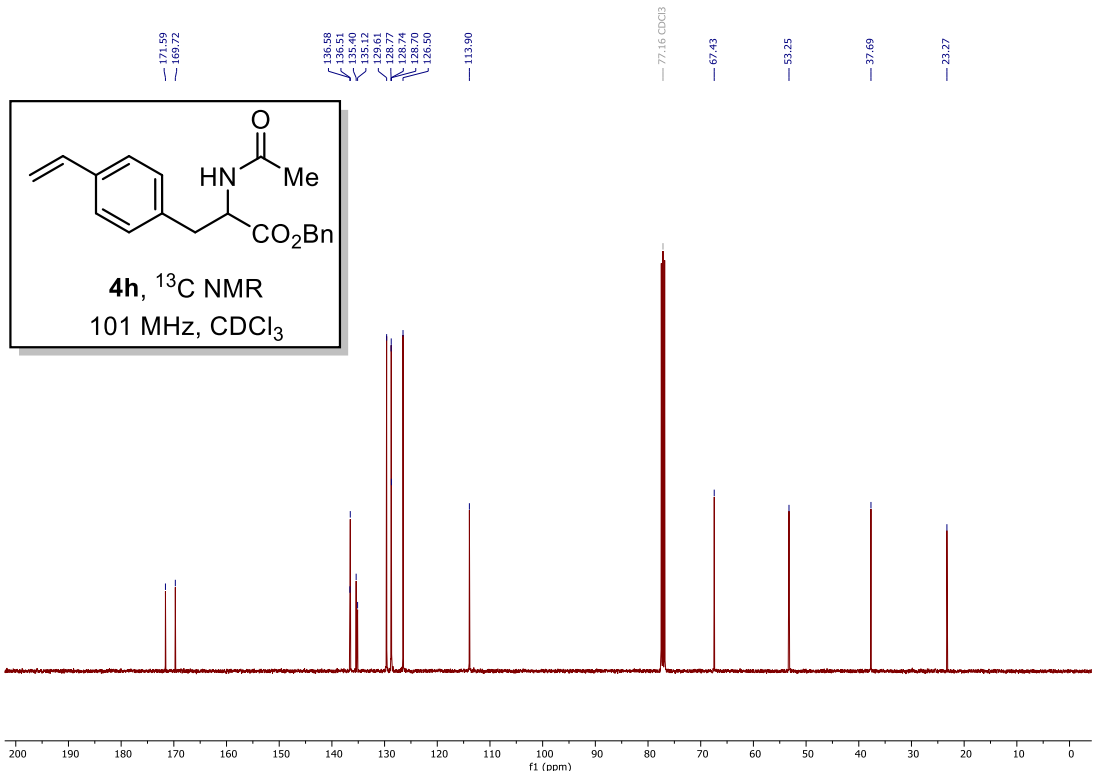
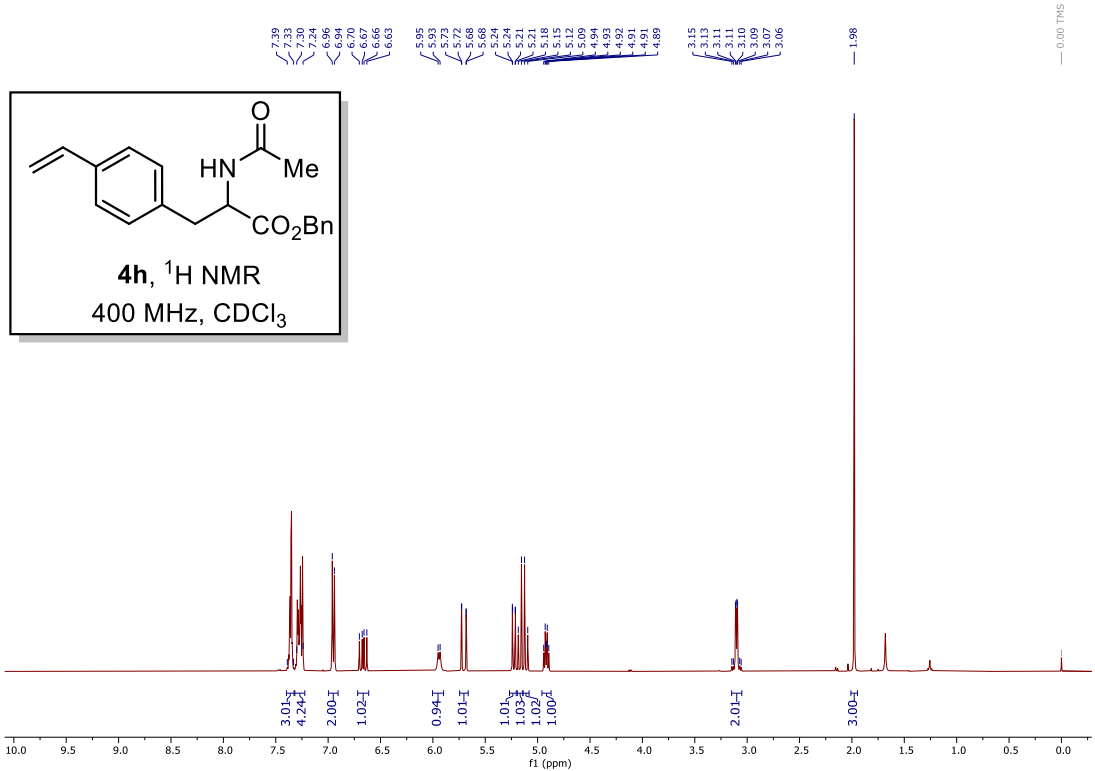


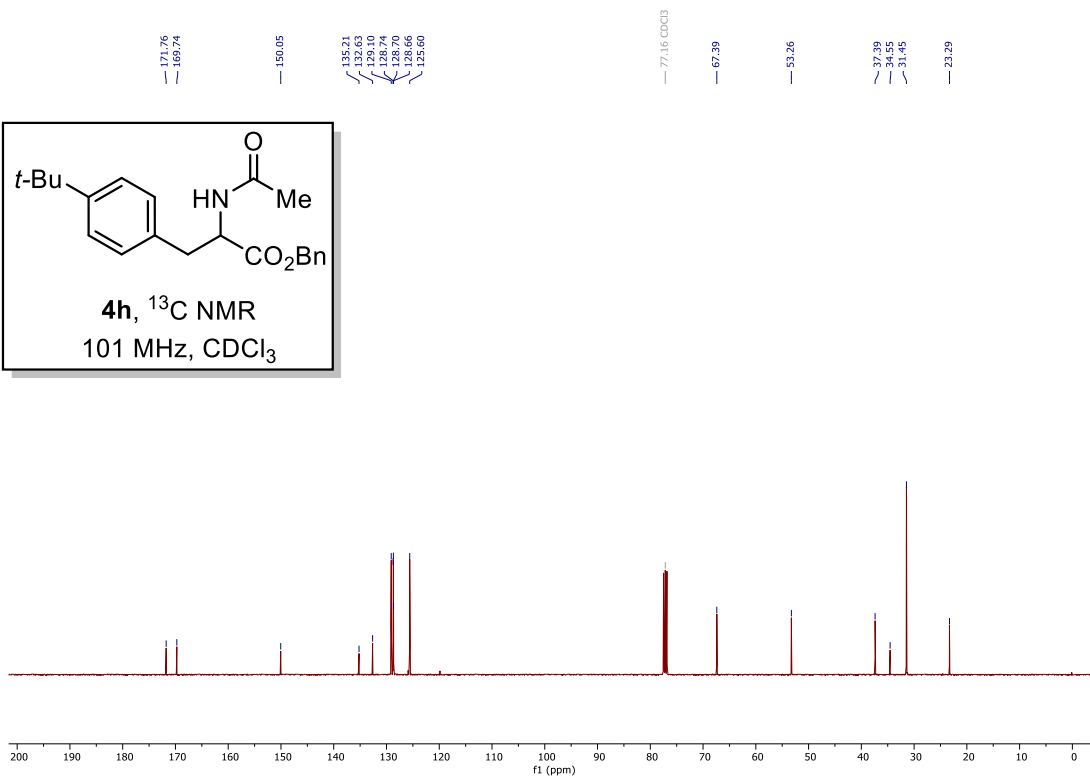
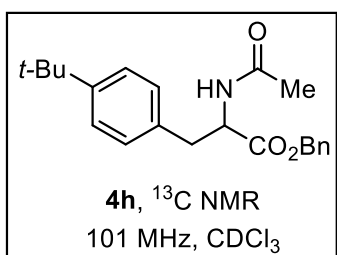
