

# Apoptosis inducing 1,3,4-oxadiazole conjugates of Capsaicin: Their *in vitro* anti-proliferative and *in silico* studies

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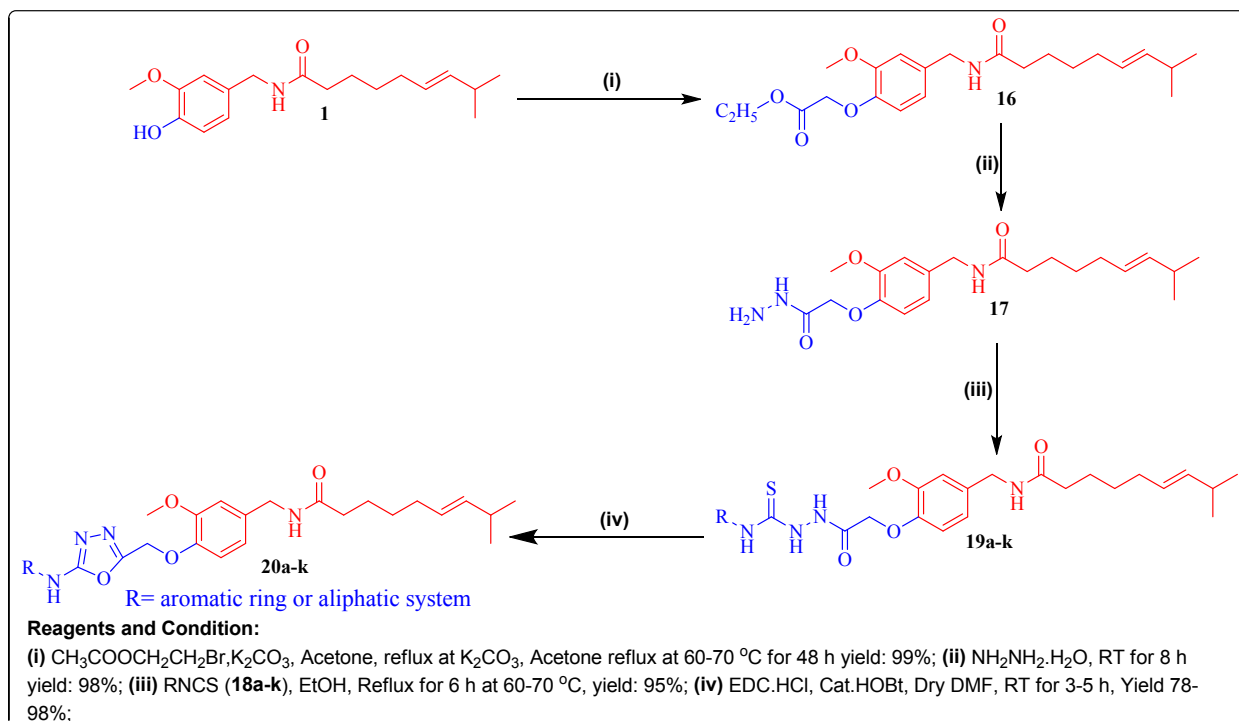
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## 1. Chemistry

### 1.1. Scheme S-1. Synthesis of compound 20a-k



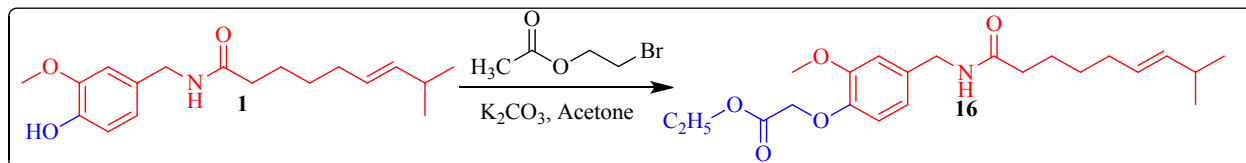
**Scheme S-1**

### 1.2. General information

All the chemicals and reagents used in the study were purchased from Merck (India), Spectrochem, and Sigma Aldrich which were of reagent grade. TLC was performed for indication of reaction completion on 0.25 mm silica gel 60-F254 plates. UV light was used to visualize spots. Melting points of all the synthesized compounds were measured using Buchi labortechnik AG 9230 automated melting point apparatus (Switzerland); IR spectra were recorded on Bruker ALPHA FT-IR spectrometer (Germany),  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were determined on a Bruker (300 MHz and 75MHz) spectrometer and chemical shifts were expressed as ppm against TMS as internal reference. Solvent peak of  $\text{CDCl}_3$  in  $^1\text{H}$  NMR was observed at 7.26 ppm and in  $^{13}\text{C}$  observed at 77.28-76.77 ppm. A residual  $\text{DMSO-d}_5$   $^1\text{H}$  NMR signal is observed at 2.52 ppm. Mass spectra's were recorded on ESI-MS. CHNS elemental analysis of synthesized compounds has been done on Vario EL III Elementar Equipment All compounds prepared in this paper are novel and confirmed with spectral data. All the synthesized compounds were recrystallized in methanol and were purified by column chromatography.

### 1.3. Synthesis of compound 16-20a-k.

#### 1.3.1. Synthesis of (*E*)-ethyl 2-(2-methoxy-4-((8-methylnon-6-enamido)methyl)phenoxy)acetate (16)

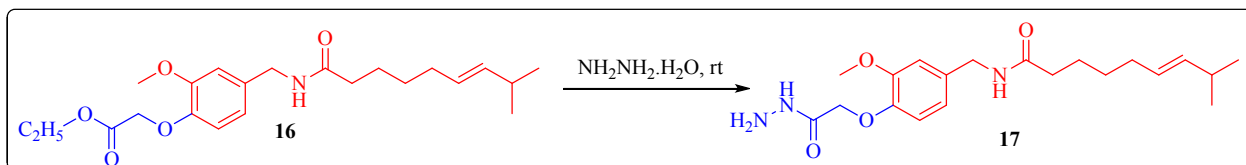


1g of capsaicin **1** was dissolved in 10 mL of dry acetone. Then 0.54 g of activated potassium carbonate was added to the reaction mixture and was allowed to stir for 10 min. after 10 min, 0.5 mL (1.2 equiv.) of bromoethyl acetate was added and refluxed at 60-70 °C for 48 h. Completion of reaction was monitored by doing TLC (4:6 ratio of ethylacetate:hexane) and anisaldehyde was used as spraying agent for detection of TLC. After completion, reaction mixture was poured into ice and white colour solid was produced which was further filtered and dried under vacuum.

<sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  (ppm): 8.24 (s, 1H), 6.87 (s, 1H), 6.79 (d, *J* = 8 Hz, 1H), 6.71 (s, *J* = 8.4 Hz, 1H), 5.40-5.29 (m, 2H), 4.71 (s, 1H), 4.18-4.10 (m, 4H), 3.75 (s, 3H), 2.11 (t, *J* = 7.2 Hz, 2H), 1.96-1.91 (m, 2H), 1.55-1.48 (m, 2H), 1.33-1.19 (m, 6H), 0.93 (d, *J* = 6.8 Hz, 4H), 0.85 (d, *J* = 6.4 Hz, 2H).

MS (ESI) *m/z*: [M-1]<sup>+</sup> 390.1.

#### 1.3.2. Synthesis of (*E*)-N-(4-(2-hydrazinyl-2-oxoethoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (17)

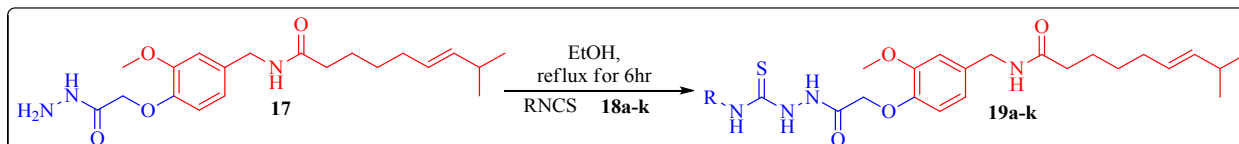


To 2 ml of hydrazine hydrate, 1.4g of compound **16** was added and reaction mixture was stirred for 8 h at room temperature. Completion of reaction was examined by TLC (6:4 ratio of ethylacetate:hexane). A white colour solid was produced within the reaction mixture which was further carried to filter off and dried under high vacuum pump.

$^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 9.14 (s, 1H), 8.42 (t,  $J = 5.6$  Hz, 1H), 6.86 (d,  $J = 8.4$  Hz, 2H), 6.72 (d,  $J = 8$  Hz, 1H), 4.42 (s, 2H), 4.33 (s, 2H), 4.18 (d,  $J = 6.0$  Hz, 2H), 3.75 (s, 3H), 2.11 (t,  $J = 7.2$  Hz, 2H), 1.53-1.48 (m, 3H), 1.23 (s, 6H), 0.84 (d,  $J = 6.8$  Hz, 6H).

MS (ESI)  $m/z$ :  $[\text{M}+3]^+$  380.4.

### 1.3.3. Synthesis of *(E)*-N-(3-methoxy-4-(2-oxo-2-(2-(phenylcarbamothioyl)hydrazinyl)ethoxy)benzyl)-8-methylnon-6-enamide (**19a-k**)

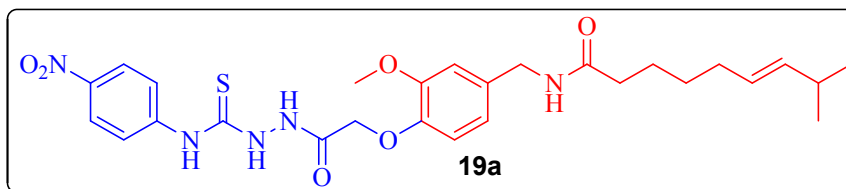


#### General procedure A

To ethanolic solution (absolute) of compound **17**, different substituted aryl and alkyl isothiocyanates (**18a-k**) were added to it then the reaction mixture was refluxed for 6 h at 60-70 °C. Solid precipitate was formed in the reaction mixture was filtered of and washed with cold alcohol to yield compounds (**19a-k**). TLC was run in TEF 4:5:1 which indicate the completion of the reaction.

#### 1.3.3.1. Synthesis of *(E)*-N-(3-methoxy-4-(2-(2-((4-nitrophenyl)carbamothioyl)hydrazinyl)-2-oxoethoxy)benzyl)-8-methylnon-6-enamide (**19a**)

To 10 mL of absolute ethanol, 0.5 g of compound **17** was dissolved, followed by addition of 0.47 g (1.2

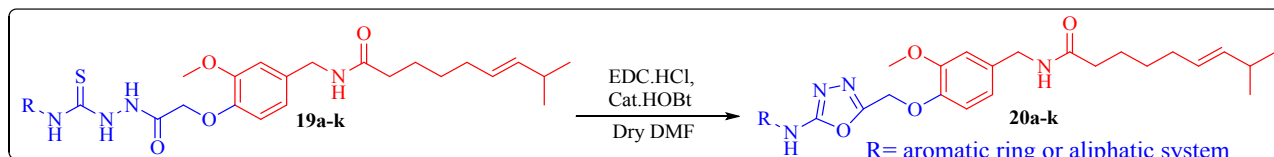


equivalents) of 1-isothiocyanato-4-nitrobenzene **18a**. Reaction mixture was refluxed for 6 h at 60-70 °C and after completion of reaction; solid precipitate was formed in reaction mixture. These Solid precipitate was filtered off and dried to afford compound **19a**.

$^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  (ppm): 11.66 (s, 1H), 10.24 (s, 1H), 10.09 (s, 1H), 9.93 (s, 1H), 8.23 (d,  $J = 8$  Hz, 2H), 7.89 (d,  $J = 8$  Hz, 2H), 6.89 (s, 2H), 6.74 (d,  $J = 8$  Hz, 1H), 4.61 (s, 2H), 4.19 (d,  $J = 8$  Hz, 2H), 3.71 (s, 3H), 2.11 (t,  $J = 8$  Hz, 2H), 1.53-1.48 (m, 3H), 1.39-1.36 (m, 1H), 1.24 (s, 6H), 0.84 (d,  $J = 8$  Hz, 6H).

MS (ESI)  $m/z$ :  $[\text{M}+1]^+$  558.5.

### 1.3.4. Synthesis of (*E*)-*N*-(3-methoxy-4-((5-(phenylamino)-1,3,4-oxadiazol-2-yl)methoxy)benzyl)-8-methylnon-6-enamide (20a-k)

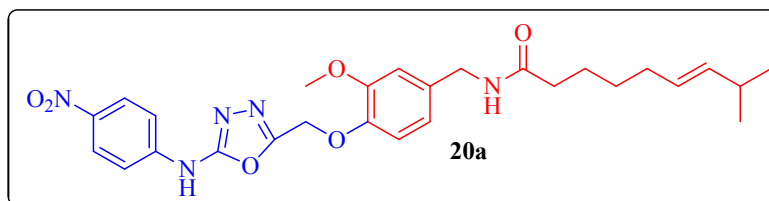


#### **General procedure B**

To 5 mL of dry dimethylformamide (DMF), 0.5g of compound **19a-k** and 0.52 g (3 equivalents) of *N*-(3-Dimethylaminopropyl)-*N'*-ethylcarbodiimide hydrochloride (EDC.HCl) with catalytic amount of Hydroxybenzotriazole (HOBt) were added to it. Reaction mixture was stirred at room temperature for 3-5 h under inert atmosphere. After the completion of reaction monitored by TLC (TEF, 4:5:1), the reaction mixture was poured onto ice. The precipitate formed was filtered off, washed with excess of water, dried under high vacuum and recrystallized in ethanol to afford the title compound **20a-k** in pure form.

#### 1.3.4.1. Synthesis of (*E*)-*N*-(3-methoxy-4-((5-((4-nitrophenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)benzyl)-8-methylnon-6-enamide (20a)

According to general procedure A,  
To 5 mL of dry dimethylformamide (DMF), 0.5g of compound **19a** and 0.52 g (3



equivalents) of *N*-(3-Dimethylaminopropyl)-*N'*-ethylcarbodiimide hydrochloride (EDC.HCl) with catalytic amount of Hydroxybenzotriazole (HOBt) were added to it. Reaction mixture was stirred at room temperature for 3-5 h under inert atmosphere. After the completion of reaction monitored by TLC (TEF, 4:5:1), the reaction mixture was poured onto ice. The precipitate formed was filtered off, washed with excess of water, dried under high vacuum and recrystallized in ethanol to afford the title compound **20a-k** in pure form.

Creamish yellow; yield: 91%.

Mp: 189.0 °C.

IR  $\text{cm}^{-1}$ : 3487, 3437, 3384, 3335, 3285, 3231, 3200, 3049, 2990, 2924, 1668, 1639, 1590, 1513, 1459, 1419, 1325, 1266, 1230, 1196, 1142, 1107, 1014, 955, 846, 770, 738, 679, 619, 546.

$^1\text{H}$  NMR (300 MHz,  $\text{DMSO}-d_6$ )  $\delta$  (ppm): 10.97 (s, 1H), 8.21 (d,  $J = 8.4$  Hz, 2H), 7.76 (d,  $J = 8.4$  Hz, 2H), 7.09 (s, 1H), 6.99 (d,  $J = 8.1$  Hz, 1H), 6.88 (s, 1H), 6.80 (d,  $J = 8.1$  Hz, 1H), 5.22 (s,

2H), 4.34 (d,  $J = 5.1$  Hz, 2H), 3.84 (s, 3H), 2.21 (t,  $J = 7.5$  Hz, 2H), 1.64 (t,  $J = 6$  Hz, 2H), 1.54-1.43 (m, 1H), 1.28 (s, 6H), 0.85 (d,  $J = 6.3$  Hz, 6H).

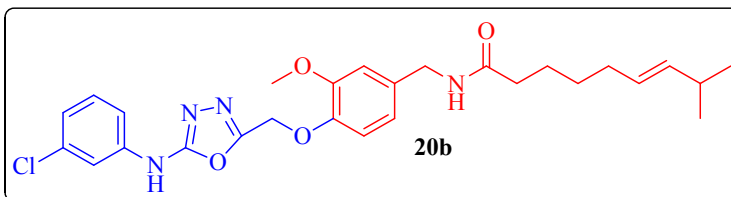
$^{13}\text{C}$  NMR (75 MHz, DMSO- $d_6$ )  $\delta$ : 173.2, 160.2, 156.6, 149.9, 145.6, 144.4, 141.7, 134.4, 125.0, 124.4, 120.6, 119.6, 116.6, 115.7, 111.7, 61.3, 55.6, 42.6, 36.3, 29.4, 29.2, 27.7, 27.0, 25.7, 22.5, 14.0.

HRMS (ESI)  $m/z$ :  $[\text{M} + \text{H}]^+$  Calcd for  $\text{C}_{27}\text{H}_{34}\text{N}_5\text{O}_6$  524.2509; Found 524.2513.

Anal. Calcd for  $\text{C}_{27}\text{H}_{33}\text{N}_5\text{O}_6$ : C, 61.94; H, 6.35; N, 13.38; O, 18.33%, Found: C, 61.62; H, 6.24; N, 13.28 %.

#### 1.3.4.2. Synthesis of (*E*)-N-(4-((5-((3-chlorophenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (20b)

According to general procedure A, Compound **19b** was cyclized under coupling conditions with EDC.HCl and HOBT to yield desired compound **20b**.



White solid, yield: 89%.

Mp: 158.5 °C.

IR  $\text{cm}^{-1}$ : 3425, 3387, 3274, 3228, 3120, 3093, 3056, 3006, 2955, 2922, 1649, 1596, 1570, 1544, 1511, 1475, 1420, 1262, 1223, 120, 1082, 1021, 907, 848, 793, 740, 700, 642, 580.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 7.60-7.57 (m, 1H), 7.43-7.36 (m, 2H), 7.04 (d,  $J = 7.8$  Hz, 1H), 6.96 (d,  $J = 8.4$  Hz, 1H), 6.81 (s, 1H), 6.74 (d,  $J = 6.3$  Hz, 1H), 5.78 (s, 1H), 5.20 (s, 2H), 4.36 (d,  $J = 5.7$  Hz, 2H), 3.83 (s, 3H), 2.21 (t,  $J = 7.5$  Hz, 2H), 1.76 (s, 2H), 1.28-1.22 (m, 7H), 0.85 (d,  $J = 6.6$  Hz, 6H).

HRMS (ESI)  $m/z$ :  $[\text{M} + \text{H}]^+$  Calcd for  $\text{C}_{27}\text{H}_{34}\text{ClN}_4\text{O}_4$  513.2269; Found 513.2272.

Anal. Calcd for  $\text{C}_{27}\text{H}_{33}\text{ClN}_4\text{O}_4$ : C, 63.21; H, 6.48; Cl, 6.91; N, 10.92; O, 12.47%, Found: C, 63.17; H, 6.41; N, 10.83 %.

#### 1.3.4.3. Synthesis of (*E*)-N-(4-((5-((4-chlorophenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (20c)

According to general procedure A, Compound **19c** was cyclized under coupling conditions with EDC.HCl and HOBT to yield desired compound **20c**.

White solid, yield: 98%.

Mp: 175.9 °C.

IR  $\text{cm}^{-1}$ : 3457, 3419, 3387, 3323, 3223,

3235, 3197, 3120, 3024, 2995, 2926,

1651, 1621, 1572, 1515, 1462, 1416, 1317, 1267, 1224, 1139, 1018, 963, 843, 806, 754, 707, 671, 612.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 8.31 (d,  $J = 7.5$  Hz, 1H), 7.56 (d,  $J = 7.5$  Hz, 1H), 7.47 (s, 1H), 7.40-7.35 (m, 1H), 7.02-6.94 (m, 1H), 6.86 (s, 1H), 6.79 (d,  $J = 7.5$  Hz, 1H), 5.70 (s, 1H), 5.23 (s, 2H), 4.38 (d,  $J = 5.1$  Hz, 2H), 3.86 (s, 3H), 2.20 (t,  $J = 6.6$  Hz, 2H), 1.65 (s, 2H), 1.53-1.48 (m, 1H), 1.28 (s, 6H), 0.85 (d,  $J = 6.3$  Hz, 6H).

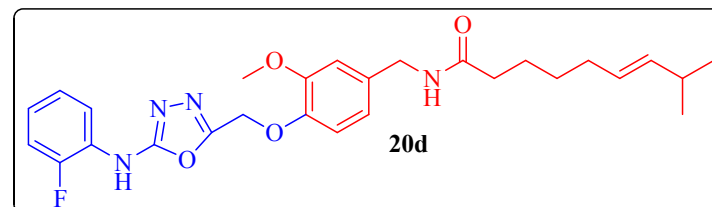
$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 172.9, 156.8, 146.0, 135.0, 133.8, 132.4, 128.8, 124.1, 120.0, 118.6, 115.7, 111.9, 111.4, 61.3, 55.9, 43.3, 38.9, 36.8, 29.6, 29.3, 27.9, 27.2, 25.7, 22.6.

HRMS (ESI)  $m/z$ :  $[\text{M} + \text{H}]^+$  Calcd for  $\text{C}_{27}\text{H}_{34}\text{ClN}_4\text{O}_4$  513.2269; Found 513.2270.

Anal. Calcd for  $\text{C}_{27}\text{H}_{33}\text{ClN}_4\text{O}_4$ : C, 63.21; H, 6.48; Cl, 6.91; N, 10.92; O, 12.47%, Found: C, 63.17; H, 6.31; N, 10.83%.

#### 1.3.4.4. Synthesis of (*E*)-*N*-(4-((5-((2-fluorophenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (20d)

According to general procedure A, Compound **19d** was cyclized under coupling conditions with EDC.HCl and HOBT to yield desired compound **20d**.



White solid, yield: 78%.

Mp: 141.4 °C.

IR  $\text{cm}^{-1}$ : 3473, 3432, 3375, 3321, 3260, 3201, 3122, 3048, 2922, 1651, 1639, 1581, 1557, 1518, 1460, 1426, 1389, 1271, 1221, 1134, 1029, 851, 814, 730, 690, 645, 618, 560.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 8.13 (t,  $J = 8.1$  Hz, 1H), 7.25 (s, 1H), 7.13-7.05 (m, 2H), 7.02-6.90 (m, 2H), 6.77 (s, 1H), 6.70 (d,  $J = 7.8$  Hz, 1H), 5.67 (s, 1H), 5.13 (s, 2H), 4.29 (d,  $J = 5.7$  Hz, 2H), 3.77 (s, 1H), 2.13 (t,  $J = 7.2$  Hz, 2H), 1.91 (q,  $J = 6.6$  Hz, 1H), 1.61-1.56 (m, 4H), 1.04-1.31 (m, 2H), 1.31-1.21 (m, 2H), 0.87 (d,  $J = 6.6$  Hz, 3H), 0.78 (d,  $J = 6.6$  Hz, 3H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 173.3, 160.9, 159.7, 156.3, 150.0, 145.9, 133.7, 119.8, 119.4, 119.3, 116.0, 115.8, 115.6, 111.8, 61.25, 55.8, 43.2, 38.9, 36.8, 29.6, 29.3, 27.9, 27.2, 25.8, 22.6; HRMS (ESI)  $m/z$ :  $[\text{M} + \text{H}]^+$  Calcd for  $\text{C}_{27}\text{H}_{34}\text{FN}_4\text{O}_4$  497.2564; Found 497.2565.

Anal. Calcd for  $\text{C}_{27}\text{H}_{33}\text{FN}_4\text{O}_4$ : C, 65.31; H, 6.70; F, 3.83; N, 11.28; O, 12.89%, Found: C, 65.28; H, 6.63; N, 11.19%.

#### 1.3.4.5. Synthesis of (*E*)-*N*-(4-((5-((4-fluorophenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (20e)

According to general procedure A, Compound **19e** was cyclized under coupling conditions with EDC.HCl and HOBT to yield desired compound **20e**.

White solid; yield: 98%.

Mp: 154.3 °C.

IR  $\text{cm}^{-1}$ : 3472, 3438, 3385, 3322,

3263, 3205, 3123, 3050, 2928,

1649, 1626, 1579, 1549, 1511, 1455, 1420, 1388, 1266, 1218, 1137, 1020, 843, 808, 722, 683, 634, 610, 552.

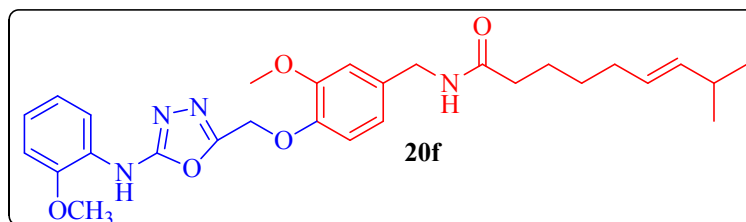
$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 8.26 (s, 1H), 7.46 (q,  $J = 4.2$  Hz, 2H), 7.02 (t,  $J = 8.4$  Hz, 2H), 6.90 (d,  $J = 8.1$  Hz, 1H), 6.78 (s, 1H), 6.70 (d,  $J = 8.4$  Hz, 1H), 5.93 (s, 1H), 5.15 (s, 1H), 4.34 (d,  $J = 5.4$  Hz, 1H), 3.78 (s, 3H), 2.21 (t,  $J = 7.2$  Hz, 2H), 1.65 (t,  $J = 6.6$  Hz, 2H), 1.53-1.41 (m, 1H), 1.26 (s, 1H), 0.84 (d,  $J = 6.6$  Hz, 6H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 173.3, 160.9, 159.7, 156.3, 150.0, 145.9, 133.7, 119.8, 119.4, 119.3, 116.0, 115.8, 115.6, 111.8, 61.25, 55.8, 43.2, 38.9, 36.8, 29.6, 29.3, 27.9, 27.2, 25.8, 22.6; HRMS (ESI)  $m/z$ :  $[\text{M} + \text{H}]^+$  Calcd for  $\text{C}_{27}\text{H}_{34}\text{FN}_4\text{O}_4$  497.2564; Found 497.2568.

Anal. Calcd for  $\text{C}_{27}\text{H}_{33}\text{FN}_4\text{O}_4$ : C, 65.31; H, 6.70; F, 3.83; N, 11.28; O, 12.89%, Found: C, 65.27; H, 6.64; N, 11.21%.

#### 1.3.4.6. Synthesis of (*E*)-*N*-(3-methoxy-4-((5-((2-methoxyphenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)benzyl)-8-methylnon-6-enamide (20f)

According to general procedure A, Compound **19f** was cyclized under coupling conditions with EDC.HCl





and HOBt to yield desired compound **20f**.

White solid; Yield 95%.

Mp: 104.7 °C.

IR  $\text{cm}^{-1}$ : 3491, 3413, 3390, 3353, 3307, 3223, 3194, 3108, 3054, 2998, 2923, 1629, 1577, 1520, 1460, 1421, 1391, 1250, 1217, 1137, 1018, 976, 918, 849, 805, 736, 670, 642, 581.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 8.12 (d,  $J = 3.3$  Hz, 1H), 7.55 (s, 1H), 7.01 (d,  $J = 6.9$  Hz, 3H), 6.91-6.88 (m, 1H), 6.84 (s, 1H), 6.78 (d,  $J = 7.8$  Hz, 1H), 5.70 (s, 1H), 5.21 (s, 2H), 4.37 (d,  $J = 5.4$  Hz, 2H), 3.87 (d,  $J = 16.8$  Hz, 6H), 2.20 (t,  $J = 7.8$  Hz, 2H), 1.67-1.62 (t,  $J = 6.3$  Hz, 2H), 1.54-1.43 (m, 1H), 1.28 (s, 6H), 0.84 (d,  $J = 6.6$  Hz, 6H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 172.97, 160.5, 156.4, 150.2, 147.0, 146.1, 133.6, 126.9, 122.8, 121.3, 120.1, 117.1, 115.7, 111.9, 110.0, 61.4, 55.9, 55.8, 43.3, 38.9, 36.8, 29.6, 29.3, 27.9, 27.2, 25.8, 22.6.

HRMS (ESI)  $m/z$ :  $[\text{M} + \text{H}]^+$  Calcd for  $\text{C}_{28}\text{H}_{37}\text{N}_4\text{O}_5$  509.2764; Found 509.2766.

Anal. Calcd for  $\text{C}_{28}\text{H}_{36}\text{N}_4\text{O}_5$ : C, 66.12; H, 7.13; N, 11.02; O, 15.73%, Found: C, 66.06; H, 7.08; N, 10.98%.

### 1.3.4.7. Synthesis of (*E*)-N-(3-methoxy-4-((5-((4-methoxyphenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)benzyl)-8-methylnon-6-enamide (**20g**)

According to general procedure A, Compound **19g** was cyclized under coupling conditions with EDC.HCl and HOBt to yield desired compound **20g**.

White solid, yield: 94%.

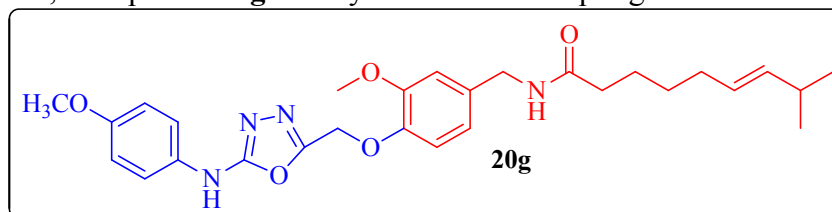
Mp: 154.9 °C.

IR  $\text{cm}^{-1}$ : 3463, 3424, 3380, 3273, 3192, 3138, 3063, 3007, 2924, 1650, 1621, 1576, 1510, 1451, 1420, 1383, 1312, 1239, 1175, 1015, 965, 836, 691, 634, 581, 557, 535.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 7.37 (d,  $J = 8.7$  Hz, 2H), 7.05-6.98 (m, 2H), 6.88 (d,  $J = 9.0$  Hz, 1H), 6.83 (s, 1H), 6.76 (d,  $J = 8.4$  Hz, 1H), 5.71 (s, 1H), 5.18 (s, 2H), 4.37 (d,  $J = 5.7$  Hz, 2H), 3.82 (d,  $J = 6$  Hz, 6H), 2.20 (t,  $J = 7.2$  Hz, 2H), 1.65-1.60 (m, 2H), 1.52-1.48 (m, 1H), 1.25 (s, 6H), 0.85 (d,  $J = 6.6$  Hz, 6H).

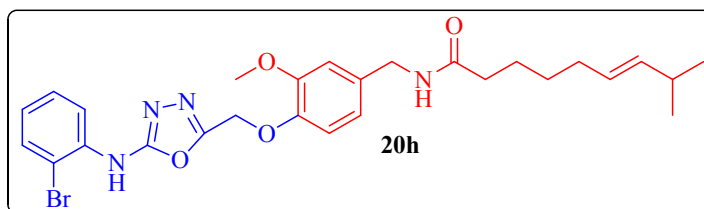
MS (ESI)  $m/z$ :  $[\text{M}+1]^+$  509.

Anal. Calcd for  $\text{C}_{28}\text{H}_{36}\text{N}_4\text{O}_5$ : C, 66.12; H, 7.13; N, 11.02; O, 15.73%, Found: C, 66.07; H, 7.06; N, 10.99%.



### 1.3.4.8. Synthesis of (*E*)-*N*-(4-((5-((2-bromophenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (20h)

According to general procedure A, Compound **19h** was cyclized under coupling conditions with EDC.HCl and HOBT to yield desired compound **20h**.



