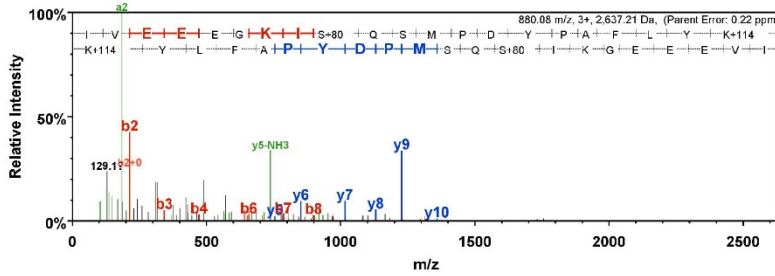




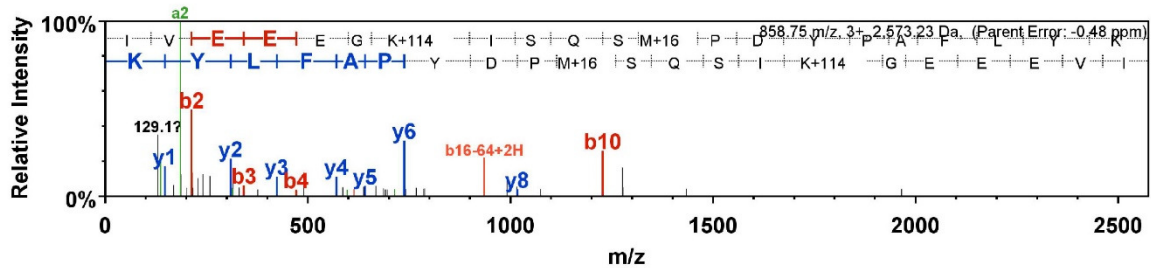
**A** AtRLP44\_GFP (100%), 58,037.7 Da  
 AtRLP44\_GFP  
 2 exclusive unique peptides, 5 exclusive unique spectra, 255 total spectra, 293/523 amino acids (56% coverage)

MTRSHRLLLL	LLLIFQTAQR	LTTADPNDEA	CLKNLRQNLE	DPASNLRNWT
NSVFSNPPCSG	FTSYLPGATC	NNGRIYKLSL	TNLSLRGSIS	PFLSNCTNLQ
SLDLSSNQIS	GVIPPEIQYL	VNLAVLNLSS	NHLSGEITPQ	LALCAYLNVI
DLHDNELSGQ	IQPQLGLLAR	LSAFDVSNNK	LSGQIPTYLS	NRTGNFPRFN
ASSFIGNKGL	YGYPLQEMMM	KSKGLSVMAI	VGIGLGSGLA	SLMISFTGVC
LWLRITEKKI	VEEEEGKISQS	MPDYPAFLYK	VVISMVSKGE	ELFTGVVPII
VELDGDVNGH	KFSVSGEGEG	DATYGLTLTK	FICTTGKLPV	PWPTLVTTLT
YGVQCFSTRYP	DHMKQHDFFK	SAMPEGYVQE	RTIFFKDDGN	YKTRAEVKFE
GDTLVNRIEL	KGIDFKEDGN	ILGHKLEYNY	NSHNVYIMAD	KQKNGIKVNF
KIRHNIEDGS	VQLADHYQQN	TPIGDGPVLL	PDNHVLSLTS	ALSKDPNEKR
DHMLVLEFVT	AAGITLGMDE	LYK		

**B** (K)IVEEEGKISQMPDYPAFLYk(V)



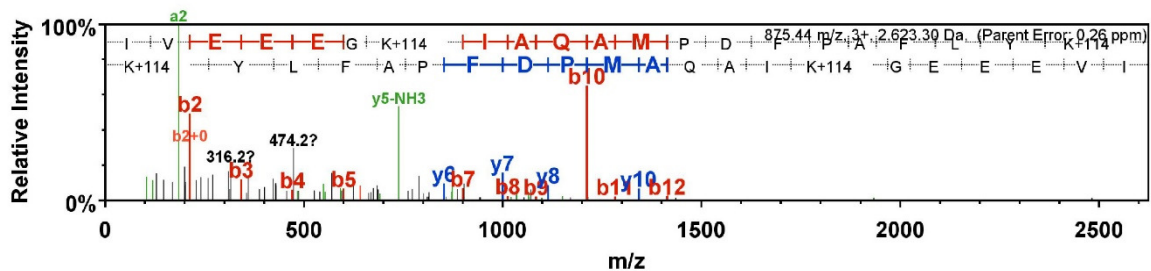
**C** (K)IVEEEGKISQSmPDYPAFLYK(V)



