# **PLOS ONE**

# Prevalence and distribution of Cryptosporidium spp. among diarrheic calves in the Republic of Korea --Manuscript Draft--

Manuscript Number:	PONE-D-21-26307
Article Type:	Research Article
Full Title:	Prevalence and distribution of Cryptosporidium spp. among diarrheic calves in the Republic of Korea
Short Title:	Different subtypes of C. parvum in calves with diarrhea
Corresponding Author:	Kyoung-Seong Choi Kyungpook National University Sangju, KOREA, REPUBLIC OF
Keywords:	Cryptosporidium; pre-weaned calves; diarrhea; C. parvum; gp60 subtypes
Abstract:	Cryptosporidium spp. are protozoan parasites that cause diarrhea in humans and animals worldwide. Data on the prevalence of Cryptosporidium spp. and its subtypes among calves in the Republic of Korea (KOR) are sparse. Hence, our study aimed to investigate the prevalence and the association between the age of calf and the identified Cryptosporidium spp. and to determine the genotypes/subtypes of Cryptosporidium spp. in pre-weaned calves with diarrhea in the KOR. A total of 460 diarrheic fecal samples were collected from calves aged 1–60 days and screened for Cryptosporidium spp. by the 18S rRNA gene. Species identification was determined by the sequencing analysis of the 18S rRNA gene, and C. parvum -positive samples were subtyped by sequence analysis of the 60-kDa glycoprotein (gp60) gene. Sequencing analysis based on the 18S rRNA gene revealed the presence of three Cryptosporidium spp., namely, C. parvum (n = 72), C. ryanae (n = 12), and C. bovis (n = 2). Co-infection by these species was not observed. The infection rate was the highest in calves aged 11–20 days (26.1%, 95% CI 17.1–35.1), whereas the lowest rate was observed in calves aged 21–30 days (7.7%, 95% CI 0.0–16.1). The prevalence of C. parvum was detected exclusively in calves aged ≤20 days, and the highest infection rate of C. ryanae was seen in calves ≥31 days of age. The occurrence of C. parvum (χ 2 = 25.300, P = 0.000) and C. ryanae (χ 2 = 18.020, P = 0.001) was significantly associated with the age of the calves. Eleven different subtypes belonging to the family IIa were recognized: IIaA14G1R1, IIaA14G3R1, IIaA15G2R1, IIaA16G4R1, IIaA17G3R1, IIaA17G4R1, IIaA19G3R1, and IIaA19G4R1. Except for two (IIaA18G3R1 and IIaA19G3R1, and IIaA19G3R1, and IIaA19G4R1. Except for two (IIaA18G3R1 and IIaA19G3R1) subtypes, nine subtypes were first identified in calves with diarrhea in the KOR. IIaA18G3R1 was the most frequently detected subtype (72.2% of calves), followed by IIaA17G3R1 (5.6%), and IIaA15G2R1 (4.2%) and IIaA19G4R1 (4.2%). These results suggest t
Order of Authors:	Dong-Hun Jang
	Hyung-Chul Cho
	Seung-Uk Shin
	Eun-Mi Kim
	Yu-Jin Park
	Sunwoo Hwang
	Jinho Park
	Kyoung-Seong Choi
Additional Information:	

### Question Response **Financial Disclosure** Kyoungg-Seong CHoi : This research was supported by the Korea Institute of Planning and Evaluation for Enter a financial disclosure statement that Technology in Food, Agriculture, and Forestry (IPET) (Grant No. 321016-01-1-HD020). The funders had no role in study design, data collection and analysis, decision to describes the sources of funding for the publish, or preparation of the manuscript. work included in this submission. Review the submission guidelines for detailed requirements. View published research articles from PLOS ONE for specific examples. This statement is required for submission and will appear in the published article if the submission is accepted. Please make sure it is accurate. Unfunded studies Enter: The author(s) received no specific funding for this work. **Funded studies** Enter a statement with the following details: · Initials of the authors who received each · Grant numbers awarded to each author · The full name of each funder • URL of each funder website · Did the sponsors or funders play any role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript? . NO - Include this sentence at the end of your statement: The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. • YES - Specify the role(s) played. \* typeset Competing Interests No: The authors have declared that they no competing interests exit. Use the instructions below to enter a competing interest statement for this submission. On behalf of all authors, disclose any competing interests that could be perceived to bias this work—acknowledging all financial support

and any other relevant financial or non-

financial competing interests.