White solid, yield: 83%.

Mp: 145.5 °C.

IR  $\text{cm}^{-1}$ : 3450, 3410, 3371, 3320, 3239, 3194, 3133, 3089, 3055, 2918, 1628, 1592, 1514, 1451, 1384, 1263, 1222, 1141, 1025, 846, 808, 749, 683, 593, 565.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 7.71 (d,  $J = 1.2$  Hz, 1H), 7.22-7.19 (m, 3H), 6.94 (d,  $J = 7.8$  Hz, 1H), 6.81 (s, 1H), 6.73 (d,  $J = 8.1$  Hz, 1H), 5.82 (s, 1H), 5.19 (s, 2H), 4.36 (d,  $J = 5.4$  Hz, 2H), 3.82 (s, 3H), 2.20 (t,  $J = 7.2$  Hz, 2H), 1.43 (t,  $J = 6.9$  Hz, 2H), 1.28-1.25 (m, 7H), 0.85 (d,  $J = 6.3$  Hz, 6H).

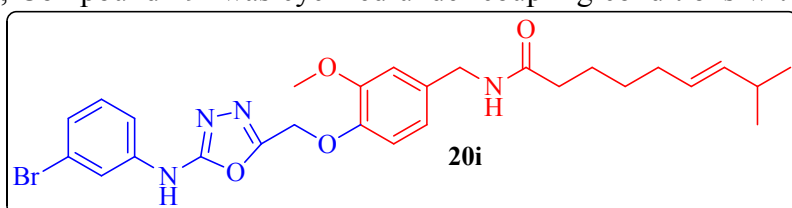
$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 173.3, 160.3, 156.6, 150.1, 145.9, 138.7, 133.7, 130.6, 128.3, 126.1, 123.0, 122.5, 120.4, 119.8, 116.1, 115.7, 111.8, 61.2, 55.8, 43.2, 38.9, 36.8, 29.6, 27.9, 27.2, 25.8, 22.6, 14.0.

HRMS (ESI)  $m/z$ :  $[\text{M}]^+$  Calcd for  $\text{C}_{27}\text{H}_{33}\text{BrN}_4\text{O}_4$  556.1685; Found 556.1688.

Anal. Calcd for  $\text{C}_{27}\text{H}_{33}\text{BrN}_4\text{O}_4$ : C, 58.17; H, 5.97; Br, 14.33, N, 10.05; O, 11.48 %, Found: C, 58.07; H, 5.90; N, 10.01%.

### 1.3.4.9. Synthesis of (*E*)-*N*-(4-((5-((3-bromophenyl)amino)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (20i)

According to general procedure A, Compound **19i** was cyclized under coupling conditions with EDC.HCl and HOBT to yield desired compound **20i**.



White solid, Yield 93%.

Mp: 145.7 °C.

IR  $\text{cm}^{-1}$ : 3482, 3453, 3416, 3379, 3332, 3275, 3275, 3237, 3169, 3072, 3039, 2925, 1644, 1621, 1571, 1543, 1471, 1418, 1387, 1333, 1266, 1221, 1137, 1096, 1026, 849, 805, 772, 716, 657, 615, 555.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 7.71 (s, 1H), 7.44-7.42 (m, 1H), 7.20 (d,  $J = 4.5$  Hz, 2H), 6.94 (d,  $J = 8.1$  Hz, 1H), 6.80 (s, 1H), 6.72 (d,  $J = 9.0$  Hz, 1H), 5.85 (s, 1H), 5.19 (s, 2H), 4.36 (d,  $J = 5.1$  Hz, 2H), 3.82 (s, 3H), 2.21 (t,  $J = 7.5$  Hz, 2H), 1.67-1.60 (m, 2H), 1.52-1.43 (m, 1H), 1.25 (s, 6H), 0.85 (d,  $J = 6.3$  Hz, 6H).

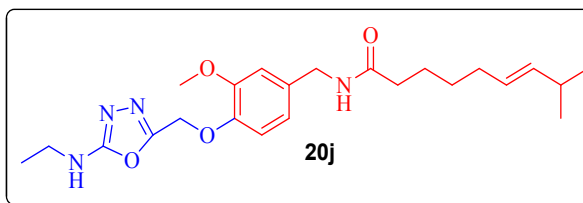
$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 173.3, 160.3, 156.6, 150.1, 145.9, 138.7, 133.7, 130.6, 128.3, 126.1, 123.0, 122.5, 120.4, 119.8, 116.1, 115.7, 111.8, 61.2, 55.8, 43.2, 38.9, 36.8, 29.6, 27.9, 27.2, 25.8, 22.6, 14.0.

MS (ESI)  $m/z$ :  $[\text{M}]^+ 557.3$ ,  $[\text{M}+1]^+ 558$ ,  $[\text{M}+2]^+ 559.1$ .

Anal. Calcd for  $\text{C}_{27}\text{H}_{33}\text{BrN}_4\text{O}_4$ : C, 58.17; H, 5.97; Br, 14.33, N, 10.05; O, 11.48 %, Found: C, 58.11; H, 5.94; N, 9.98%.

#### 1.3.4.10. Synthesis of (*E*)-*N*-(4-((5-(ethylamino)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (20j)

According to general procedure A, Compound **19j** was cyclized under coupling conditions with EDC.HCl and HOBT to yield desired compound **20j**.



White solid, yield: 87%.

Mp: 106 °C.

IR  $\text{cm}^{-1}$ : 3483, 3458, 3413, 3379, 3343, 3307, 3259, 3212, 3143, 3107, 2924, 1631, 1557, 1516, 1462, 201, 1260, 1221, 1145, 1090, 1026, 855, 798, 721, 689, 649, 554.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 6.94 (d,  $J = 7.8$  Hz, 1H), 6.85 (s, 1H), 6.78 (d,  $J = 7.2$  Hz, 1H), 5.80 (s, 1H), 5.33 (s, 1H), 4.38 (d,  $J = 5.4$  Hz, 2H), 4.27-4.20 (m, 2H), 3.83 (s, 3H), 2.20 (t,  $J = 7.2$  Hz, 2H), 1.65 (t,  $J = 5.4$  Hz, 2H), 1.54-1.48 (m, 1H), 1.42 (t,  $J = 7.2$  Hz, 3H), 1.27 (s, 6H), 0.85 (d,  $J = 6.6$  Hz, 6H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 173.1, 168.2, 150.3, 148.0, 145.6, 134.0, 120.0, 115.9, 111.9, 62.0, 55.7, 43.3, 39.9, 38.9, 36.8, 29.6, 29.3, 27.9, 27.2, 25.8, 22.6, 13.5.

MS (ESI)  $m/z$ :  $[\text{M}+1]^+ 431.2$ .

Anal. Calcd for  $\text{C}_{23}\text{H}_{34}\text{N}_4\text{O}_4$ : C, 64.16; H, 7.96; N, 13.01; O, 14.86 %, Found: C, 64.08; H, 7.91, N, 12.99%.

### 1.3.4.11. Synthesis of (*E*)-*N*-(4-((5-(butylamino)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (**20k**)

According to general procedure A, Compound **19k** was cyclized under coupling conditions with EDC.HCl and HOBT to yield desired compound **20k**.

Light grey; solid, yield: 91%.

Mp: 145.6 °C.

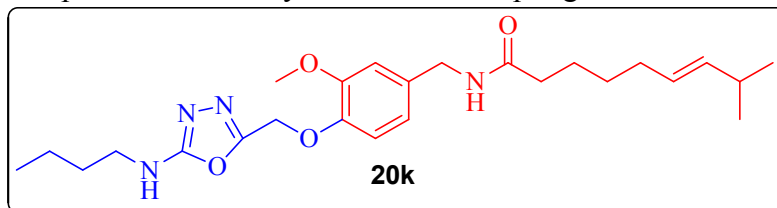
IR  $\text{cm}^{-1}$ : 3449, 3341, 3281, 3161, 3108, 3077, 3003, 2921 2848, 1636, 1558, 1502, 1460, 1355, 1262, 1213, 1147, 1113, 1029, 997, 860, 791, 747, 656, 599, 546.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 6.93 (d,  $J = 8.1$  Hz, 1H), 6.85 (s, 1H), 6.77 (d,  $J = 7.2$  Hz, 1H), 5.81 (s, 1H), 5.09 (s, 2H), 4.38 (d,  $J = 5.1$  Hz, 2H), 4.15 (t,  $J = 7.5$  Hz, 2H), 3.83 (s, 3H), 2.22 (t,  $J = 7.2$  Hz, 2H), 1.82 (t,  $J = 6.3$  Hz, 2H), 1.65 (t,  $J = 5.7$  Hz, 2H), 1.54-1.37 (m, 3H), 1.28 (s, 6H), 0.85 (d,  $J = 6.6$  Hz).

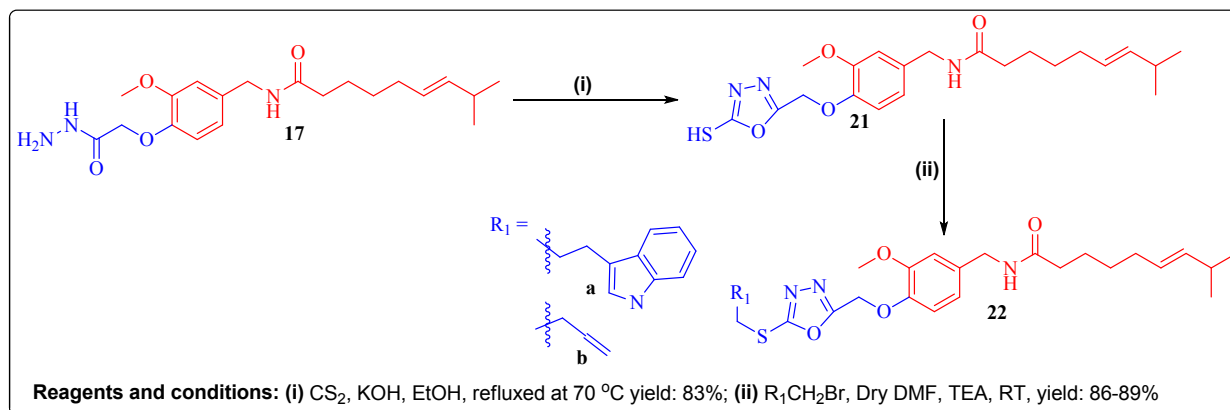
$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 173.1, 168.4, 150.3, 148.1, 145.6, 134.0, 120.0, 116.0, 111.9, 62.0, 55.7, 44.5, 43.3, 39.9, 38.9, 36.8, 30.2, 29.6, 29.3, 27.9, 27.2, 25.8, 22.6, 20.0, 13.6.

MS (ESI)  $m/z$ :  $[\text{M}+3]^+$  461.1.

Anal. Calcd for  $\text{C}_{23}\text{H}_{34}\text{N}_4\text{O}_4$ : C, 65.48; H, 8.35; N, 12.22; O, 13.96 %, Found: C, 65.41; H, 8.29; N, 12.18%.

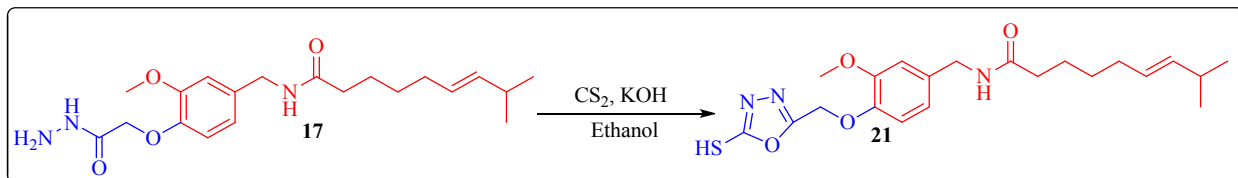


### 1.4. Scheme S-2. Synthesis of compound **21**, **22a-b**.



Scheme S-2

#### 1.4.1. Synthesis of (*E*)-*N*-(4-((5-mercapto-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (**21**)



To ethanolic solution of carbon disulphide (0.21g), 0.16g of potassium hydroxide was added and the reaction mixture was stirred at room temperature for 30 min. After 30 min, 0.5g of capsaicin hydrazide **17** was added to the reaction mixture and was refluxed at 70 °C. The completion of reaction was monitored by TLC (9:1 ratio of ethyl acetate:methanol). Reaction mixture was concentrated under rota-evaporator and the concentrated reaction mixture was poured into ice. White solid precipitates were formed and filtered under vacuum pump and dried.

White solid, yield: 83%.

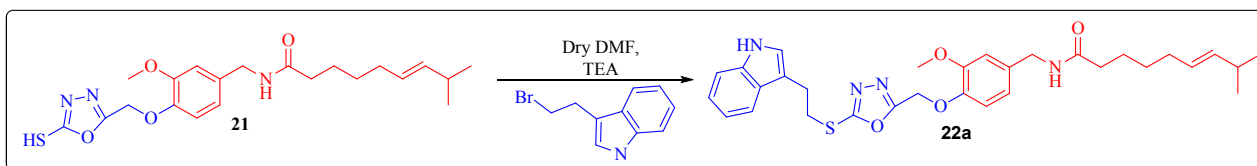
Mp: 160.3 °C.

IR  $\text{cm}^{-1}$ : 3459, 3395, 3311, 3281, 3224, 3189, 3093, 2955, 2927, 1681, 1638, 1514, 1481, 1377, 1265, 1232, 1143, 1025, 911, 800, 707, 682, 637, 602, 540.

$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 9.51 (s, 1H), 6.89-6.76 (m, 3H), 5.60 (s, 1H), 5.33-5.18 (m, 2H), 4.60 (s, 2H), 4.31 (s, 2H), 3.82 (s, 3H), 2.14 (t,  $J = 6.9$  Hz, 2H), 1.93-1.88 (m, 1H), 1.59-1.47 (m, 6H), 0.87 (d,  $J = 6.3$  Hz, 3H), 0.63 (d,  $J = 6$  Hz, 3H).

MS (ESI)  $m/z$ :  $[\text{M}-1]^+$  418.4.

#### 1.4.2. Synthesis of (*E*)-*N*-(4-(((2-(1H-indole-3-yl)ethyl)thio)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (**22a**)



0.2 g of compound **21** was added to dry DMF, 0.12 g of 3-(2-bromoethyl)indole and 9.4  $\mu\text{L}$  of triethylamine were added to the reaction mixture and was stirred at room temperature until the reaction get complete. Completion of reaction was monitored by TLC (7:3 ratio of ethyl

acetate:hexane). Reaction mixture was poured into ice. A white solid precipitates were formed and filtered and dried under vacuum.

White solid, yield: 86%.

Mp: 109.4 °C.

IR  $\text{cm}^{-1}$ : 3470, 3431, 3385, 3323, 3171, 3122, 3086, 3057, 2932, 1643, 1544, 1513, 1456, 1420, 1318, 1252, 1216, 1143, 1026, 964, 923, 843, 808, 765, 736, 656, 600, 558.

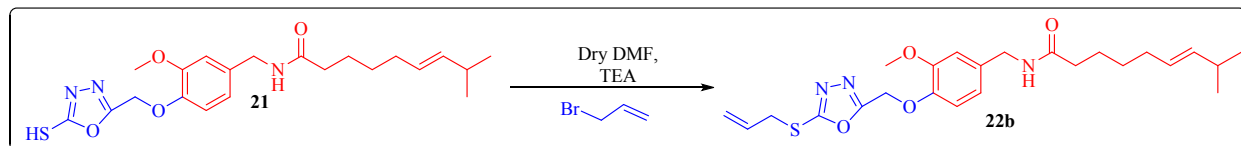
$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 8.27 (s, 1H), 7.63 (d,  $J = 7.8$  Hz, 1H), 7.35 (d,  $J = 7.8$  Hz, 1H), 7.21-7.12 (m, 2H), 7.02 (s, 1H), 6.95 (d,  $J = 8.1$  Hz, 1H), 6.83 (s, 1H), 6.76 (d,  $J = 8.4$  Hz, 1H), 5.72 (s, 1H), 5.41-5.18 (m, 3H), 4.36 (d,  $J = 5.7$  Hz, 2H), 3.82 (s, 3H), 3.54 (t,  $J = 6.9$  Hz, 2H), 3.26 (t,  $J = 7.2$  Hz, 2H), 2.20 (t,  $J = 7.2$  Hz, 2H), 2.01-1.95 (m, 1H), 1.70-1.60 (m, 3H), 1.43-1.25 (m, 4H), 0.94 (d,  $J = 6.6$  Hz, 4H), 0.85 (d,  $J = 6.6$  Hz, 2H).

$^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$ : 172.9, 166.1, 163.3, 150.2, 146.0, 138.1, 136.2, 133.8, 126.9, 126.4, 122.1, 119.9, 119.5, 118.6, 115.9, 113.1, 111.8, 111.3, 61.3, 55.8, 43.2, 38.9, 36.6, 33.2, 32.2, 30.9, 29.6, 29.2, 27.9, 27.2, 25.5, 25.2, 22.6.

HRMS (ESI)  $m/z$ :  $[\text{M} + \text{H}]^+$  Calcd for  $\text{C}_{31}\text{H}_{39}\text{N}_4\text{O}_4\text{S}$  563.2692; Found 563.2692.

Anal. Calcd for  $\text{C}_{31}\text{H}_{38}\text{N}_4\text{O}_4\text{S}$ : C, 66.17; H, 6.81; N, 9.96; O, 11.37; S, 5.70 %, Found: C, 66.10; H, 6.76; N, 9.91; S, 5.67%.

### 1.4.3. Synthesis of (*E*)-*N*-(4-((5-(allylthio)-1,3,4-oxadiazol-2-yl)methoxy)-3-methoxybenzyl)-8-methylnon-6-enamide (22b)



0.2 g of compound **21** was added to dry DMF, 0.07 g of allylbromide and 9.4  $\mu\text{L}$  of triethylamine was added to the reaction mixture and was stirred at room temperature until the reaction get complete. Completion of reaction was monitored by TLC (7:3 ratio of ethyl acetate:hexane). Reaction mixture was poured into ice. A white solid precipitates were formed and filtered and dried under vacuum.

White solid, yield: 89%.

Mp: 84.7 °C.

IR  $\text{cm}^{-1}$ : 3478, 3407, 3364, 3292, 3124, 3085, 3005, 2960, 2925, 1633, 1519, 1469, 1419, 1384, 1269, 1221, 1169, 1140, 1026, 979, 918, 849, 796, 730, 700, 667, 632, 585, 532.

$^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm): 6.97 (d,  $J = 8.1$  Hz, 1H), 6.84 (s, 1H), 6.77 (d,  $J = 8.1$  Hz, 1H), 6.03-5.90 (m, 1H), 5.67 (s, 1H), 5.41-5.31 (m, 2H), 5.24 (s, 2H), 4.37 (d,  $J = 5.7$  Hz, 2H), 3.85 (m, 4H), 2.21 (t,  $J = 7.5$  Hz, 2H), 1.99 (q,  $J = 7.2$  Hz, 1H), 1.71-1.59 (m, 4H), 1.28-1.25 (m, 5H), 0.95 (d,  $J = 6.9$  Hz, 3H), 0.85 (d,  $J = 6.6$  Hz, 3H).

HRMS (ESI)  $m/z$ :  $[\text{M} + \text{H}]^+$  Calcd for  $\text{C}_{24}\text{H}_{35}\text{N}_3\text{O}_4\text{S}$  460.2270; Found 460.2279.

Anal. Calcd for  $\text{C}_{24}\text{H}_{33}\text{N}_3\text{O}_4\text{S}$ : C, 62.72; H, 7.24; N, 9.14; O, 13.92; S, 6.98 %, Found: C, 62.69; H, 7.19; N, 9.09; S, 6.94%.

## 2. Biology

### 2.1. *In vitro* anti-proliferative assay

#### 2.1.1. *In vitro* anti-proliferative activity at single dose

All the synthesized compounds were screened for their anti-proliferative activity against a panel of 60 cancer cell line at National Cancer Institute, Bethesda, MD, USA as per the standard procedure given at <http://www.dtp.nci.nih.gov><sup>1</sup>. RPMI 1640 medium (5% fetal bovine serum and 2 mM L-glutamine) was used to grow the human tumor cell lines. All the tumor cells were incubated into 96-well microtiter plate. Then this plated was placed for incubation at 37 °C for 24 hour. After that two plates of each cell line were fixed with TCA *in situ* and optical density was measured at this point which represented the cell population of each cell line at the time of compound addition (ODzero). On the other hand, all the tested compounds were dissolved in DMSO to yield 400-fold desired final concentration and stored at -80°C. These frozen compounds were thawed and their aliquot part was diluted to  $10^{-4}$  M concentration with the medium containing 50  $\mu\text{g}/\text{mL}$  of gentamicin at the time of compound addition. Control sample was made with DMSO only. 100  $\mu\text{l}$  of the tested compounds from the aliquot parts were added to appropriate 96-well microtiter plate containing 100  $\mu\text{l}$  of medium ensuing in the required final drug concentrations of  $10^{-5}$  M and 0 M (control). After addition of tested compounds, 96-well microtiter plate was incubated for 48 h at 100%, 5%  $\text{CO}_2$ , 95% air, 100% relative humidity. Cold TCA was used to stop the assay for adherent cells. Further on 50 mL of 50% (w/v) TCA was used to fix the cell and incubated for 1 h at 4 °C. The supernatant was removed, and the 96-well microtiter plates were rinsed five times with water and air dried. 100 mL solution of protein binding dye, Sulforhodamine B (SRB) was made at 0.4% (w/v) in 1% acetic acid and was added to each well of the plates. These Plates were placed at room temperature for incubation for 10

minutes then were washed with 1% acetic acid five times to remove unbound dye. Then the plates were treated with 10 Mm trizma base, so that unbound dye was solubilized with trizma base. The absorbance was measured at a wavelength of 515 nm on an automated plate reader and results for each tested compounds were calculated as the percent of tumor growth of the treated cells in comparison with the untreated control cells. Optical density (OD) was recored for SRB-derived color just before exposing the cells to the test compound (ODzero) and after 48 h exposure to the test compound (OD test) or the control vehicle (OD ctrl).



**Table S1:** Growth percentage against NCI panel of 60 human cancer cell lines at 10  $\mu$ M of all the synthesized compounds

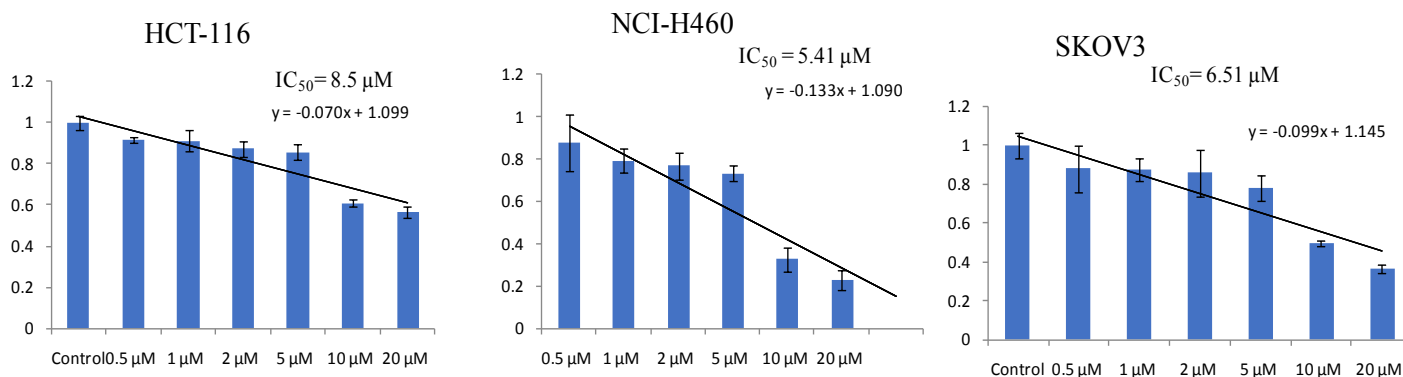
Sub panel cancer cell line	Growth percentage													
	20a	20b	20c	20d	20e	20f	20g	20h	20i	20j	20k	21	22a	22b
<b>Leukemia</b>														
CCRF-CEM	78.85	83.38	91.92	82.95	92.76	91.99	nt	97.26	81.76	97.54	91.34	99.62	<b>34.67</b>	90.64
HL-60(TB)	101.03	89.41	89.08	89.70	99.69	81.09	98.57	89.55	79.77	98.36	86.64	103.37	<b>50.23</b>	98.26
K-562	59.62	70.62	87.28	60.36	70.24	67.95	84.49	81.01	65.02	91.59	85.73	93.12	<b>40.71</b>	70.34
MOLT-4	84.22	88.02	89.91	82.05	103.71	76.74	98.19	86.25	83.54	103.67	86.36	Nt	<b>29.16</b>	83.83
RPMI-8226	52.34	74.91	84.57	64.91	84.21	70.71	93.01	88.54	70.31	95.52	76.77	99.31	Nt	Nt
SR	64.20	74.53	84.76	67.28	89.46	76.06	95.22	80.43	67.67	89.02	71.54	95.22	<b>49.30</b>	80.53
<b>Non-small cell lung cancer</b>														
A549/ATCC	<b>49.02</b>	87.33	102.17	105.30	97.53	99.28	99.76	100.52	95.38	97.00	100.32	101.17	79.42	98.75
EKVX	74.70	88.18	101.13	77.14	93.21	85.03	91.18	90.94	78.09	96.74	98.76	96.09	57.36	94.12
HOP-62	<b>0</b>	90.21	88.26	56.40	87.34	79.11	106.83	99.73	75.58	98.27	78.66	110.73	92.77	82.37
NCI-H226	62.14	66.28	85.01	64.53	66.49	79.74	85.73	83.50	<b>50.41</b>	101.46	84.32	100.61	67.25	91.66
NCI-H23	62.14	77.86	90.57	63.80	82.89	92.25	92.09	87.06	66.01	93.67	93.14	100.87	66.57	96.56
NCI-H322M	54.87	101.74	82.91	86.07	83.98	97.37	85.45	94.14	98.04	107.20	90.56	104.13	88.69	105.55
NCI-H460	<b>33.48</b>	97.70	103.15	80.34	97.70	104.67	105.96	93.22	97.55	102.79	102.28	106.50	89.80	105.48
NCI-H522	72.97	73.96	85.69	60.29	78.59	80.95	84.61	76.81	68.60	83.70	83.69	99.00	59.35	89.50
<b>Colon cancer</b>														
COLO 205	70.91	113.34	105.02	104.24	98.70	125.25	123.61	113.16	109.93	110.12	112.79	103.84	71.22	110.75
HCC-2998	83.57	100.28	102.82	90.82	94.58	105.27	103.51	99.78	100.83	102.90	100.87	107.49	97.97	112.22
HCT-116	<b>24.55</b>	78.21	95.48	53.45	84.11	86.76	98.15	64.86	67.59	87.16	95.61	102.79	69.99	98.01
HCT-15	81.17	95.64	103.31	90.10	92.38	88.33	94.13	98.87	90.22	96.85	96.64	97.59	68.56	95.32
HT29	71.71	98.66	99.81	89.95	93.76	102.87	109.88	97.63	97.66	104.84	109.45	113.25	69.68	109.65
KM12	71.65	95.09	98.42	78.80	96.75	95.47	92.43	82.88	90.31	98.25	100.69	101.75	73.28	103.88
SW-620	67.32	105.60	103.77	98.94	97.91	105.15	103.10	94.64	95.44	109.92	101.45	100.81	84.70	96.35
<b>CNS cancer</b>														
SF-268	79.65	88.42	98.18	<b>45.48</b>	87.38	90.77	89.30	90.35	92.63	97.15	102.42	100.57	78.89	97.08
SF-295	64.90	89.63	98.07	<b>48.03</b>	88.34	88.79	89.97	93.76	84.25	97.13	102.61	100.13	68.78	101.54
SF-539	70.24	103.25	94.45	<b>48.16</b>	86.63	93.38	98.35	90.97	88.44	103.92	99.53	101.54	80.83	101.59
SNB-19	54.05	99.28	96.48	58.55	86.86	94.57	96.21	90.61	99.55	103.83	94.52	97.90	77.05	95.53
SNB-75	<b>48.71</b>	80.51	81.58	57.78	68.43	94.51	80.03	95.25	83.48	86.23	88.42	83.81	83.14	83.94
U251	<b>48.58</b>	93.59	103.48	83.96	99.62	101.74	101.89	88.97	101.81	98.29	101.12	102.68	80.52	100.05
<b>Melanoma</b>														
LOX IMVI	55.24	92.37	92.05	83.41	90.96	95.63	92.06	94.00	88.92	93.19	92.01	100.41	55.24	95.91
M14	69.64	98.83	98.52	99.73	90.34	95.41	98.88	101.43	93.30	101.59	94.96	92.91	Nt	Nt
MDA-MB-435	66.39	104.85	103.53	100.01	99.57	103.20	102.02	105.39	101.19	104.46	100.39	100.51	90.22	98.38
SK-MEL-2	71.89	103.66	106.15	111.03	94.60	101.56	104.40	101.46	104.61	103.70	104.03	108.93	86.68	94.98
SK-MEL-28	78.87	98.50	94.43	85.18	90.32	98.06	99.09	99.16	107.28	103.96	97.23	110.77	104.42	106.07
SK-MEL-5	<b>46.26</b>	97.12	99.88	86.57	96.15	98.00	101.19	100.83	89.84	102.38	99.66	Nt	105.37	108.83
UACC-257	77.56	98.76	105.06	98.18	99.66	96.70	102.09	106.47	105.03	105.33	99.62	105.61	100.67	103.40
UACC-62	<b>39.61</b>	76.93	76.09	65.48	73.92	81.95	78.65	83.60	67.54	94.98	84.54	97.76	67.51	81.27
<b>Ovarian cancer</b>														
IGROV1	53.49	99.02	89.70	58.44	88.25	97.77	86.79	91.27	92.89	102.86	103.25	97.96	73.71	95.03
OVCAR-3	77.31	97.45	103.80	68.29	94.73	89.64	97.64	104.92	87.99	105.82	98.26	110.88	88.32	106.81
OVCAR-4	<b>11.25</b>	85.83	96.58	<b>0.69</b>	84.23	101.24	95.28	97.29	85.10	103.05	94.12	99.42	71.67	99.46
OVCAR-5	100.97	93.32	94.10	77.67	90.36	89.67	94.23	92.26	91.08	103.14	93.11	105.20	101.99	102.84
OVCAR-8	<b>24.96</b>	99.66	100.55	<b>45.17</b>	100.44	100.61	97.55	98.66	98.03	105.88	99.16	101.33	71.26	96.48
NCI/ADR-RES	79.62	96.09	100.02	<b>43.35</b>	93.66	89.81	95.71	90.56	87.04	102.01	94.80	103.36	56.51	90.53
SK-OV-3	<b>11.82</b>	88.19	78.61	<b>48.61</b>	85.35	107.43	86.46	97.96	82.58	91.51	94.35	103.46	85.74	107.07
<b>Renal cancer</b>														
786-0	<b>22.25</b>	89.17	97.65	<b>11.98</b>	85.60	96.22	94.33	95.33	95.60	95.46	103.39	104.31	89.26	98.36
A498	56.81	77.11	114.46	92.53	97.63	102.89	111.00	101.93	77.79	108.94	110.55	105.56	73.28	93.70
ACHN	<b>41.17</b>	92.42	91.36	56.05	88.30	95.65	90.34	95.45	85.14	101.38	100.13	100.49	76.55	97.61
CAKI-1	<b>33.60</b>	Nt	Nt	Nt	Nt	Nt	Nt	Nt	Nt	Nt	Nt	Nt	Nt	Nt
RXF 393	54.11	109.40	119.47	68.76	105.35	102.60	101.82	96.06	70.61	120.40	117.01	85.08	54.00	Nt
SN 12C	<b>49.04</b>	102.91	89.04	70.47	86.50	105.33	97.41	97.94	95.72	104.29	95.08	97.66	83.06	95.06
TK-10	84.10	102.60	146.30	97.67	100.39	103.22	129.89	118.10	104.24	106.57	111.63	106.13	91.69	107.78
UO-31	58.74	77.39	68.08	52.79	68.66	75.24	64.27	74.38	64.01	84.23	79.13	76.93	<b>45.03</b>	77.23
<b>Prostate cancer</b>														
PC-3	<b>37.64</b>	93.71	93.53	72.50	92.51	84.37	103.14	103.24	87.90	103.73	96.62	98.96	56.85	93.63
DU-145	62.03	101.73	103.12	88.70	98.76	98.97	100.78	101.15	89.79	109.00	103.03	105.86	94.93	103.35
<b>Breast cancer</b>														
MCF7	55.23	93.28	95.59	66.88	90.88	87.12	88.37	82.71	83.70	95.22	90.76	95.31	58.79	98.26
MDA-MB-231/ATCC	67.10	88.44	84.33	63.87	81.41	99.24	81.96	92.54	78.30	103.58	92.62	102.36	70.64	97.59
HS 578T	63.17	92.30	100.61	83.68	91.01	98.03	93.97	96.68	93.67	98.58	95.40	94.96	81.05	89.11
BT-549	108.01	95.78	96.92	76.95	80.99	85.59	88.24	100.93	76.43	102.29	93.67	113.07	75.03	103.85
T-47D	<b>48.58</b>	66.28	79.31	65.02	69.56	76.11	92.81	85.60	58.86	81.04	88.82	97.17	<b>42.98</b>	93.68
MDA-MB-468	53.75	78.84	105.40	58.92	88.47	99.71	105.45	96.37	48.07	104.24	87.42	111.40	<b>51.81</b>	98.31
<b>Mean GP</b>	59.69	91.19	95.99	71.72	89.51	93.21	96.03	93.94	85.48	99.24	95.63	101.05	72.88	96.78