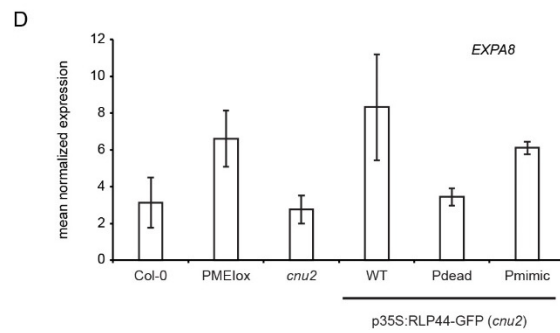
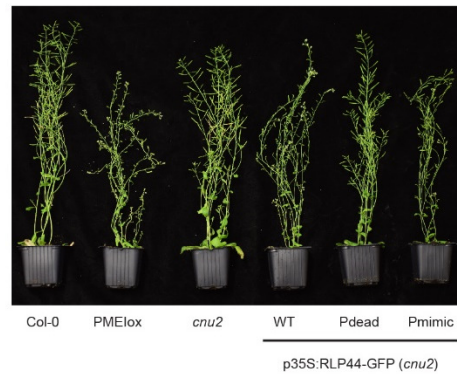
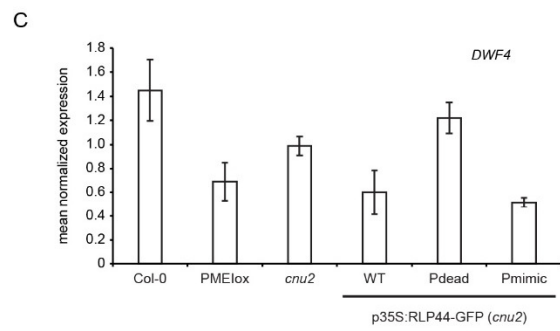
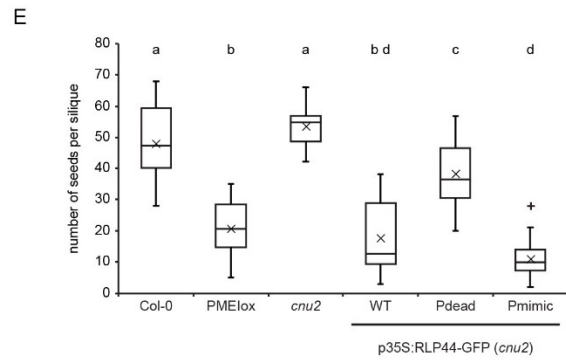
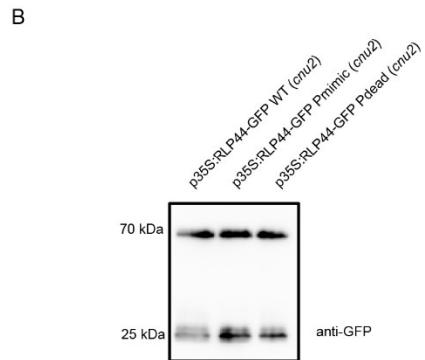
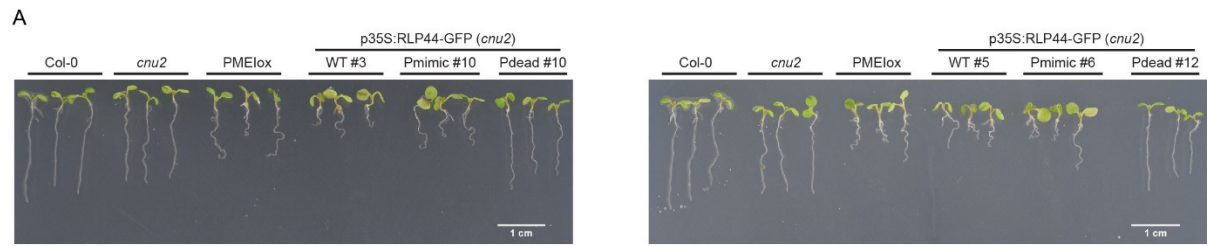
**D** AtRLP44pdead\_GFP (100%), 57,959.7 Da  
 AtRLP44pdead\_GFP  
 2 exclusive unique peptides, 4 exclusive unique spectra, 200 total spectra, 261/523 amino acids (50% coverage)

