

Figure S1. Genome-wide cisplatin sensitivity screen, related to Figure 1

(A & B). Scatter plots of the PD5 (A) and PD10 (B) cisp screens showing log fold changes (on the x-axes) plotted against negative log of the p-values (y-axes) generated by MAGeCK. A few of the top scoring genes are indicated. (C) EdgeR camera plots showing the distribution of guide RNAs. Statistics indicates the log fold changes from positive (on the left) to negative (on the right) of the plots. (D-F) qPCR depletion analysis of cells expressing shRNAs against the indicated proteins. (G) Top panel shows an EdgeR camera plot of the distribution of gRNAs against *SCAI* at PD5. Bottom panel shows MAGeCK scores of the top 10 scoring genes at PD5.

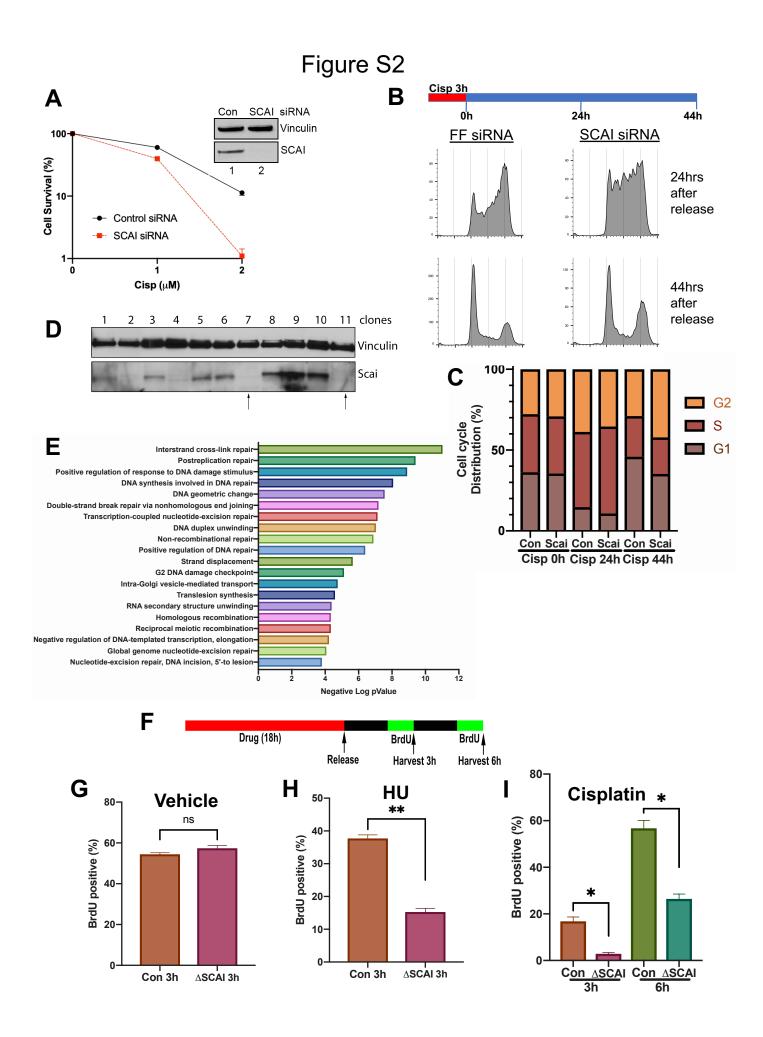


Figure S2. SCAI is important for proper repair of ICLs, related to Figures 1 & 2.