nt = not tested; GP = Growth Percentage

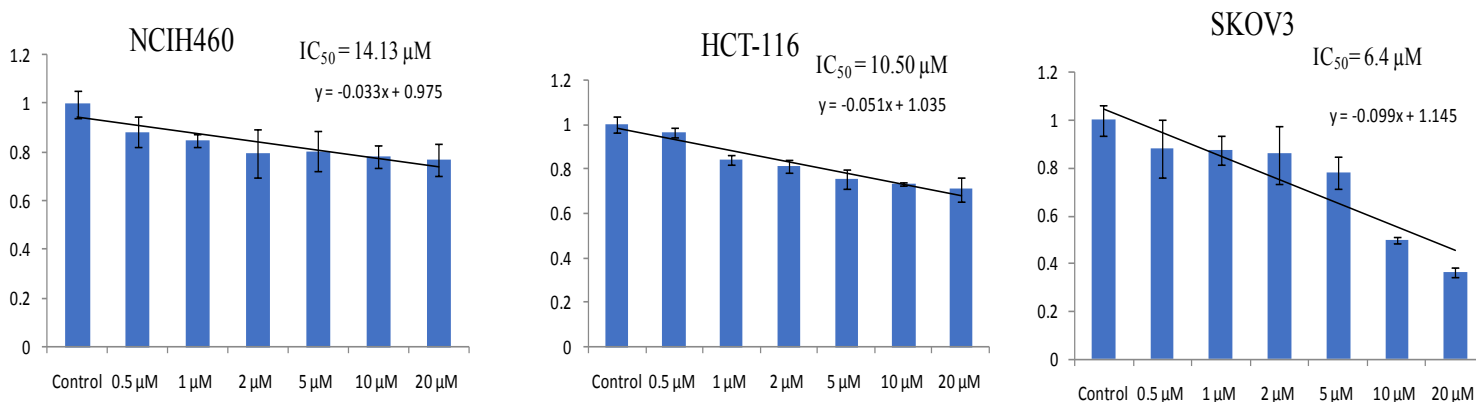
### 2.1.2. Crystal violet assay

The cell viability of compound **20a** against HCT-116 cancer cell line, NCI-H460 cancer cells, SKOV3 cancer cell line and normal PNT2 cells (normal prostatic epithelial cells) was measured by crystal violet assay. Approx.  $4 \times 10^3$  were seeded in each well of 96 well plates. After 24 h, the cells were treated with different concentrations of compound **20a** and further incubated for 72 h. Following this, the media was removed and cells were stained with 0.4% crystal violet solution (prepared in 50% methanol) and the cells were stained for 30 minutes at room temperature. Plates were then washed gently with tap water and air dried at room temperature without lid for 24 h. Next, 100  $\mu$ l of methanol was added and incubated at room temperature for 15 minutes on bench rocker, followed by measuring the absorbance of dissolved dye at 570 nm. Cell viability was calculated as fold change in absorbance values of compound **20a** treated cells with respect to control<sup>2</sup>.

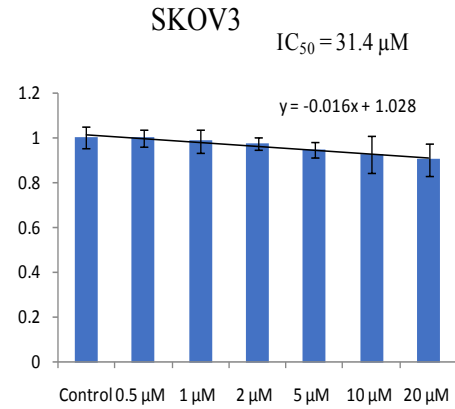
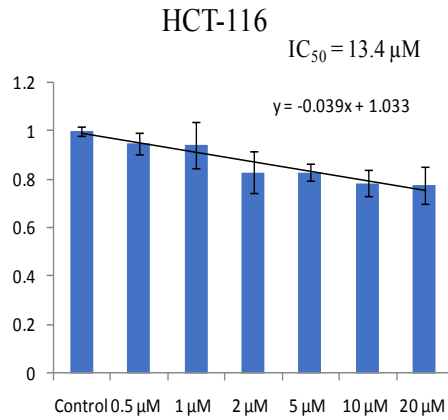
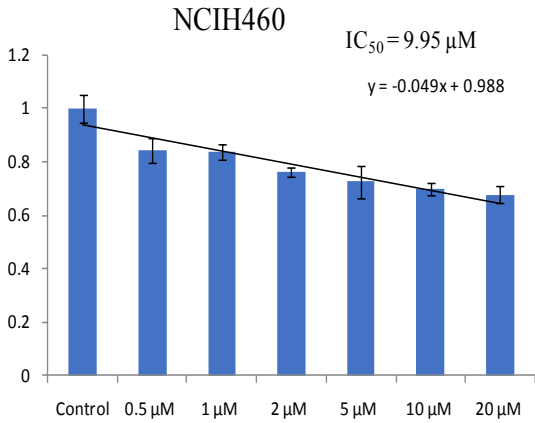
#### Dose response curves for compound 20a



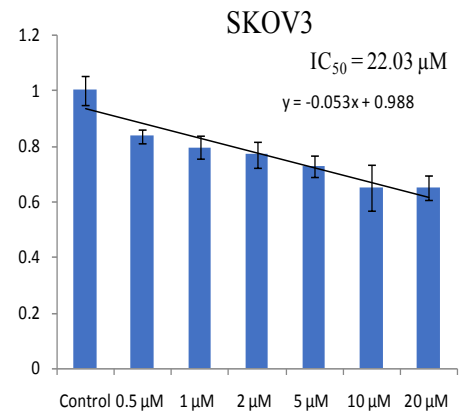
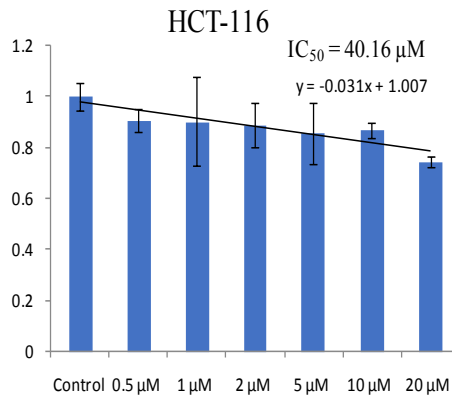
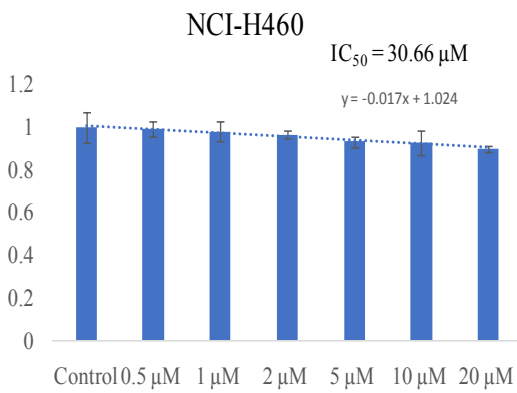
#### Dose response curves for compound 20d



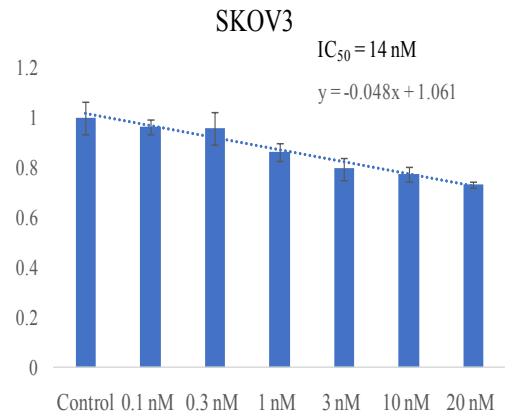
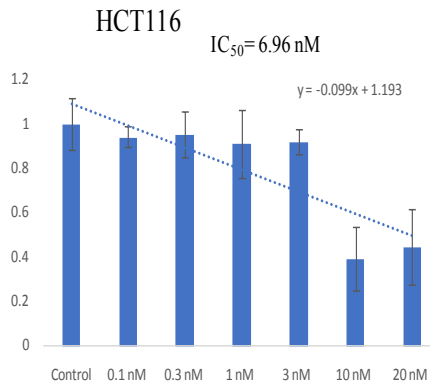
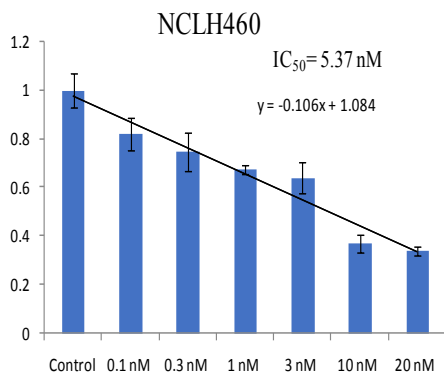
## Dose response curves for compound 22a



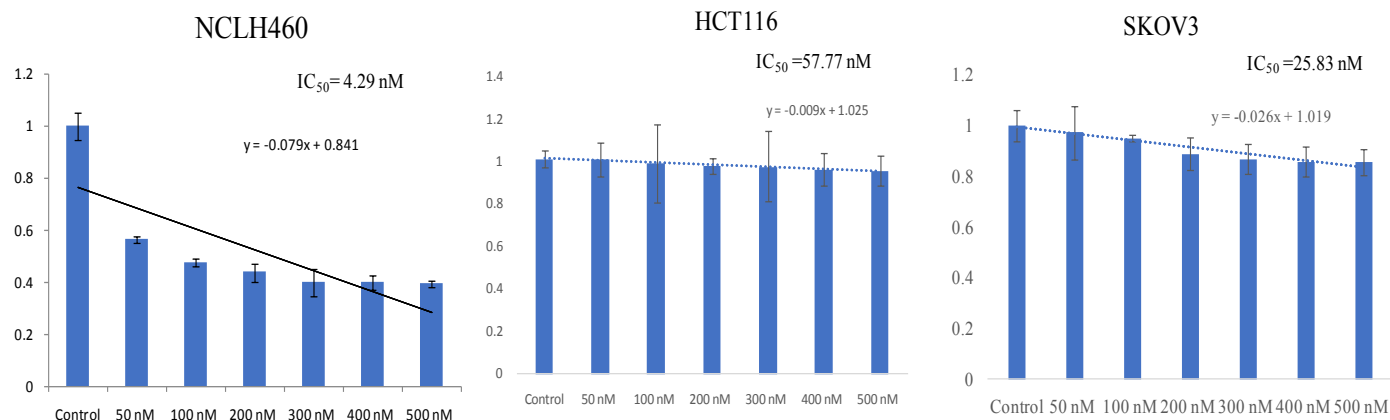
## Dose response curve for Capsaicin



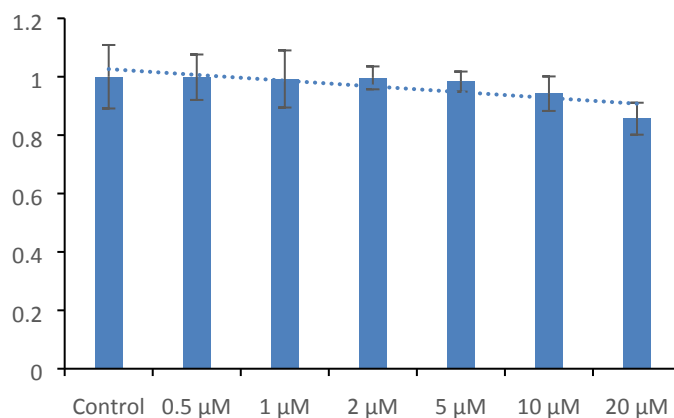
## Dose response curve for Combretastatin



## Dose response curve for Doxorubicin



## Dose response curve for Compound 20a against normal PNT2 cells (normal prostatic epithelial cells)



### 2.1.3. Colony formation assay

NCI-H460 cells were seeded in six-well plate at a density of 500 cells per well and caused adherence of the cells (left for overnight), these cells were treated with different concentration of **20a**. After every 72 h, the media was changed and the cells were treated with **20a** and these experiments lasted for 15 days. After the development of colony, the wells were washed with PBS, fixed with 3.7% formaldehyde for 20 min, washed with PBS and then were stained using 0.4% crystal violet followed by rinsing the wells with PBS 4 to 5 times. The colonies were counted by using Image J software <sup>3</sup>.

### 2.2. Evaluation of Mechanistic Studies

### 2.2.1. Flow cytometry for assessing intracellular ROS

The levels of intracellular reactive oxygen species (ROS) were measured following **20a** treatment. After treatment, the cells were incubated for 24h followed by trypsinization of cells and re-suspension in pre-warmed PBS containing probe CM-H<sub>2</sub>DCFDA for 30 minutes in the dark at 37°C. The fluorescence intensity was measured using flow cytometer <sup>4</sup>.

### 2.2.2. Cell cycle analysis NCI-H460 cells

Effect of **20a** on the cell cycle of cell line was evaluated by using flow cytometry at 5 μM, 10 μM, and 20 μM, concentrations. These cells were seeded into 6 well plates and placed for incubation at 37 °C for 24 h which lead to attachment of cells with 50-60% confluency. Further on it was treated with vehicle control 0.1% DMSO and **20a** (0.5, 1, 2, 5 and 10 μM) and incubated for 72 h. The collected cells were harvested and fixed with 75% ice cold ethanol at 4°C. Excess of ethanol was removed and cells were washed with PBS. Cells were treated and stained with RNase (50 U/mL) and Propidium Iodide (20 μg/mL) staining solution for 20 min at room temperature in dark and was further subjected for flow cytometry (Bectone Dickinson, FACS Verse) <sup>5</sup>.

### 2.2.3. Cell apoptosis assay

To assess the effect of compound **20a** treatment on apoptosis, Annexin V-FITC and Propidium iodide staining was performed and the cells were analyzed using FACS verse (BD). Briefly, NCI-H460 cells were treated with compound **20a** at 5 μM, 10μM and 20μM and the cells were incubated for 72 h at 37°C and 5% CO<sub>2</sub>. Next, the cells were harvested and incubated with 1X binding buffer containing Annexin-V FITC and propidium for 30 minutes. The samples were acquired for cell death analysis using FACS *verse* (BD) <sup>6</sup>.

### 2.2.4. Wound healing assay

NCI-H460 cells (3 x10<sup>5</sup>) were seeded and grown to each well of 6 well plates until mono-layer was formed. Then by using 200 μL pipette tip, a scratch was given in confluent mono layer. Cells were washed, to remove non-adherent cell debris. Cells were further treated with 5μM, 10 μM and 20 μM of compound **20a**. Cell migration across the wound area were captured with an inverted microscope and photographed for indicated time (0, and 24 h). The percentage wound healed area was calculated after normalizing the wound area <sup>7</sup>.

### 2.2.5. Western blotting

NCI-H460 Cells treated with compound **20a** were lysed after 72 h of incubation period with 2X SDS lysis buffer containing 0.5 M Tris-HCl, pH 6.8, glycerol, 10 % (w/v) SDS, protease inhibitor cocktail. The lysates were sonicated and centrifuged at 16000 g for 20 min following which supernatant was collected and subjected to protein estimation by Pierce™ BCA assay kit (ThermoFisher). 30-40 µg of proteins were separated on SDS gel and transferred to PVDF membranes. Membranes were exposed to blocking buffer containing 5 % skim milk and probed with specific primary antibodies at 4 °C overnight. Next, the blots were washed three times for 5 minutes with Tris buffer solution with tween-20 (TBST) and incubated with HRP-conjugated secondary antibody for 1 h. The blots were washed three times with 1x TBST and the bands were detected by ECL Elistar ETA ultra 20 (Cyanagen) <sup>8</sup>.

### 2.26. RNA isolation and quantitative PCR

TRIzol reagent was used to isolate total RNA from the control and drug treated NCI-H460 cells. Following that, cDNA was synthesized from 2g of total RNA using a cDNA synthesis kit (Takara), according to the manufacturer's instructions. **Table S4** lists below the primers used against the genes under investigation.

**Table S2:** Lists the primers used against the genes under investigation.

GENE	FORWARD PRIMER SEQUENCE	REVERSE PRIMER SEQUENCE
GAPDH	5'-GTCAAGGCTGAGAACGGGAA-3'	5'-AGTGGCTCCATTCACCGC-3'
VEGFR2	5'-GGAACCTCACTATCCGCAGA-3'	5'-CCGCCGTGCCTACTAGAATA-3'

The PCR cycling schedule was 10 minutes at 95°C, followed by 40 cycles of 15 seconds at 95°C, 30 seconds at 60°C, and a melt curve with a single reaction cycle at 95°C for 15 seconds, 60°C for 1 minute, and dissociation at 95°C for 15 seconds. The resulting Ct values were then adjusted to the housekeeping gene actin. The relative expression of genes was determined using the 2<sup>-ΔΔct</sup> method.

### 2.27. Molecular docking

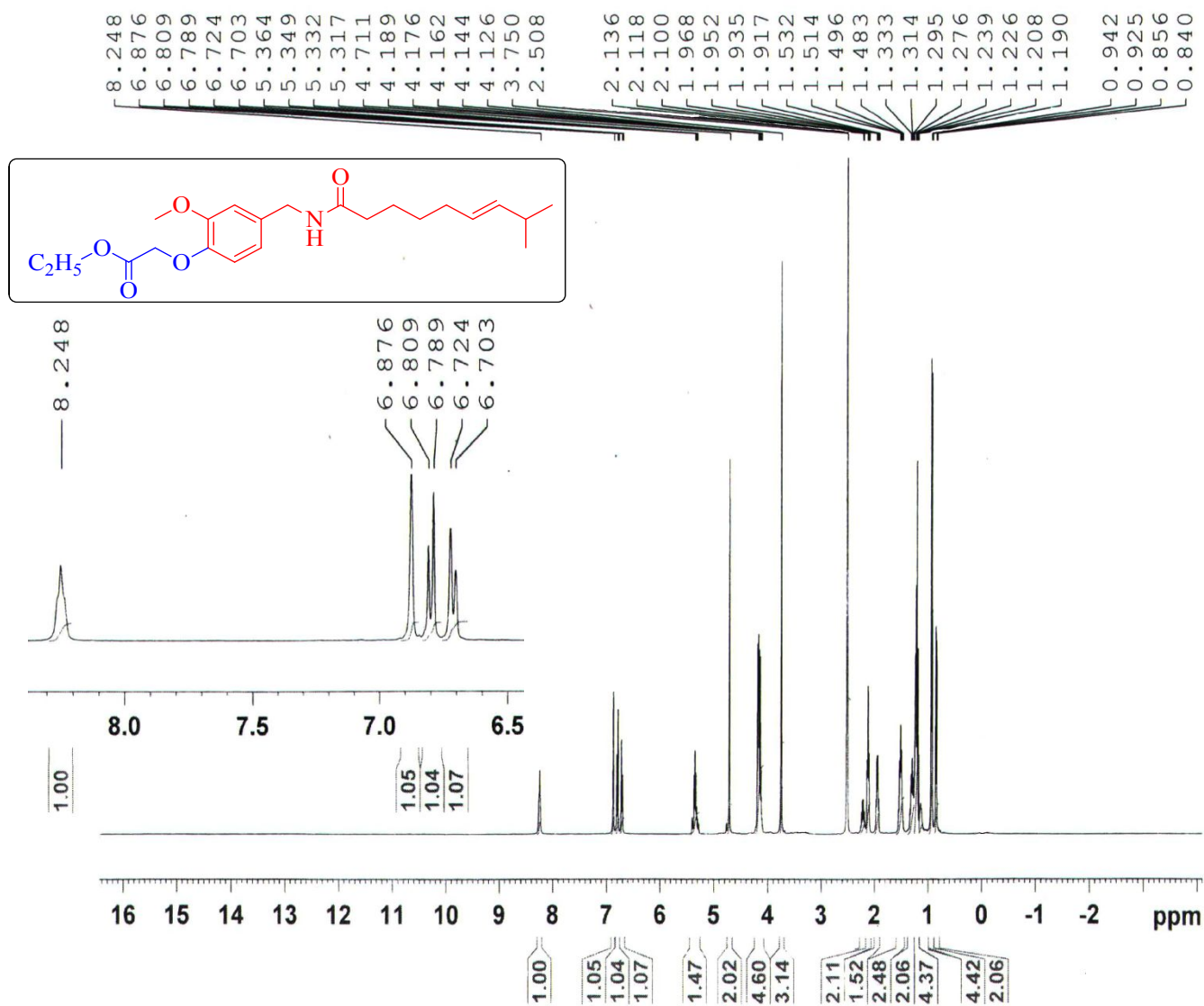
To view binding pattern of all the synthesized compounds for their anti-proliferative activity, molecular docking was carried out at the VEGFR2 kinase receptor catalytic ligand binding site (PDBID: 2QU5). Docking simulations of Compounds 20a-k, 21, 22a-b were performed using Maestro, version 9.6 implemented from Schrodinger software suite. The ligands were sketched in 3D format using build panel and were prepared for docking using ligprep application. The protein for docking study was obtained from Protein data bank (PDB ID: 2QU5) and prepared by removing solvent, adding hydrogen and further minimization in the presence of bound ligand (276) using protein preparation wizard. Grids for molecular docking were generated with bound co-crystallized ligand. For the validation of docking parameters, the co-crystal ligand (276) was re-docked at the catalytic site of protein and the RMSD between co-crystal and re-docked pose was found to be 0.405 Å. All the synthesized compounds were docked using Glide extra-precision (XP) mode, with up to three poses saved per molecule <sup>9</sup>.

### 3. References

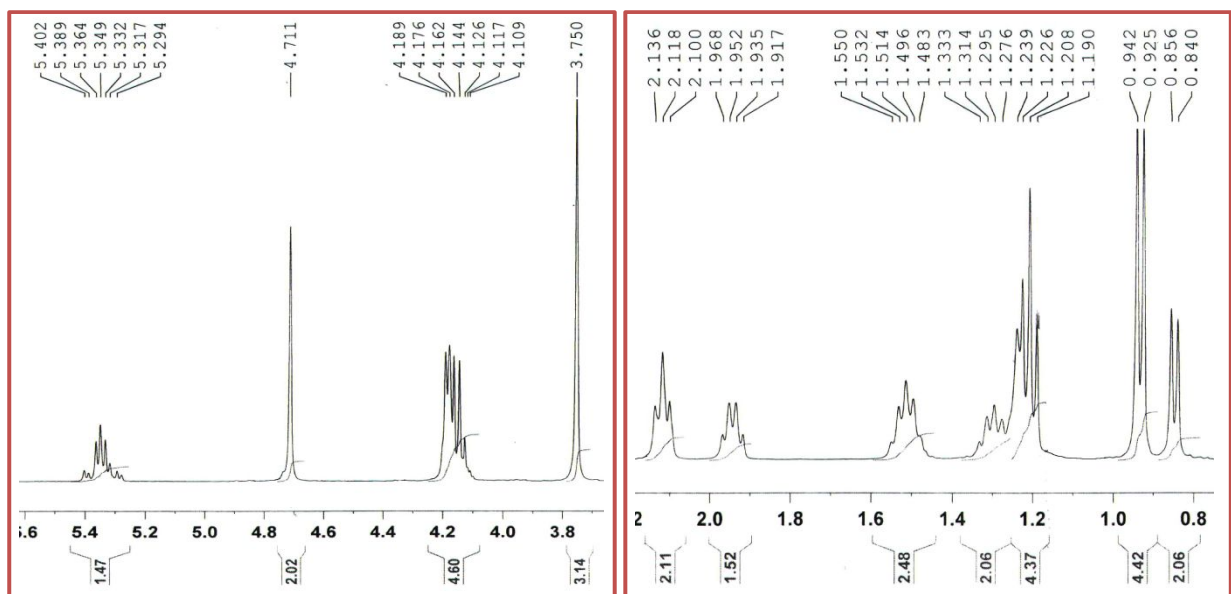
- (1) Janganati, V.; Ponder, J.; Thakkar, S.; Jordan, C. T.; Crooks, P. A. Succinamide derivatives of melampomagnolide B and their anti-cancer activities. *Bioorg Med Chem* **2017**, *25*, 2017.
- (2) Feoktistova, M.; Geserick, P.; Leverkus, M. Crystal Violet Assay for Determining Viability of Cultured Cells. *Cold Spring Harbor protocols* **2016**, *2016*, 2016.
- (3) Franken, N. A. P.; Rodermond, H. M.; Stap, J.; Haveman, J.; van Bree, C. Clonogenic assay of cells in vitro. *Nature Protocols* **2006**, *1*, 2006.
- (4) Wu, D.; Yotnda, P. Production and detection of reactive oxygen species (ROS) in cancers. *J Vis Exp* **2011**, 2011.
- (5) Mustafa, M.; Abdelhamid, D.; Abdelhafez, E. M. N.; Ibrahim, M. A. A.; Gamal-Eldeen, A. M.; Aly, O. M. Synthesis, antiproliferative, anti-tubulin activity, and docking study of new 1,2,4-triazoles as potential combretastatin analogues. *European journal of medicinal chemistry* **2017**, *141*, 2017.
- (6) Ma, L.; Peng, H.; Li, K.; Zhao, R.; Li, L.; Yu, Y.; Wang, X.; Han, Z. Luteolin exerts an anticancer effect on NCI-H460 human non-small cell lung cancer cells through the induction of Sirt1-mediated apoptosis. *Molecular medicine reports* **2015**, *12*, 2015.
- (7) Li, W.; Yin, Y.; Shuai, W.; Xu, F.; Yao, H.; Liu, J.; Cheng, K.; Xu, J.; Zhu, Z.; Xu, S. Discovery of novel quinazolines as potential anti-tubulin agents occupying three zones of colchicine domain. *Bioorganic chemistry* **2018**, *83*, 2018.
- (8) Gour, R.; Ahmad, F.; Prajapati, S. K.; Giri, S. K.; Lal Karna, S. K.; Kartha, K. P. R.; Pokharel, Y. R. Synthesis of novel S-linked dihydroartemisinin derivatives and evaluation of their anticancer activity. *European journal of medicinal chemistry* **2019**, *178*, 2019.
- (9) Potashman, M. H.; Bready, J.; Coxon, A.; DeMelfi, T. M., Jr.; DiPietro, L.; Doerr, N.; Elbaum, D.; Estrada, J.; Gallant, P.; Germain, J.; Gu, Y.; Harmange, J. C.; Kaufman, S. A.; Kendall, R.; Kim, J. L.; Kumar, G. N.; Long, A. M.; Neervannan, S.; Patel, V. F.; Polverino, A.; Rose, P.; Plas, S.; Whittington, D.; Zanon, R.; Zhao, H. Design, synthesis, and evaluation of orally active benzimidazoles and benzoxazoles as vascular endothelial growth factor-2 receptor tyrosine kinase inhibitors. *J Med Chem* **2007**, *50*, 2007.



