

Peer Review Information

Journal: Nature Structural and Molecular Biology

Manuscript Title: GPCR activation mechanisms across classes and macro/microscales

Corresponding author name(s): David Gloriam

Reviewer Comments & Decisions:

Decision Letter, initial version:
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2nd Jun 2021

Dear David,

Thank you again for submitting your manuscript "GPCR activation mechanisms across classes and macro/microscales". I apologize for the delay in responding, which, as you already know, resulted from the difficulty in obtaining suitable referee reports. Nevertheless, we now have comments (below) from the 2 reviewers who evaluated your paper. In light of those reports, we remain interested in your study and would like to see your response to the comments of the referees, in the form of a revised manuscript.

You will see that the reviewers (both experts in GPCR structure/function) are positive about the interest and quality of the study, and reviewer 2 recommends publication as is. However, reviewer 1 made several useful suggestions to improve the presentation of the findings and requested experimental validation of key results, which we agree would greatly strengthen the manuscript.

We are committed to providing a fair and constructive peer-review process. Do not hesitate to contact us if there are specific requests from the reviewers that you believe are technically impossible or unlikely to yield a meaningful outcome.

Please be sure to address all concerns of the referees in full in a point-by-point response and highlight all changes in the revised manuscript text file. If you have comments that are intended for editors only, please include those in a separate cover letter.

We expect to see your revised manuscript within 12 weeks. If you cannot send it within this time, please contact us to discuss an extension; we would still consider your revision, provided that no similar work has been accepted for publication at NSMB or published elsewhere.

As you already know, we put great emphasis on ensuring that the methods and statistics reported in our papers are correct and accurate. As such, if there are any changes that should be reported, please submit an updated version of the Reporting Summary along with your revision.

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We look forward to seeing the revised manuscript and thank you for the opportunity to review your work.

Kind regards,
Florian

Florian Ullrich, Ph.D.
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Nature Structural & Molecular Biology
ORCID 0000-0002-1153-2040

Reviewers' Comments:

Reviewer #1:

Remarks to the Author:

Hauser et al. used a newly developed online GPCR structure analysis platform to investigate the GPCR activation mechanisms. The authors analyzed all available 488 structures from different GPCR classes and presented a molecular map for GPCR activation, ligand binding, and G protein coupling. The newly developed online GPCR structure analysis platform is useful and easy to navigate. Using this platform, the authors provided macro and micro switches for GPCR activation. The results suggested new activation switches along with activation switches that have been suggested by other studies. Overall, the manuscript suggests novel findings that would provide valuable information for the structural mechanism of GPCR activation. There are, however, a few issues that should be addressed to improve the manuscript.

Major comments

1. Although they have provided the detailed information as supplementary data, It would be nice if the authors provide more detailed information about activation structures (agonist-bound or G protein-bound or arrestin-bound states) analyzed in the manuscript text.
2. Would the analysis results provide different macro or micro switches that discern the G protein binding and arrestin binding or selectivity for G protein subtypes?
3. It has been suggested that there are sequential conformational changes during GPCR activation and G protein coupling, and the complex structures are final stage structures (DOI: 10.1016/j.cell.2019.04.022, 10.1016/j.cell.2019.04.021). Please consider discussing this issue along

with the analysis results of the manuscript.

4. Please provide experimental evidence for the newly suggested GPCR activation switches.

Minor comments

1. On page 4 and Fig. 3a class F, should it be 52% and 48%, not 62% (line 25) and 38%?
2. Please consider rewriting a sentence on page 5 line 15, "We next ... pairs confirms."

Reviewer #2:

Remarks to the Author:

This manuscript by Kooistra et al., Babu and Gloriam provides an integrated and comprehensive analysis of the movements and transduction pathways shared and different among the four major GPCR families as they transit from inactivated to activated states. A great strength of the work is their use of distance pairs, rather than more traditional superposition of structures and RMSDs, to find conserved interactions and switches among inactive- and active-stabilizing residue pairs, and switch residues implicated in both. A key contribution will be the reduction to groups of these residues that define the states among each of the GPCR families. From these studies both granular analyses of transduction pathways emerge, as do more general features (e.g., that residue rotamer switches are rare, the centrality of TM Helix 3, the ubiquitousness of helix rotation, often over rotamer changes, and the greater similarities of among receptors within families than within active vs inactive states). Many of these will provide guidance to the community to drive specific research questions (e.g., what state is my receptor in, how to I drive that/interrupt that by mutation, what is such-and-so ligand doing...?). I thus find this a strong manuscript that will interest the community, and I support publication. As I have seen it previously (and liked it then), and since most of my key critiques have been addressed, I think the manuscript can be published as is.