(A) CSAs showing survival of control or SCAI siRNA-treated HeLa cells upon treatment with the indicated doses of cisp. Data are normalized to untreated cells for each siRNA treatment condition. Mean + SEM survival of two independent experiments are shown. Western blots showing SCAI knockdown shown on right. (B) Top. Schematic showing details of the cell cycle arrest experiment. Cisp or vehicle alone was added for 3h and then washed out. Bottom. Cell cycle distribution showing increased accumulation of SCAI siRNA treated cells in S and G2 phases following treatment with 2 µM cisp. (C) Cell cycle profile of experiment in (B). (D) Western blot analyses showing SCAI expression in various SCAI knockout clones. (E) GO term enrichment analyses showing the enriched pathways among the high scoring hits from Figure 1C. (F). Schematic for the experiments in G-I showing the BrdU incorporation assay protocol. Cells were treated with the indicated agent for 18 h, washed and released for 2.5 or 5.5 h and labeled for 30 m with BrdU. (G - I). WT and SCAI null U2OS cells were treated with vehicle (G), or HU (H) or cisp (I) and analyzed for BrdU incorporation by flow cytometry 3 h or 6 h after release as indicated. Data in G-I shows mean and SEM of two independent experiments. Student ttest, ns= p>0.05, *= p<0.05, **= p<0.01. Con represents WT cells.

Figure S3

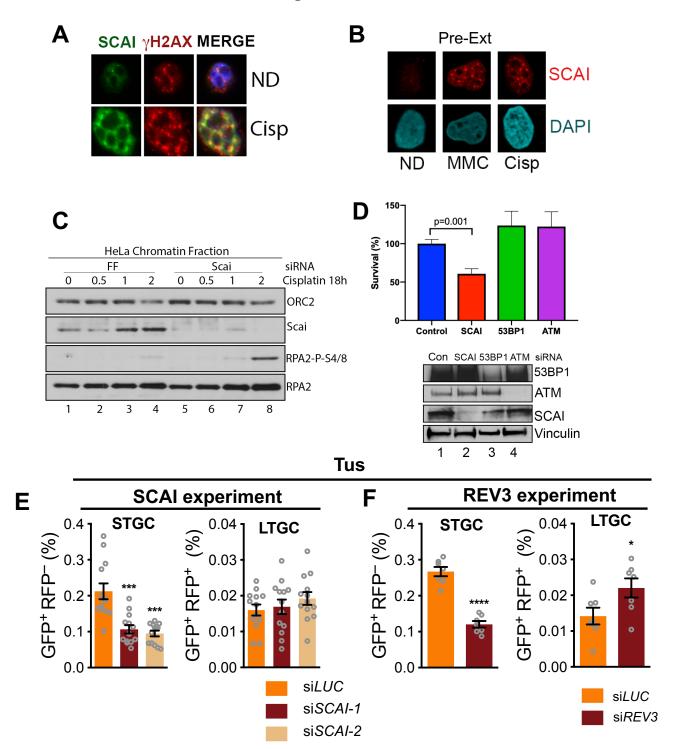


Figure S3. SCAI is recruited to ICL sites and promotes HR, related to Figure 3.

(A) IF images showing recruitment of U2OS cells expressing GFP-tagged SCAI to foci that colocalize with H2AX after treatment with 1 μ M Cisp for 2 h. ND – no drug (B) U2OS cells expressing FLAG-SCAI pre-extracted as described in experimental procedures. treated with 50 nM MMC or 1 µM Cisp for 8 h before IF analyses. ND-no drug. (C) HeLa cells were transfected with either firefly control (FF) or SCAI siRNA for 48 h then treated with vehicle or the indicated doses of cisp for 18 h prior to chromatin fractionation and immunoblotting against the indicated proteins. (D) U2OS cells were transfected with control or siRNAs against the indicated genes for 48 h, then treated with vehicle or 3 µM cisp for 24 h and analyzed using cell titer viability assays 48 h afterwards. Data is mean + SEM of two independent experiments. Student t-test. Bottom panel shows western blot analysis of lysates from the experiment. (E) STGC (left panel) and LTGC (right panel) frequencies of Tus-stalled forks in mouse 6xTer/HR cells co-transfected with Tus and either control siRNA or two different mouse siRNAs against SCAI. Data represent the mean + SEM. Welch's test, ***=p<0.001. (F) STGC (left panel) and LTGC (right panel) frequencies of Tus-stalled forks in mouse 6xTer/HR cells co-transfected with Tus and either control siRNA or siRNAs against mouse REV3. Data represent the mean + SEM. Welch's test, *=p<0.05, ****=p<0.0001.

