

Supplemental information

Polyclonal F(ab')₂ fragments of equine antibodies raised against the spike protein neutralize SARS-CoV-2 variants with high potency

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SUPPLEMENTAL INFORMATION

Polyclonal F(ab')₂ fragments of equine antibodies raised against recombinant SARS-CoV-2 spike protein neutralize the P.1 and P.2 strains with high potency

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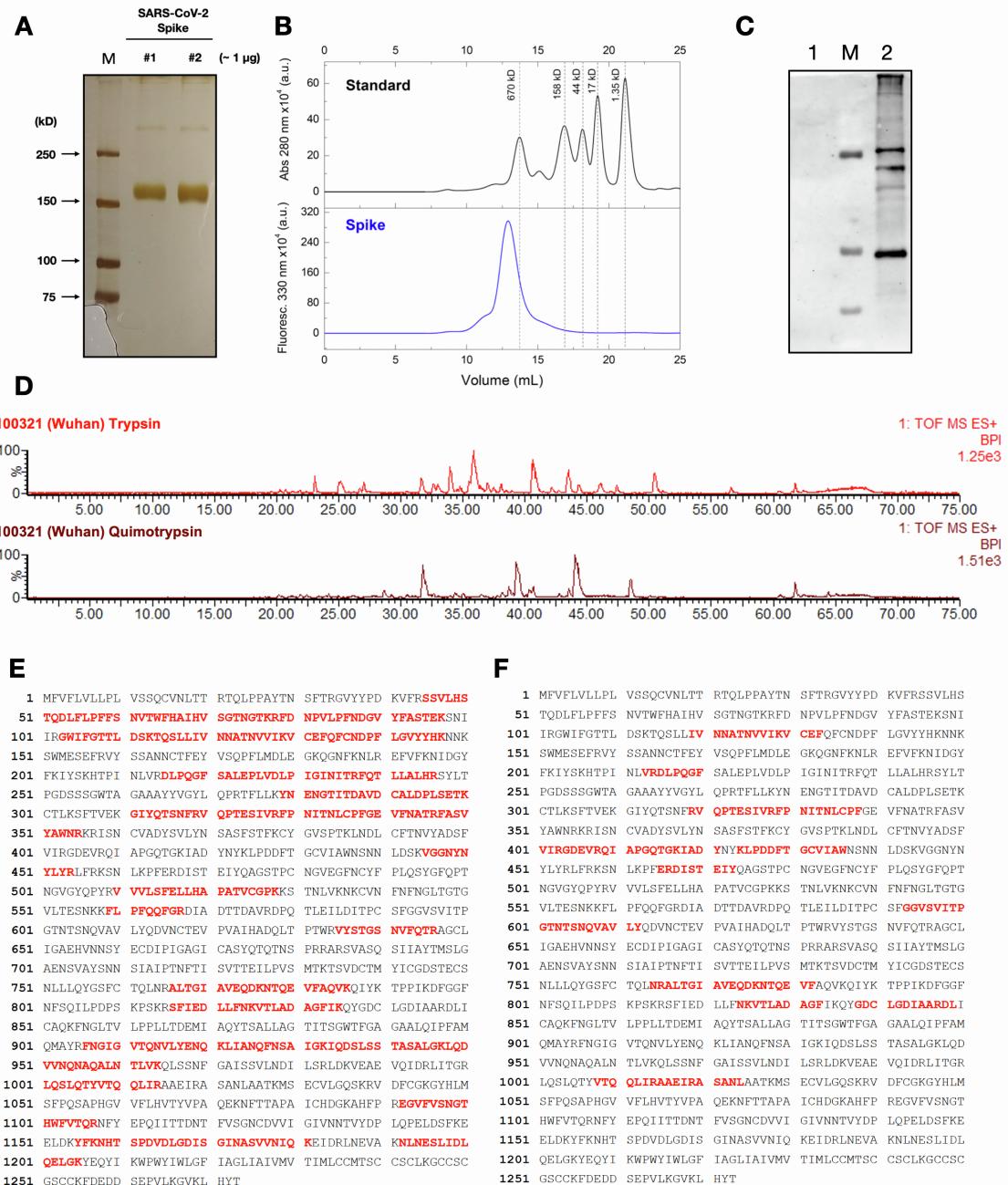


Figure S1. Quality checks of SARS-CoV-2 spike protein (related to Figure 1). (A) Image of a silver-stained 7% SDS-PAGE gel showing two representative batches of SARS-CoV-2 spike preparation (two batches for purposes of comparison). (B) Line plots showing size-exclusion chromatograms using a Superose 6 10/300 column. The fluorescence intensity at 330 nm of the spike protein was recorded as a function of the retention volume. The absorbance at 280 nm of a molecular mass standard (black line) is shown for comparison. (C) Western-blot membrane for detection of host-cell proteins (HCP) from HEK293 cells using a specific antibody (#ABIN1113182, Antibodies-Online.com, 1:1.000). Lane 1, Purified SARS-CoV-2 spike preparation (15 µL of sample, containing 13 µg of protein, was applied to the gel); Lane 2, Molecular-mass marker (Bio-Rad, 1:1000 dilution). (D) Mass spectra of Q100321 (Wuhan) Trypsin and Q100321 (Wuhan) Quimotrypsin. (E) Sequence alignment of the SARS-CoV-2 spike protein. (F) Sequence alignment of the SARS-CoV-2 spike protein.

