

Tryptophan (W) at position 37 of murine IL-12/IL-23 p40 is mandatory for binding to IL-12Rβ1 and subsequent signal transduction

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Supporting Information

Figure S1: Alignment of murine and human p40.

Figure S2: Dose-response analysis of HIL-23 variants on cellular proliferation of Ba/F3-gp130-mIL-12Rβ1-mIL-23R cells

Figure S3: Purification of cytokines using Strep-Tactin[®]XT 4Flow[®] columns.

Figure S4: Far-UV CD spectroscopy of HIL-23 and HIL-12 variants

Figure S5: Receptor binding of HIL-23 and HIL-12 variants.

Figure S6: Interaction of cytokines with respective receptors.

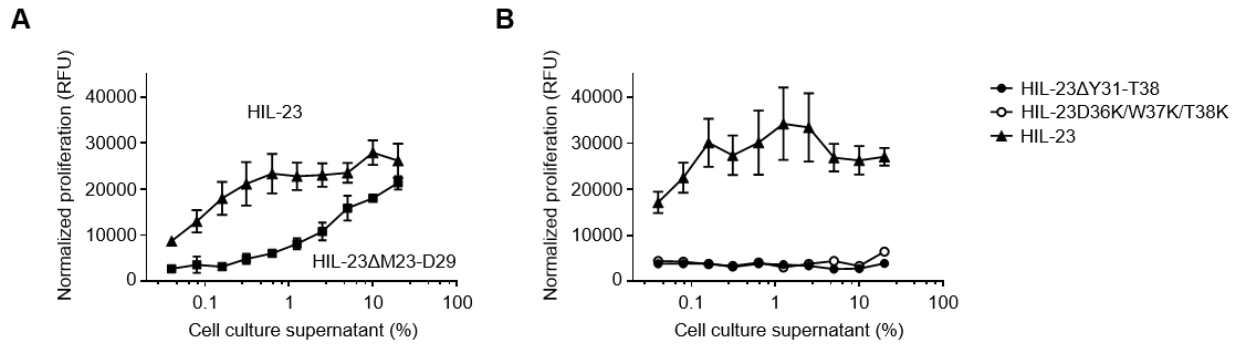


Figure S2: Dose-response analysis of HIL-23 variants on cellular proliferation of Ba/F3-gp130-mIL-12R β 1-mIL-23R cells. The cells were cultured for 3 days in the presence of increasing concentrations of the indicated cytokines (0.04 to 20% conditioned cell culture supernatant of transfected CHO-K1 cells). The results of one representative experiment of three (A) or two (B) are shown. Error bars represent S.D. for technical replicates.