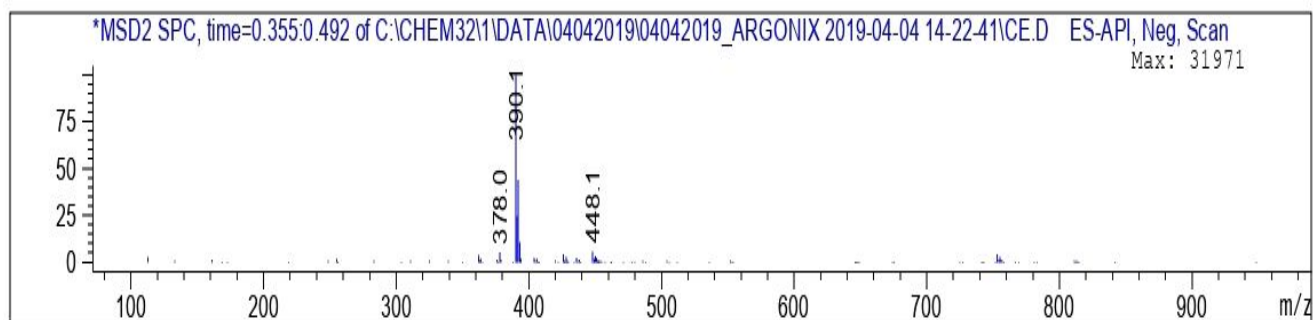
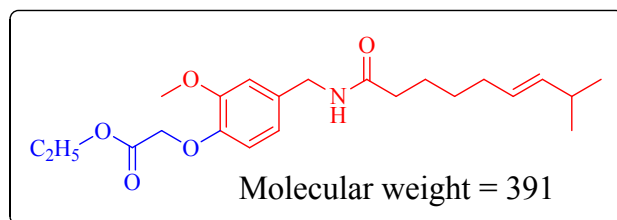
4. Scanned copies of  $^1\text{H}$ ,  $^{13}\text{C}$  NMR and Mass spectra's of the representative compounds:



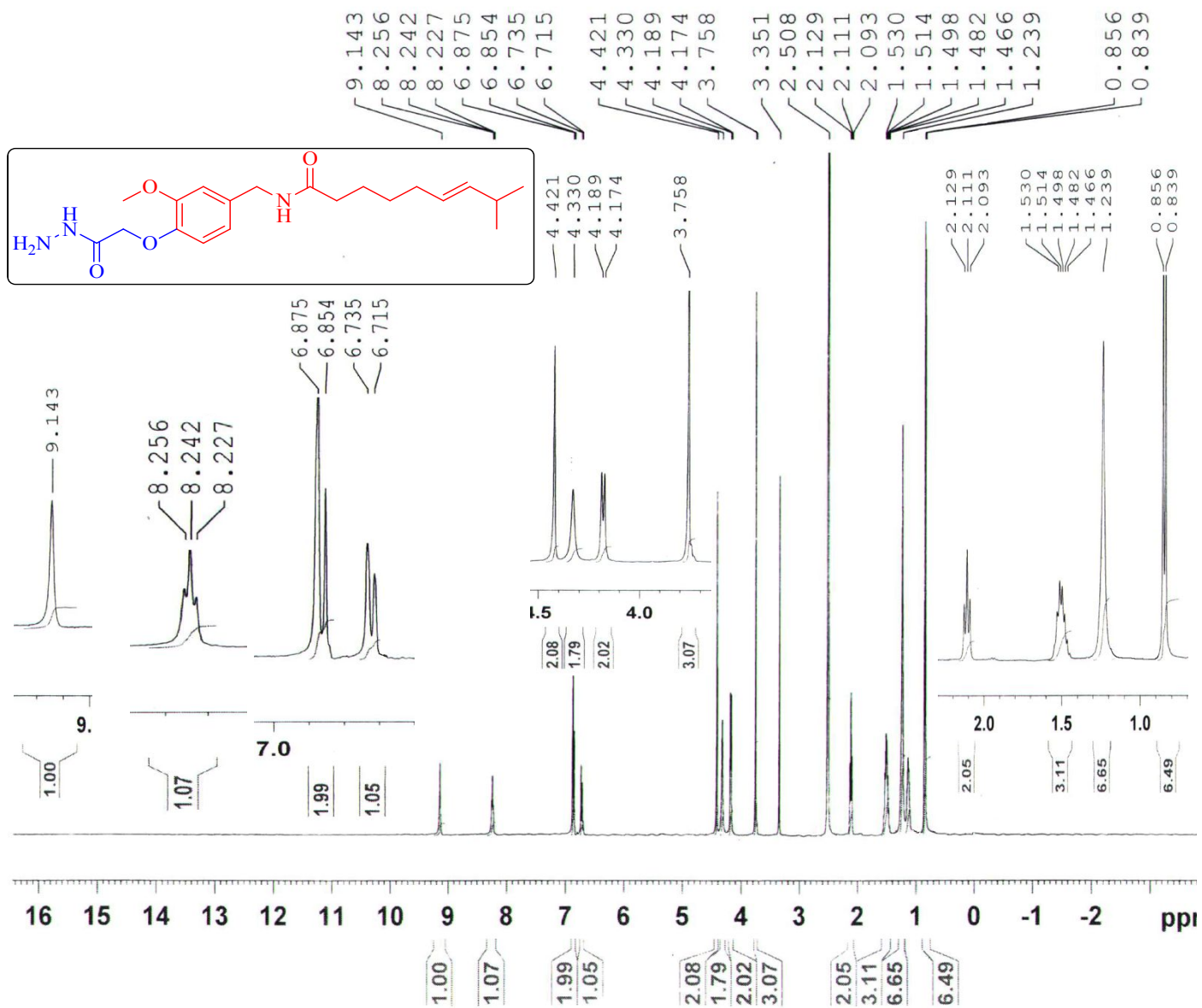
$^1\text{H}$  NMR of Compound 16



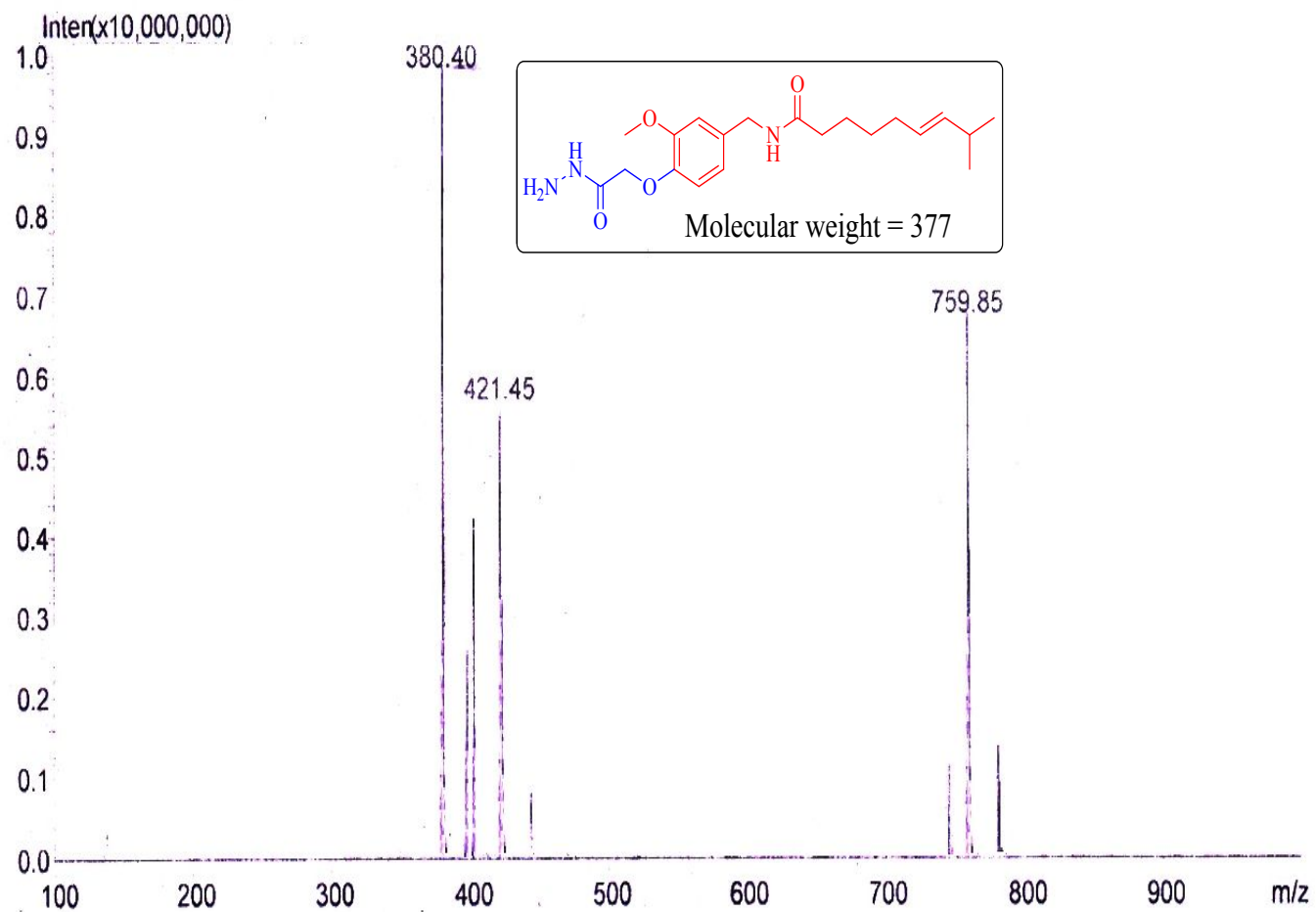
**<sup>1</sup>H NMR of Compound 16**



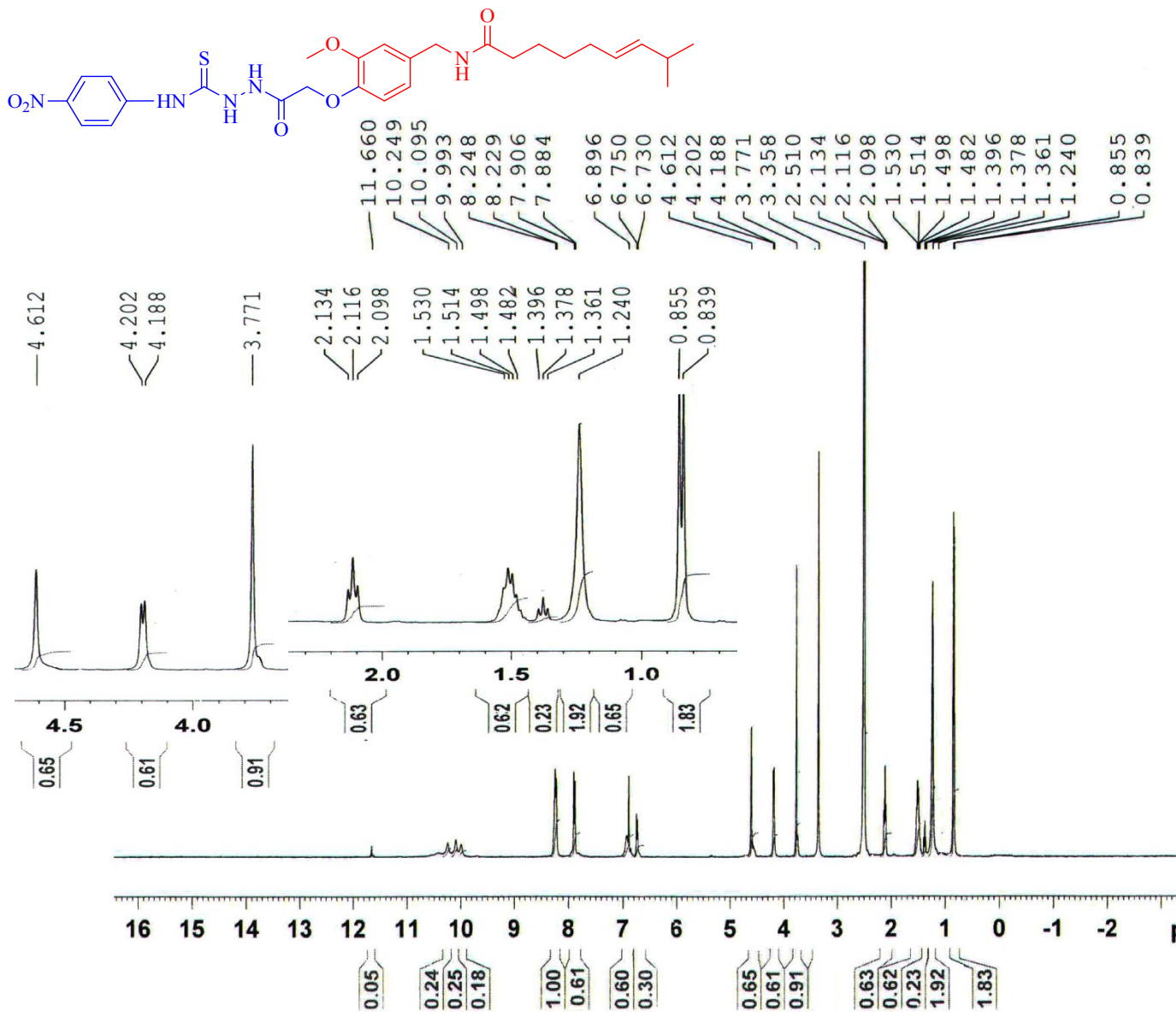
**ESI-MS of compound 16**



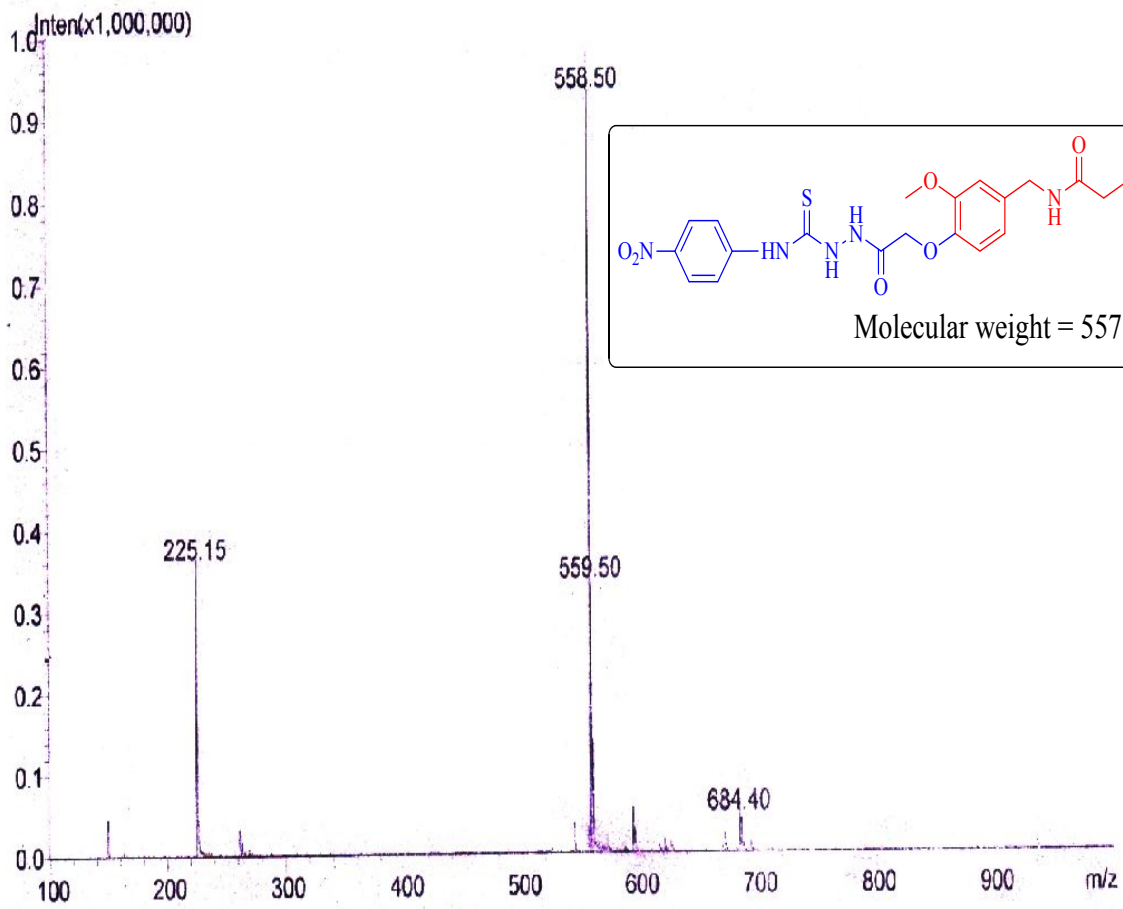
**<sup>1</sup>H NMR of Compound 17**



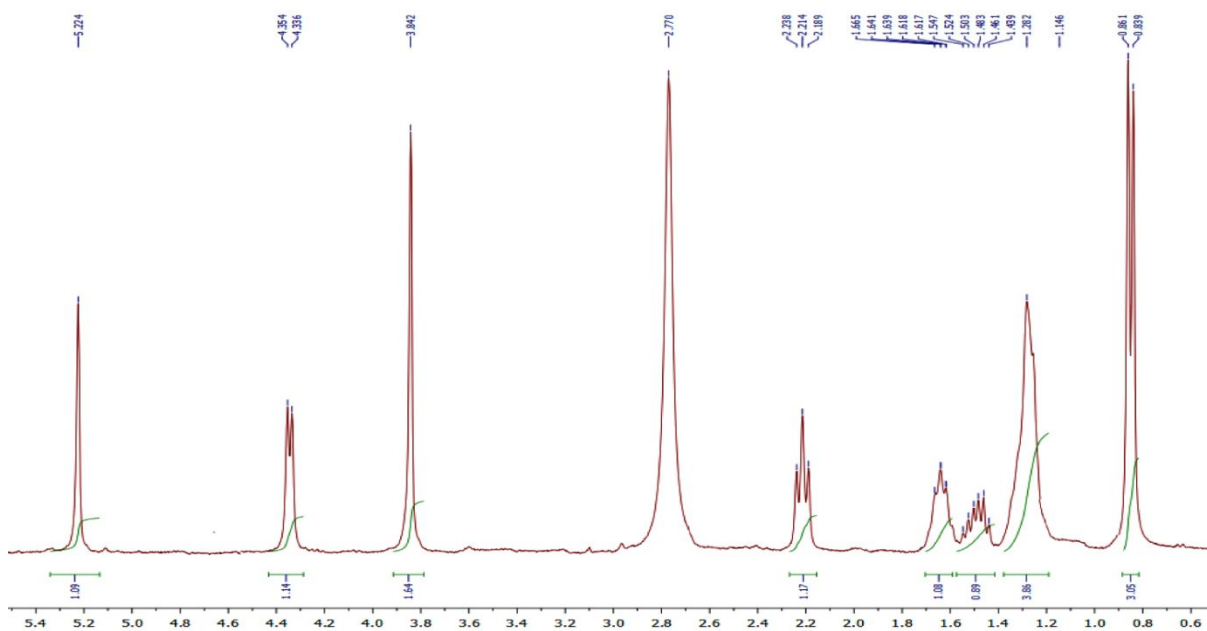
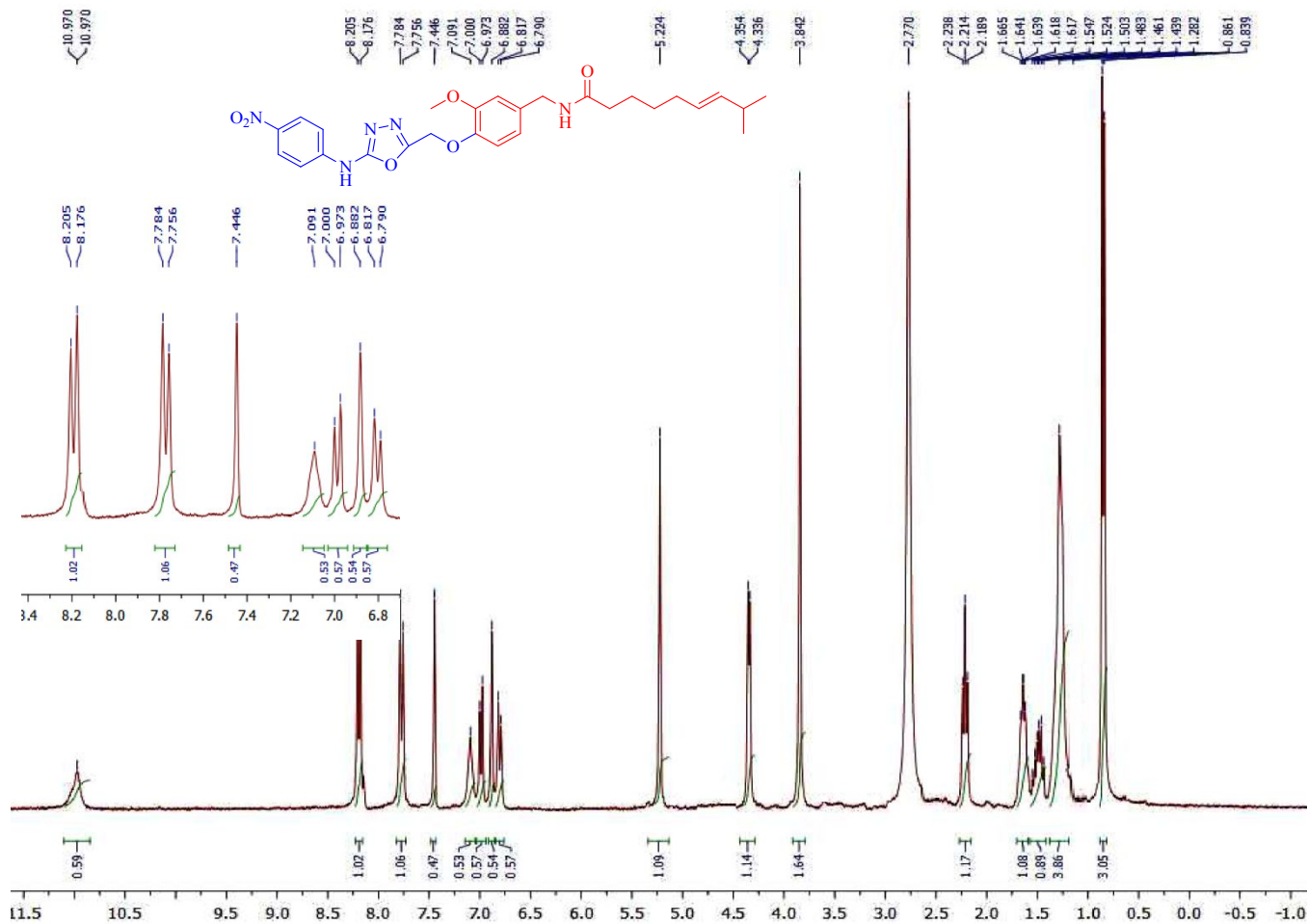
ESI-MS of compound 17



**<sup>1</sup>H NMR of Compound 19a**

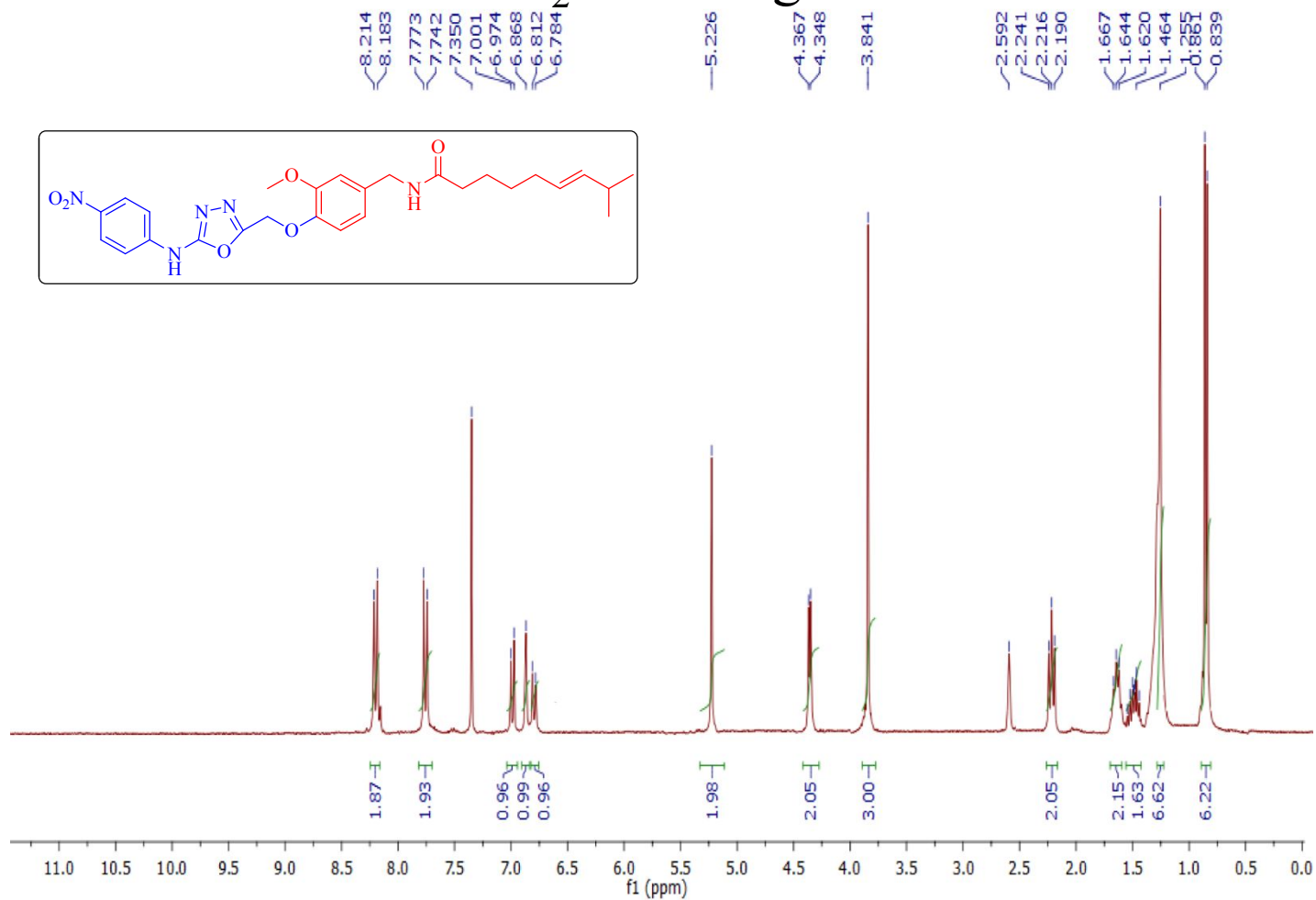


ESI-MS of compound 19a



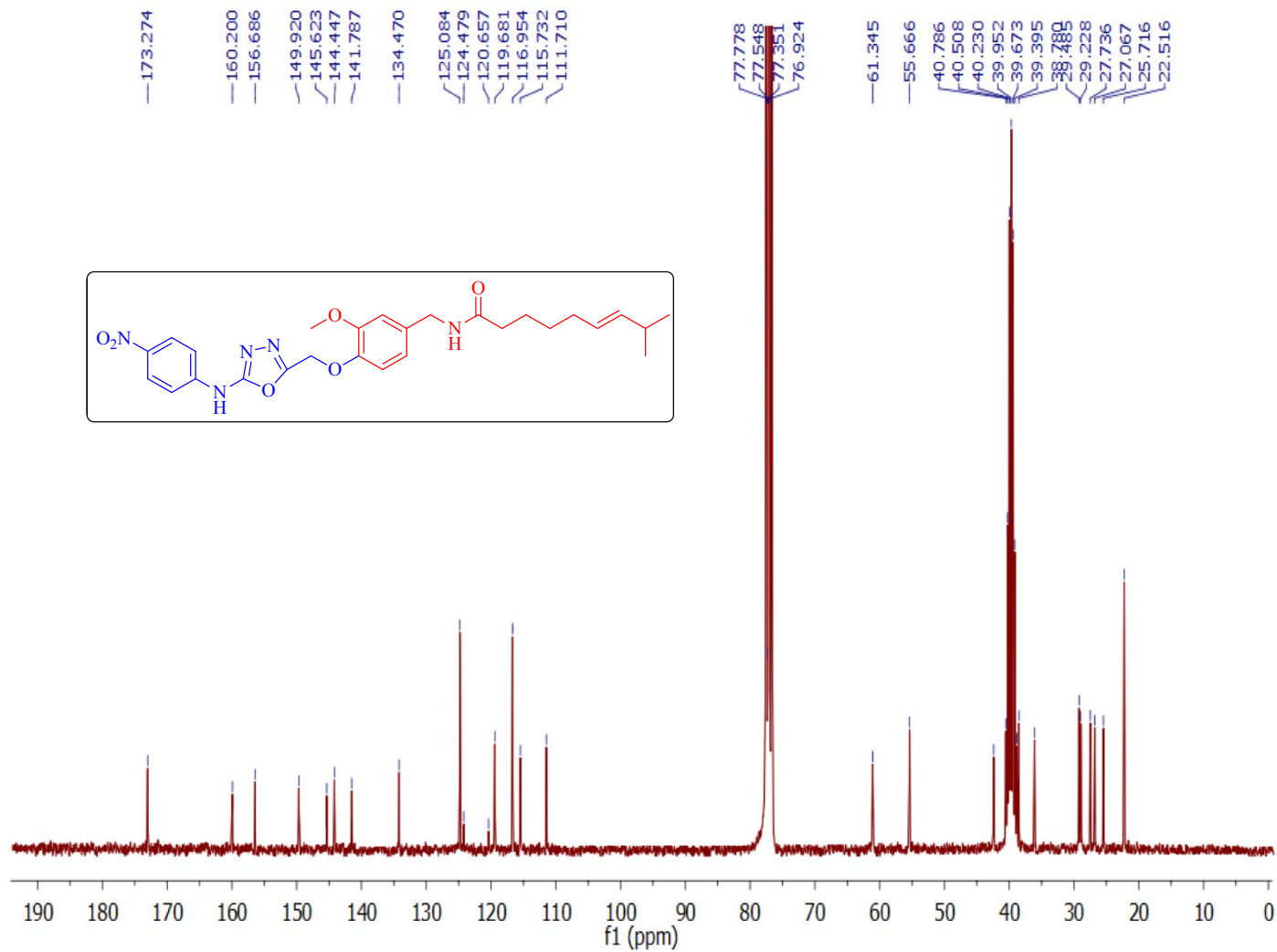
**<sup>1</sup>H NMR of Compound 20a**

# D<sub>2</sub>O exchange

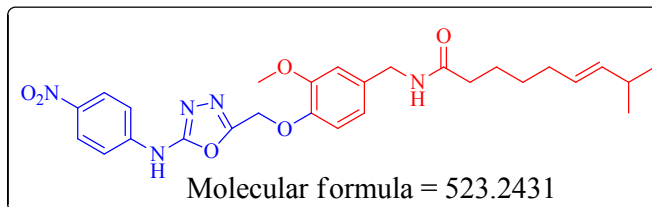


<sup>1</sup>H NMR of Compound 20a





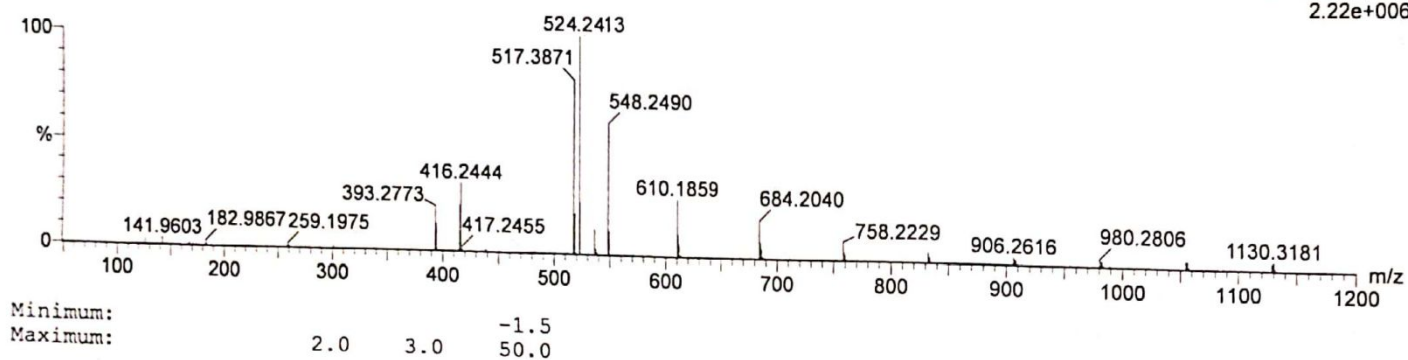
<sup>13</sup>C NMR of Compound 20a



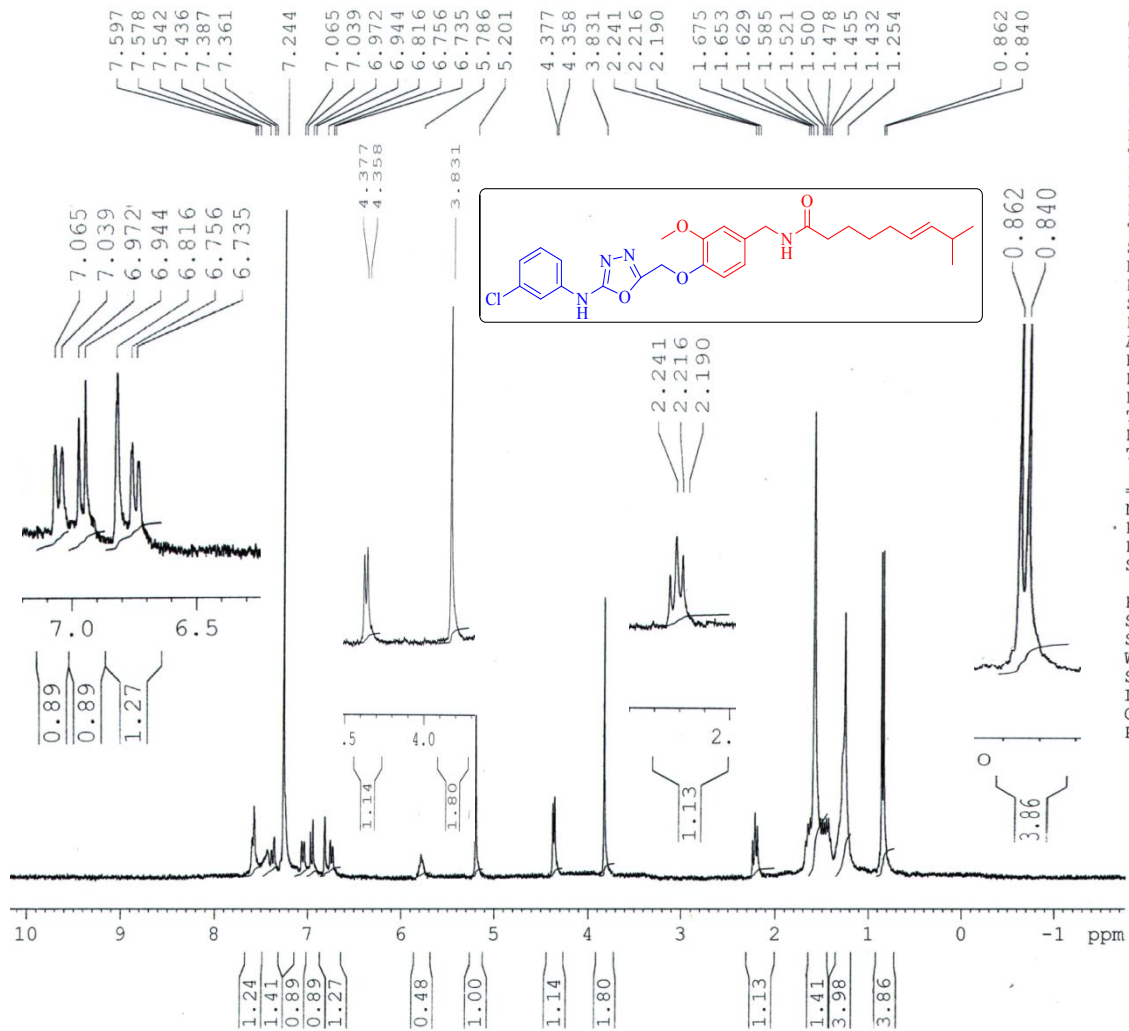
240821\_05 12 (0.259) Cm (12:13)