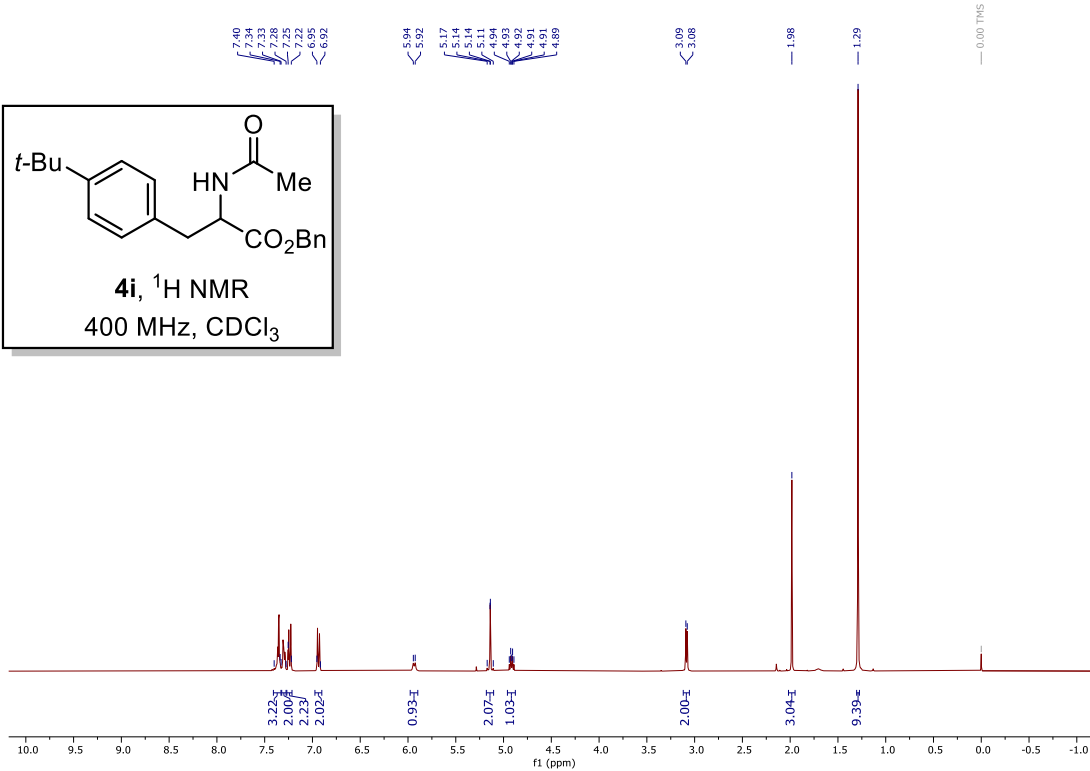
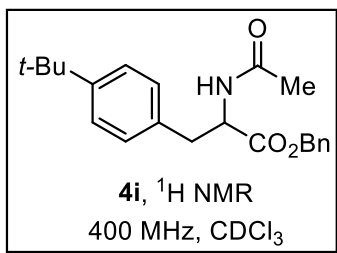


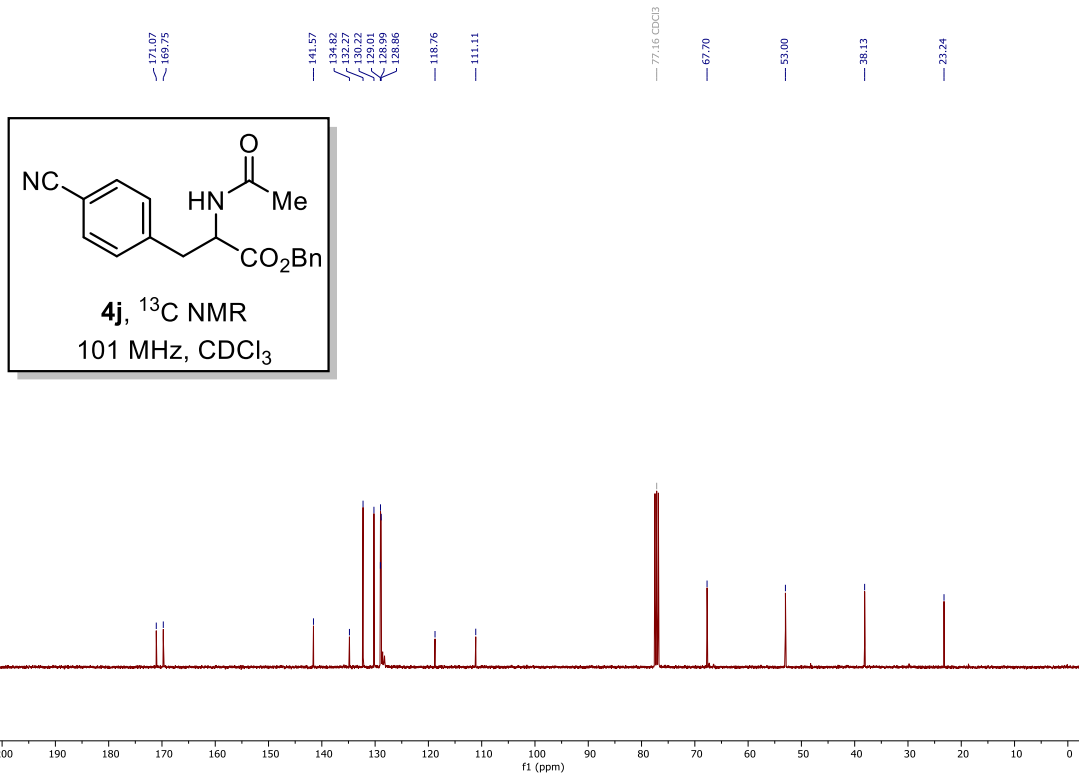
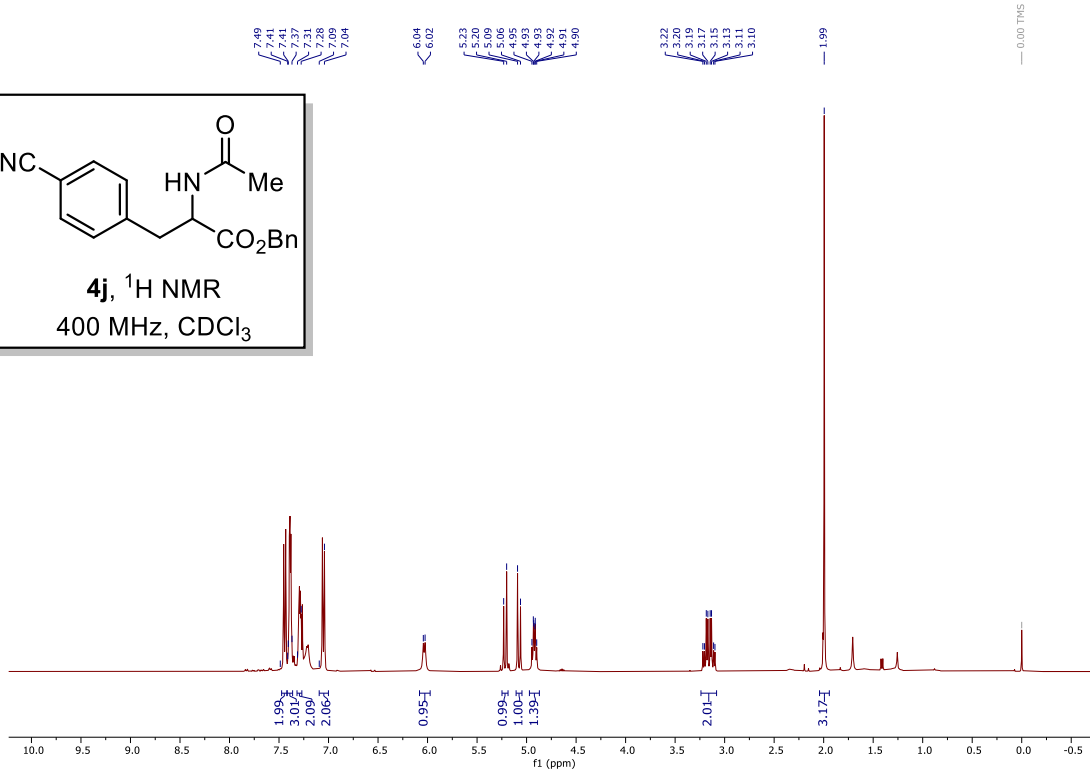