MTRSHRLLLL	LLLIFQTAQR	LTTADPNDEA	CLKNLRQNLE	DPASNLRNWT
NSVFSNPPCSG	FTSYLPGATC	NNGRIYKLSL	TNLSLRGSIS	PFLSNCTNLQ
SLDLSSNQIS	GVIPPEIQYL	VNLAVLNLSS	NHLSGEITPQ	LALCAYLNVI
DLHDNELSGQ	IQPQLGLLAR	LSAFDVSNNK	LSGQIPTYLS	NRTGNFPRFN
ASSFIGNKGL	YGYPLQEMMM	KSKGLSVMAI	VGIGLGSGLA	SLMISFTGVC
LWLRITAEKKI	VEEEEGKIAQA	MPDFPAFLYK	VVISMVSKGE	ELFTGVVPII
VELDGDVNGH	KFSVSGEGEG	DATYGLTLTK	FICTTGKLPV	PWPTLVTTLT
YGVQCFSTRYP	DHMKQHDFFK	SAMPEGYVQE	RTIFFKDDGN	YKTRAEVKFE
GDTLVNRIEL	KGIDFKEDGN	ILGHKLEYNY	NSHNVYIMAD	KQKNGIKVNF
KIRHNIEDGS	VQLADHYQQN	TPIGDGPVLL	PDNHVLSLTS	ALSKDPNEKR
DHMLVLEFVT	AAGITLGMDE	LYK		

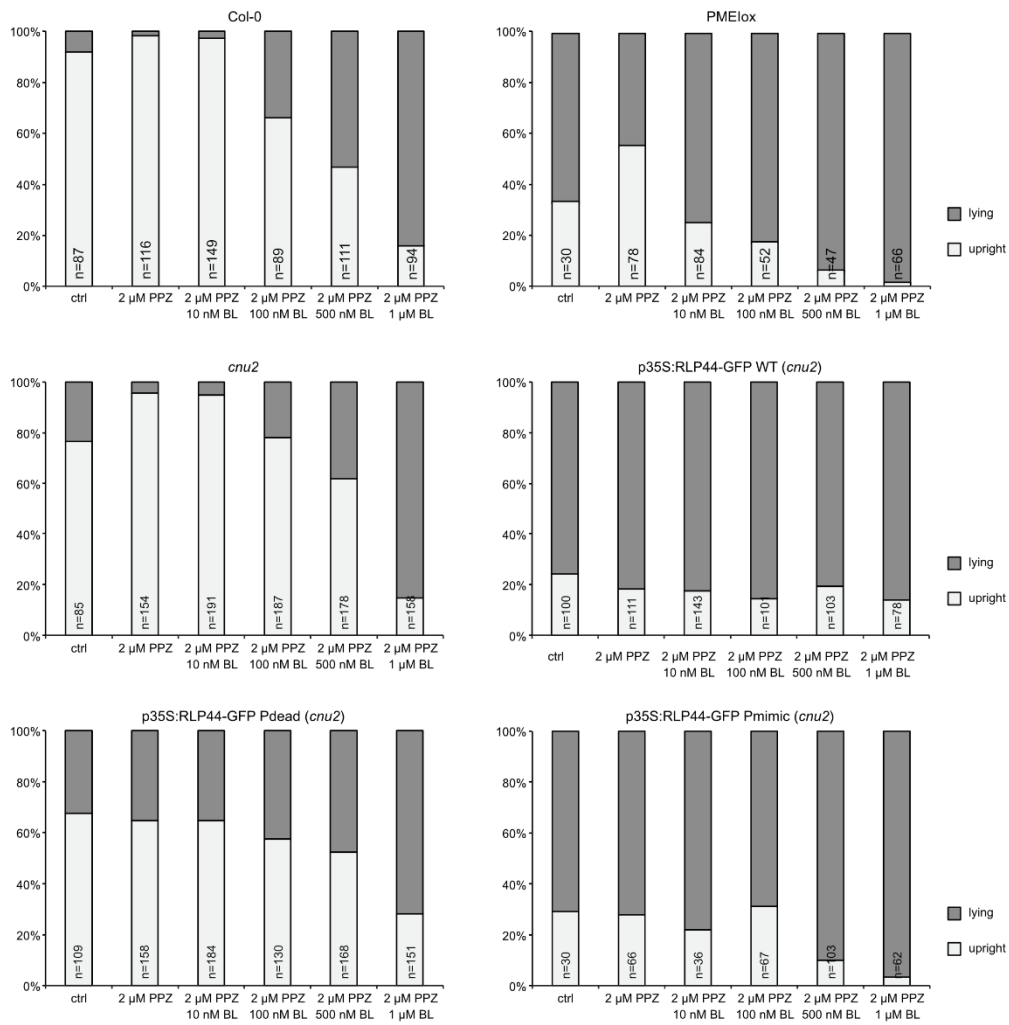
**E** (K)IVEEEGKIAQAMPDFPAFLYk(V)