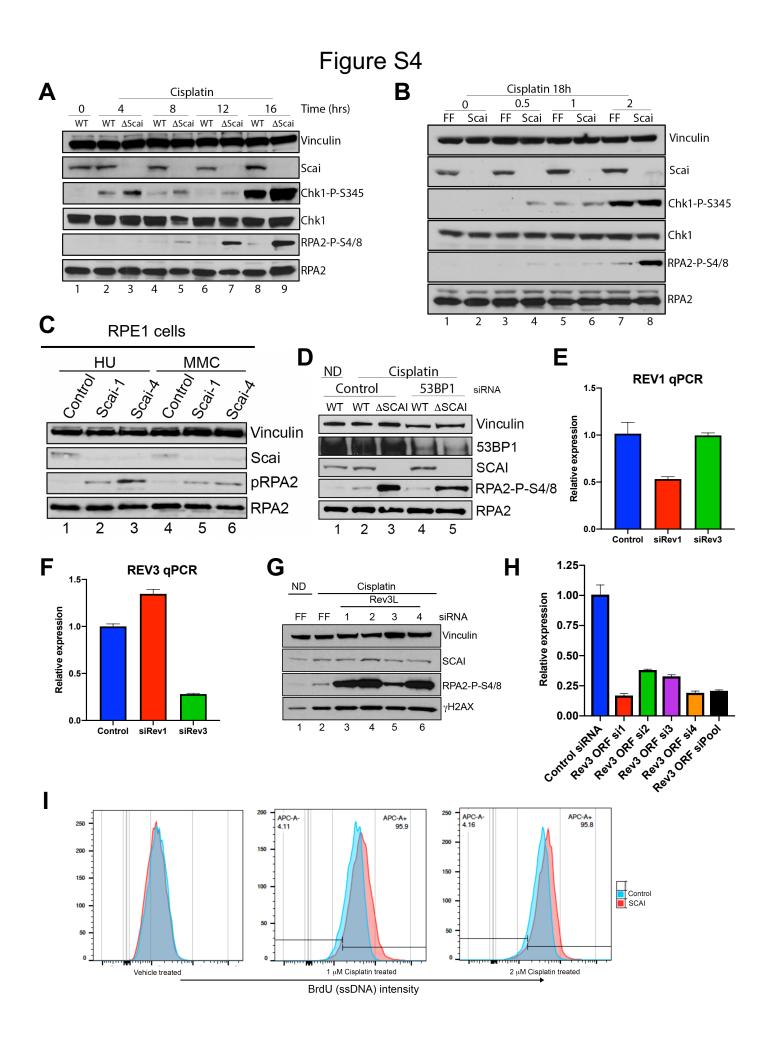
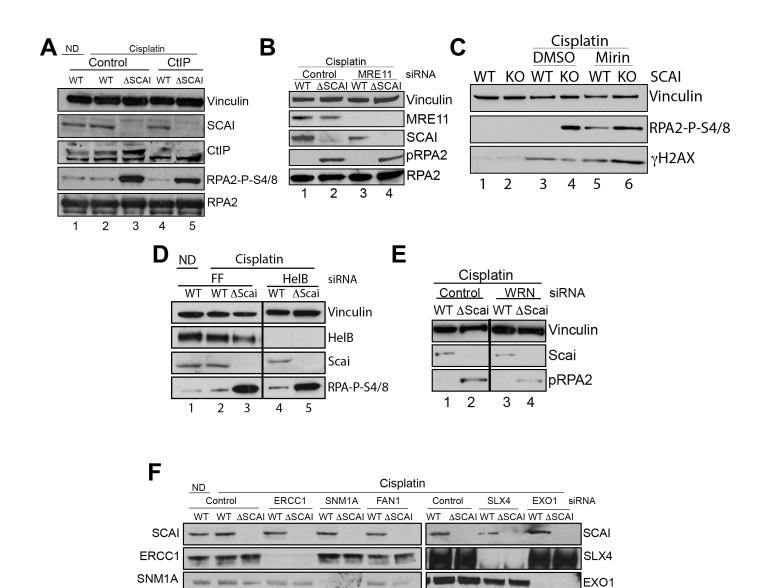


Figure S4. Protexin limits ssDNA accumulation, related to Figure 4.