This statement is required for submission and will appear in the published article if the submission is accepted. Please make sure it is accurate and that any funding sources listed in your Funding Information later in the submission form are also declared in your Financial Disclosure statement.

View published research articles from *PLOS ONE* for specific examples.

#### NO authors have competing interests

Enter: The authors have declared that no competing interests exist.

#### Authors with competing interests

Enter competing interest details beginning with this statement:

I have read the journal's policy and the authors of this manuscript have the following competing interests: [insert competing interests here]

#### \* typeset

#### Ethics Statement

Enter an ethics statement for this submission. This statement is required if the study involved:

- Human participants
- Human specimens or tissue
- · Vertebrate animals or cephalopods
- · Vertebrate embryos or tissues
- · Field research

Write "N/A" if the submission does not require an ethics statement.

General guidance is provided below.

Consult the <u>submission guidelines</u> for detailed instructions. Make sure that all information entered here is included in the Methods section of the manuscript.

All animal procedures were conducted according to ethical guidelines for the use of animal samples, and were approved by the Jeonbuk National University (Institutional Animal Care and Use Committee Decision No. CBNU 2020-052). All procedures and possible consequences were explained to the managers of the surveyed farm, and written consent was obtained.

#### Format for specific study types

# Human Subject Research (involving human participants and/or tissue)

- Give the name of the institutional review board or ethics committee that approved the study
- Include the approval number and/or a statement indicating approval of this research
- Indicate the form of consent obtained (written/oral) or the reason that consent was not obtained (e.g. the data were analyzed anonymously)

# Animal Research (involving vertebrate animals, embryos or tissues)

- Provide the name of the Institutional Animal Care and Use Committee (IACUC) or other relevant ethics board that reviewed the study protocol, and indicate whether they approved this research or granted a formal waiver of ethical approval
- Include an approval number if one was obtained
- If the study involved non-human primates, add additional details about animal welfare and steps taken to ameliorate suffering
- If anesthesia, euthanasia, or any kind of animal sacrifice is part of the study, include briefly which substances and/or methods were applied

#### Field Research

Include the following details if this study involves the collection of plant, animal, or other materials from a natural setting:

- · Field permit number
- Name of the institution or relevant body that granted permission

#### **Data Availability**

Authors are required to make all data underlying the findings described fully available, without restriction, and from the time of publication. PLOS allows rare exceptions to address legal and ethical concerns. See the PLOS Data Policy and FAQ for detailed information.

Yes - all data are fully available without restriction

A Data Availability Statement describing where the data can be found is required at submission. Your answers to this question constitute the Data Availability Statement and will be published in the article, if accepted.

**Important:** Stating 'data available on request from the author' is not sufficient. If your data are only available upon request, select 'No' for the first question and explain your exceptional situation in the text box.

Do the authors confirm that all data underlying the findings described in their manuscript are fully available without restriction?

Describe where the data may be found in full sentences. If you are copying our sample text, replace any instances of XXX with the appropriate details.

- If the data are held or will be held in a public repository, include URLs, accession numbers or DOIs. If this information will only be available after acceptance, indicate this by ticking the box below. For example: All XXX files are available from the XXX database (accession number(s) XXX, XXX.).
- If the data are all contained within the manuscript and/or Supporting Information files, enter the following:
   All relevant data are within the manuscript and its Supporting Information files.
- If neither of these applies but you are able to provide details of access elsewhere, with or without limitations, please do so. For example:

Data cannot be shared publicly because of [XXX]. Data are available from the XXX Institutional Data Access / Ethics Committee (contact via XXX) for researchers who meet the criteria for access to confidential data.

The data underlying the results presented in the study are available

All relevant data are within the manuscript and its Supporting Information files.