Author Rebuttal to Initial comments

We thank all Reviewers for the insightful and positive comments which have improved the manuscript and online resources. We respond to each comment below. In addition, we have also incorporated recent structural templates for classes C and F (class A and B1 already had strong coverage) and updated all associated figures and text accordingly. We look forward to your opinion on the revised manuscript.

Reviewer #1:

Hauser et al. used a newly developed online GPCR structure analysis platform to investigate the GPCR activation mechanisms. The authors analyzed all available 488 structures from different GPCR classes and presented a molecular map for GPCR activation, ligand binding, and G protein coupling.

The newly developed online GPCR structure analysis platform is useful and easy to navigate. Using this platform, the authors provided macro and micro switches for GPCR activation. The results suggested new activation switches along with activation switches that have been suggested by other studies. Overall, the manuscript suggests novel findings that would provide valuable information for the structural mechanism of GPCR activation. There are, however, a few issues that should be addressed to improve the manuscript.

We thank the reviewer for the nice summary of the paper and its scope providing both macro and micro switches for GPCR activation, including many new activation switches (not reported for classes B1, C and F before). Furthermore, we highly appreciate the confirmation of that the novel findings provide valuable information about the structural mechanism of GPCR activation, and of the utility of the online structure analysis platform.

Major comments

1. Although they have provided the detailed information as supplementary data, It would be nice if the authors provide more detailed information about activation structures (agonist-bound or G protein-bound or arrestin-bound states) analyzed in the manuscript text.

We have added more detailed information about active structures in all places where specific receptors are discussed or where the type of complex could be expected to affect the conclusions. As it is not possible to list all complexed proteins and ligands for all active state receptors investigated, for the other places of the text we have added additional references to the relevant supplementary data. All representative active state structures used for the analyses are in the G protein-bound active state. No arrestin-bound states have gone into the analysis of the activation state comparisons.

2. Would the analysis results provide different macro or micro switches that discern the G protein binding and arrestin binding or selectivity for G protein subtypes?

This particular paper focuses on the common activation mechanisms that govern the stabilization of an inactive and active state, respectively. It solely uses G protein-bound templates, as these are at the top of the signaling cascade and have a shared structural scaffold allowing comparable active receptor

states, which is a prerequisite of our comparative structure analysis approach. Other studies have described molecular mechanisms (DOI: 10.1038/s41586-018-0077-3, 10.1038/s41594-018-0071-3) and motifs (DOI: 10.1038/s41467-019-09204-y, DOI: 10.1016/j.cell.2017.07.002) involved in GPCR-mediated arrestin activation.

It would indeed be intriguing to investigate G protein family specific determinants of binding and activation. However, there are not sufficient templates yet to do that in a comprehensive comparative fashion across the GPCR classes and G protein families. Such an analysis would, conversely to this, not identify common activation determinants for a given GPCR class but those unique to a G protein family. Such an analysis would therefore have a larger focus on determinants of G protein binding selectivity, which has also been studied previously (e.g. DOI: 10.1038/nature22070) using a sequence-based approach. We suggest that structure-based comparisons across GPCR classes and G protein families are revisited when the coverage by structural templates is improved.

3. It has been suggested that there are sequential conformational changes during GPCR activation and G protein coupling, and the complex structures are final stage structures (DOI: 10.1016/j.cell.2019.04.022, 10.1016/j.cell.2019.04.021). Please consider discussing this issue along with the analysis results of the manuscript.