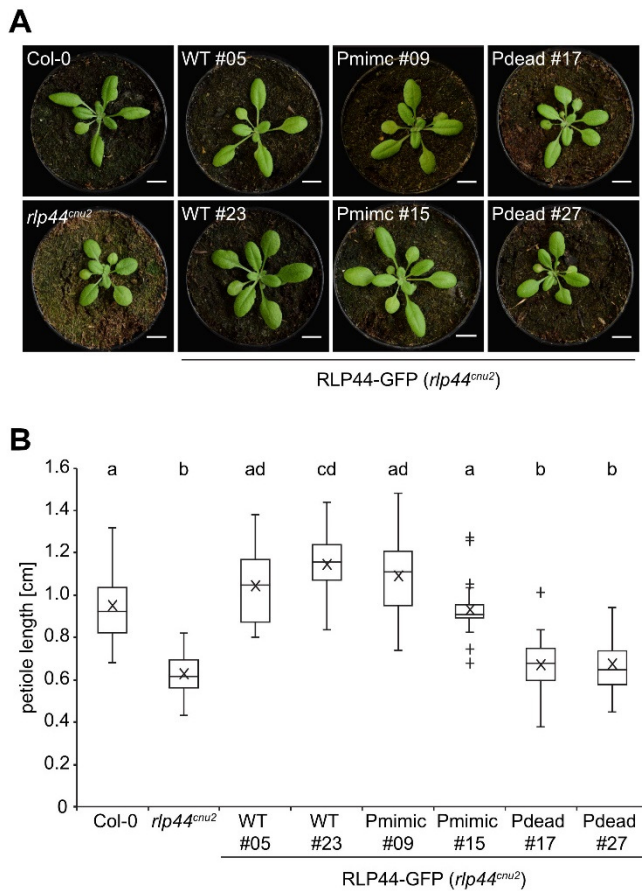
**Fig. S2. Proteomics analysis of RLP44-GFP during transient expression in *N. benthamiana* and trypsin digestion.** Peptide coverage (yellow) and modified amino acids (green) are indicated. Lower panes show spectra of peptides with phosphorylation of S268 and ubiquitination as indicated by double glycine remnant of K266. **(A)** RLP44-GFP WT peptide coverage. **(B)** RLP44-GFP WT peptide with phosphorylation of S268. **(C)** RLP44-GFP WT peptide with ubiquitination of K266. **(D)** RLP44-GFP Pdead peptide coverage. **(E)** RLP44-GFP Pdead peptide with ubiquitination of K266.



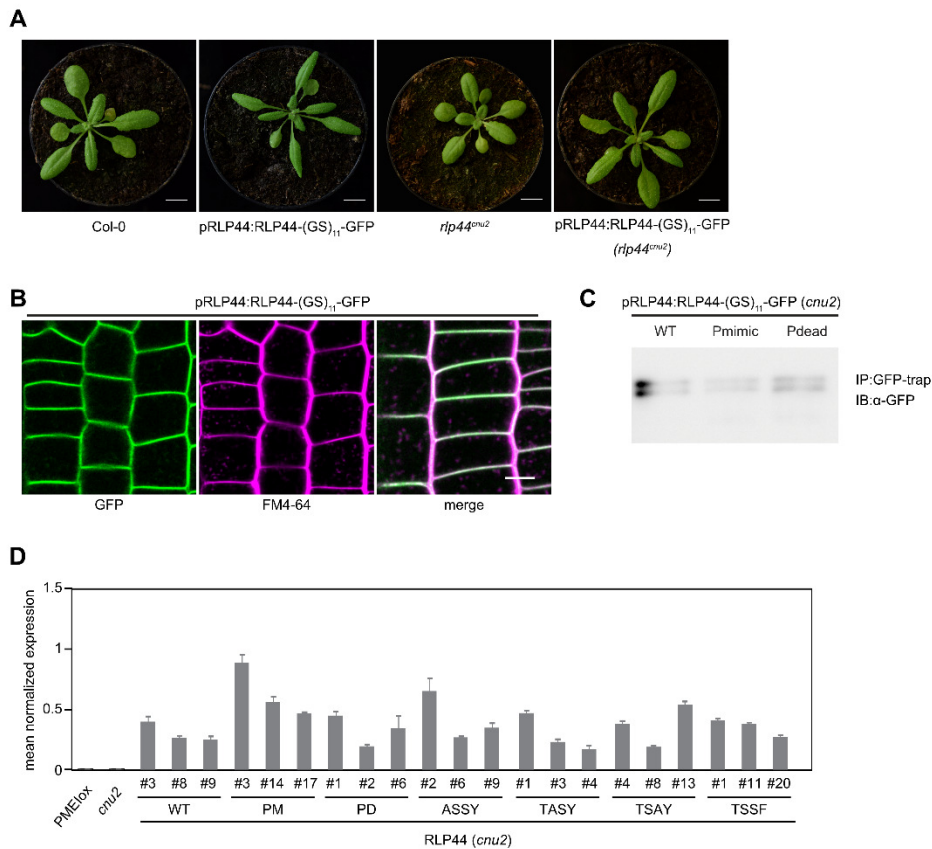
**Fig. S3. Complementation of *gnu2* by RLP44-GFP WT and RLP44-GFP Pmimic, but not by RLP44-GFP Pdead.** (A) Expression of RLP44-GFP WT and Pmimic, but not of Pdead, is able to complement the PMElox suppressor mutant *gnu2* and leads to recovery of the PMElox root waving phenotype in seedlings and contorted leaf arrangement in adult plants (see also Fig. 1). (B) Lines with comparable RLP44-GFP expression levels were selected based on Western Blot analysis with anti-GFP antiserum. (C) Quantitative Real-Time PCR analysis of the BR signalling marker gene *DWF4*, n=3. (D) Quantitative Real-Time PCR analysis of the BR signalling marker gene *EXPANSIN8*, n=3. (E) Analysis of seed yield in Col-0, PMElox, *gnu2*, and *gnu2* complementation lines. Boxes indicate range from 25<sup>th</sup> to 75<sup>th</sup> percentile, horizontal line indicates the median, whiskers indicate data points within 1.5 times the interquartile range. Markers above whiskers indicate outliers, n = 18. Lettering indicates statistically significant difference according to Tukey's post-hoc test following one-way ANOVA.