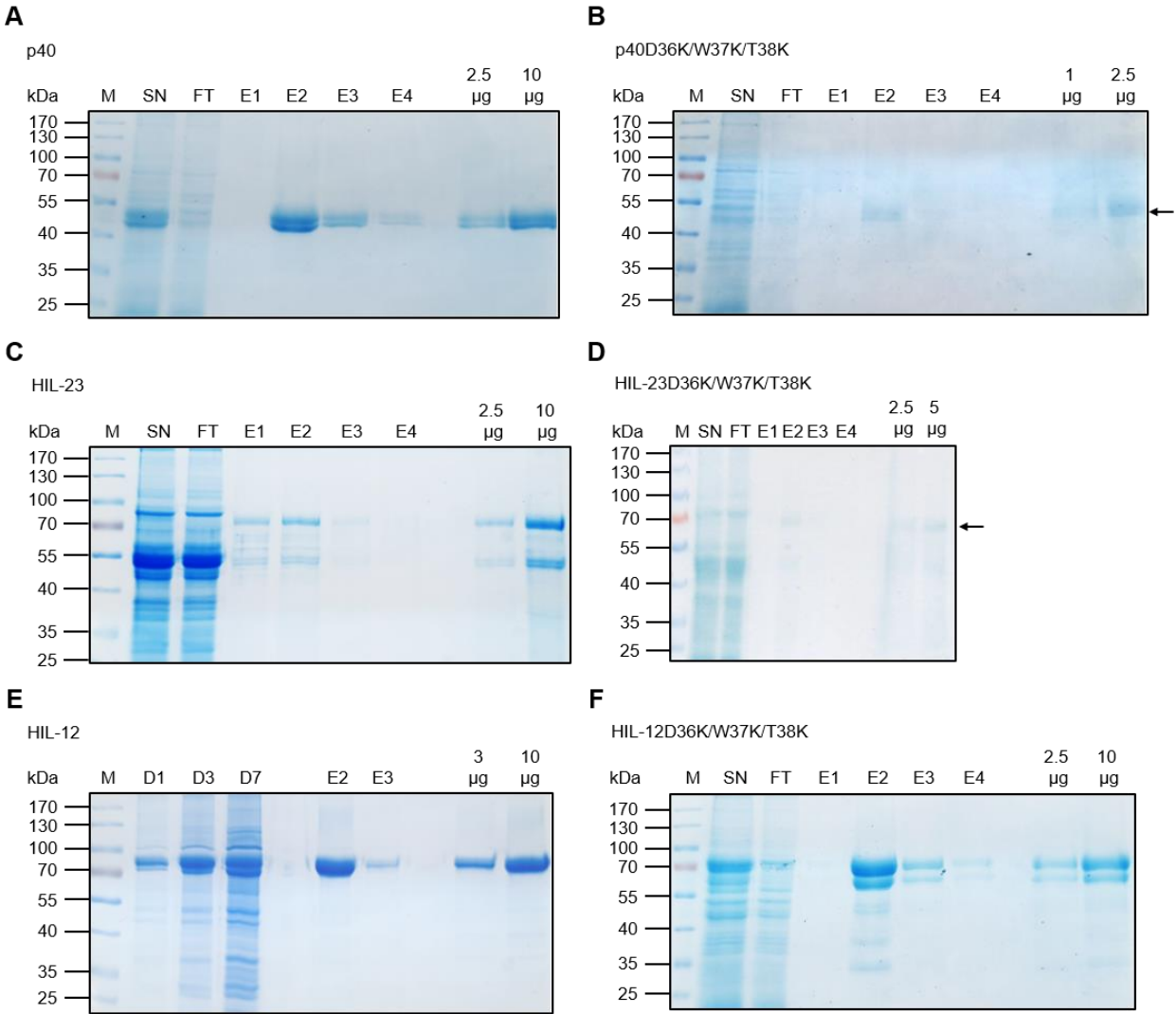


Figure S3: Purification of cytokines using Strep-Tactin[®]XT 4Flow[®] columns. Purity of the Expi cell produced murine p40, p40D36K/W37K/T38K, HIL-12, HIL-12D36K/W37K/T38K, HIL-23 and HIL-23D36K/W37K/T38K was analyzed by SDS-PAGE on reducing gels via Coomassie brilliant blue staining. M, molecular weight marker; SN, cell culture supernatant; FT, flow through; E1-E4, elution fraction 1 to 4; D1, D3, D7, cell culture supernatant of Expi cells from day 1, 3 or 7.