Indeed, during GPCR activation more transient intermediate states can form and the complex structures in our analysis represent the stable nucleotide-free GPCR-G protein “final stage” complexes. To reflect this, we have added the following text to the Discussion:

It has furthermore been suggested that there are sequential conformational changes during GPCR activation and G protein coupling with transient intermediate states facilitating the transition of the extensive conformational rearrangement, which is not captured by currently available complex structures. (10.1016/j.cell.2019.04.022, 10.1016/j.cell.2019.04.021). The proposed intermediate state complexes may require additional state determinants beyond the ones identified herein. Hence, going forward, it will be important to combine structural studies with biophysical investigations such as FRET-based systems (<https://doi.org/10.1038/nature22354>), DEER- (10.1073/pnas.2013904117), NMR (10.1073/pnas.2009786117) or even mass spectroscopy (10.1016/j.sbi.2021.03.014) for monitoring specific interactions in more infrequent conformations.

4. Please provide experimental evidence for the newly suggested GPCR activation switches.

We have added mutagenesis data generated by Franziska M. Heydenreich, Dmitry B. Veprintsev and Michel Bouvier. This confirms that mutations of predicted state-changing residues have a larger effect on potency (but not on efficacy) than do predicted non-state changing residues. We also provide a structural mapping of the tested mutations showing a clustering in the transduction pathway bridging the ligand and G protein sites.

Minor comments

1. On page 4 and Fig. 3a class F, should it be 52% and 48%, not 62% (line 25) and 38%?

We thank the Reviewer for spotting this error. It has now been corrected.

2. Please consider rewriting a sentence on page 5 line 15, "We next ... pairs confirms."

This sentence has been corrected to "*We next investigated single helix rearrangements across 13 receptor inactive/active state structure pairs.*".

Reviewer #2:

Remarks to the Author:

This manuscript by Kooistra et al., Babu and Gloriam provides an integrated and comprehensive analysis of the movements and transduction pathways shared and different among the four major GPCR families as they transit from inactivated to activated states. A great strength of the work is their use of distance pairs, rather than more traditional superposition of structures and RMSDs, to find conserved interactions and switches among inactive- and active-stabilizing residue pairs, and switch residues implicated in both. A key contribution will be the reduction to groups of these residues that define the states among each of the GPCR families. From these studies both granular analyses of transduction pathways emerge, as do more general features (e.g., that residue rotomer switches are rare, the centrality of TM Helix 3, the ubiquitousness of helix rotation, often over rotomer changes, and the

greater similarities of among receptors within families than within active vs inactive states). Many of these will provide guidance to the community to drive specific research questions (e.g., what state is my receptor in, how to I drive that/interrupt that by mutation, what is such-and-so ligand doing...?). I thus find this a strong manuscript that will interest the community, and I support publication. As I have seen it previously (and liked it then), and since most of my key critiques have been addressed, I think the manuscript can be published as is.

We thank Reviewer 2 for reviewing this manuscript again, which has indeed been revised after review by another journal. We are very pleased to get the solid confirmation of the value of the new approaches, and that the analysis will be able to guide future studies seeking to answer yet unsolved questions about receptor activation.

Decision Letter, first revision:

12th Aug 2021

Dear David,

Thank you for submitting your revised manuscript "GPCR activation mechanisms across classes and macro/microscales" (NSMB-A44783A). It has now been seen by one of the original referees and their comments are below. The reviewer finds that the paper has improved in revision, and therefore we'll be happy in principle to publish it in Nature Structural & Molecular Biology, pending minor revisions to comply with our editorial and formatting guidelines.

We are now performing detailed checks on your paper and will send you a checklist detailing our editorial and formatting requirements in about a week. Please do not upload the final materials and make any revisions until you receive this additional information from us.

To facilitate our work at this stage, we would appreciate if you could send us the main text as a word file. Please make sure to copy the NSMB account (cc'ed above).

Thank you again for your interest in Nature Structural & Molecular Biology Please do not hesitate to contact me if you have any questions.

Kind regards,
Florian

Florian Ullrich, Ph.D.
Associate Editor
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Reviewer #1 (Remarks to the Author):

The authors resolved all the concerns that I raised.

Final Decision Letter:

22nd Sep 2021

Dear David,

We are now happy to accept your revised paper "GPCR activation mechanisms across classes and macro/microscales" for publication as a Article in Nature Structural & Molecular Biology.

Acceptance is conditional on the manuscript's not being published elsewhere and on there being no announcement of this work to the newspapers, magazines, radio or television until the publication date in Nature Structural & Molecular Biology.

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Kind regards,
Florian

Florian Ullrich, Ph.D.
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