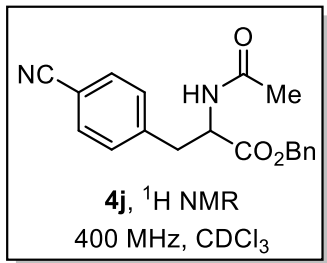


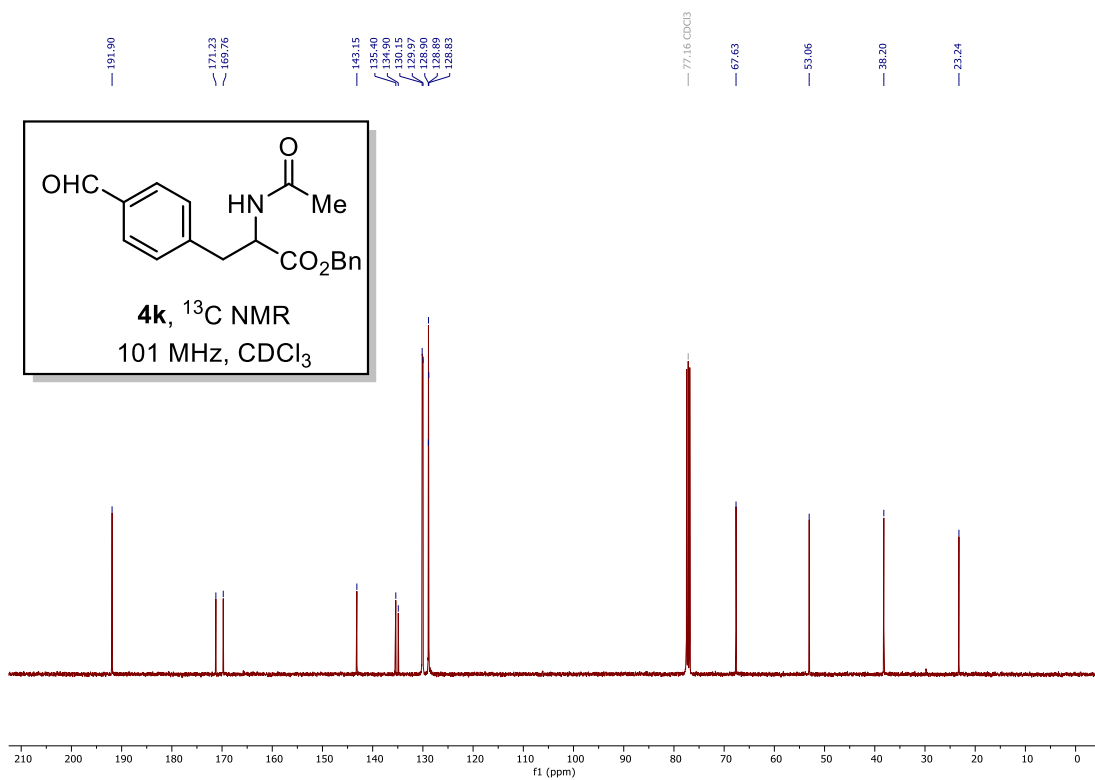
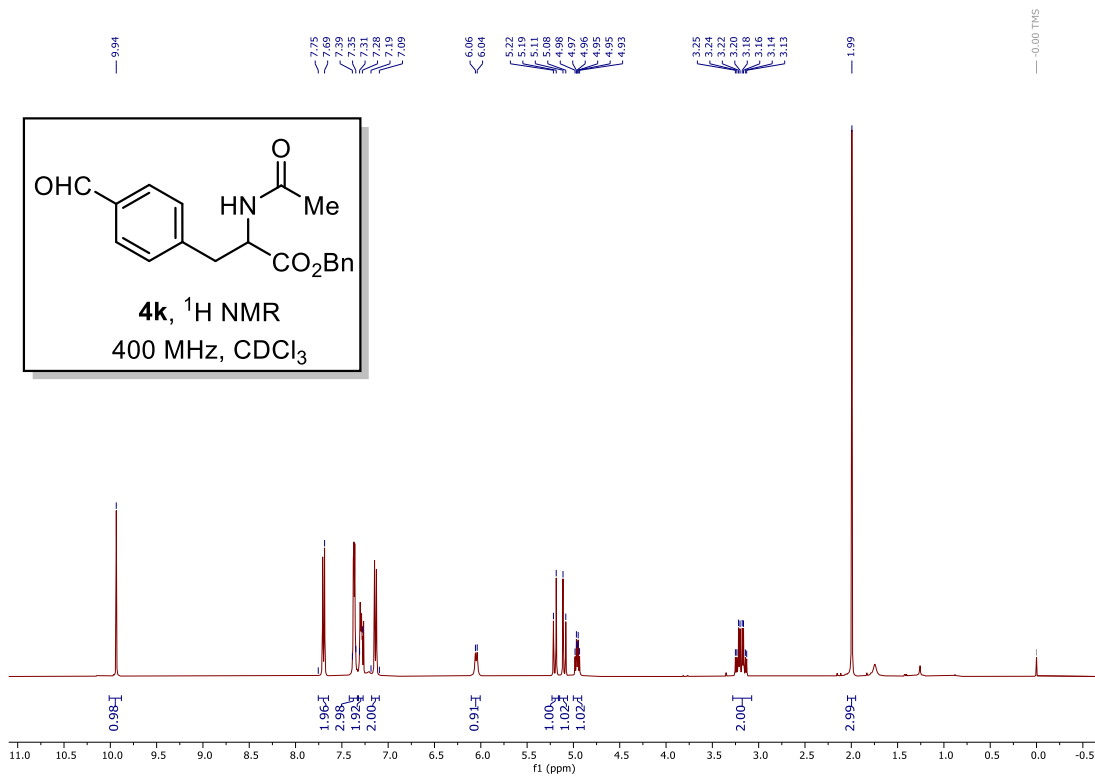


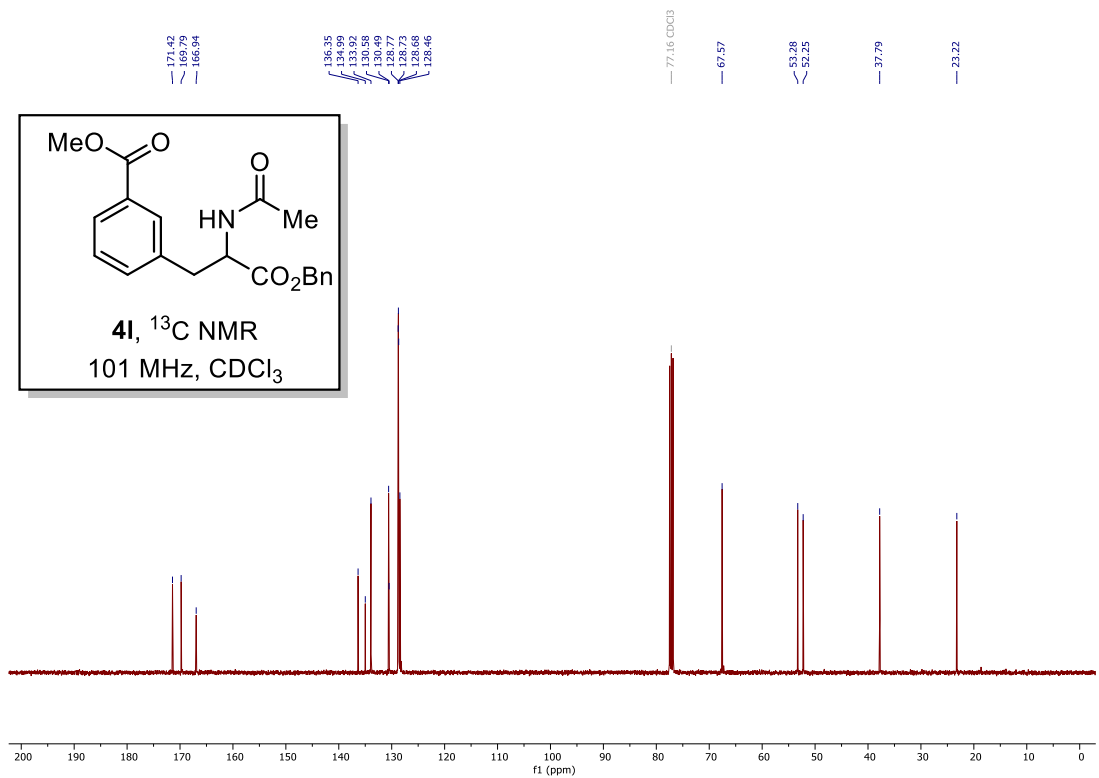