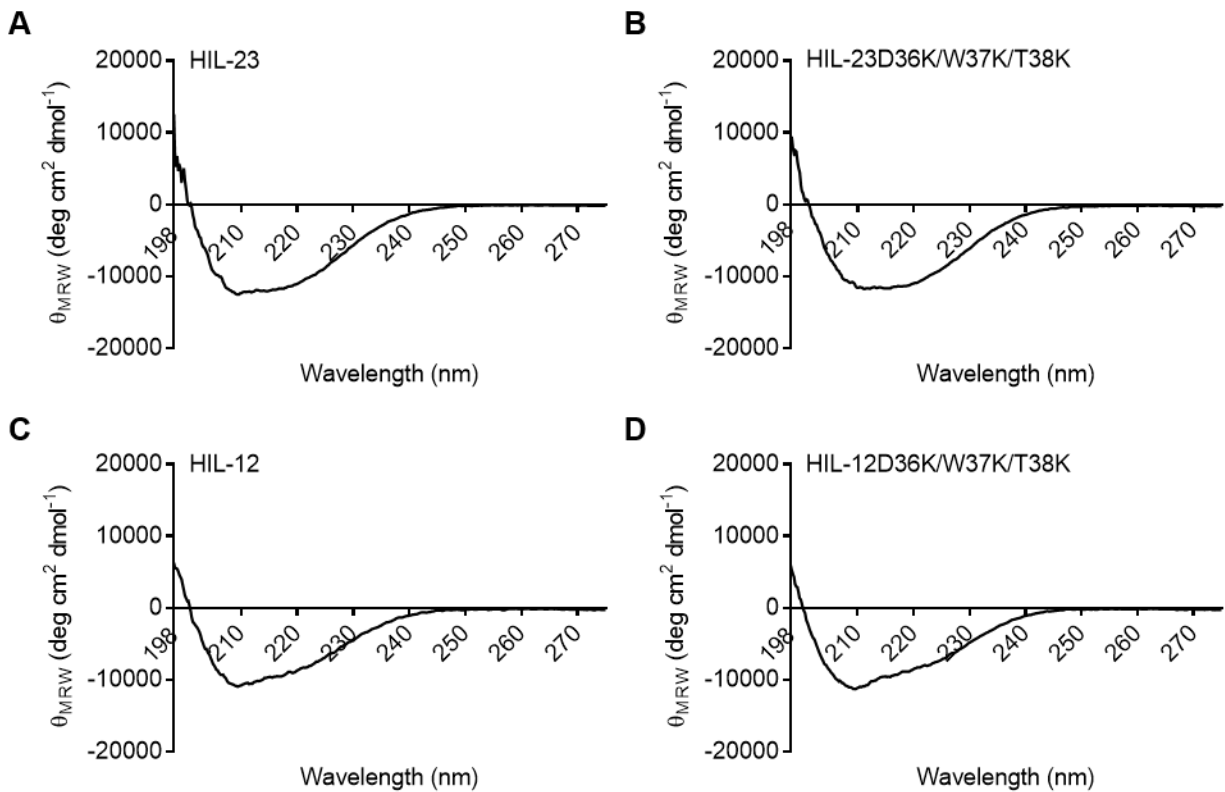


Figure S4: Far-UV CD spectroscopy of HIL-23 and HIL-12 variants. The far-UV spectra of HIL-23 (A) and HIL-23D36K/W37K/T38K (B), or HIL-12 (C) and HIL-12D36K/W37K/T38K (D) indicate that the wild type proteins and their KKK mutants have a highly similar overall secondary structure. Y-axis, θ_{MRW} in $\text{deg} \times \text{cm}^2 \times \text{dmol}^{-1}$; x-axis, wavelength in nm.

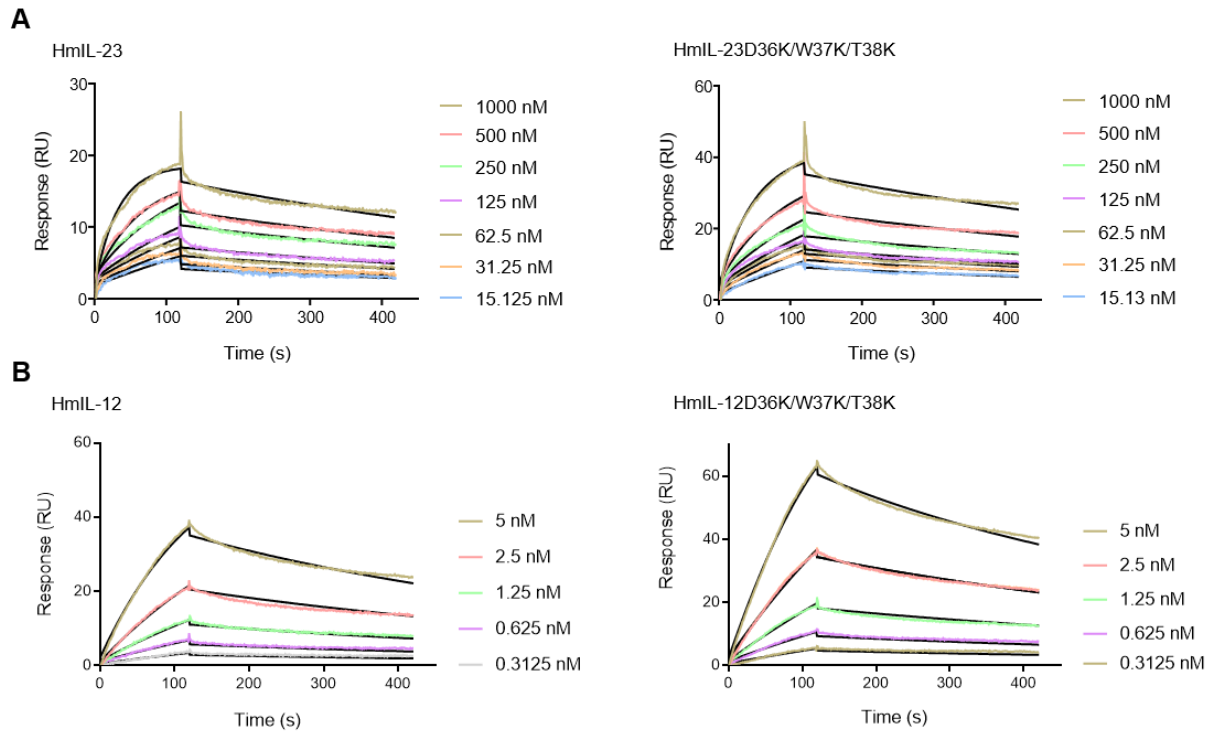


Figure S5. Receptor binding of HIL-23 and HIL-12 variants. A, surface plasmon resonance analysis of HIL-23 binding to IL-23R-Fc. IL-23R-Fc was captured on a ProtA chip and increasing concentrations of HIL-23 and HIL-23D36K/W37K/T38K were injected (15-1000 nM). B, surface plasmon resonance analysis of HIL-12 binding to IL-12R β 2-Fc. IL-12R β 2-Fc was captured on a ProtA chip and increasing concentrations of HIL-12 and HIL-12D36K/W37K/T38K were injected (0.3-5 nM). Sensorgrams in response units (RU) over time are depicted as colored lines, fit data are displayed as black lines.