Xevo G2-XS QTOF YFC2015

24-Aug-2021  
13:01:33  
1: TOF MS ES+  
2.22e+006



### HRMS of compound 20a



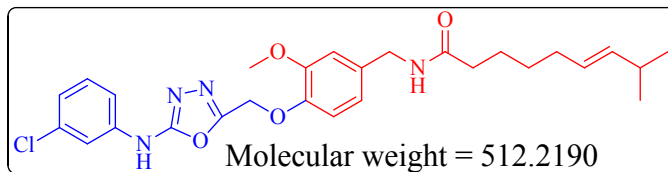
Current Data Parameters  
 NAME 27March2018  
 EXPNO 45  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20190409  
 Time\_ 10.22  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 16  
 DS 2  
 SWH 6172.839 Hz  
 FIDRES 0.094190 Hz  
 AQ 5.3084660 sec  
 RG 362  
 DW 81.000 usec  
 DE 6.50 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 10.70 usec  
 PL1 -2.00 dB  
 SFO1 300.1318534 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1300055 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

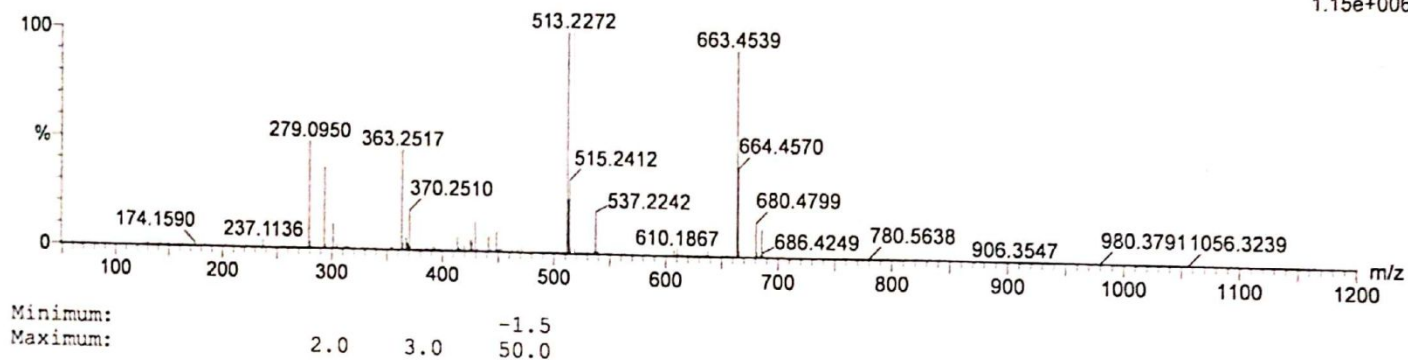
**<sup>1</sup>H NMR of Compound 20b**



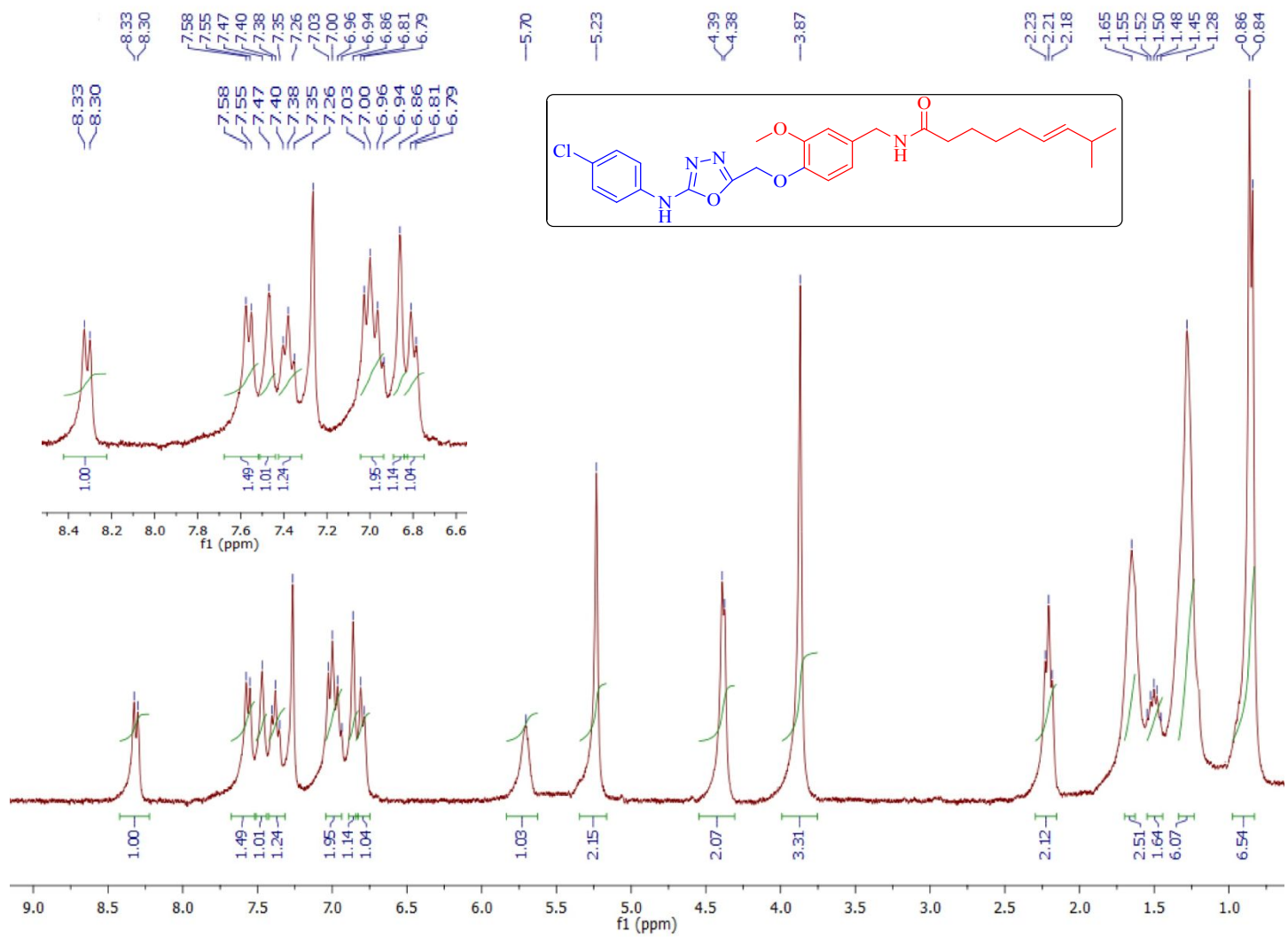
170921\_05 10 (0.225) Cm (10)

Xevo G2-XS QTOF YFC2015

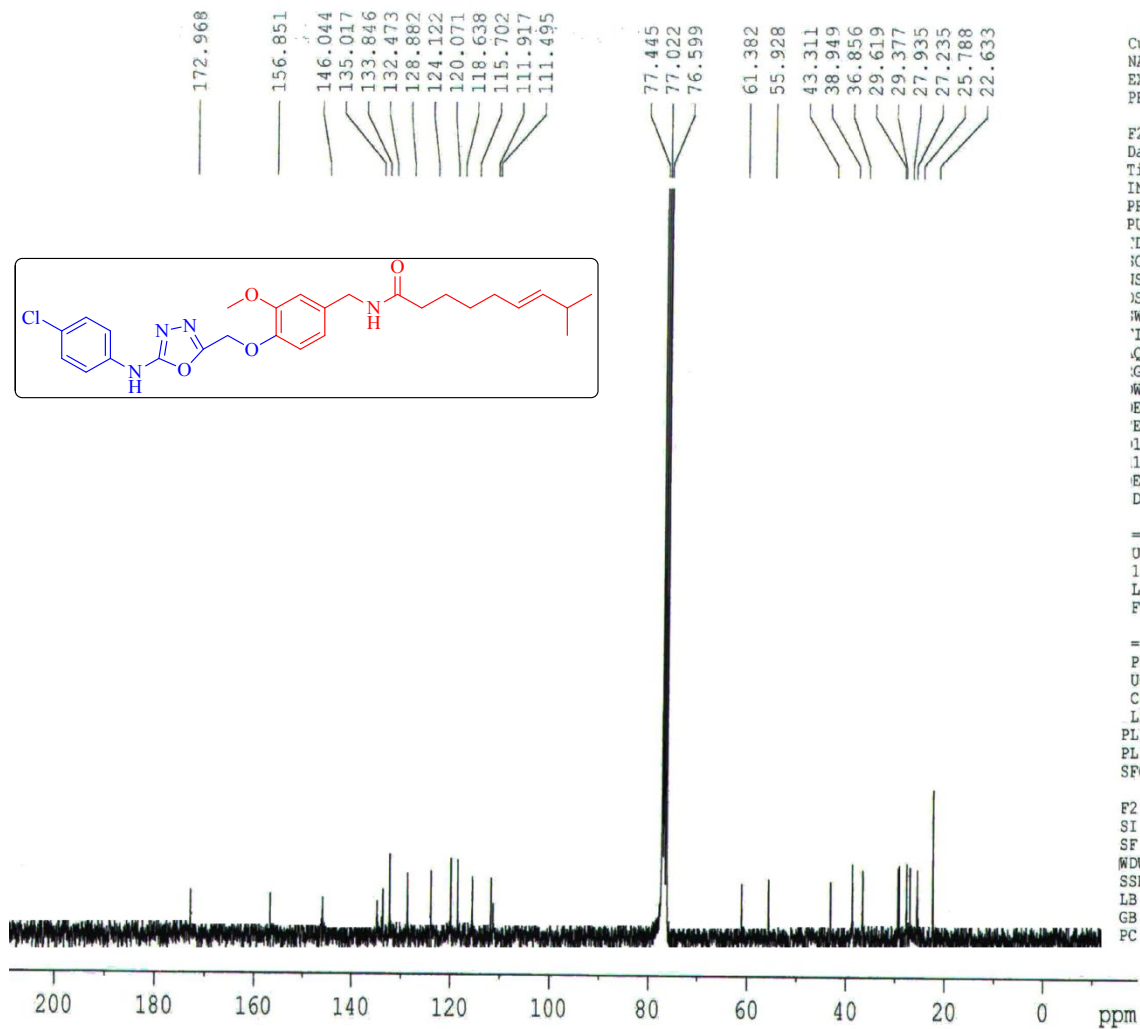
11:51:21  
1: TOF MS ES+  
1.15e+006



### HRMS of compound 20b



**<sup>1</sup>H NMR of Compound 20c**



Current Data Parameters  
 NAME 27March2018  
 EXPNO 38  
 PROCNO 1

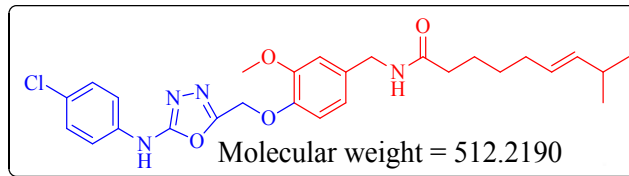
F2 - Acquisition Parameters  
 Date 20190408  
 Time 13.59  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zgpg30  
 ID 65536  
 SOLVENT CDCL3  
 IS 900  
 IS 4  
 WH 17985.611 Hz  
 IDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 SG 32768  
 SW 27.800 usec  
 SE 6.50 usec  
 TE 300.0 K  
 d1 2.00000000 sec  
 d11 0.03000000 sec  
 ELTA 1.89999998 sec  
 D0 1

===== CHANNEL f1 =====  
 UC1 13C  
 L1 10.10 usec  
 L1 -2.00 dB  
 FO1 75.4752953 MHz

===== CHANNEL f2 =====  
 PDPRG2 waltz16  
 UC2 1H  
 CPD2+ , 115.00 usec  
 L2 -2.00 dB  
 PL12 18.50 dB  
 PL13 22.50 dB  
 SFO2 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677490 MHz  
 WDW EM  
 SSB 0  
 LB 2.00 Hz  
 GB 0  
 PC 1.40

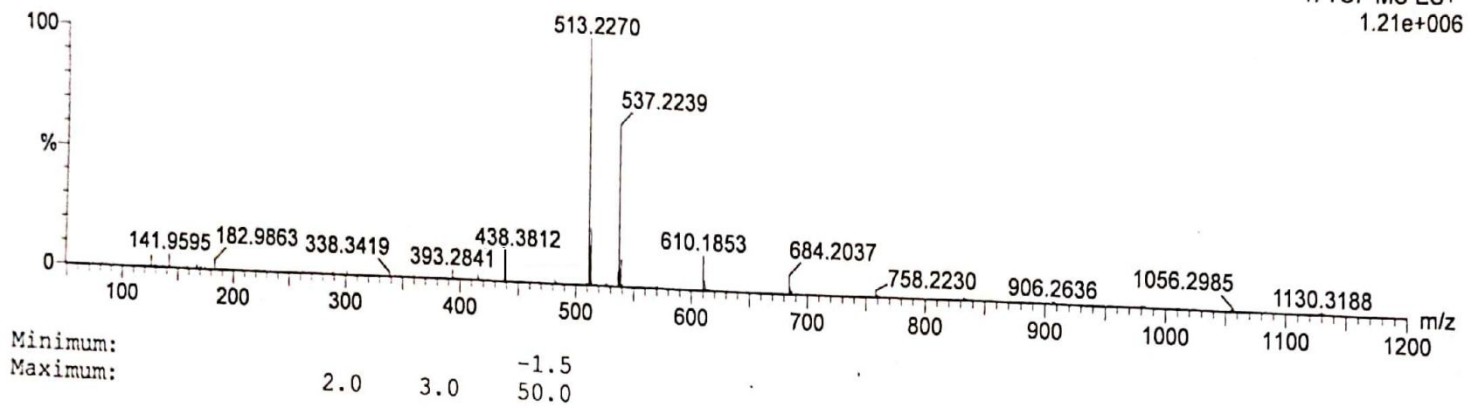
<sup>13</sup>C NMR of Compound 20c



240821\_08 14 (0.293) Cm (14:15)

Xevo G2-XS QTOF YFC2015

24-Aug-2021  
13:09:15  
1: TOF MS ES+  
1.21e+006



**HRMS of Compound 20c**

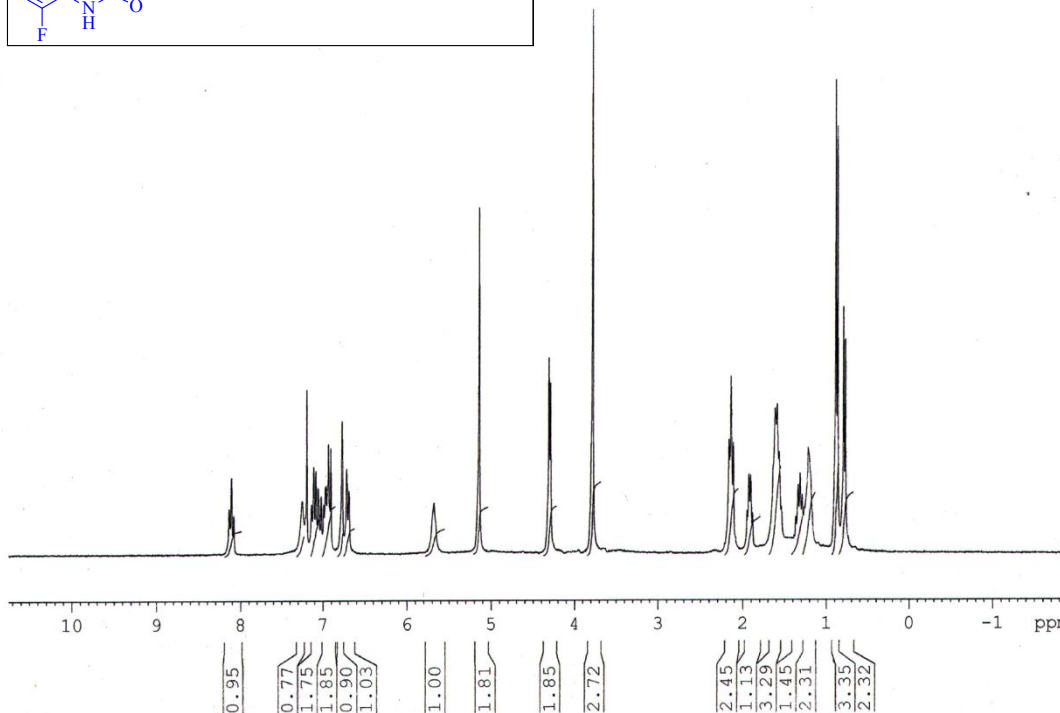
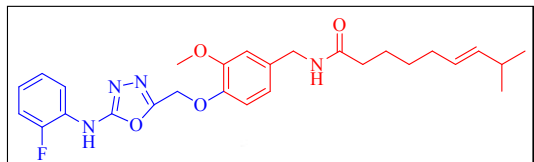
8.136  
8.109  
8.082  
7.250  
7.193  
7.136  
7.111  
7.084  
7.057  
7.020  
6.989  
6.968  
6.933  
6.906  
6.770  
6.715  
6.689

5.678

5.139  
4.307  
4.288  
3.775  
2.160  
2.136  
2.111  
1.946  
1.924  
1.903  
1.880  
1.610  
1.585  
1.560

1.408  
1.363  
1.337  
1.312  
1.287  
1.213

0.885  
0.863  
0.791  
0.769  
0.000

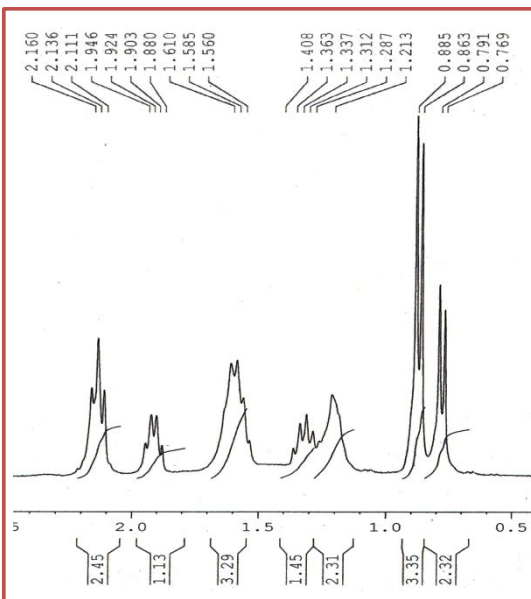
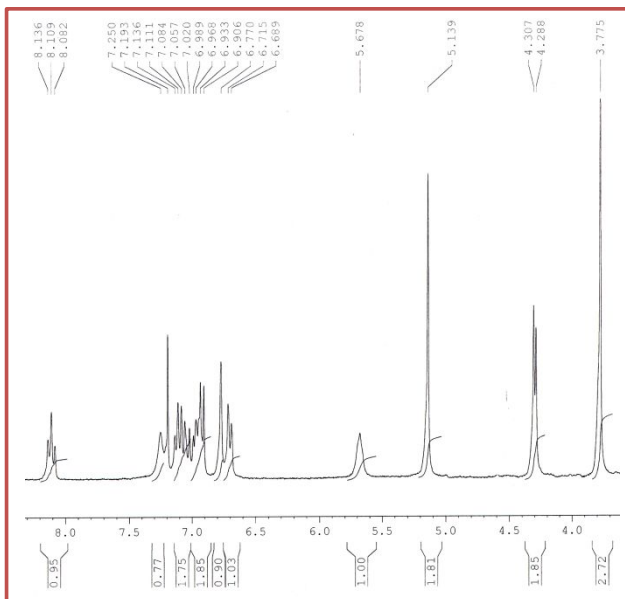


Current Data Parameters  
NAME 28 JUI12019  
EXPNO 26  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190729  
Time 15.07  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 12  
DS 2  
SWH 6172.839 Hz  
FIDRES 0.094190 Hz  
AQ 5.3084660 sec  
RG 228.1  
DW 81.000 usec  
DE 6.50 usec  
TE 300.0 K  
D1 1.00000000 sec  
TDO 1

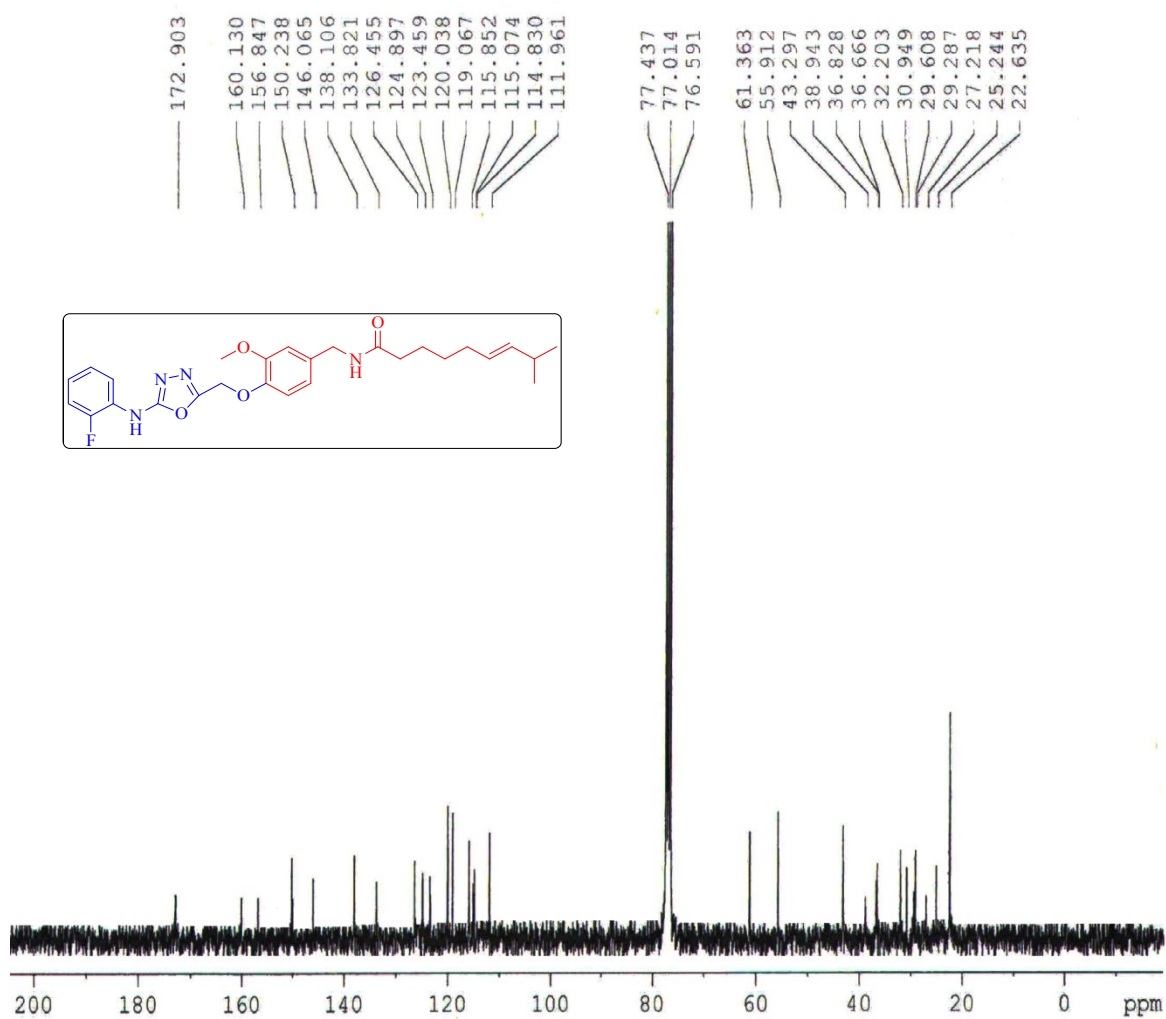
===== CHANNEL f1 =====  
NUC1 1H  
P1 10.70 usec  
PL1 -2.00 dB  
SFO1 300.1318534 MHz

F2 - Processing parameters  
SI 32768  
SF 300.1300257 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



**<sup>1</sup>H NMR of Compound 20d**





Current Data Parameters  
 NAME 28 JULY2019  
 EXPNO 27  
 PROCNO 1

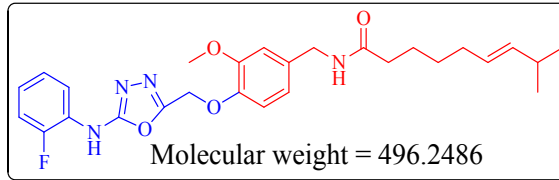
F2 - Acquisition Parameters  
 Date 20190729  
 Time 16.32  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1123  
 DS 4  
 SWH 17985.611 Hz  
 FIDRES 0.274439 Hz  
 AQ 1.8219508 sec  
 RG 32768  
 DW 27.800 usec  
 DE 6.50 usec  
 TE 300.0 K  
 D1 2.0000000 sec  
 d11 0.0300000 sec  
 DELTA 1.89999998 sec  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 10.10 usec  
 PL1 -2.00 dB  
 SFO1 75.4752953 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 115.00 usec  
 PL2 -2.00 dB  
 PL12 18.50 dB  
 PL13 22.50 dB  
 SFO2 300.1312005 MHz

F2 - Processing parameters  
 SI 32768  
 SF 75.4677490 MHz  
 WDW EM  
 SSB 0  
 LB 2.00 Hz  
 GB 0  
 PC 1