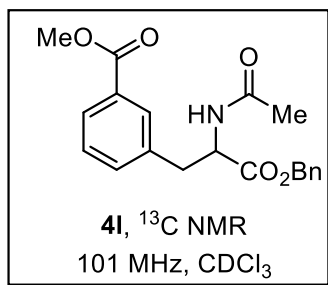
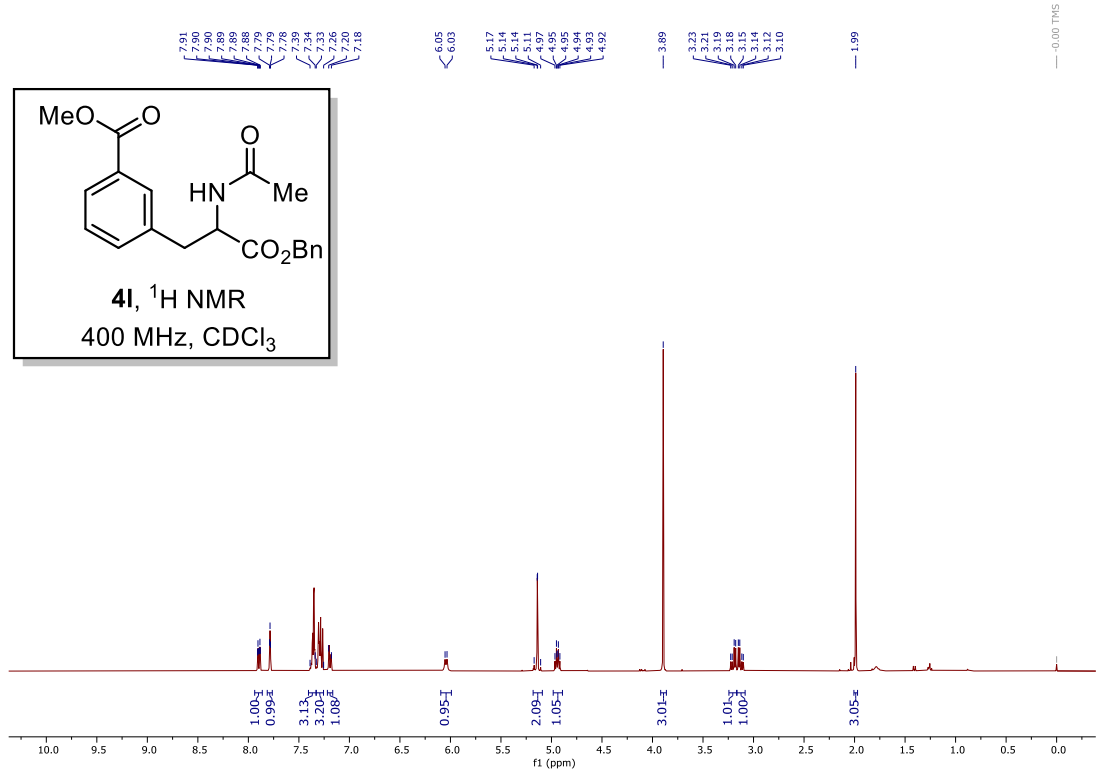
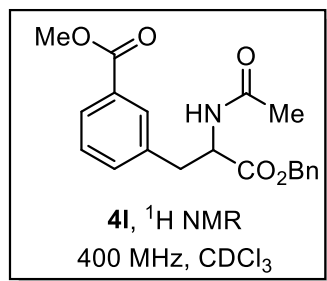


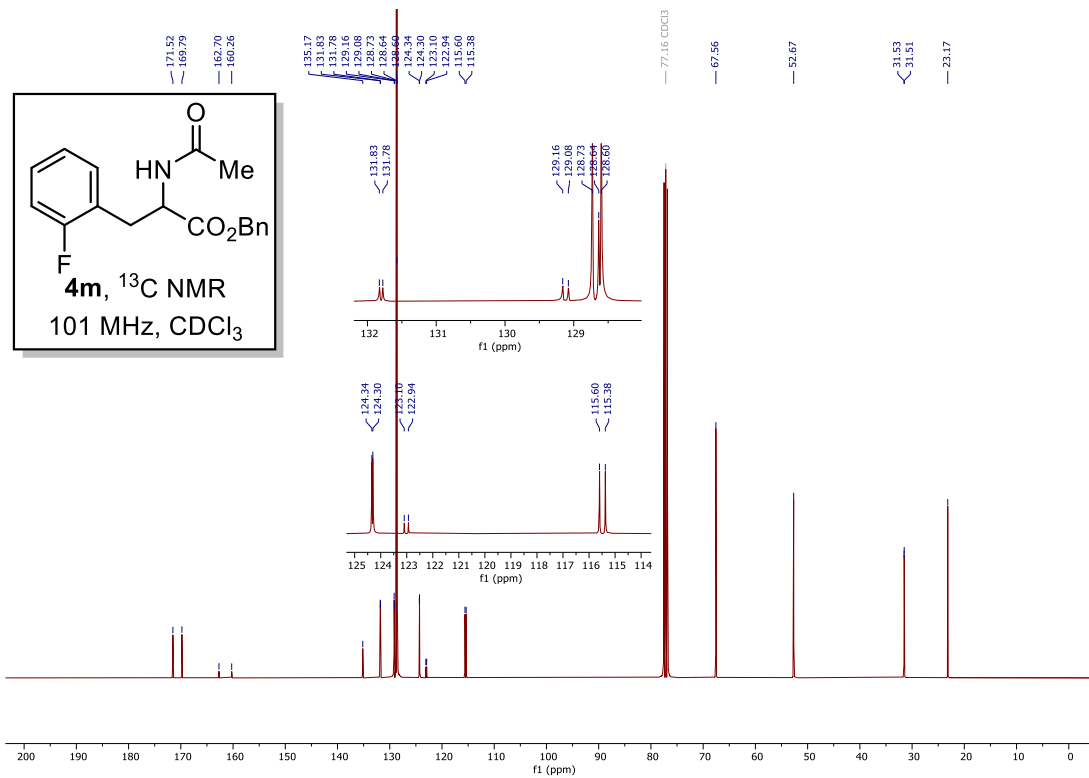