(A) Time course of pRPA. Control WT U2OS cells or SCAI null U2OS cells were treated with 2 µM of cisp for the indicated duration. Cells were then processed for western blotting against the indicated antibodies. (B) U2OS cells were transfected with control (FF) or SCAI siRNAs for 48 h, then treated with vehicle or the indicated doses of cisp for 18 h prior to western blot analyses using antibodies against the indicated proteins. (C) RPE1 cells were treated with control or indicated siRNAs against SCAI for 60 h. Cells were then treated with 1 mM HU or 1 µM MMC for 18 h before immunoblotting. (D) WT U2OS or SCAI null U2OS cells were transfected with control siRNAs or siRNA to 53BP1 for 72 h. Cells were then treated with 2 μ M Cisp for 18 h, harvested and processed as above. (E & F) gPCR showing knockdown of REV1 and REV3 respectively. (G) U2OS cells were transfected with firefly (FF) siRNAs or the indicated siRNAs to REV3 for 72 h. Cells were harvested and processed for immunoblotting against the indicated proteins. (H) qPCR of RNA samples from the experiment in (G). (I) Control or SCAI siRNA treated cells were stained with BrdU for 10 h followed by treatment with vehicle or indicated doses of cisp for 6 h. BrdU intensity was assayed using flow cytometry under non-denaturing conditions.

Figure S5



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Figure S5. Protexin protects replication forks from nucleases, related to Figure 5.

(A) WT and *SCAI*-null U2OS cells were treated with control or CtIP siRNAs for 72 h. Cells were then treated with vehicle or 1.5 μ M cisp for 18 h before immunoblotting. (B) WT and *SCAI*-null U2OS cells were treated with control or MRE11 siRNAs for 72 h. Cells were then treated with vehicle or 1.5 μ M cisp for 18 h before immunoblotting. (C) WT and *SCAI*-null U2OS cells were treated with cisp for 8 h and then treated with vehicle (DMSO) or 10 μ M mirin for 10 h prior to harvest. Cisp was left on throughout the mirin (or DMSO) treatment. Cells were harvested and processed for immunoblotting as above. (D) WT U2OS or *SCAI*-null U2OS cells were treated with 2 μ M Cisp for 18 h, harvested and processed for immunoblotting. All lanes are from the same blots – irrelevant lanes were cropped out. (F) Additional blots against the indicated proteins from the experiment in Figure 5E.