**Fig. S4. RLP44-GFP WT and Pmimic, but not RLP44-GFP Pdead restore agravitropic growth of *cnu2* along the surface of agar plates in the dark as observed in PMElox.** Endogenous brassinosteroids were depleted with PPZ treatment to test sensitivity to increasing amounts of epi-brassinolide (BL)



**Fig. S5. RLP44-GFP Pdead is unable to complement the petiole phenotype of *rlp44<sup>cnu2</sup>*.** (A) Rosette phenotype of Col-0, *rlp44<sup>cnu2</sup>*, and *rlp44<sup>cnu2</sup>* complementation lines in long day conditions. (B) Quantification of petiole length in lines depicted in A). Boxes indicate range from 25<sup>th</sup> to 75<sup>th</sup> percentile, horizontal line indicates the median, whiskers indicate data points within 1.5 times the interquartile range. Markers above whiskers indicate outliers, n = 21. Lettering indicates statistically significant difference according to Tukey's post-hoc test following one-way ANOVA.



**Fig. S6. Overall charge, rather than specific phosphosites modulate RLP44 function.** (A) RLP44 overexpression phenotype (see Wolf et al., 2014) of hyperphosphorylated RLP44 under control of its own promoter. (B) Plasma membrane localization of hyperphosphorylated pRLP44:RLP44-(GS)<sub>11</sub>-GFP. (C) RLP44 Pdead and Pmimic mutants with serine-rich linker show slower migrating band similar to the WT version, suggesting linker phosphorylation. (D) Expression of *RLP44* in transgenic lines evaluated in Figure 10 using qPCR (n=3 for each line).



**Table S1.** Mutants and transgenic lines used in this study.

Mutant/transgenic line	
<i>rlp44<sup>cnu2</sup></i>	Wolf <i>et al.</i> , 2014
<i>cnu2</i>	Wolf <i>et al.</i> , 2014
<i>bri1-null</i>	Jaillais <i>et al.</i> , 2011a
<i>pskr1-3 pskr2-1</i>	Kutschmar <i>et al.</i> , 2009
<i>p35S:RLP44-GFP WT (cnu2)</i>	This study
<i>p35S:RLP44-GFP Pdead (cnu2)</i>	This study
<i>p35S:RLP44-GFP Pmimic (cnu2)</i>	This study
<i>p35S:RLP44-GFP WT</i>	This study
<i>p35S:RLP44-GFP Pdead</i>	This study
<i>p35S:RLP44-GFP Pmimic</i>	This study
<i>p35S:RLP44-GFP WT (rlp44<sup>cnu2</sup>)</i>	This study
<i>p35S:RLP44-GFP Pdead (rlp44<sup>cnu2</sup>)</i>	This study
<i>p35S:RLP44-GFP Pmimic (rlp44<sup>cnu2</sup>)</i>	This study
<i>p35S:RLP44-GFP WT (bri1-null)</i>	Holzwart <i>et al.</i> , 2018
<i>p35S:RLP44-GFP Pdead (bri1-null)</i>	This study
<i>p35S:RLP44-GFP Pmimic (bri1-null)</i>	This study
<i>p35S:RLP44-GFP WT (pskr1-3 pskr2-1)</i>	This study
<i>p35S:RLP44-GFP Pdead (pskr1-3 pskr2-1)</i>	This study
<i>pESTR:amiT-TML</i>	Gadeyne <i>et al.</i> , 2014
<i>p35S:RLP44-GFP WT (pESTR:amiT-TML)</i>	This study
<i>p35S:RLP44-GFP Pdead (pESTR:amiT-TML)</i>	This study
<i>pRLP44:RLP44 WT (cnu2)</i>	This study
<i>pRLP44:RLP44 Pdead (cnu2)</i>	This study
<i>pRLP44:RLP44 Pmimic (cnu2)</i>	This study
<i>pRLP44:RLP44 T256A (cnu2)</i>	This study
<i>pRLP44:RLP44 S268A (cnu2)</i>	This study
<i>pRLP44:RLP44 S270A (cnu2)</i>	This study
<i>pRLP44:RLP44 Y274F (cnu2)</i>	This study
<i>pRLP44:RLP44-(GS)11-GFP</i>	Holzwart <i>et al.</i> , 2018
<i>pRLP44:RLP44-(GS)11-GFP WT (cnu2)</i>	This study
<i>pRLP44:RLP44-(GS)11-GFP Pmimic (cnu2)</i>	This study
<i>pRLP44:RLP44-(GS)11-GFP (Pdead (cnu2))</i>	This study