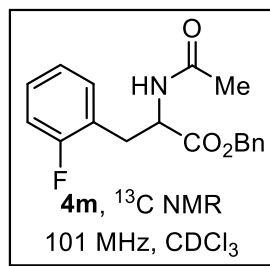
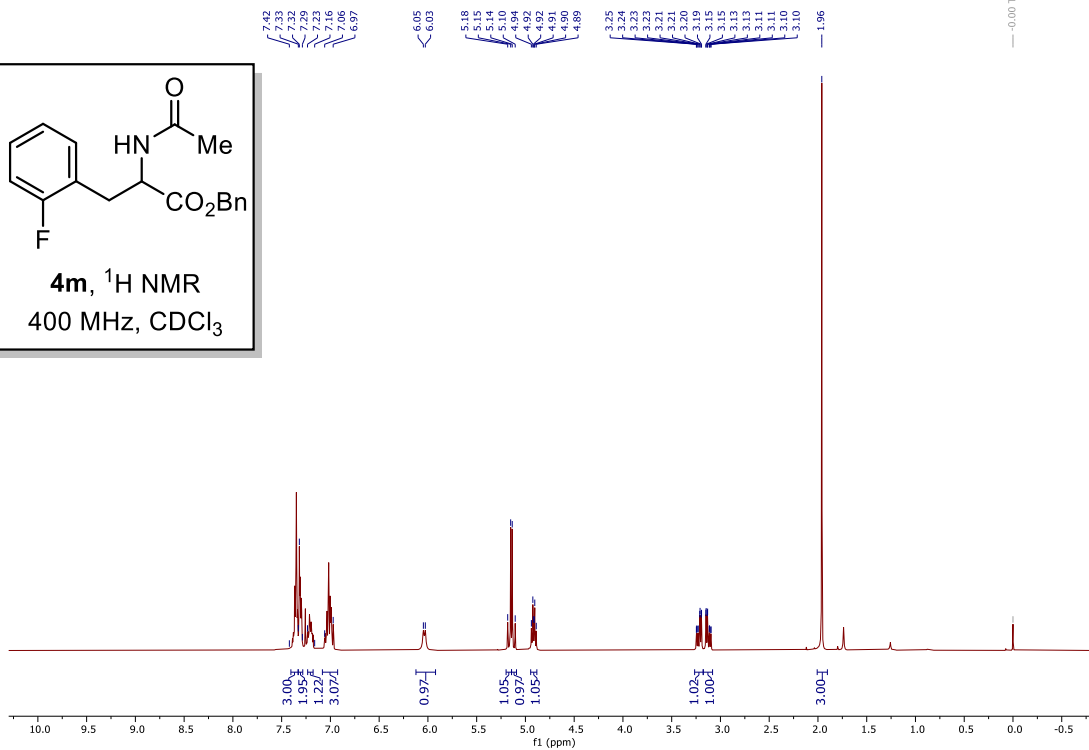
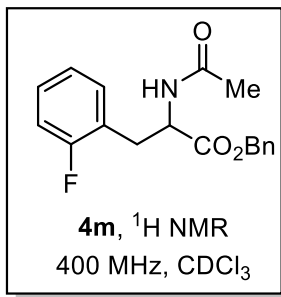


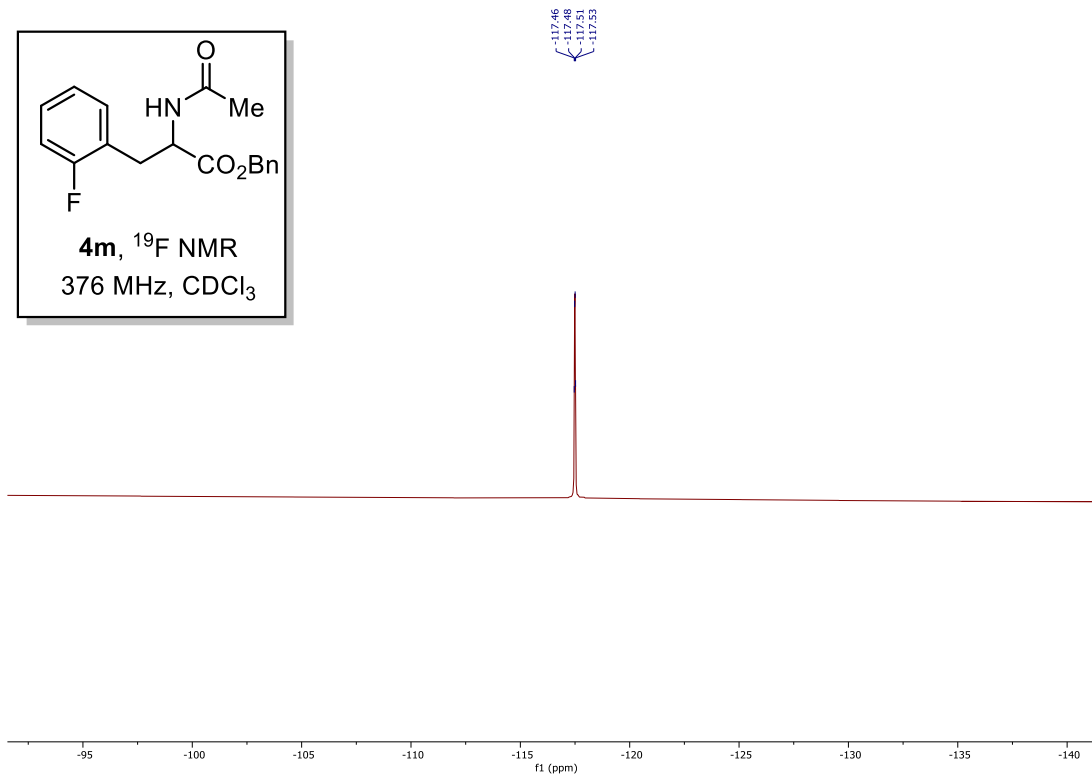