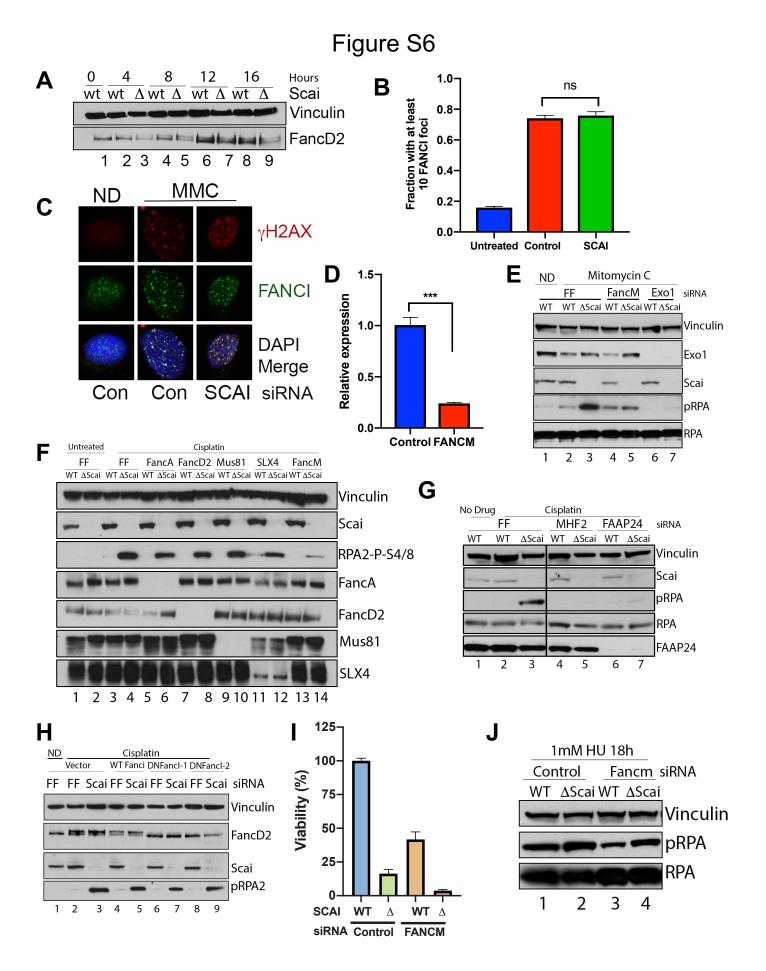


Figure S6. FANCM drives ssDNA accumulation following Protexin loss independently of FA pathway activation, related to Figures 5 & 6.

(A) Western blot analyses showing time course of FANCD2 ubiquitination following 2 μM cisp treatment in WT and SCAI-null cells. (B & C) U2OS cells transfected with the indicated siRNAs for 60 h were treated with vehicle or 1 µM MMC for 6 h prior to IF analyses for FANCI and γ -H2AX foci. Quantification of the experiment is shown in B. At least 50 cells were counted for each condition. Analysis by student t-test, ns - p-value < 0.05. (D) U2OS cells were transfected with control or FANCM siRNAs for 72 h and processed for qPCR analysis. (E) WT and SCAI null cells were treated with siRNA against the indicated proteins and control (FF) for 72 h. Cells were then treated with vehicle (ND) or 100 nM MMC for 16 h and analyzed by immunoblotting. (F) WT and SCAI-null U2OS cells were treated with control (FF) or siRNA against the indicated proteins for 72 h. Cells were then treated with vehicle or 1.5 μ M cisp for 18 h before immunoblotting. (G) WT or SCAI-null U2OS cells were transfected with control (FF), MHF2 or FAAP24 siRNAs for 72 h. Cells were then treated with 2 μ M Cisp for 18 h, harvested and processed for immunoblotting. All lanes are from the same blots – irrelevant lanes were cropped out. (H) U2OS cells expressing vector or the indicated ORFs were treated with control (FF) or SCAI siRNA for 72 h before treatment with vehicle (ND) or 1.5 µM cisp for 18 h and analysis by immunoblotting. (I) WT and SCAI-null U2OS cells were treated with control or siRNAs to FANCM for 48 h before 16 h treatment with vehicle or 2 µM cisp. CSAs showing dependence on FANCM for survival. Viability is relative to vehicle-treated cells. Mean and SEM of two independent experiments are shown. (J) WT or SCAI-null U2OS cells were transfected with control or FANCM siRNAs for 72 h. Cells were then treated with 1mM HU for 18 h, harvested and processed for immunoblotting.