#1610374); Lane 3 supernatant of HEK293 cells as positive control for HCP (15 µL of sample was applied to the gel). **(D)** Ion counts as a function of elution time of tryptic (red) and quimotryptic (dark red) peptides from a C18 column. Mapping of **(E)** tryptic and **(F)** quimotryptic peptides against the SARS-CoV-2 spike sequence comprises a total coverage of ca. 44%.

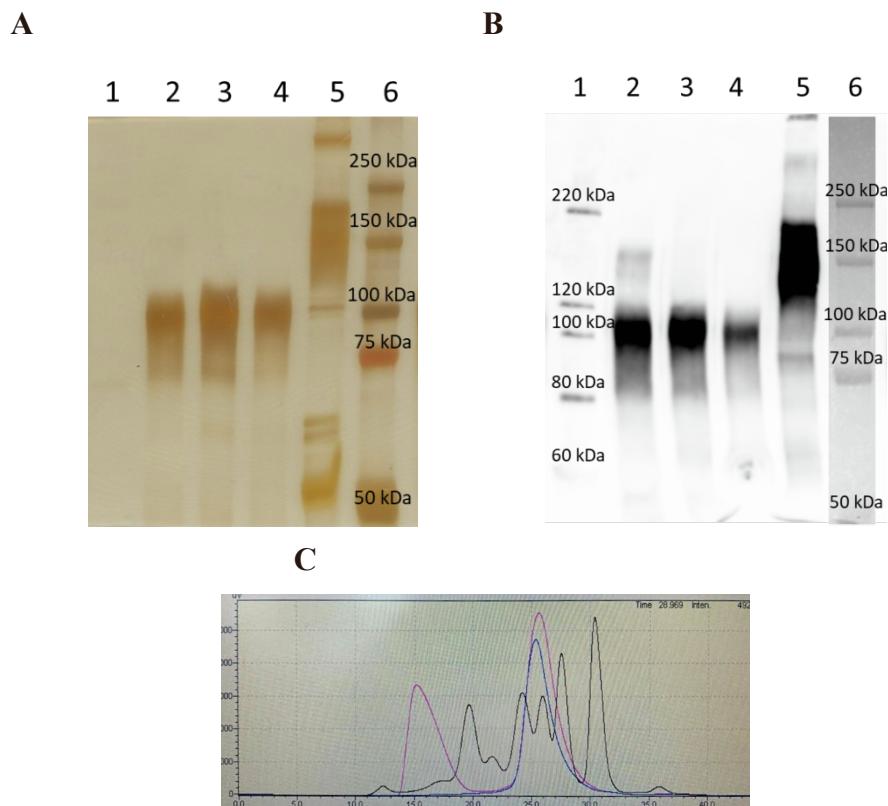


Figure S2. Silver-stained SDS-PAGE (A), chemiluminescent Western blot (B) and Gel Filtration of equine hyperimmune F(ab')₂ anti-SARS-CoV-2 (related to Figures 3C and 4). **A, B:** 1. MagicMark XP MW Markers (Thermo Fisher); 2. Concentrated hyperimmune anti-SARS-CoV-2 F(ab')₂ (1:200); 3. Ampouled antivenom F(ab')₂ (1:200); 4. Ampouled anti-rabies F(ab')₂ (1:200); 5. Equine hyperimmune plasma (1:200); 6. Precision Plus Protein Dual Color Markers (Bio-Rad). **C:** Size-exclusion chromatograms using a Superose 6 10/300 column. Blue line: Industrial concentrated hyperimmune anti-SARS-CoV-2 F(ab')₂; Rose Line: Pre-industrial lot of anti-SARS-CoV-2 F(ab')₂; Black line: Molecular markers.