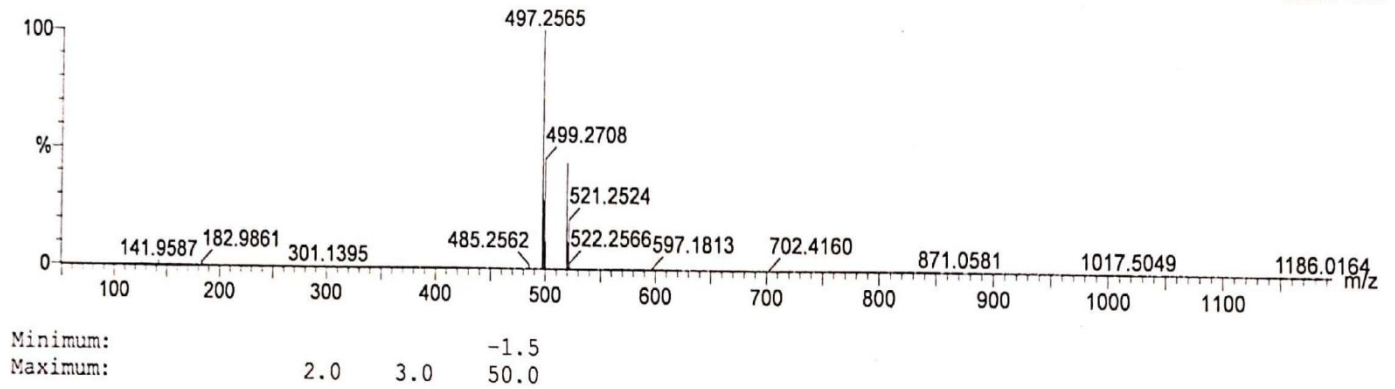
<sup>13</sup>C NMR of Compound 20d



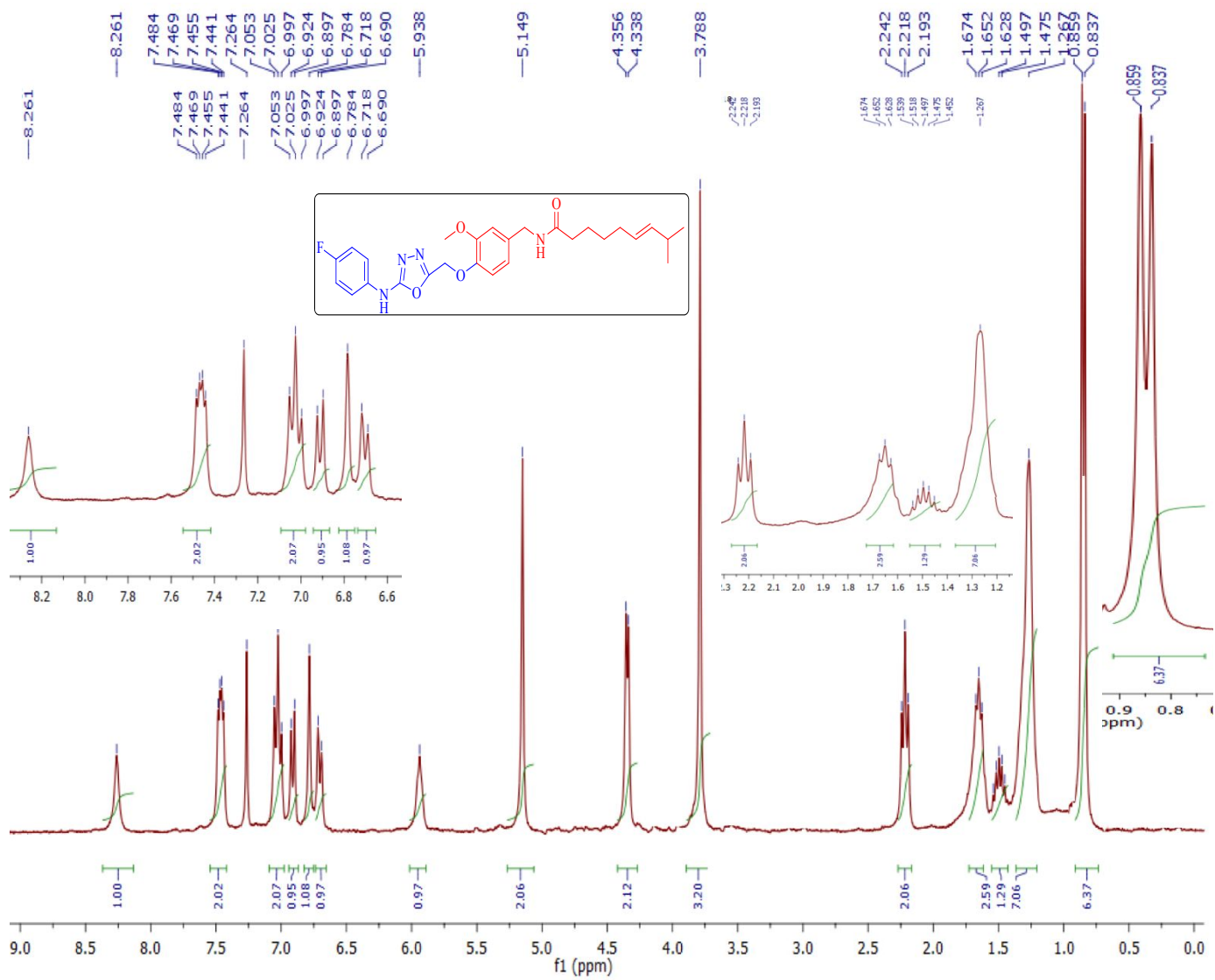
Xevo G2-XS QTOF YFC2015

11:48:46  
1: TOF MS ES+  
2.84e+006

170921\_04 18 (0.380) Cm (18)

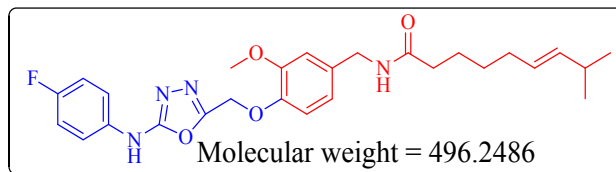


### HRMS of Compound 20d



**<sup>1</sup>H NMR of Compound 20e**

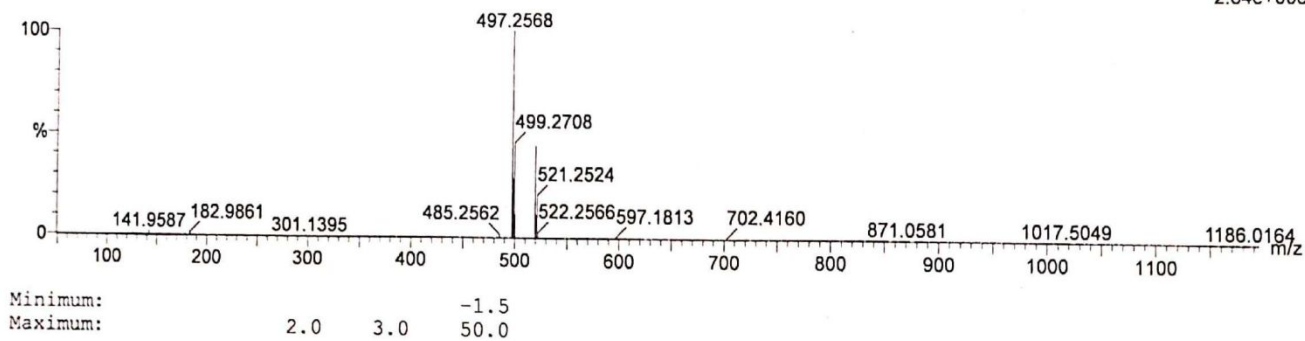




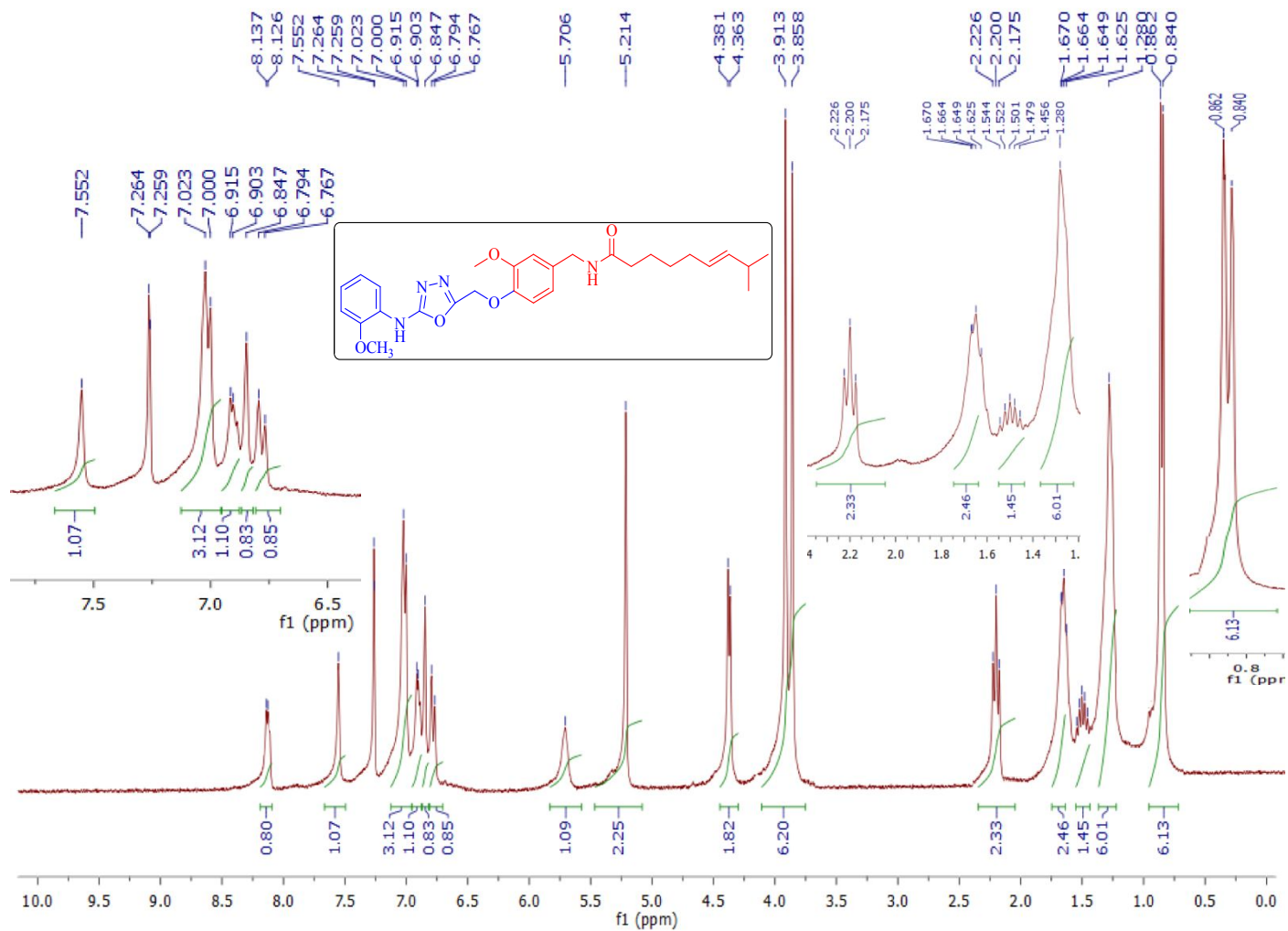
Xevo G2-XS QTOF YFC2015

170921\_04 18 (0.380) Cm (18)

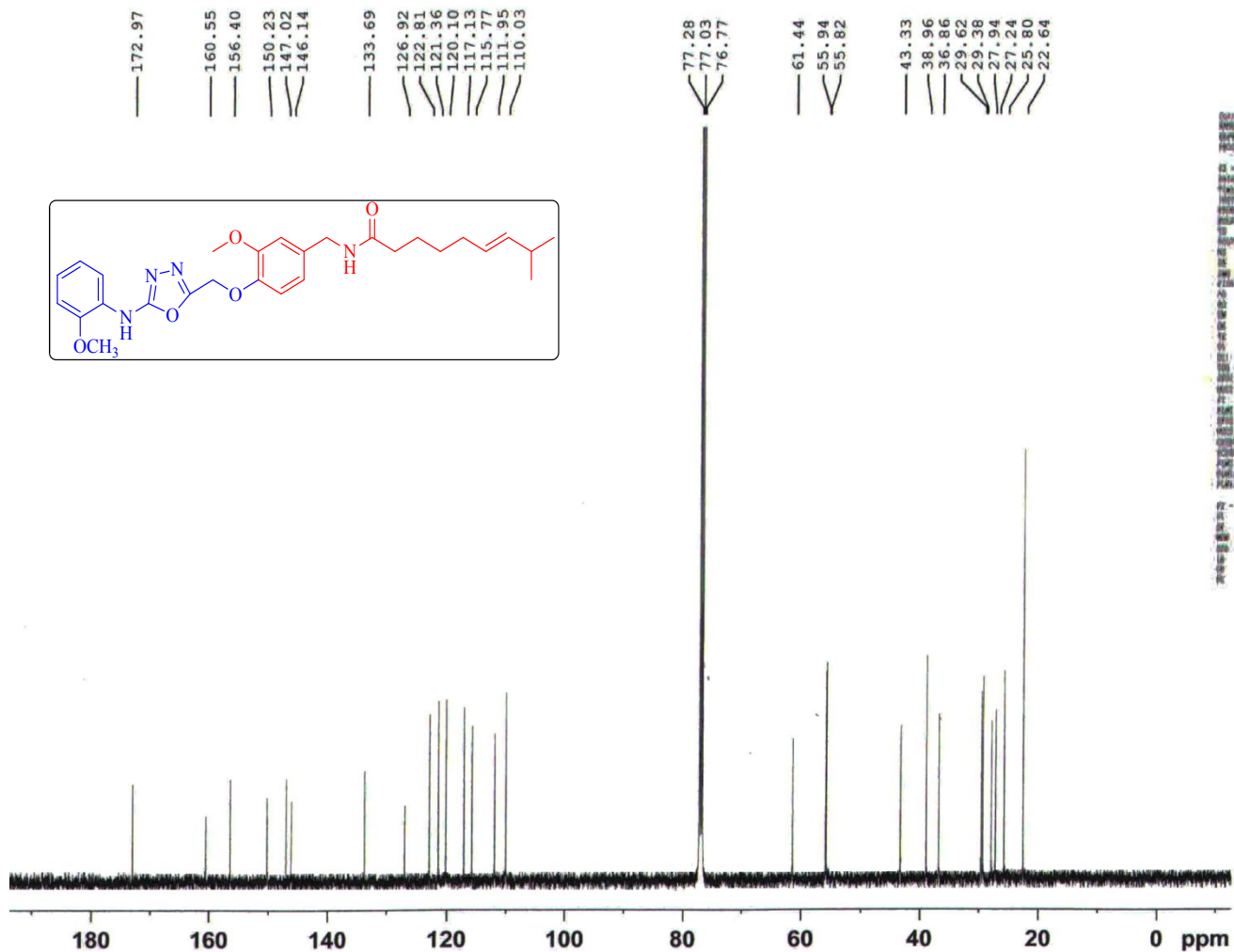
11:48:46  
1: TOF MS ES+  
2.84e+006



**HRMS of Cmpound 20e**

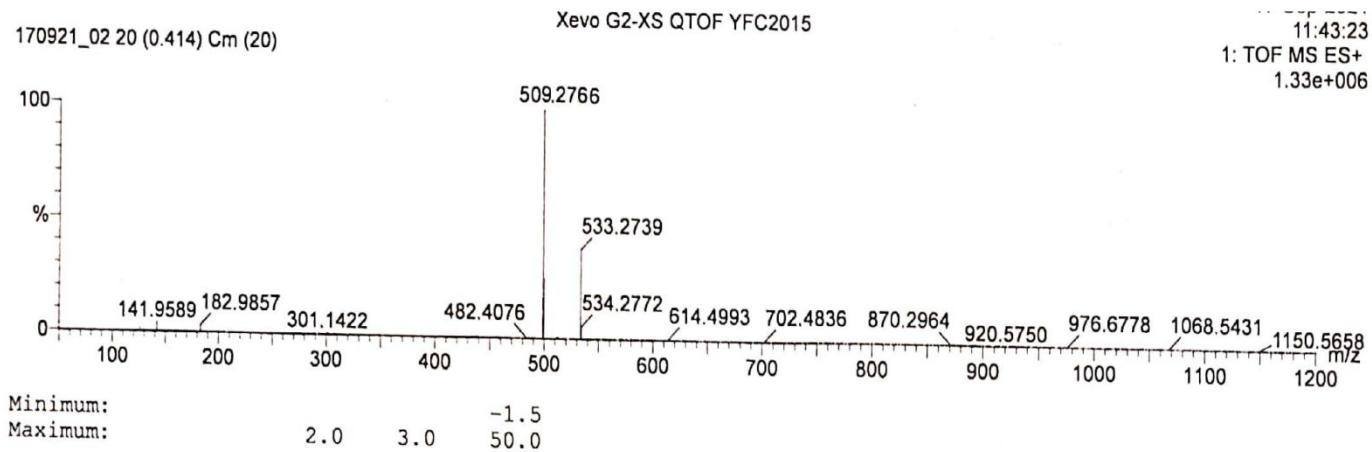
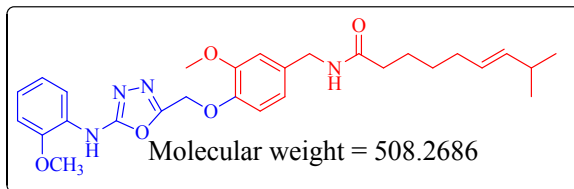


**<sup>1</sup>H NMR of Compound 20f**



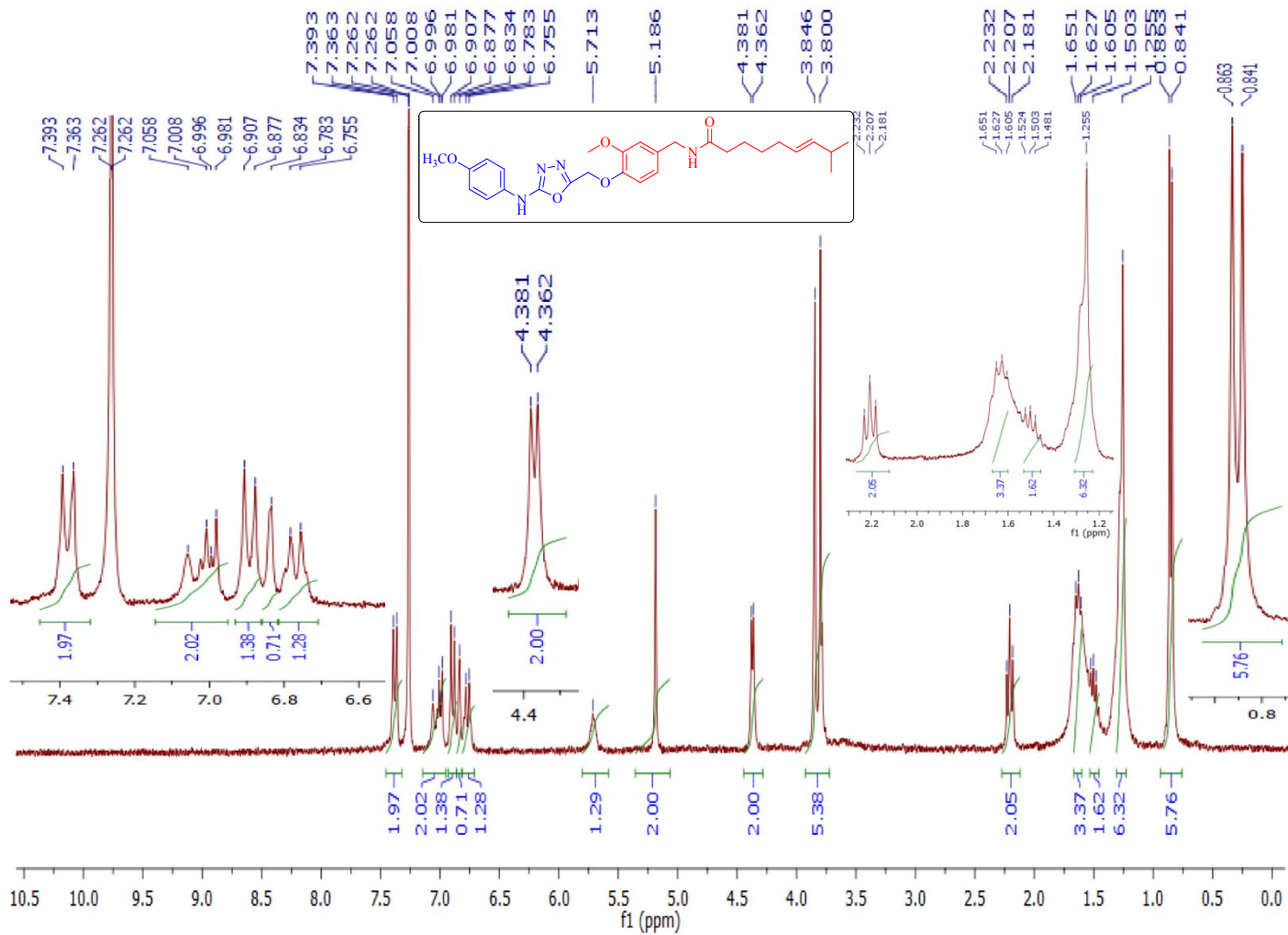
Name: 20f  
 Date: 11/11/2019  
 Time: 14:14:14  
 Sample: 20f  
 Solvent: CDCl3  
 Concentration: 100 mg/ml  
 Acquisition: 129.62 MHz  
 F2 - Acquisition Parameters  
 Date\_ 11/11/2019  
 Time\_ 14:14:14  
 Name\_ 20f  
 Solvent\_ CDCl3  
 Concentration\_ 100 mg/ml  
 Acquisition\_ 129.62 MHz  
 F2 - Processing Parameters  
 Date\_ 11/11/2019  
 Time\_ 14:14:14  
 Name\_ 20f  
 Solvent\_ CDCl3  
 Concentration\_ 100 mg/ml  
 Acquisition\_ 129.62 MHz

<sup>13</sup>C NMR of Compound 20f

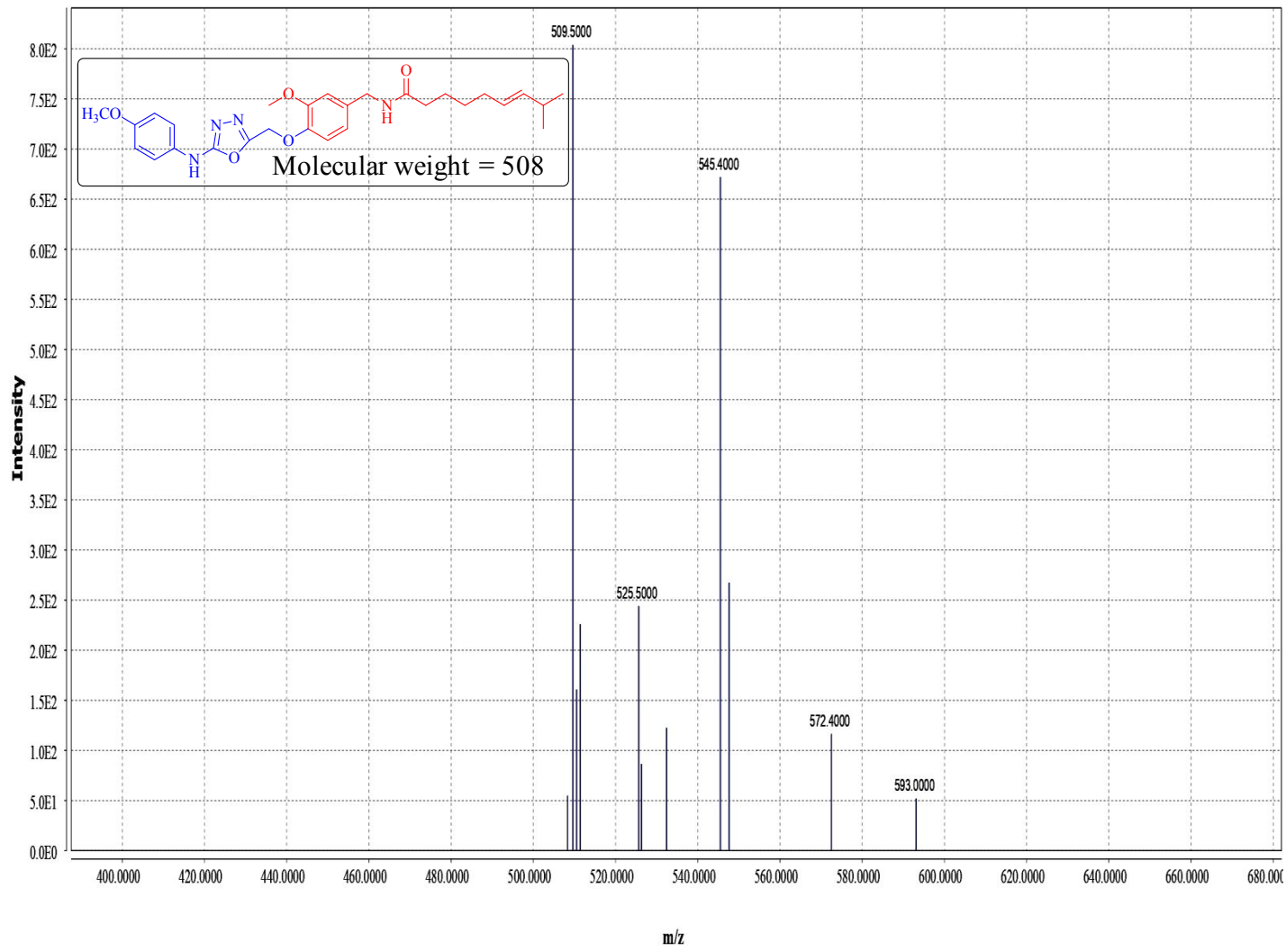


**HRMS of compound 20f**

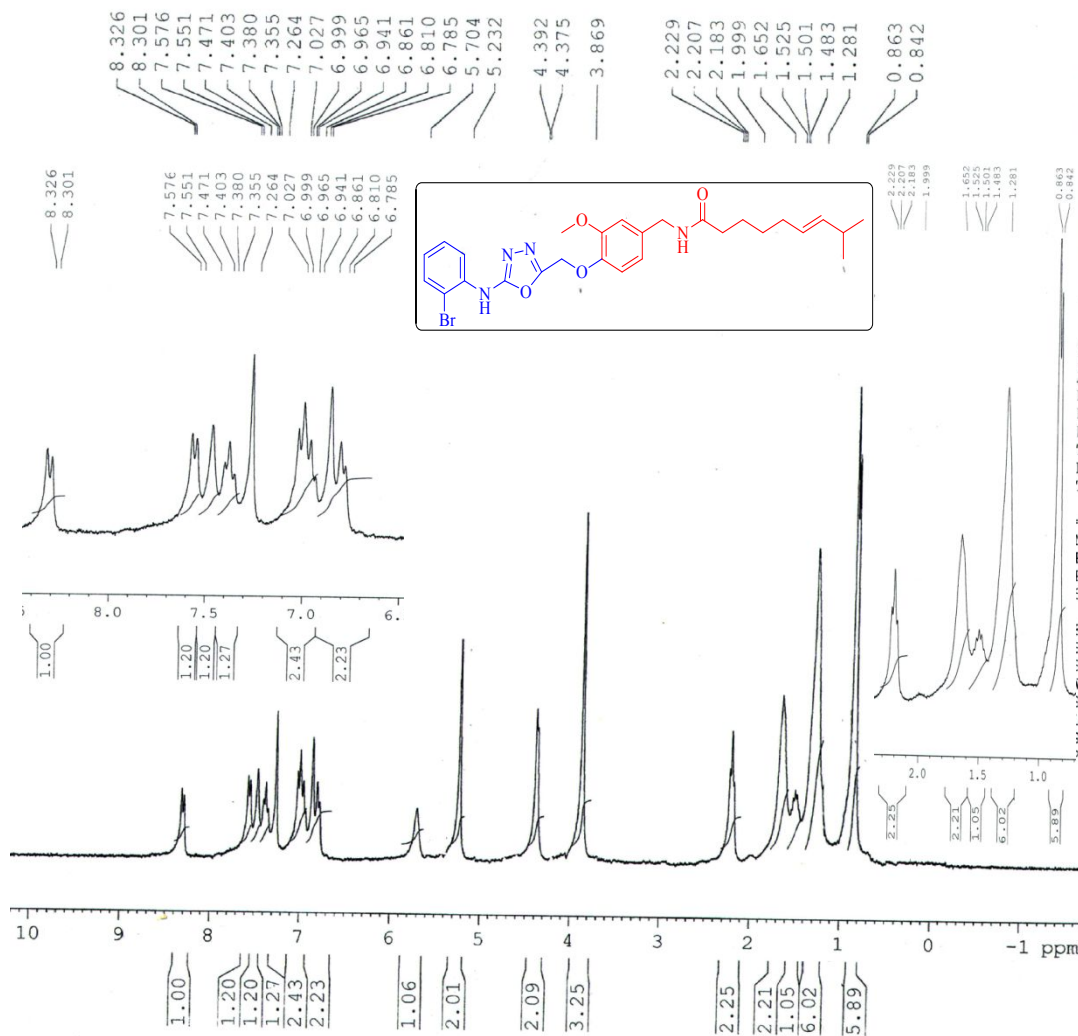




**<sup>1</sup>H NMR of Compound 20g**



ESI-MS of compound 20g



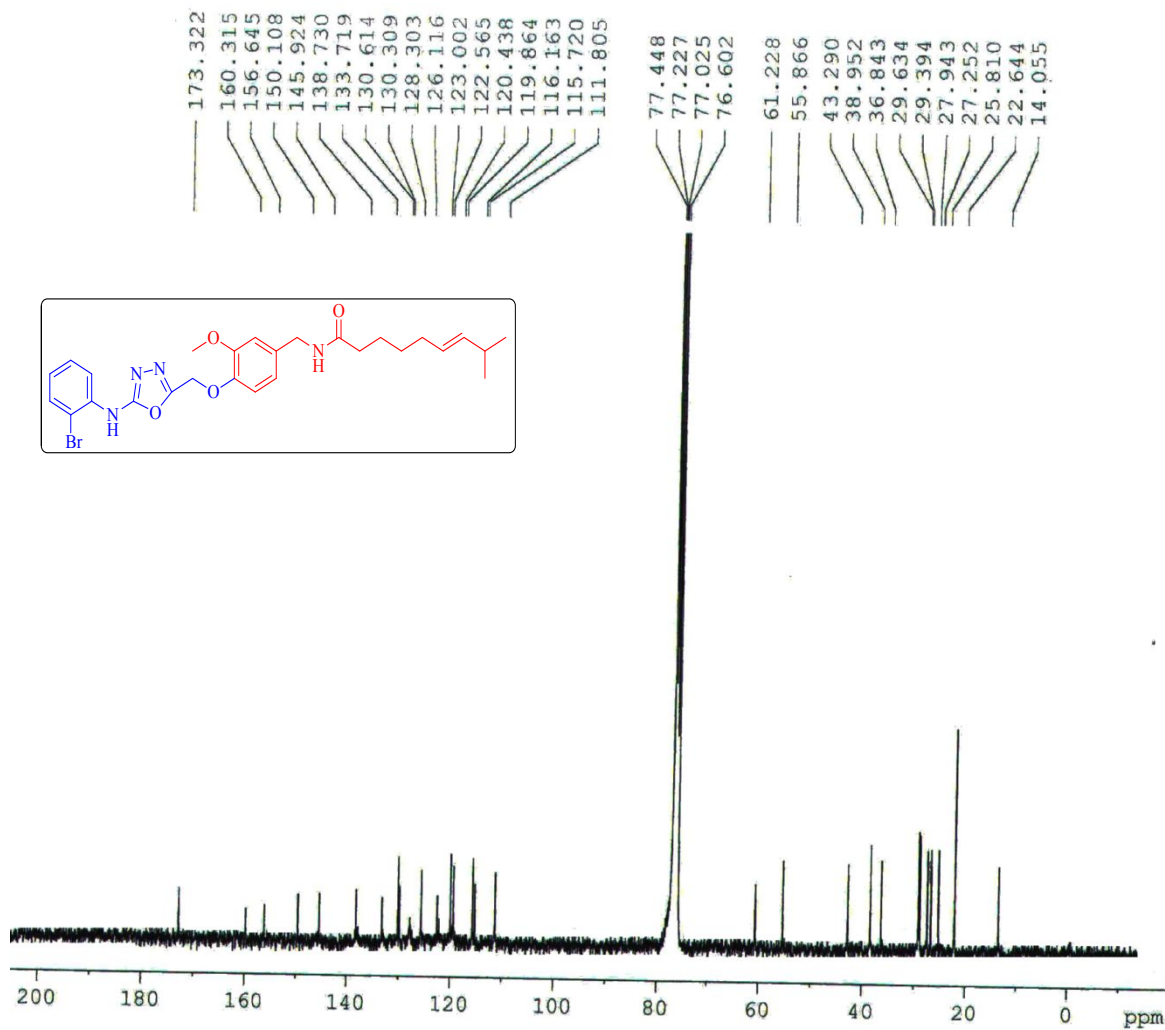
Current Data Parameters  
 NAME 27March2018  
 EXPNO 36  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20190408  
 Time\_ 10.18  
 INSTRUM spect  
 PROBHD 5 mm PABBO BB-  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 16  
 DS 2  
 SWH 6172.839 Hz  
 FIDRES 0.094190 Hz  
 AQ 5.3084660 sec  
 RG 322.5  
 DW 81.000 usec  
 DE 6.50 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TDO 1