<ul> <li>from (include the name of the third party and contact information or URL).</li> <li>This text is appropriate if the data are owned by a third party and authors do not have permission to share the data.</li> <li>* typeset</li> </ul>	
Additional data availability information:	Tick here if the URLs/accession numbers/DOIs will be available only after acceptance of the manuscript for publication so that we can ensure their inclusion before publication.

Prevalence and distribution of *Cryptos* 

2 Prevalence and distribution of Cryptosporidium spp. among diarrheic calves in the

3 Republic of Korea

4

- 5 Dong-Hun Jang<sup>1</sup>, Hyung-Chul Cho<sup>1</sup>, Seung-Uk Shin<sup>1</sup>, Eun-Mi Kim<sup>1</sup>, Yu-Jin Park<sup>2</sup>,
- 6 Sunwoo Hwang<sup>1</sup>, and Jinho Park<sup>3</sup>, Kyoung-Seong Choi<sup>1,2\*</sup>

7

- 8 <sup>1</sup>Department of Animal Science and Biotechnology, College of Ecology and Environmental
- 9 Science, Kyungpook National University, Sangju 37224, Republic of Korea
- <sup>2</sup>Department of Horse/Companion and Wild animals, College of Ecology and Environmental
- 11 Science, Kyungpook National University, Sangju 37224, Republic of Korea
- <sup>3</sup>College of Veterinary Medicine, Jeonbuk National University, Iksan 54596, Republic of Korea

13

- \*Correspondence should be addressed to:
- 15 Kyoung-Seong Choi, DVM, MS, PhD
- 16 College of Ecology and Environmental Science
- 17 Kyungpook National University
- 18 Sangju 37224, Republic of Korea
- 19 Tel: 82-54-530-1222
- 20 Fax: 82-54-530-1959
- 21 E-mail: <u>kschoi3@knu.ac.kr</u>

#### Abstract

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

Cryptosporidium spp. are protozoan parasites that cause diarrhea in humans and animals worldwide. Data on the prevalence of Cryptosporidium spp. and its subtypes among calves in the Republic of Korea (KOR) are sparse. Hence, our study aimed to investigate the prevalence and the association between the age of calf and the identified Cryptosporidium spp. and to determine the genotypes/subtypes of Cryptosporidium spp. in pre-weaned calves with diarrhea in the KOR. A total of 460 diarrheic fecal samples were collected from calves aged 1–60 days and screened for *Cryptosporidium* spp. by the 18S rRNA gene. Species identification was determined by the sequencing analysis of the 18S rRNA gene, and C. parvum-positive samples were subtyped by sequence analysis of the 60-kDa glycoprotein (gp60) gene. Sequencing analysis based on the 18S rRNA gene revealed the presence of three Cryptosporidium spp., namely, C. parvum (n = 72), C. ryanae (n = 12), and C. bovis (n = 2). Co-infection by these species was not observed. The infection rate was the highest in calves aged 11-20 days (26.1%, 95% CI 17.1-35.1), whereas the lowest rate was observed in calves aged 21-30 days (7.7%, 95% CI 0.0-16.1). The prevalence of C. parvum was detected exclusively in calves aged ≤20 days, and the highest infection rate of *C. ryanae* was seen in calves  $\geq 31$  days of age. The occurrence of C. parvum ( $\chi^2 = 25.300$ , P = 0.000) and C. ryanae  $(\chi^2 = 18.020, P = 0.001)$  was significantly associated with the age of the calves. Eleven different subtypes belonging to the family IIa were recognized: IIaA14G1R1, IIaA14G3R1, IIaA15G1R1, IIaA15G2R1, IIaA16G4R1, IIaA17G3R1, IIaA17G4R1, IIaA18G3R1, IIaA19G1R1, IIaA19G3R1, and IIaA19G4R1. Except for two (IIaA18G3R1 and IIaA15G2R1) subtypes, nine subtypes were first identified in calves with diarrhea in the KOR. IIaA18G3R1 was the most frequently detected subtype (72.2% of calves), followed by IIaA17G3R1 (5.6%),

and IIaA15G2R1 (4.2%) and IIaA19G4R1 (4.2%). These results suggest that the prevalence of *Cryptosporidium* spp. is significantly associated with calf age. Furthermore, the findings

demonstrate the high genetic diversity of *C. parvum* and the widespread occurrence of zoonotic *C. parvum* in pre-weaned calves. Hence, calves are a potential source of zoonotic transmission

with considerable public health implications.