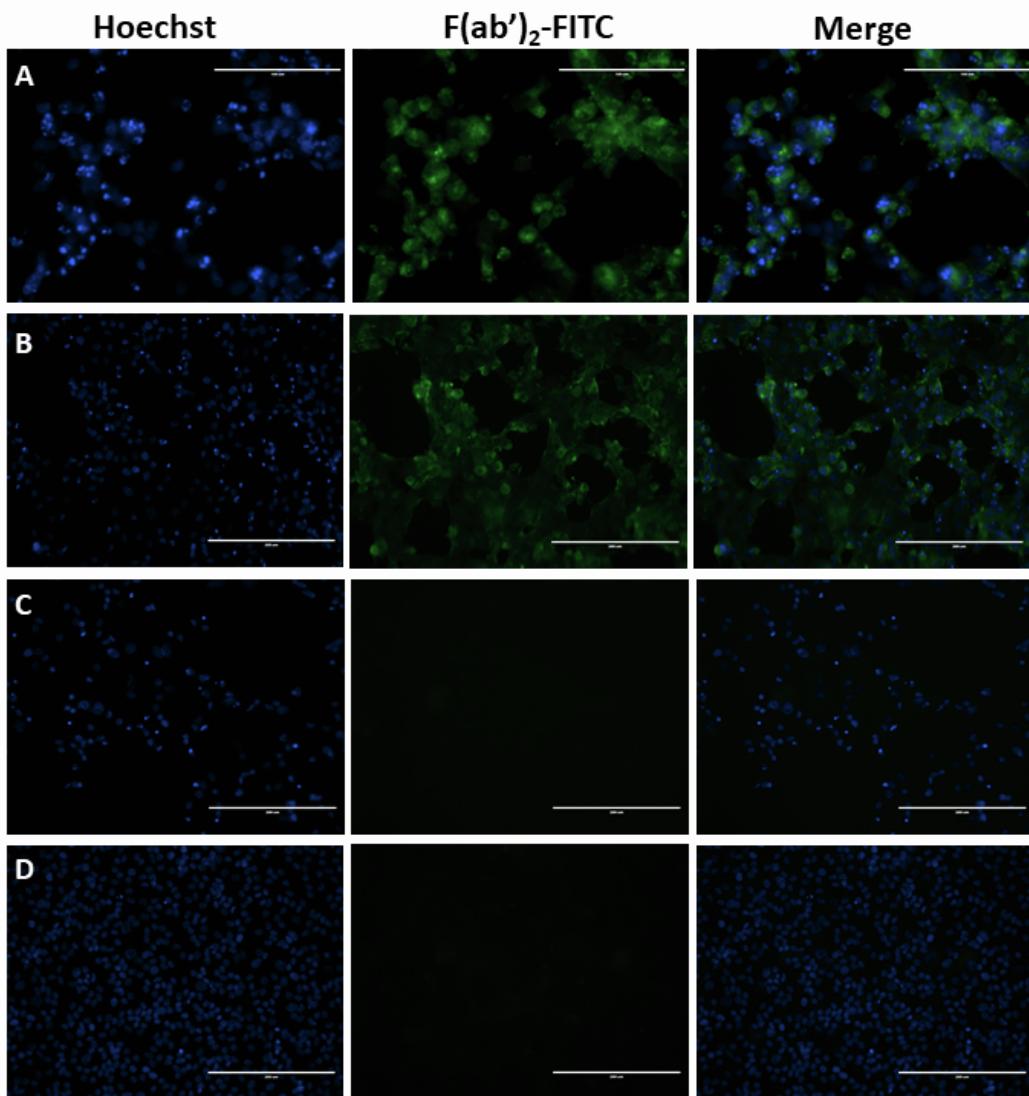


Figure S3: Equine F(ab')₂ binds specifically to SARS-CoV-2 infected cell (related to Figures 3C and 4). Vero E6 cells were infected with SARS-CoV-2 (MOI = 0.05) for 48 h and immunolabeled with F(ab')₂-FITC conjugate. **(A)** Cells infected with SARS-CoV-2, labeled with F(ab')₂ -FITC, and observed with 40x objective. **(B)** Cells infected with SARS-CoV-2 and labeled with F(ab')₂-FITC, and observed with 20x objective. **(C)** Control infected cells incubated in the absence of F(ab')₂-FITC. **(D)** Control infected cells incubated in the presence of non-labeled F(ab')₂. First column shows images in the blue channel corresponding to Hoechst labeling of cell nuclei, second column shows images obtained in the green channel for the detection of F(ab')₂-FITC fluorescence, and third column is a merge of blue and green channels. Bars on (A) 100 μ m, on all other images 200 μ m.

Table S1 – Mean R_h values corresponding to the mean hydrodynamic radius of $F(ab')_2$ (related to Figures 3C and 4). The hydrodynamic radius (R_h) of $F(ab')_2$ samples were determined by Dynamic Light Scattering with a 630-nm laser at 25°C.

Sample	Hydrodynamic Radius, R_h
	nm
Anti-SARS-CoV-2 $F(ab')_2$	10.6 +/- 0.13
Antivenom $F(ab')_2$	11.8 +/- 0.21
Anti-rabies $F(ab')_2$	12.4 +/- 0.23