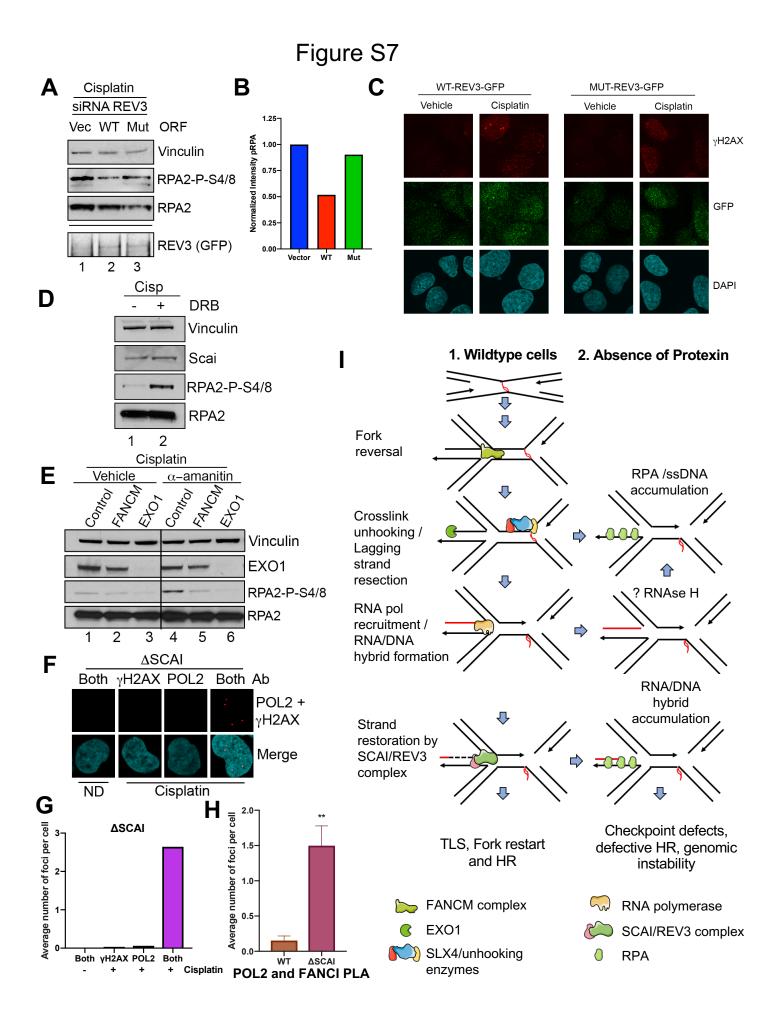


Figure S7. REV3 and POL2 polymerase roles in limiting ssDNA, related to Figure 7. (A) Top panel: U2OS cells stably expressing vector control, GFP-tagged WT REV3 or a REV3 D2781A/D2783A mutant that lacks polymerase activity were treated with control siRNA or an siRNA to the 3' UTR of endogenous REV3 as indicated for 48 h. Cells were then treated with cisp for 18 h and assayed for western blotting. In parallel, U2OS cells expressing vector control, GFP-tagged WT-REV3 or D2781A/D2783A mutant REV3 were immunoprecipitated and blotted for GFP expression (bottom panel). (B) Quantification of the pRPA intensity compared to total RPA amounts from the experiment in (A). (C) Cells expressing GFP-tagged WT- REV3 or mutant were treated with cisp for 16 h and processed for IF against GFP and γ H2AX. Representative images are shown. (D) WT U2OS cells were treated with cisp for 12 h followed by vehicle or treatment with 100 µM DRB for 3 h. Cells were blotted against indicated proteins. (E) WT U2OS cells were transfected with control siRNAs or siRNA to FANCM or EXO1 for 60 h before treatment with 2 μ M Cisp for 10 h. Cells were then treated with 10 μ g/ml α -amanitin for 6 h and processed for western blotting as above. (F) SCAI-null U2OS cells were treated with vehicle or 1 μ M cisp for 8 h and processed for PLAs using individual or both mouse γ H2AX and rabbit POL2 antibodies as indicated. Representative images are shown. (G) Quantification of the experiment in (F). (H) WT and SCAI-null U2OS were treated with 1 µM cisp for 8 h and processed for PLAs using antibodies against mouse POL2 and rabbit FANCI. Mann Whitney test, **=p<0.01. (I) Model showing potential roles of the Protexin complex in resection control and repair at an ICL stalled fork.