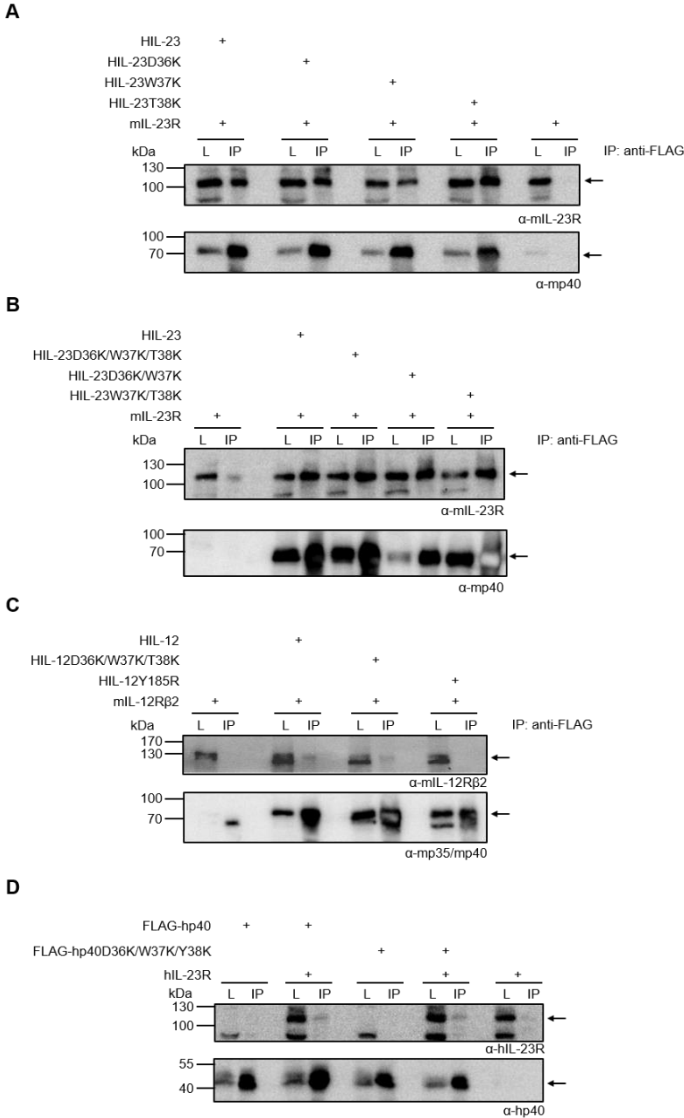


Figure S6: Interaction of cytokines with respective receptors. A, co-IP of FLAG-tagged murine HIL-23 variants (wild-type, D36K, W37K and T38K) and full-length mIL-23R. The position of mIL-23R and HIL-23 variants is indicated by arrows. One of two independent experiments is shown. B, co-IP of FLAG-tagged murine HIL-23 variants (wild-type, D36K/W37K/T38K, D36K/W37K, W37K/T38K) and full-length mIL-23. The position of mIL-23R and HIL-23 variants is indicated by arrows. One of two independent experiments is shown. C, co-IP of FLAG-tagged murine HIL-12 variants (wild-type, D36K/W37K/T38K, Y185R) and full-length mIL-12Rβ2. The position of mIL-12Rβ2 and HIL-12 variants is indicated by arrows. One of two independent experiments is shown. D, co-IP of FLAG-tagged human p40 variants (wild-type, D36K/W37K/Y38K) and full-length hIL-23R. The position of hIL-23R and hp40 variants is indicated by arrows. One of two independent experiments is shown.

References

1. Luo, J., Wu, S. J., Lacy, E. R., Orlovsky, Y., Baker, A., Teplyakov, A., Obmolova, G., Heavner, G. A., Richter, H. T., and Benson, J. (2010) Structural basis for the dual recognition of IL-12 and IL-23 by ustekinumab. *J. Mol. Biol.* **402**, 797-812
2. Glassman, C. R., Mathiharan, Y. K., Jude, K. M., Su, L., Panova, O., Lupardus, P. J., Spangler, J. B., Ely, L. K., Thomas, C., Skiniotis, G., and Garcia, K. C. (2021) Structural basis for IL-12 and IL-23 receptor sharing reveals a gateway for shaping actions on T versus NK cells. *Cell* **184**, 983-999 e924