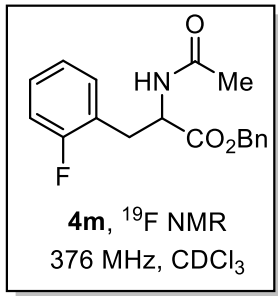


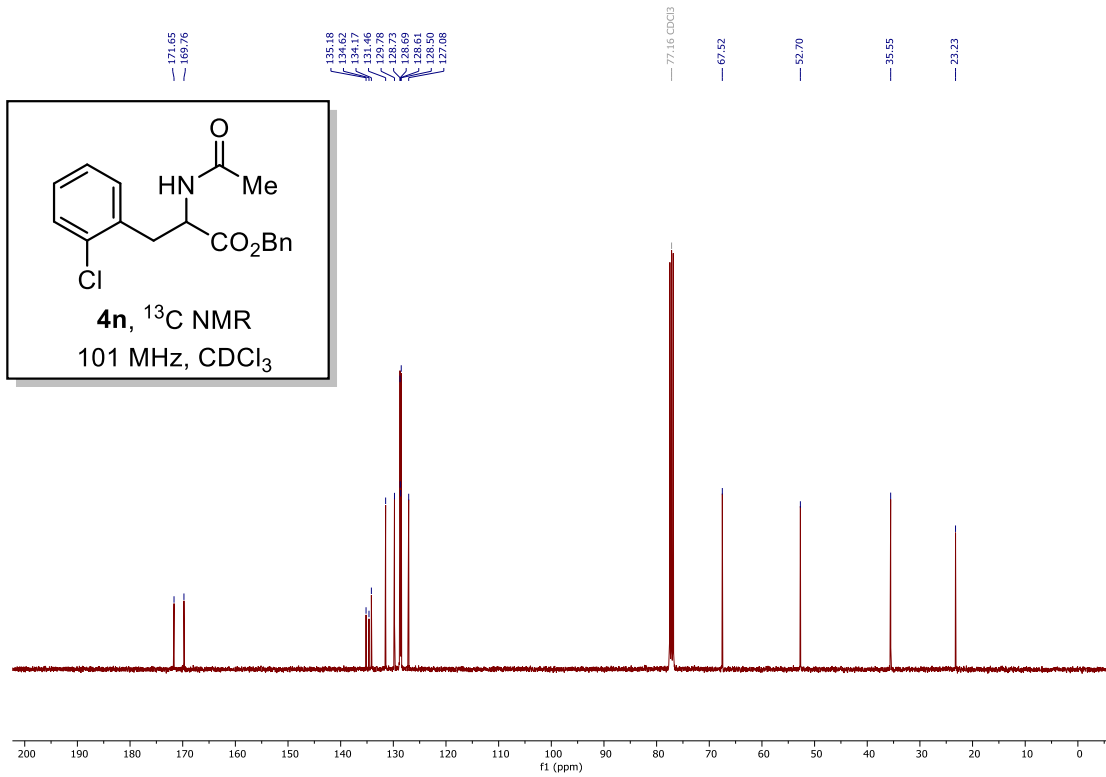
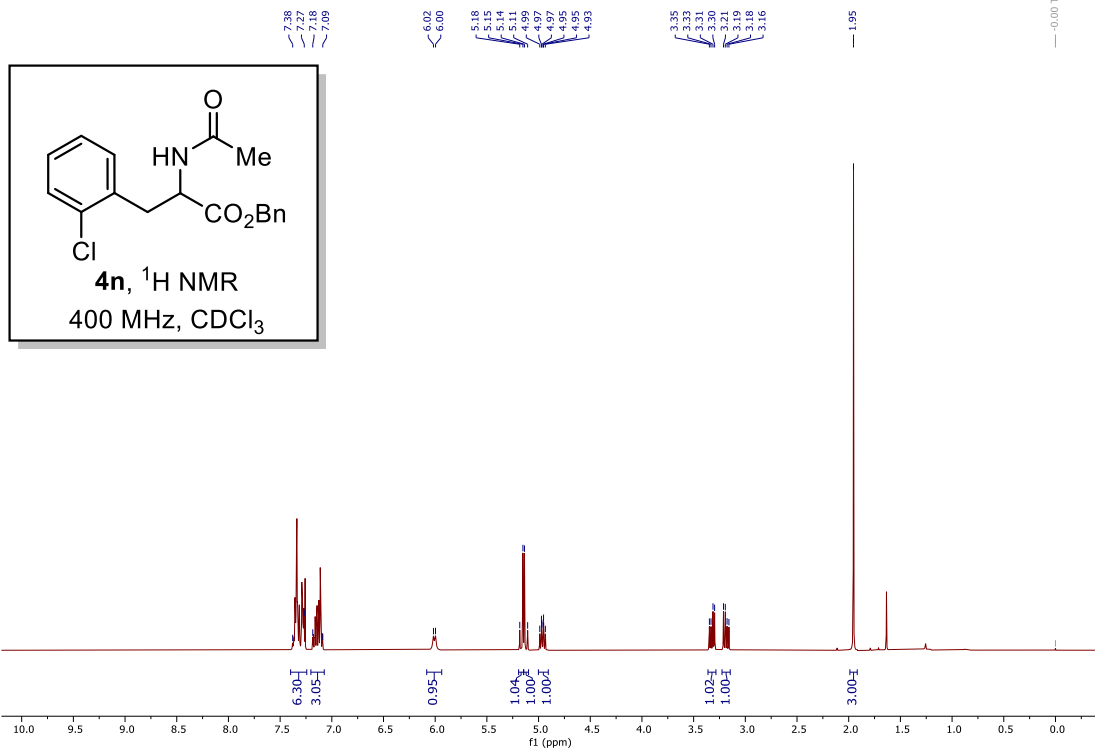


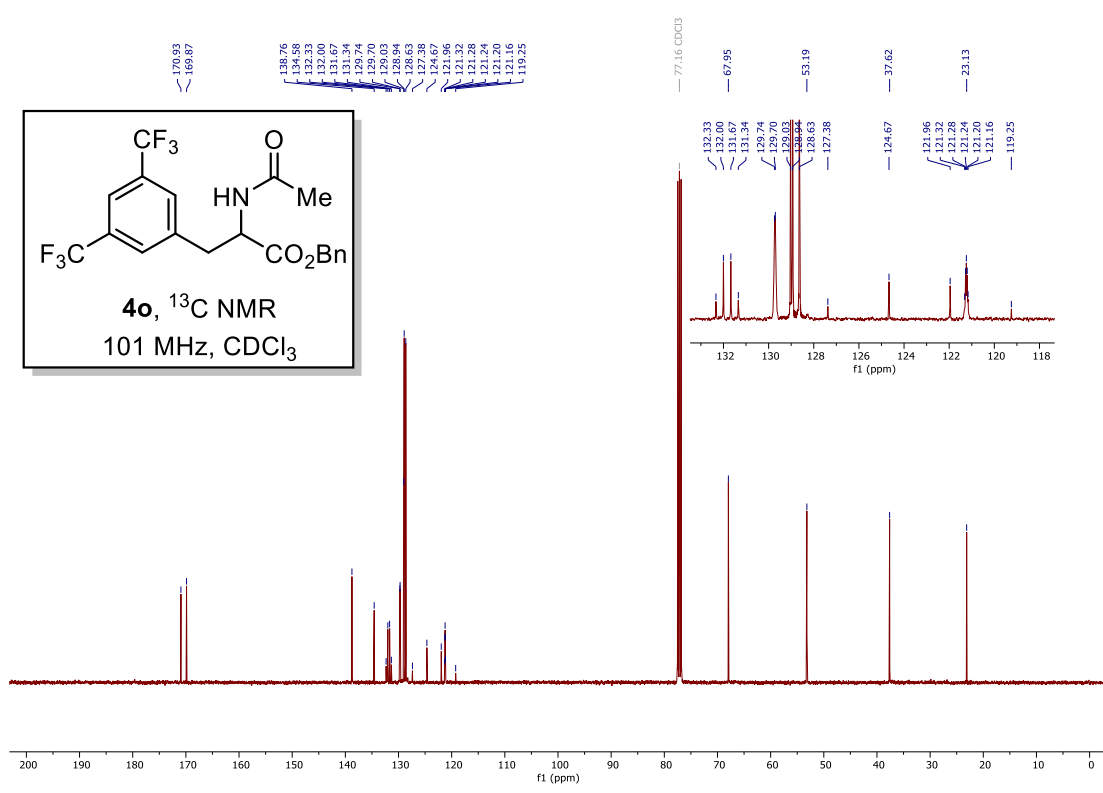