==== CHANNEL f1 =====  
 NUC1 1H  
 P1 10.70 usec  
 PL1 -2.00 dB  
 SFO1 300.1318534 MHz

F2 - Processing parameters  
 SI 32768  
 SF 300.1300033 MHz  
 YDW EM  
 SB 0  
 JB 0.30 Hz  
 WB 0  
 CB 1.00

**<sup>1</sup>H NMR of Compound 20h**



Current Data Parameters  
NAME 27March2018  
EXPNO 28  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20190328  
Time 10.21  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 15840  
DS 4  
SWH 17985.611 Hz  
FIDRES 0.274439 Hz  
AQ 1.8219508 sec  
RG 32768  
DW 27.800 usec  
DE 6.50 usec  
TE 300.0 K  
D1 2.00000000 sec  
d11 0.03000000 sec  
DELTA 1.89999998 sec  
TDQ 1

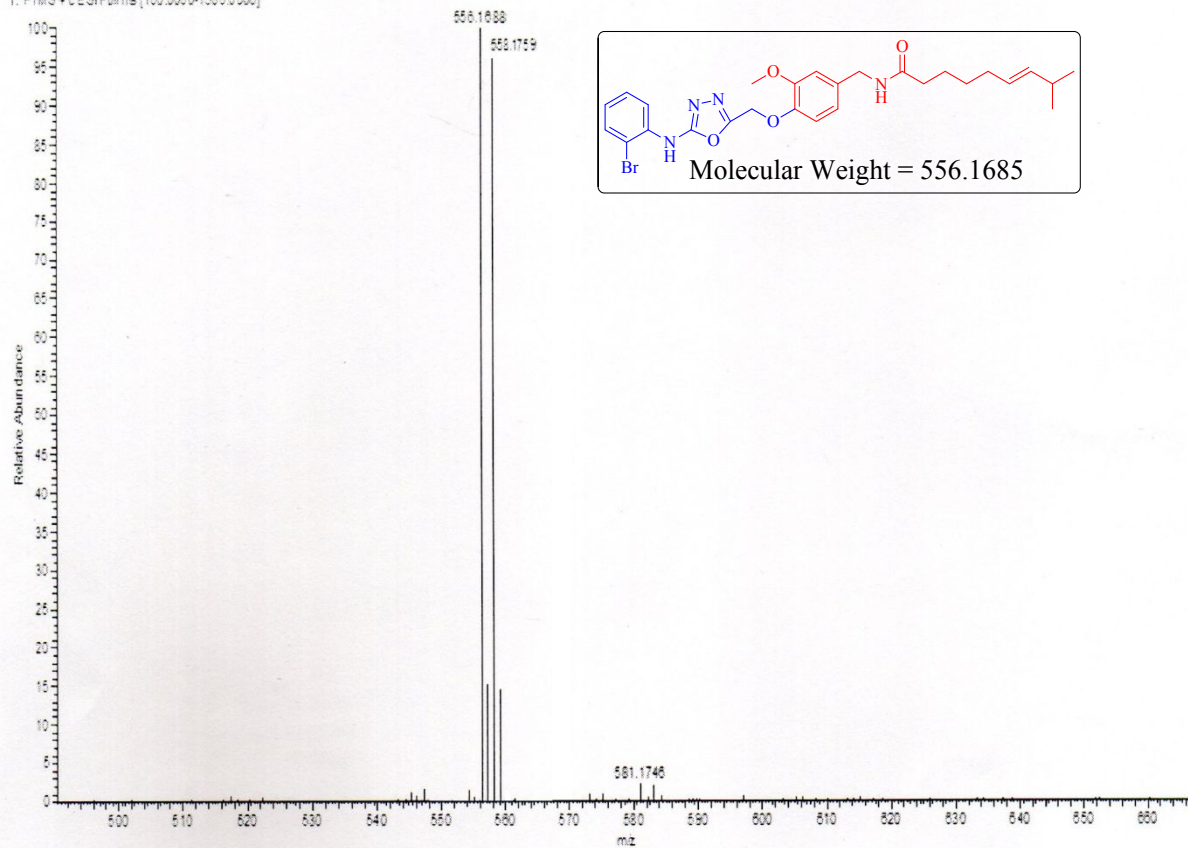
===== CHANNEL f1 =====  
NUC1 13C  
P1 10.10 usec  
PL1 -2.00 dB  
SFO1 75.4752953 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 115.00 usec  
PL2 -2.00 dB  
PL12 18.50 dB  
PL13 22.50 dB  
SFO2 300.1312005 MHz

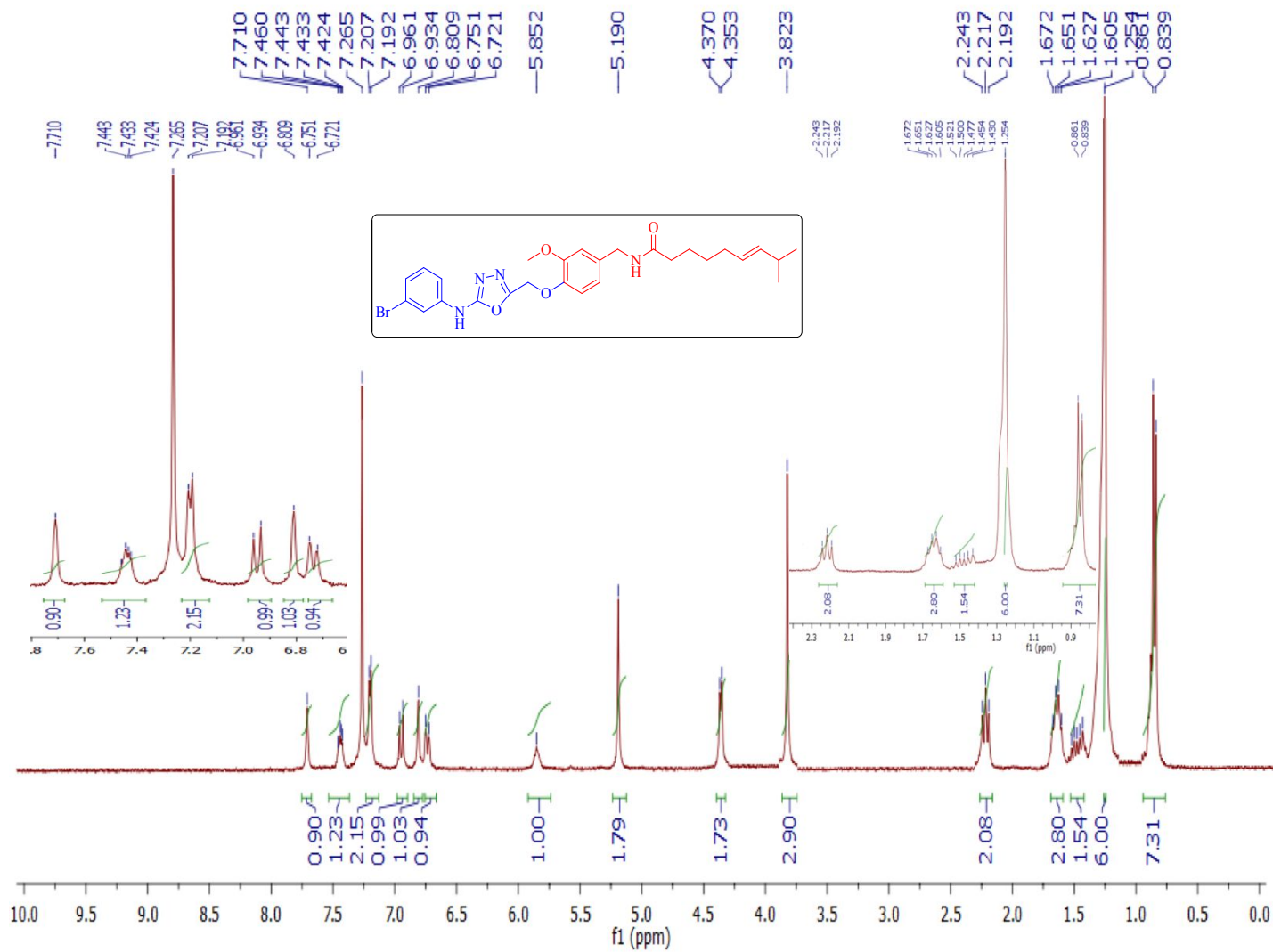
F2 - Processing parameters  
SI 32768  
SF 75.4677490 MHz  
WDW EM  
SSB 0  
LB 2.00 Hz  
GB 0  
PC 1.40

<sup>13</sup>C NMR of Compound 20h

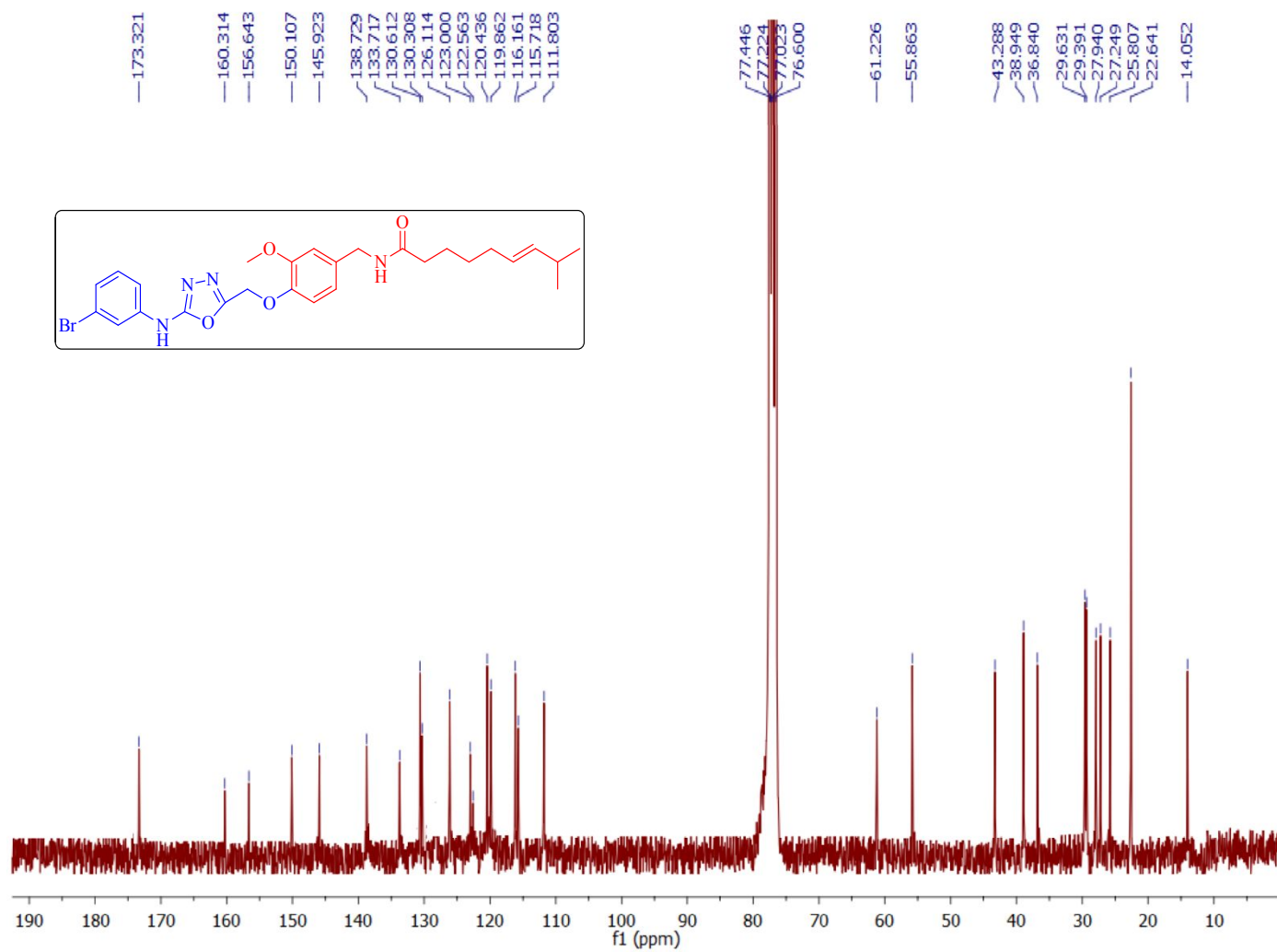
T: FTMS - cESI Full ms [100.0000-1500.0000]