**Table S2.** Oligonucleotides used in this study

Primer No.	Primer name	Sequence (5' → 3')	target
SW660	RLP44_GW_L	GGGGACAAGTTTGTACA <del>AAAAAGCAGGCTATGA</del> CAAGGAGTCACCGTTAC	At3g49750
SW670	RLP_GW_WToY_R	GGGGACCACTTTGTACAAGAAAGCTGGGTGTAA TCAGGCATAGATTGAC	At3g49750
SW666	RLP44_SDMT25_6A_F	gtttatggttgaggattgctgagaagaagattgtg	At3g49750
SW667	RLP44_SDMT25_6A_R	caacaatcttctcagcaatcctcaaccataaac	At3g49750
SW668	RLP_S268,270A_Y274F_Rneu	GGGGACCACTTTGTACAAGAAAGCTGGGT <del>agaaa</del> tcaggcatagcttgagcaatcttaccttcttcaac	At3g49750
SW672	RLP44_SDMT25_6E_F	GAGGATTGAAGAGAAGAAGATTGTTGAAGAAG	At3g49750
SW673	RLP44_SDMT25_6E_R	ATCTTCTTCTCTTCAATCCTCAACCATAAAC	At3g49750
SW671	RLP_SSY-EEE_R	GGGGACCACTTTGTACAAGAAAGCTGGGT <del>agaaa</del> tcaggcatagattgactaatc	At3g49750
SW661	RLP44_Y274F_R	GGGGACCACTTTGTACAAGAAAGCTGGGT <del>tttcatc</del> aggcatttctgttcaatcttaccttcttcaac	At3g49750
SW662	RLP44_S268A_R	GGGGACCACTTTGTACAAGAAAGCTGGGT <del>tagtaat</del> caggcatagcttgactaatcttaccttctc	At3g49750
SW663	RLP44_S270A_R	GGGGACCACTTTGTACAAGAAAGCTGGGT <del>tagtaat</del> caggcatagattgagcaatcttaccttcttcaac	At3g49750
SW2446	RLP44-if-f	tgAagcttGGTCTCaGGCTcAATGACAAGGAGTCAC CGG	At3g49750
SW2447	RLP44WT-if-r	atGGCACCCGCCCTGCTCcGTAATCAGGCATAG ATTG	At3g49750
SW2448	GAGAGA-GFP-if-f	gGAGCAGGGGCGGGTGCC	GFP
SW2449	GAGAGA-GFP-if-r	cgaGAATTcGGTCTCaCTGAttactgtacagctcgtcc	GFP
SW2450	RLP44TS-AY-if-r	atGGCACCCGCCCTGCTCcGTAATCAGGCATAG cTTG	At3g49750
SW2452	RLP44TS-SF-if-r	atGGCACCCGCCCTGCTCcGaAATCAGGCATAG ATTG	At3g49750
SW2454	RLP44AA-AF-if-r	atGGCACCCGCCCTGCTCcGaAATCAGGCATAG cTTG	At3g49750
SW2455	RLP44EE-EE-if-r	atGGCACCCGCCCTGCTCccttATCAGGCATttcTT G	At3g49750
SW3000	RLP44prom_F	GGGGACAAGTTTGTACA <del>AAAAAGCAGGCTTTTG</del> CGATATTTTTGGCTGTC	At3g49750
SW3001	RLP44stop_R	GGGGACCACTTTGTACAAGAAAGCTGGGTTTTA GTAATCAGGCATAGATTGACT	At3g49750
SW3002	RLP44_TASY_stop_R	GGGGACCACTTTGTACAAGAAAGCTGGGTTTTA GTAATCAGGCATAGATTG	At3g49750
SW3003	RLP44_PM_stop_R	GGGGACCACTTTGTACAAGAAAGCTGGGTTTTA TTCATCAGGCATTTCTTGTTCAATCTT	At3g49750
SW3004	RLP44_PD_stop_R	GGGGACCACTTTGTACAAGAAAGCTGGGTTTTA GAAATCAGGCATAGCTTGAGCAATCTT	At3g49750
SW3005	RLP44_TSAY_stop_R	GGGGACCACTTTGTACAAGAAAGCTGGGTTTTA GTAATCAGGCATAGCTTGACTAATCTT	At3g49750
SW3006	RLP44_TSSF_stop_R	GGGGACCACTTTGTACAAGAAAGCTGGGTTTTA GAAATCAGGCATAGATTGACTAATC	At3g49750
SW1179	RLP44_GG_F	AACAGGTCTCAGGCTCAATGACAAGGAGTCACC GGTTA	At3g49750
SW1205	RLP44_GG_R	AACAGGTCTCACTGAGTAATCAGGCATAGATTG AC	At3g49750
SW1367	RLP44_PD_GG_C_R	AACAGGTCTCACTGAGAAATCAGGCATAGCTTG	At3g49750