**Keywords:** Cryptosporidium, pre-weaned calves, diarrhea, C. parvum, gp60 subtypes

#### Introduction

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

Cryptosporidium spp. are protozoan parasites that cause mild-to-severe diarrhea in humans and a wide range of animals [1]. Infections with these parasites occur via the fecal-oral route either by direct contact with infected animals or by the ingestion of infective oocysts from contaminated water or food [2-5]. To date, 40 Cryptosporidium spp. have been described [6], and among them, four species, namely, C. andersoni, C. bovis, C. parvum, and C. ryanae, have been identified in cattle. The distribution of these species is known to vary according to age [4, 7]. In particular, *C. parvum* is one of the most important pathogens causing diarrhea in neonatal calves worldwide and leads to severe economic losses owing to poor growth, decreased productivity, and even death [8]. Moreover, C. parvum is the major pathogenic species that affects humans [9, 10]. Unlike C. parvum, C. bovis and C. ryanae usually infect post-weaned calves and yearlings without causing illness, and C. andersoni is mainly found in adult cattle [11-13]. The pathogenicity of *C. bovis*, and *C. ryanae* in post-weaned calves has not been established [9]. The oocysts of *C. parvum*, *C. bovis*, and *C. ryanae* are similar in size and shape. While *C. ryanae* is smaller than the others and requires molecular methods for its determination [14, 15], C. andersoni is larger in size and infects the abomasum [16]. According to the subtyping of C. parvum based on sequence analysis of the 60-kDa

According to the subtyping of *C. parvum* based on sequence analysis of the 60-kDa glycoprotein (*gp*60) gene, IIa and IId subtypes have been detected in both humans and calves and can cause zoonotic cryptosporidiosis [17]. The IIa subtype is mostly identified in calves, and IIaA15G2R1 is the predominant subtype [7] globally, including the Republic of Korea (KOR) [18]. The IId subtype is usually found in lambs and goat kids [4, 19] and has been described in calves in some countries such as Sweden, Turkey, Egypt, and China [20-23]. To date, most investigations of cryptosporidiosis in calves caused by *C. parvum* have focused on

the IIa subtype in most countries. However, there are a few studies on *C. parvum* subtypes in calves in the KOR [18, 24].

Cryptosporidium parvum infects the intestinal mucosa and accounts for over 90% of Cryptosporidium infections in neonatal calves [23]. In contrast, in pre-weaned calves, the prevalence of C. bovis and C. ryanae and their effects on causing diarrhea remain unclear. Several studies have reported that C. bovis and C. ryanae are present in pre-weaned calves [23, 25, 26] and that C. ryanae infections are particularly associated with moderate diarrhea in pre-weaned calves [23]. However, little is known about the association between C. bovis and diarrhea. In addition, a previous study has indicated that the high prevalence of C. bovis and C. ryanae in hemorrhagic diarrhea was found in the KOR [24]. Nevertheless, the pathogenicity of these organisms is still unclear. Therefore, this study aimed to investigate the prevalence of Cryptosporidium spp. in pre-weaned calves with diarrhea and to evaluate the association between the age of calf and the identified Cryptosporidium spp. Furthermore, we intended to determine genotype of Cryptosporidium spp. and subtyping of C. parvum in calves in the KOR and to assess the significance of calves as a source of human infections.

#### **Materials and methods**

#### **Ethics statement**

All animal procedures were conducted according to ethical guidelines for the use of animal samples, and were approved by the Jeonbuk National University (Institutional Animal Care and Use Committee Decision No. CBNU 2020-052). All procedures and possible consequences were explained to the managers of the surveyed farm, and written consent was obtained.