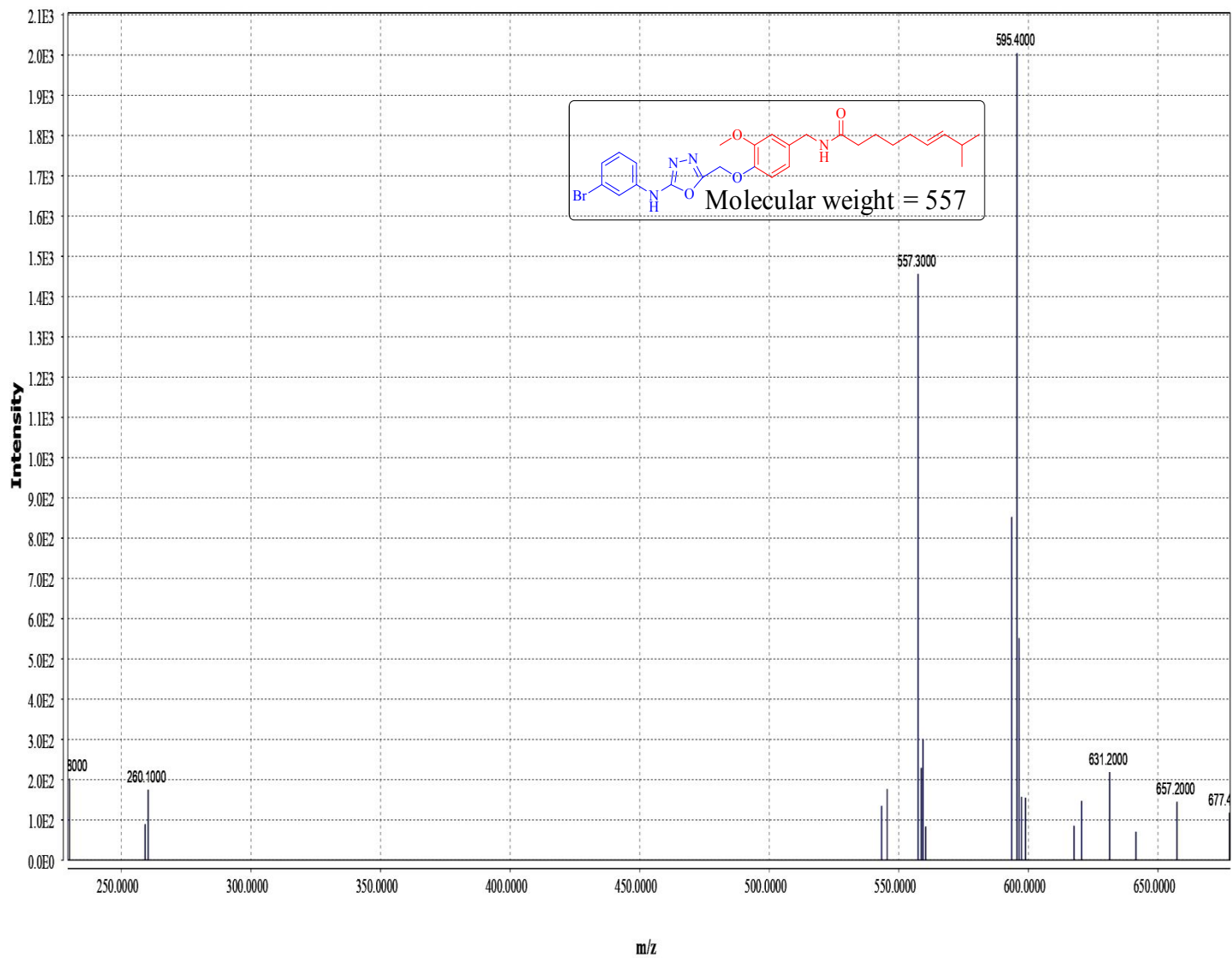
### HRMS of compound 20h



**<sup>1</sup>H NMR of Compound 20i**

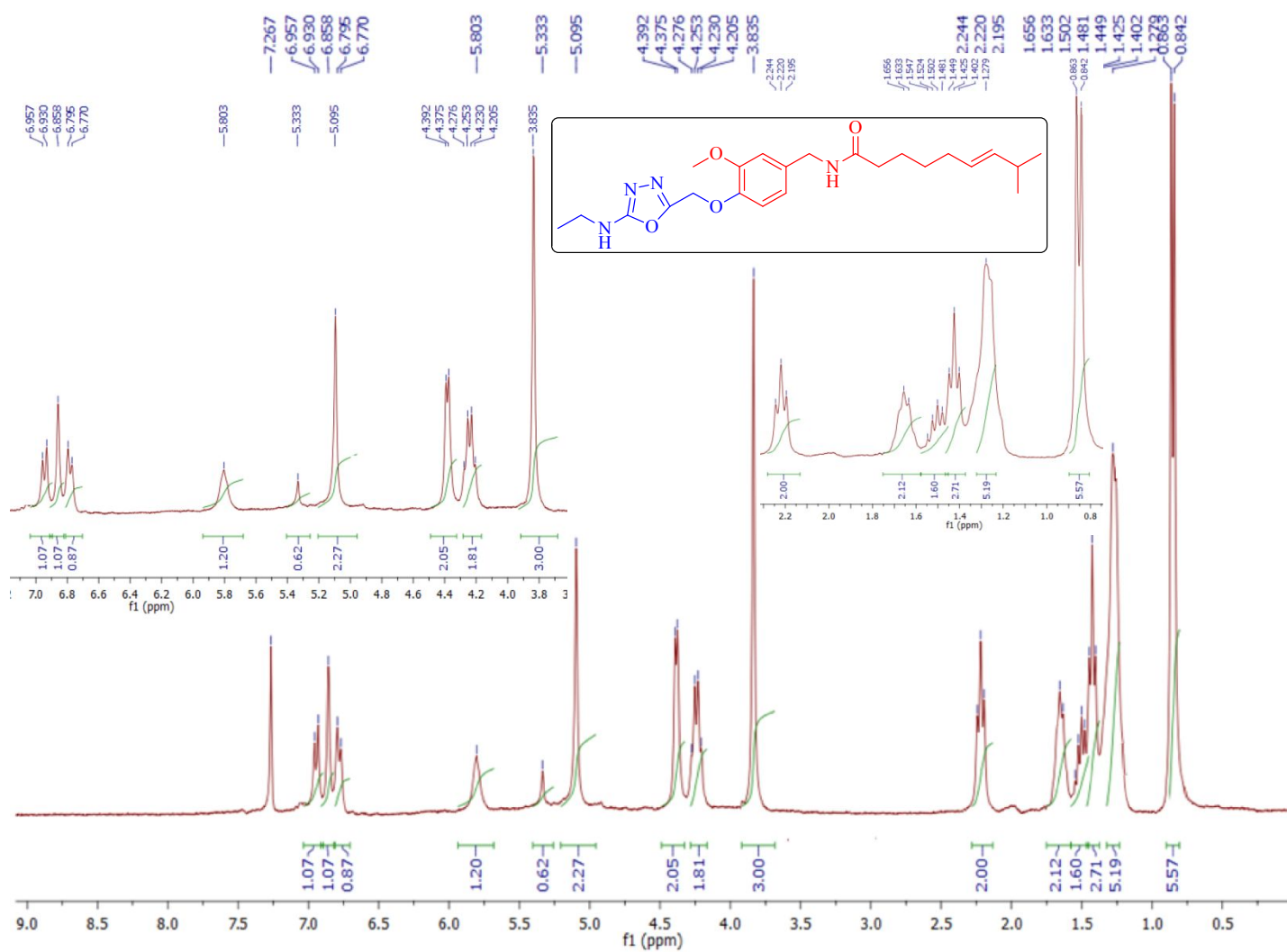


<sup>13</sup>C NMR of Compound 20i

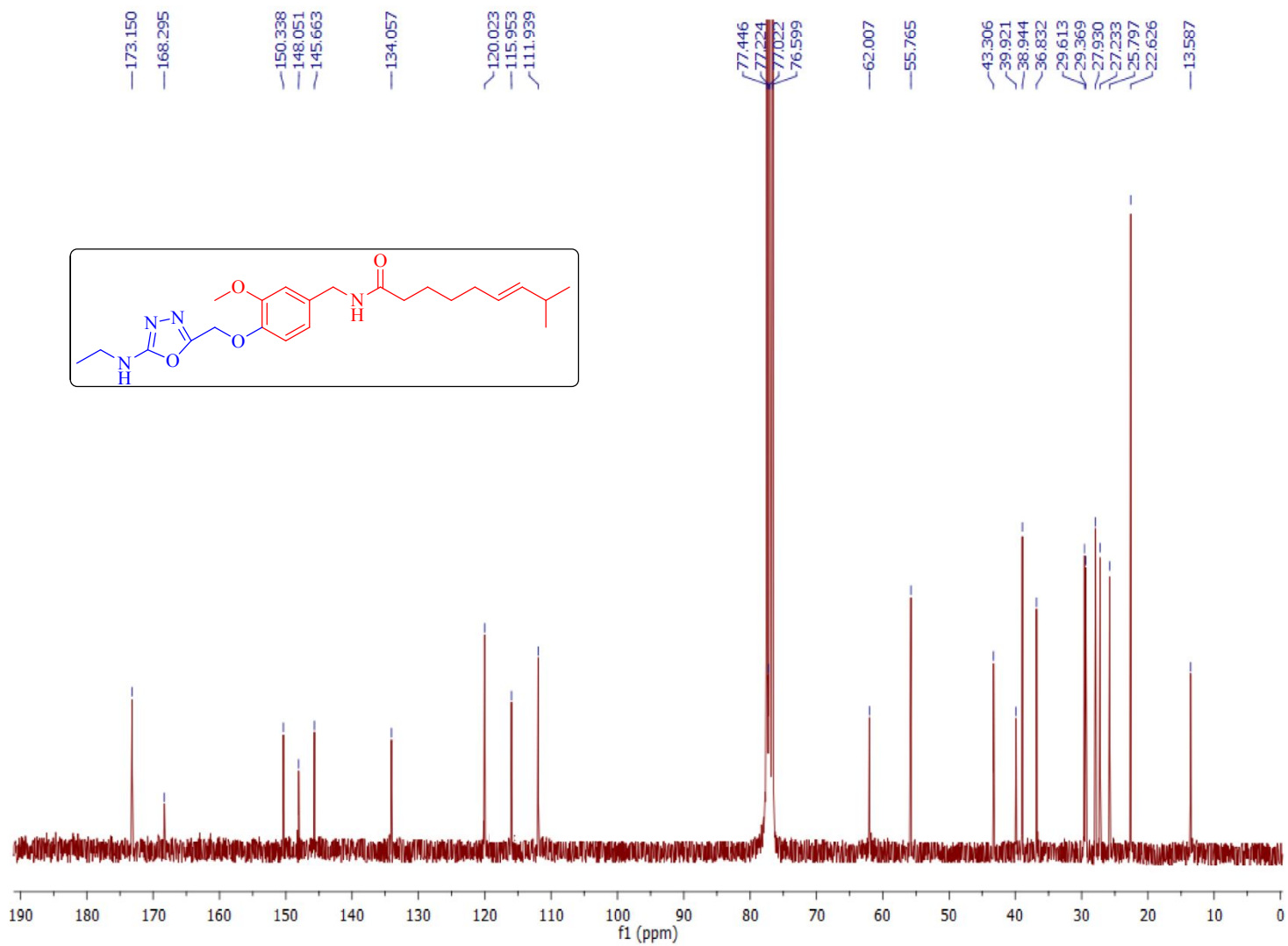


ESI-MS of compound 20i

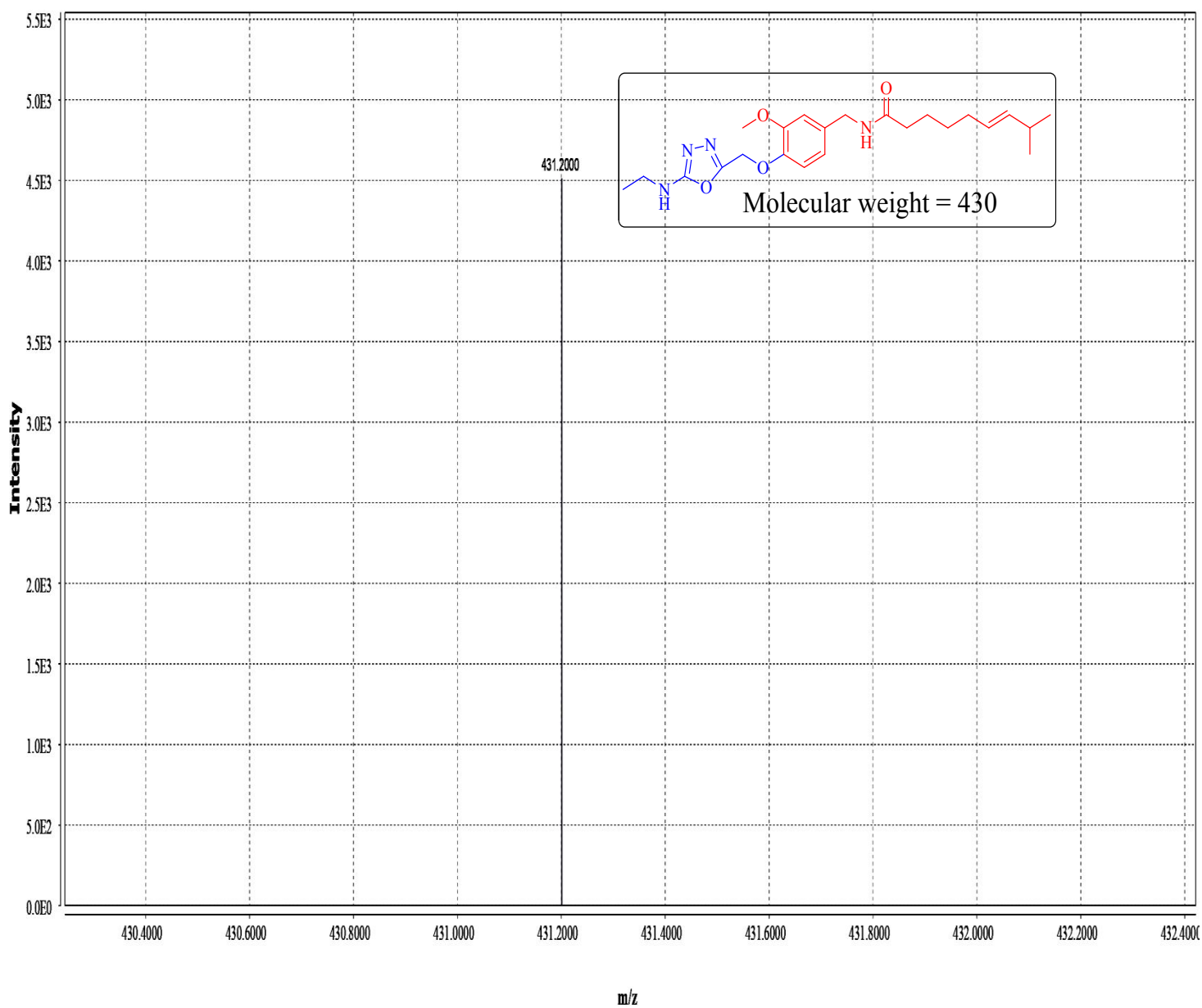




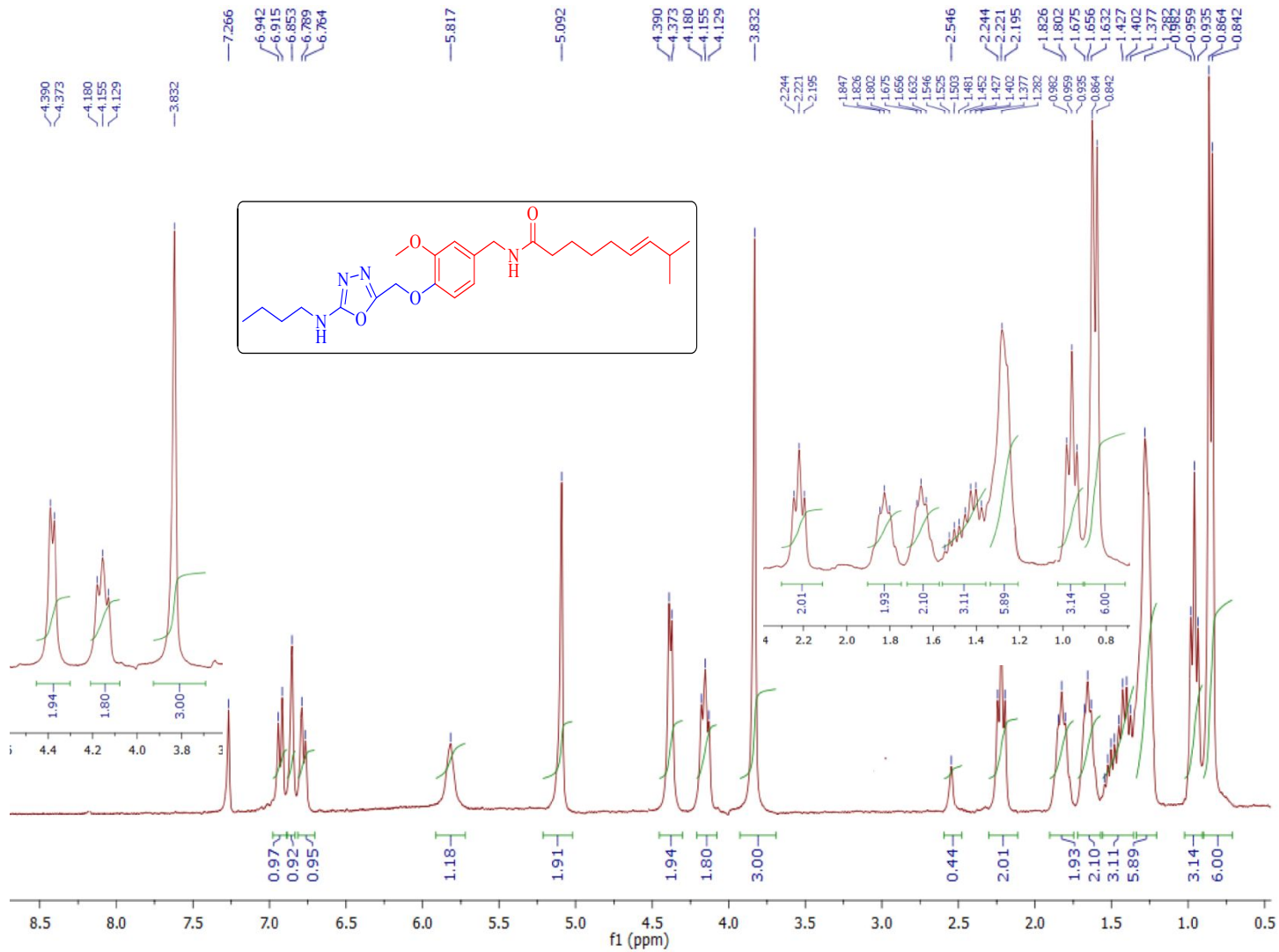
**<sup>1</sup>H NMR of Compound 20j**



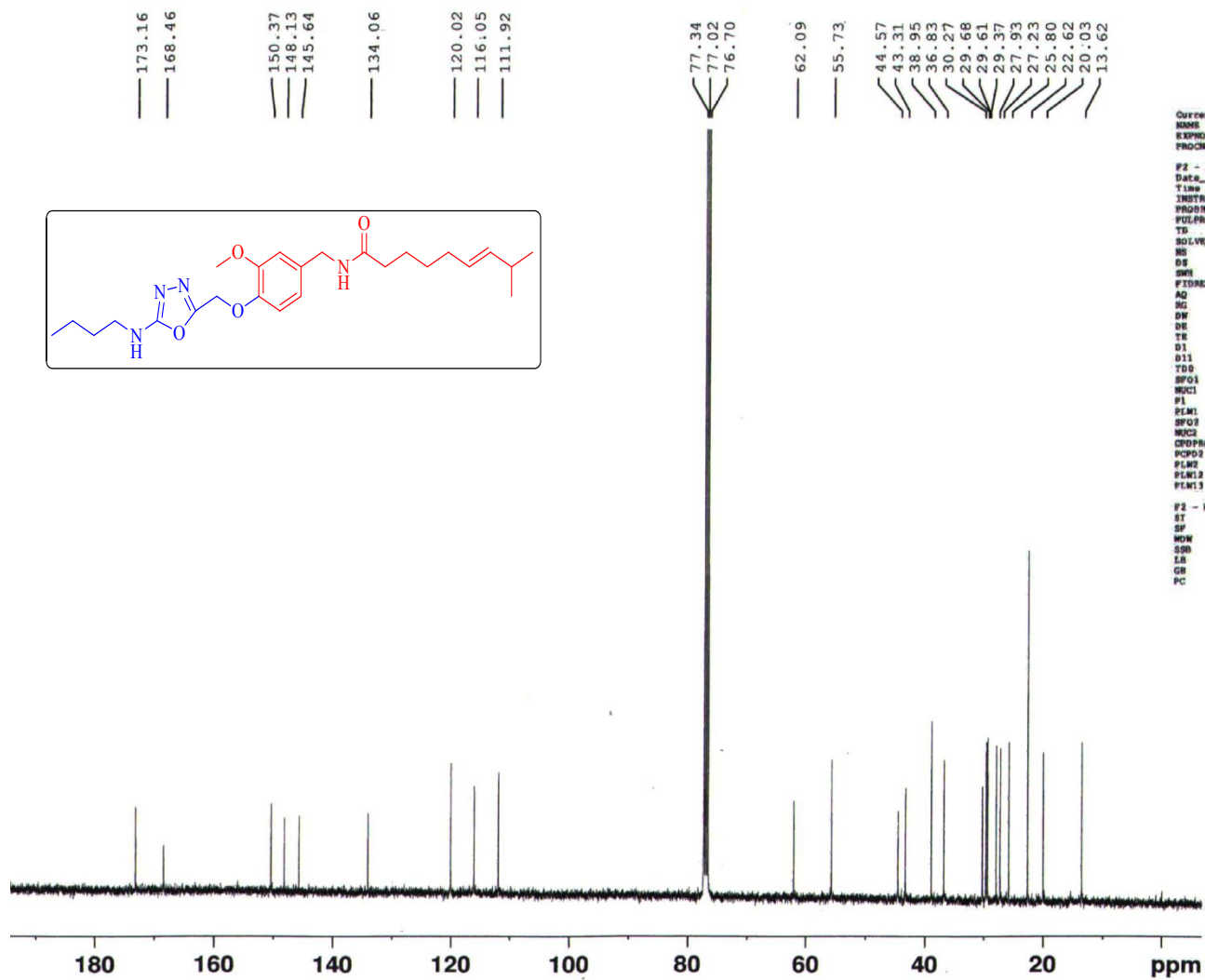
<sup>13</sup>C NMR of Compound 20j



ESI-MS of compound 20j



**<sup>1</sup>H NMR of Compound 20k**



```

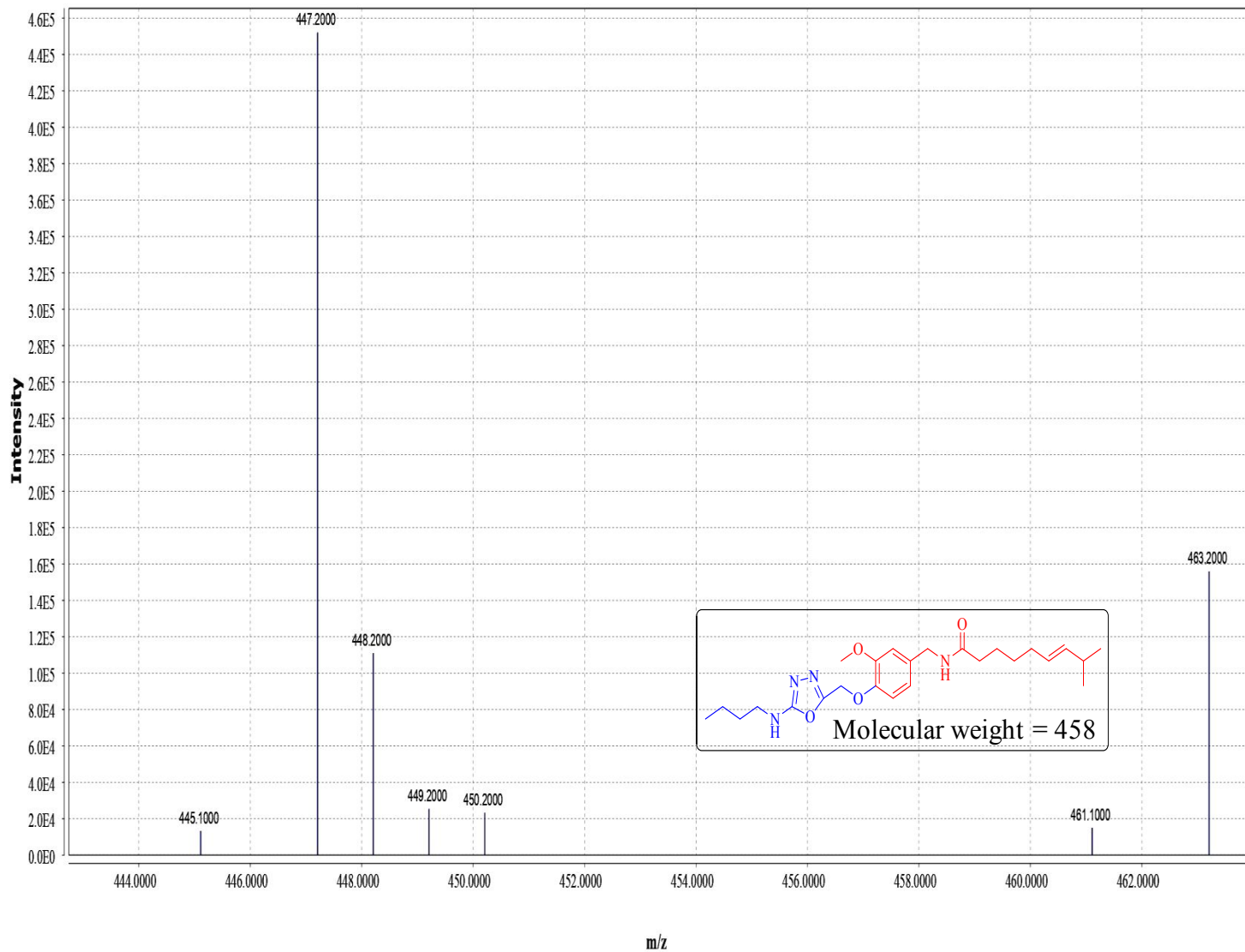
Current Data Parameters
NAME      13-c
EXPRO     357
PROCNO    1

F2 - Acquisition Parameters
Date_     20190410
Time      14.06 h
INSTRUM   spect
PROBHD    E10618_0678 (
PULPROG   zgpg30
TD        65536
SOLVENT   CDCl3
NS        1710
DS        0
SWH        24030.463 Hz
FIDRES    0.733596 Hz
AQ         1.3631488 sec
RG         209.5
DW         20.800 usec
DE         4.50 usec
TE         300.2 K
D1         1.0000000 sec
D11        0.0300000 sec
TDB        1
SFO1       100.622626 MHz
NUC1       13C
P1         9.50 usec
PL1        0.0000000 W
SFO2       400.1314001 MHz
NUC2       1H
CPDPRG2   mltz16
PCPD2     90.00 usec
PLW2      12.0000000 W
PLW12     0.29036999 W
PLW13     0.14605001 W

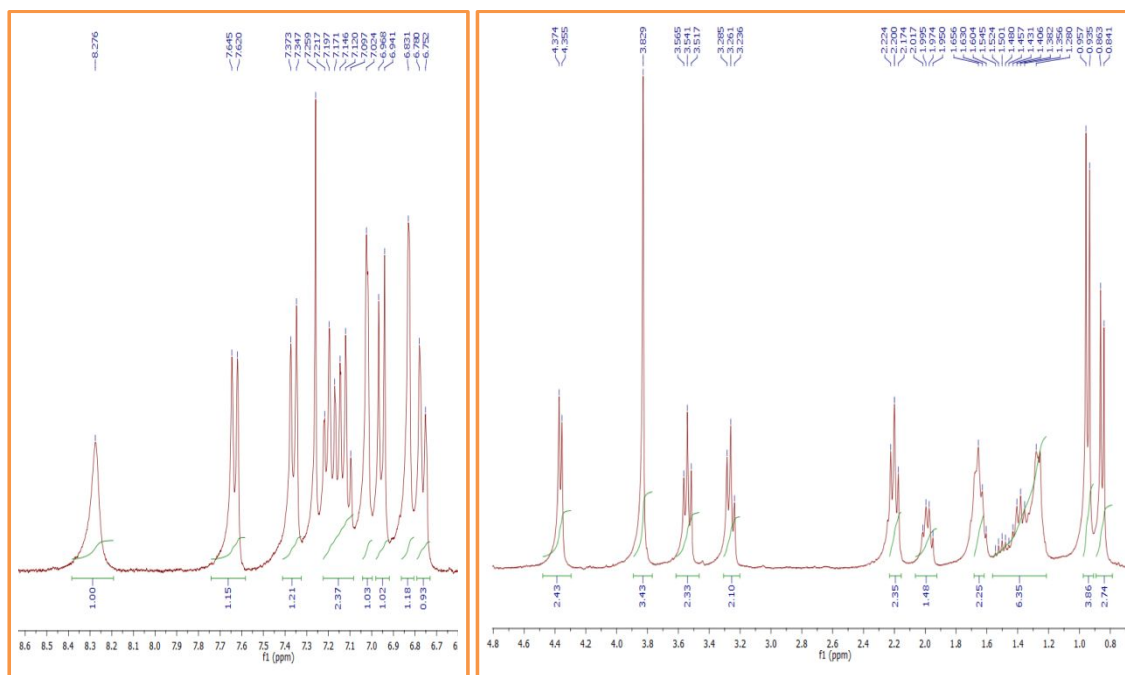
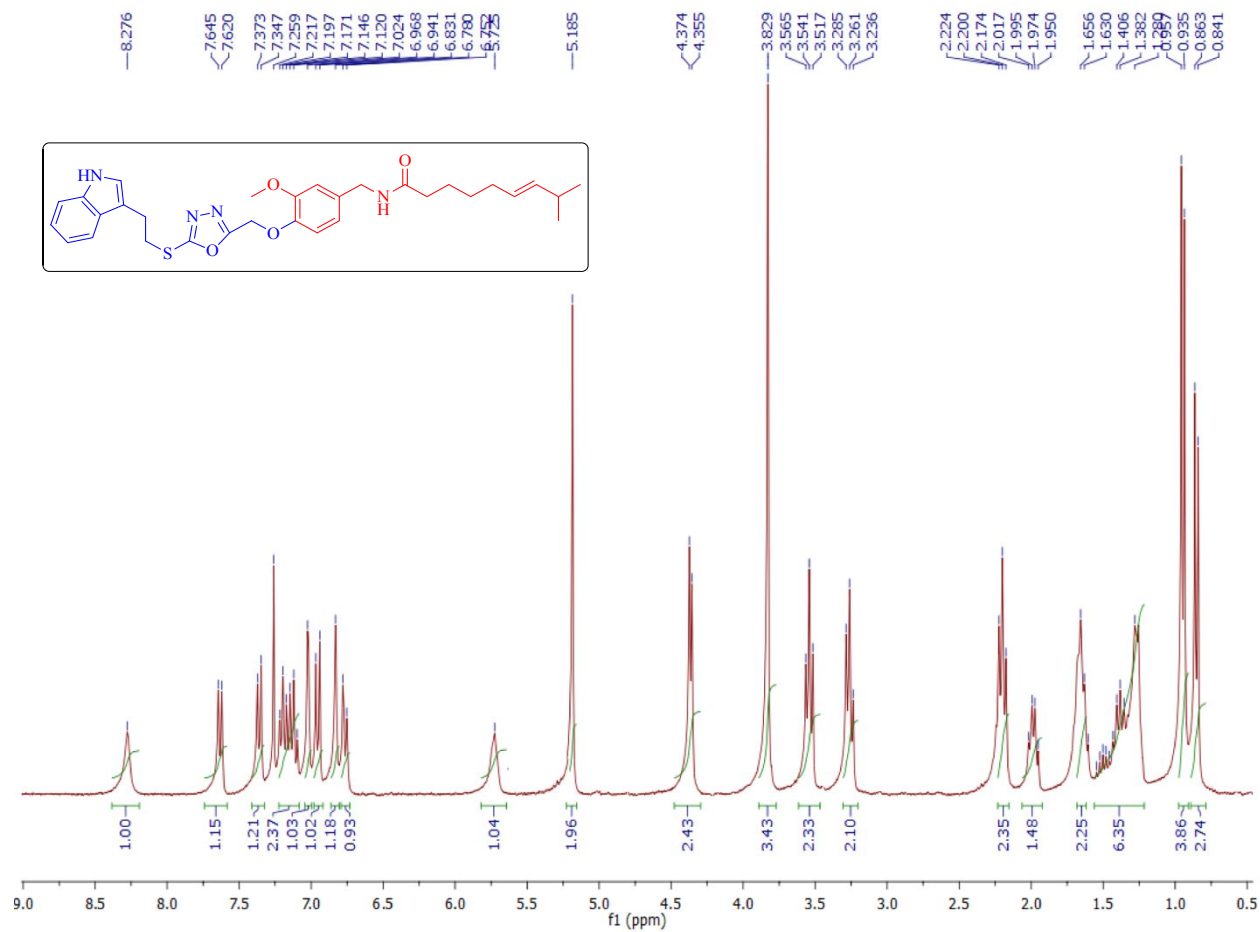
F2 - Processing parameters
SI         32768
SF         100.6127690 MHz
WDW        EM
SGB        0
LB         1.00 Hz
GB         0
PC         1.40

```

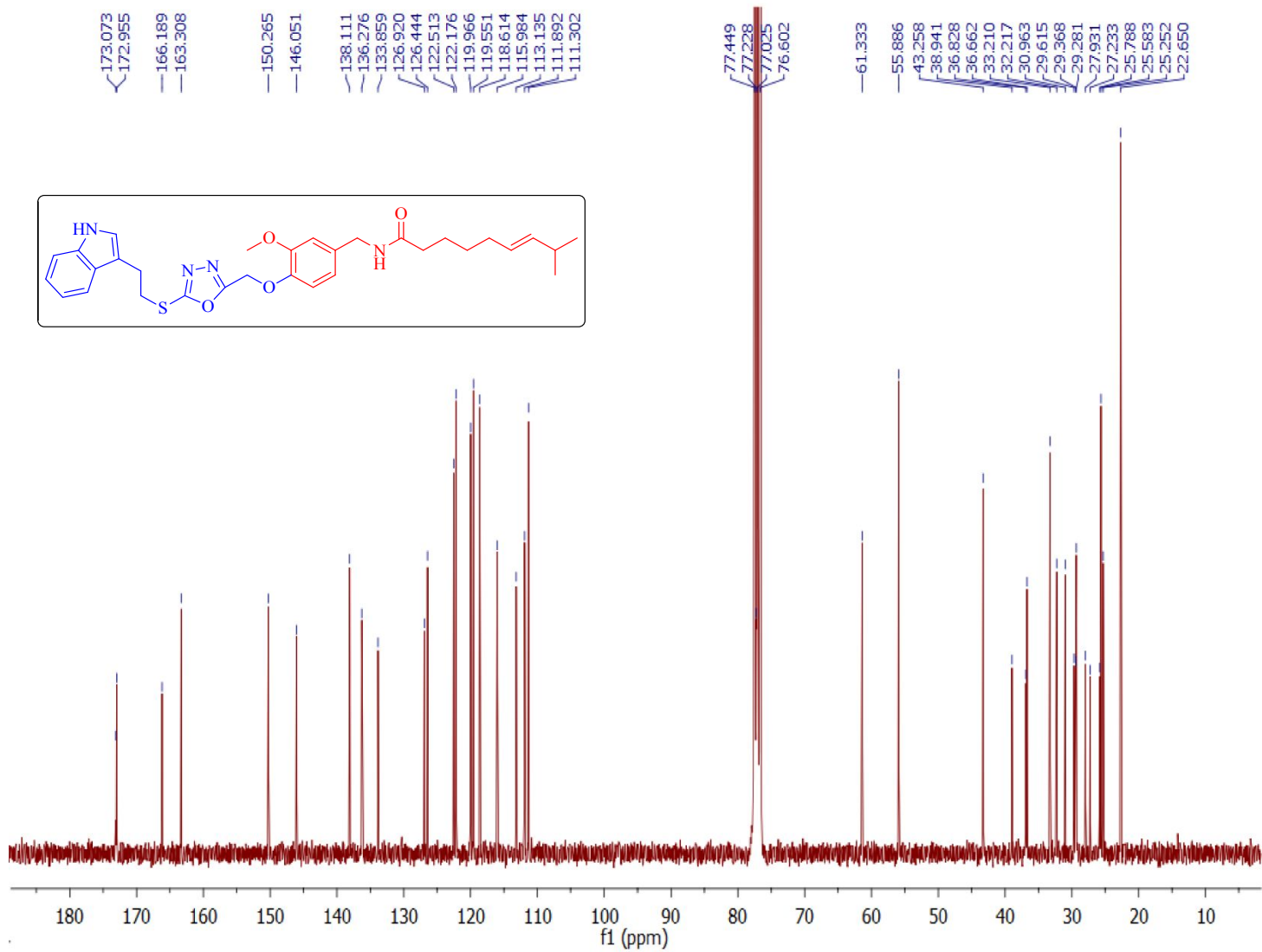
<sup>13</sup>C NMR of Compound 20k



ESI-MS of compound 20k

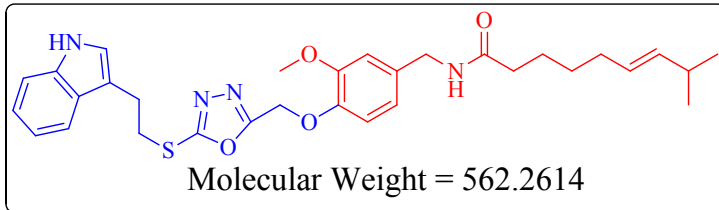


**<sup>1</sup>H NMR of Compound 22a**



<sup>13</sup>C NMR of Compound 22a

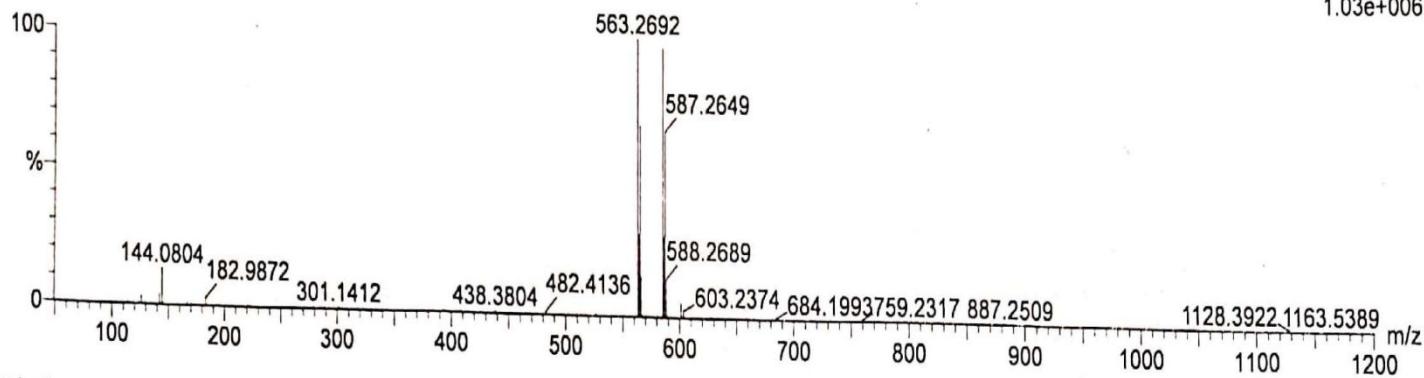




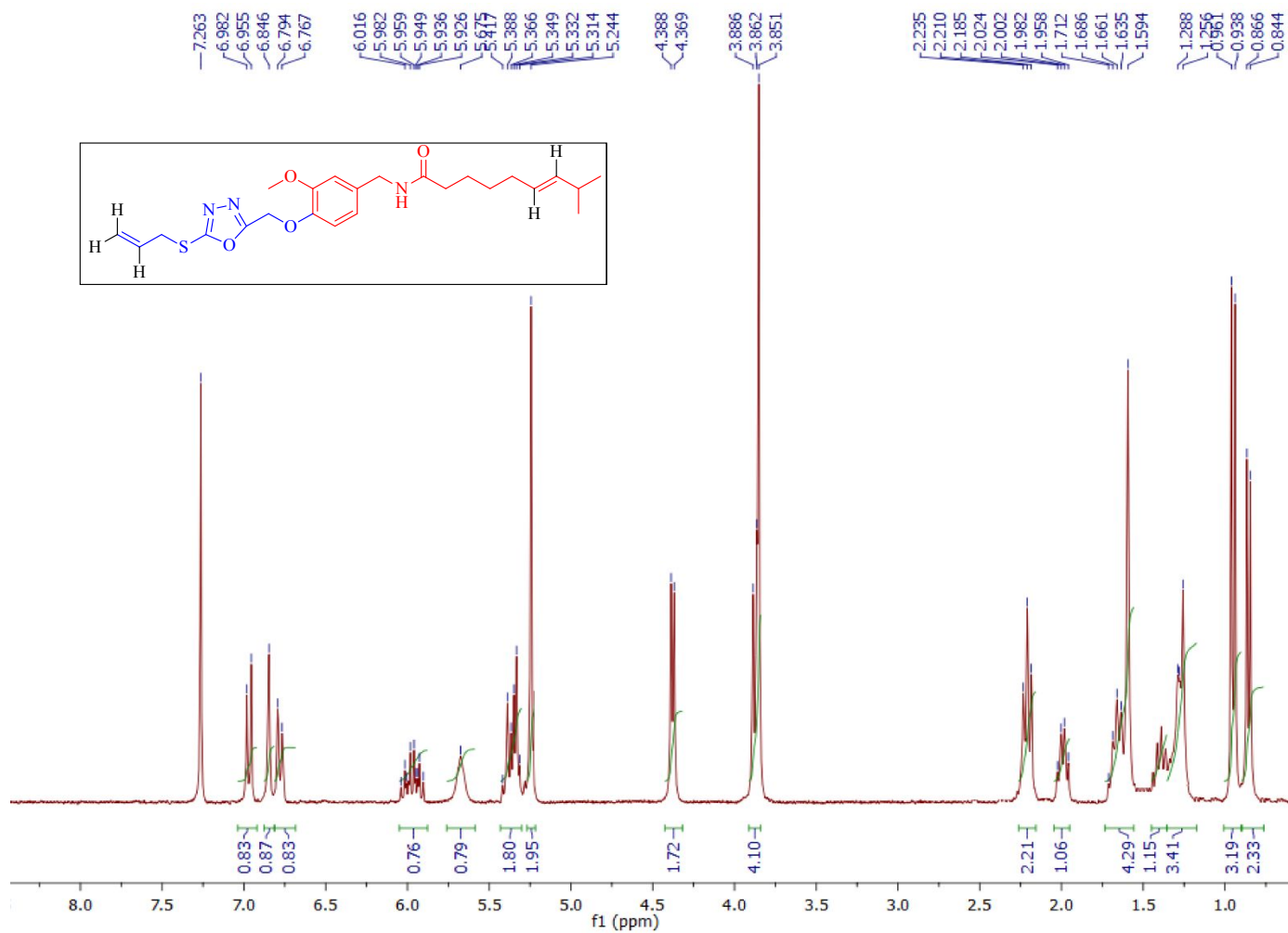
240821\_09 13 (0.276) Cm (13)

Xevo G2-XS QTOF YFC2015

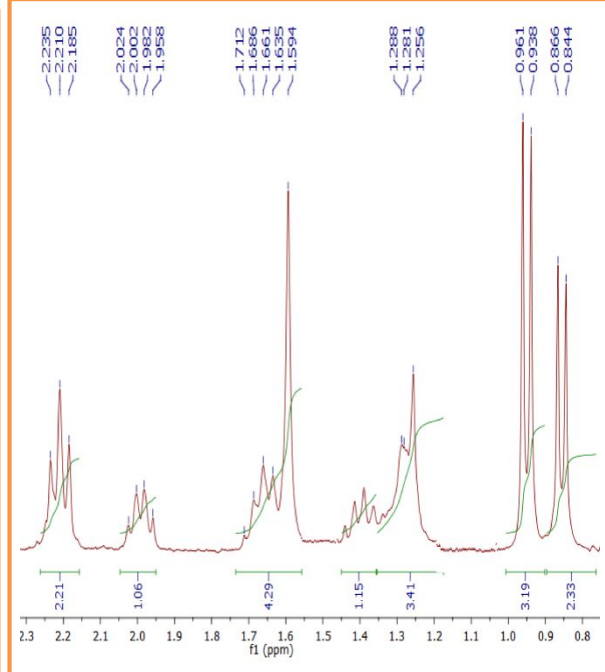
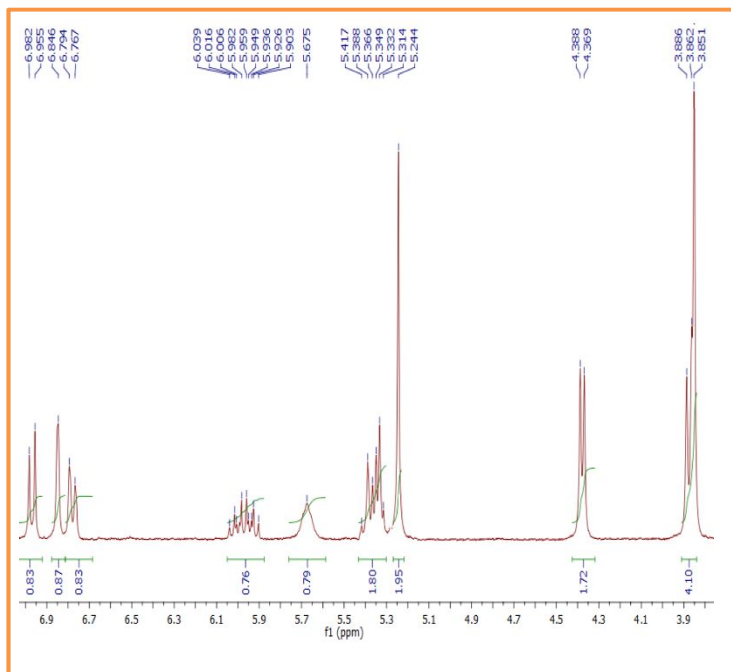
13:11:49  
1: TOF MS ES+  
1.03e+006



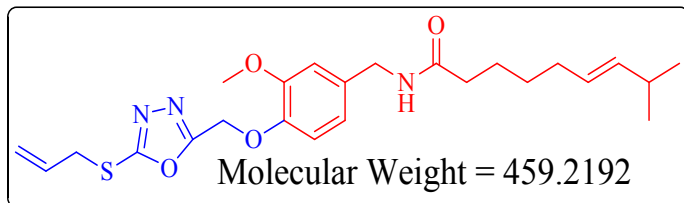
HRMS of compound 22a



**<sup>1</sup>H NMR of Compound 22b**



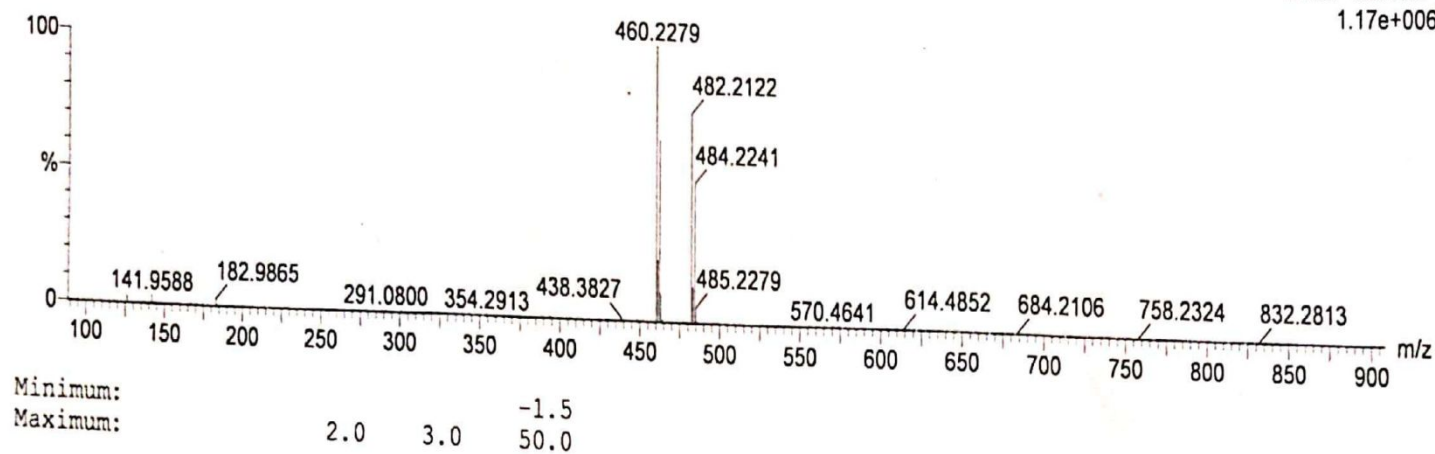
**<sup>1</sup>H NMR of Compound 22b**



240821\_06 14 (0.293) Cm (14)

Xevo G2-XS QTOF YFC2015

24-Aug-2021  
13:04:07  
1: TOF MS ES+  
1.17e+006



**HRMS of compound 22b**