SW1368	RLP44 PM GG C R	AACAGGTCTCACTGATTCATCAGGCATTTCTTG	At3g49750
SW503	RLP44- 4 CAPS F	AATCTACAAACTCTCACTCAC	At3g49750
SW504	RLP44- 4 CAPS R	CTGACCCGGATAATTCGTTATC	At3g49750
SW1377	GK-O8409	ATATTGACCATCATACTCATTGC	
SW1378	GK-134E10 F	TAGCGGAAACAAAATCAGTGG	At4g39400
SW1379	GK-134E10 R	TCGTTCCATTGAAGAGATTGG	At4g39400
SW1754	pskr1-3 F	CTCGCTTTCTGGTATGACGAG	At2g02220
SW1746	pskr1-3 R	TCCGAAACTATACACATCGCC	At2g02220
SW1984	pskr2-1 F	TTCTTAGACTGTTTGGCTCGG	At5g53890
SW1985	pskr2-1 R	GCGTTACAAACATGCAACAAG	At5g53890
SW230	LBb1.3	ATTTTGCCGATTTTCGGAAC	
SW905	attB1	ACAAGTTTGTACAAAAAGCAGGCT	
SW906	attB2	ACCACTTTGTACAAGAAAGCTGGGT	
SW1202	pGG-Bdummy F	GTATTCAGTCGACTGGTACCAAC	
SW1137	pGGA/C000 R	CAGATTGTACTGAGAGTGCACC	
SW521	AtEXP8 F	CCGAAATAACTAACCCCTCCTC	At2g40610
SW522	AtEXP8 R	TAGCCACAAGCTCCGCCAT	At2g40610
SW803	DWF4 F	CAACAGCAAACAACGGAGCG	At3g50660
SW804	DWF4 R	TCTGAACCAGCACATAGCCTTG	At3g50660
SW1015	Clath F	TCGATTGCTTGGTTTGAAGAT	At1g10730
SW1016	Clath R	GCACTTAGCGTGGACTCTGTTTGC	At1g10730
SW612	RLP44COD2 F	TCAGATTCCGCAGCAATTAG	At3g49750
SW613	RLP44COD2 R	TCCTGCAACGGATAACCATA	At3g49750
	ACT2 F	CAGTGTCTGGATCGGTGGTT	At3g18780
	ACT2 R	TGAACGATTCCTGGACCTGC	At3g18780