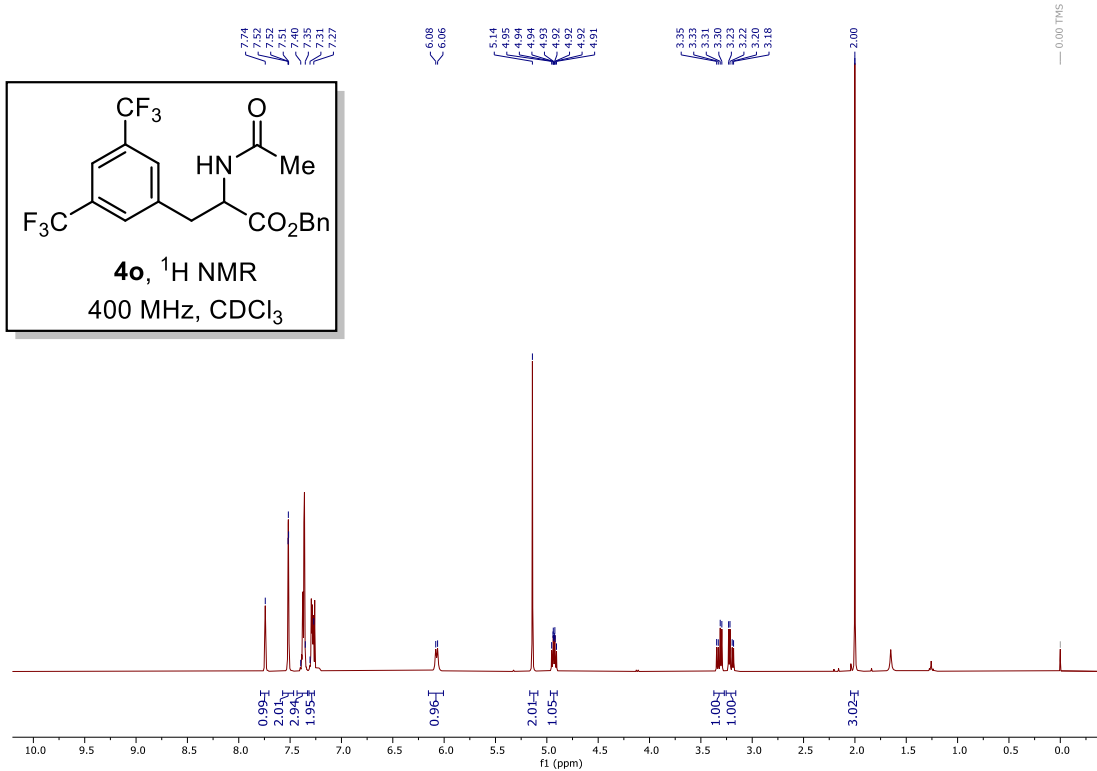


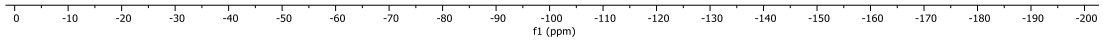
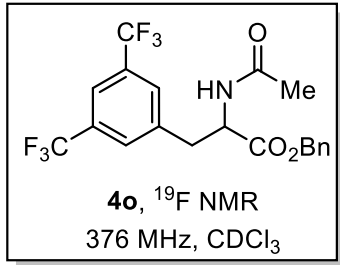


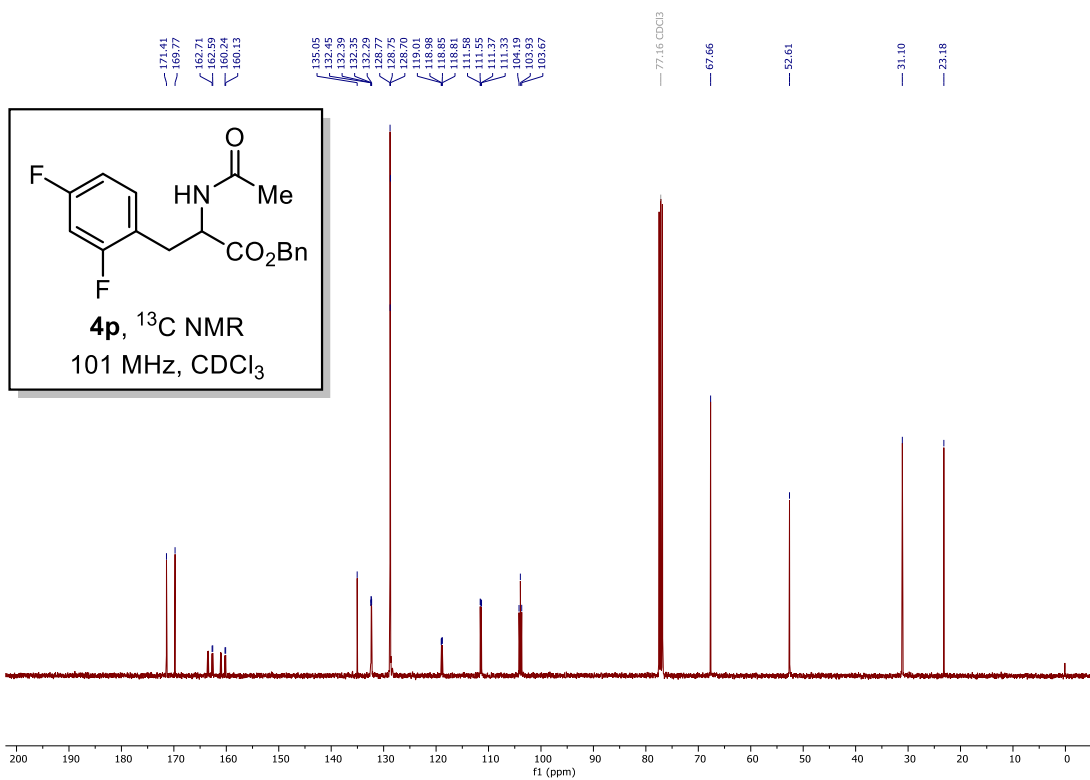
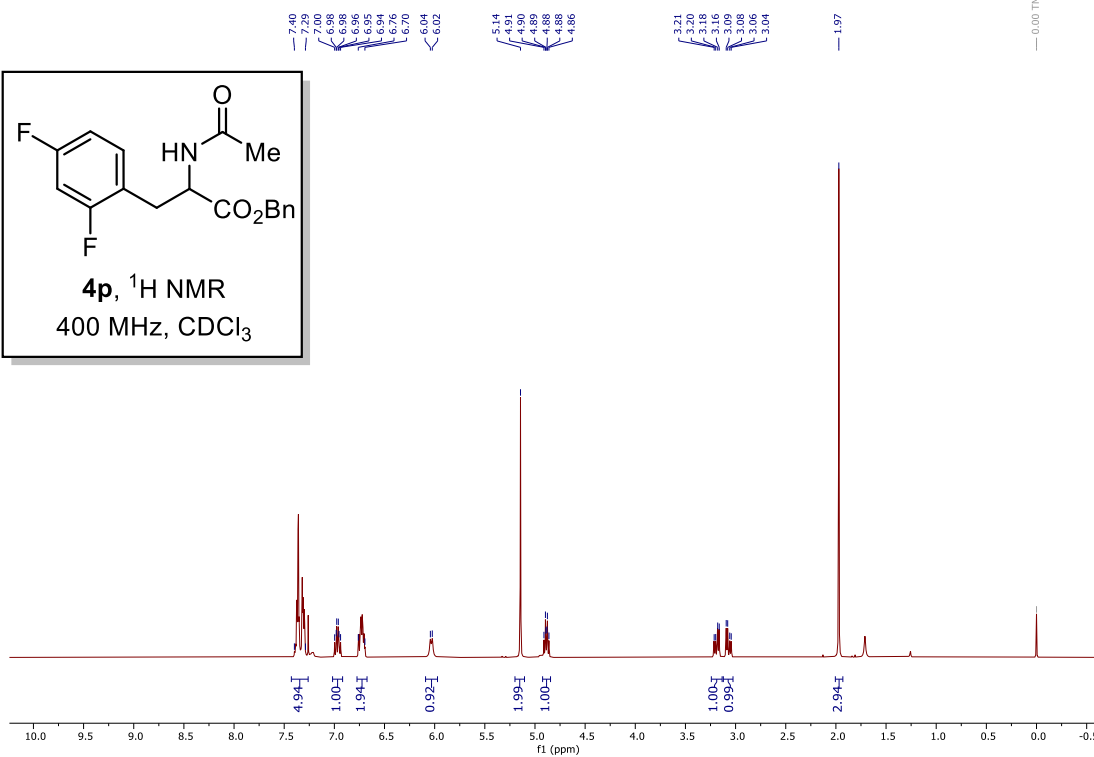
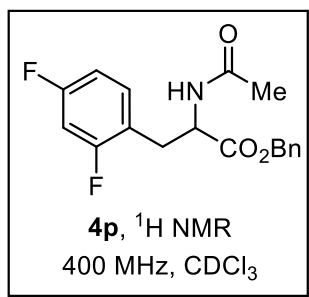


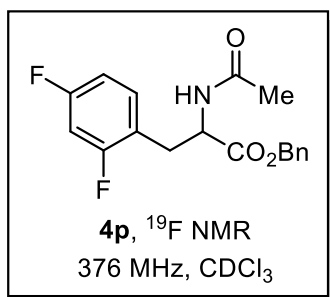












-111.19
-111.21
-111.23
-111.28
-113.12
-113.14
-113.19