**Table S3.** Overview of constructs generated with GreenGate cloning.

<b>pSW362</b>		<b>pRLP44:RLP44-(GS)<sub>11</sub>-GFP WT</b>		
	<i>Name</i>	<i>Internal name</i>	<i>Source</i>	<i>Primers</i>
"Promoter" module	pRLP44	pSW299	Holzward et al., 2018	
"N-tag" module	B-dummy	pGGB003	Lampropoulos et al., 2013	
"CDS" module	RLP44	pSW334	Holzward et al., 2018	SW1179-1205
"C-tag" module	(GS) <sub>11</sub> -GFP	PGGD001	Lampropoulos et al., 2013	
"Terminator" module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
"Resistance" module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW567</b>		<b>pRLP44:RLP44-(GS)<sub>11</sub>-GFP Pmimic</b>		
	<i>Name</i>	<i>Internal name</i>	<i>Source</i>	<i>Primers</i>
"Promoter" module	pRLP44	pSW299	Holzward et al., 2018	
"N-tag" module	B-dummy	pGGB003	Lampropoulos et al., 2013	
"CDS" module	RLP44pmimic	pSW519	This study	SW1179-1368
"C-tag" module	(GS) <sub>11</sub> -GFP	PGGD001	Lampropoulos et al., 2013	
"Terminator" module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
"Resistance" module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW566</b>		<b>pRLP44:RLP44-(GS)<sub>11</sub>-GFP Pdead</b>		
	<i>Name</i>	<i>Internal name</i>	<i>Source</i>	<i>Primers</i>
"Promoter" module	pRLP44	pSW299	Holzward et al., 2018	
"N-tag" module	B-dummy	pGGB003	Lampropoulos et al., 2013	
"CDS" module	RLP44pdead	pSW518	This study	SW1179-1367
"C-tag" module	(GS) <sub>11</sub> -GFP	PGGD001	Lampropoulos et al., 2013	
"Terminator" module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
"Resistance" module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW1027</b>		<b>pRLP44:RLP44-GAGAGA-GFP WT</b>		
"Promoter" module	pRLP44	pSW299	Holzward et al., 2018	
"N-tag" module	B-dummy	pGGB003	Lampropoulos et al., 2013	
"CDS" module	RLP44-GAGAGA-GFP	pSW999	This study	SW2446-2449
"C-tag" module	D-dummy	pGGD002	Lampropoulos et al., 2013	
"Terminator" module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
"Resistance" module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW1019</b>		<b>pRLP44:RLP44-GAGAGA-GFP Pmimic</b>		
"Promoter" module	pRLP44	pSW299	Holzward et al., 2018	
"N-tag" module	B-dummy	pGGB003	Lampropoulos et al., 2013	
"CDS" module	RLP44-GAGAGA-GFP Pmimic	pSW1018	This study	SW2446-2455, 2448+2449
"C-tag" module	D-dummy	pGGD002	Lampropoulos et al., 2013	
"Terminator" module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
"Resistance" module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW1017</b>		<b>pRLP44:RLP44-GAGAGA-GFP Pdead</b>		
"Promoter" module	pRLP44	pSW299	Holzward et al., 2018	
"N-tag" module	B-dummy	pGGB003	Lampropoulos et al., 2013	

“CDS” module	RLP44-GAGAGA-GFP Pdead	pSW1007	This study	SW2446+2454, 2448+2449
“C-tag” module	D-dummy	pGGD002	Lampropoulos et al., 2013	
“Terminator” module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
“Resistance” module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW1026</b>	<b>pRLP44:RLP44-GAGAGA-GFP ASSY</b>			
“Promoter” module	pRLP44	pSW299	Holzwardt et al., 2018	
“N-tag” module	B-dummy	pGGB003	Lampropoulos et al., 2013	
“CDS” module	RLP44-GAGAGA-GFP ASSY	pSW1000	This study	SW2446+2447, 2448+2449
“C-tag” module	D-dummy	pGGD002	Lampropoulos et al., 2013	
“Terminator” module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
“Resistance” module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW1028</b>	<b>pRLP44:RLP44-GAGAGA-GFP TASY</b>			
“Promoter” module	pRLP44	pSW299	Holzwardt et al., 2018	
“N-tag” module	B-dummy	pGGB003	Lampropoulos et al., 2013	
“CDS” module	RLP44-GAGAGA-GFP TASY	pSW1002	This study	SW2446+2447, 2448+2449
“C-tag” module	D-dummy	pGGD002	Lampropoulos et al., 2013	
“Terminator” module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
“Resistance” module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW1014</b>	<b>pRLP44:RLP44-GAGAGA-GFP TSAY</b>			
“Promoter” module	pRLP44	pSW299	Holzwardt et al., 2018	
“N-tag” module	B-dummy	pGGB003	Lampropoulos et al., 2013	
“CDS” module	RLP44-GAGAGA-GFP TSAY	pSW1004	This study	SW2446+2450, 2448+2449
“C-tag” module	D-dummy	pGGD002	Lampropoulos et al., 2013	
“Terminator” module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
“Resistance” module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	
<b>pSW1015</b>	<b>pRLP44:RLP44-GAGAGA-GFP TSSF</b>			
“Promoter” module	pRLP44	pSW299	Holzwardt et al., 2018	
“N-tag” module	B-dummy	pGGB003	Lampropoulos et al., 2013	
“CDS” module	RLP44-GAGAGA-GFP TSSF	pSW1005	This study	SW2446+2452, 2448+2449
“C-tag” module	D-dummy	pGGD002	Lampropoulos et al., 2013	
“Terminator” module	tUBQ10	pGGE009	Lampropoulos et al., 2013	
“Resistance” module	SulfR	pGGF006	Lampropoulos et al.,2013	
Destination vector		pGGZ0001	Lampropoulos et al.,2013	