#### **Sample collection**

Between August 2019 and August 2020, fresh fecal samples were collected directly from the rectum of 460 diarrheic pre-weaned calves (up to 60 days of age) by an experienced veterinarian using sterile plastic gloves in 11 different farms located in the KOR. The samples were placed in labeled sterile plastic tubes and transported to the Animal Immunology Laboratory of Kyungpook National University in a cooler with ice packs. Upon arrival, sampling date, age, animal identification number, and fecal consistency (pasty, loose, watery, or hemorrhagic) were recorded for each animal. The collected feces were mostly pasty or loose. Prior to DNA extraction, all feces were stored at  $4^{\circ}$ C for no more than 2 days. The fecal samples were divided according to age as follows; 1-10 days (n=271), 11-20 days (n=92), 21-30 days (n=39), and  $\geq 31$  days (n=58). No microscopic examination was performed.

#### DNA extraction, molecular analysis, and sequencing

DNA was extracted from 250 mg of each fecal sample using the QIAamp Fast DNA Stool Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions and

frozen at -20°C until use. The *Cryptosporidium* spp. were first tested using the 18S rRNA gene [27]. Samples that yielded positive results for *Cryptosporidium* spp. were further tested using species-specific primers [24]. After that, C. paryum was again subtyped using a nested PCR targeting the 60-kDa glycoprotein (gp60) gene [4, 28]. All positive PCR products were purified using the AccuPower PCR Purification Kit (Bioneer, Daejeon, KOR) and employed for direct sequencing (Macrogen, Daejeon, KOR). The nucleotide sequences obtained in this study were analyzed using BioEdit (version 7.2.5) and compared with the reference sequences using the Basic Local Alignment Search Tool available at the National Center for Biotechnology Information database. To determine the subtype of C. parvum, nucleotide sequences were aligned using ClustalX and then analyzed via direct comparison with reference sequences from GenBank. Since the base sequences of C. bovis and C. ryanae are similar, all positive samples of the 18S rRNA gene were separated by comparing the sequences. In this study, only samples showing a good sequencing result were considered positive for each Cryptosporidium spp. All nucleotide sequences generated in this study were deposited in the GenBank database with appropriate MZ736386-MZ736399; accession numbers (18S)rRNA: gp60: MZ736314-MZ736385).

132

133

134

135

136

137

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

#### Statistical analysis

Statistical analysis was performed using SPSS Statistics 26 software package for Windows (SPSS Inc, Chicago, IL, USA). Chi-square test was used to determine the association between the prevalence of each species and age. A *p*-value of less than 0.05 was considered statistically significant.

## Results

# Prevalence of Cryptosporidium spp.

Among the 460 diarrheic fecal samples examined, 86 (18.7%) were positive for
Cryptosporidium spp. on PCR analysis and sequencing based on the 18S rRNA gene. Three
Cryptosporidium spp. were identified in pre-weaned Korean native calves (Table 1). Of these,
C. parvum (15.7%, 72/460) was the most detected, followed by C. ryanae (2.6%, 12/460) and
C. bovis (0.4%, 2/460). Co-infection of these species was not observed. The prevalence of the
three Cryptosporidium spp. was compared according to the age groups. As shown in Table 1,
the infection rate of Cryptosporidium spp. was highest in calves aged 11-20 days (26.1%, 95%
CI 17.1–35.1), whereas the lowest infection rate was observed in calves aged 21–30 days (7.7%,
95% CI 0.0–16.1). All three <i>Cryptosporidium</i> spp. were detected only in calves aged 1–10 days
(Table 1). The association between <i>Cryptosporidium</i> spp. and age-distribution was investigated.
Interestingly, the identified Cryptosporidium spp. varied according to the age of the calves. C.
parvum infection was detected exclusively in calves ≤20 days of age (Table 2). The prevalence
peaked at the age of 11–20 days and decreased rapidly thereafter (Table 2). C. parvum infection
was significantly associated with the age of the calves ( $\chi^2 = 25.300$ , $P = 0.000$ ). Unlike $C$ .
parvum, C. ryanae was found in all age groups, and the highest infection rate was observed at
≥31 days of age (Table 2). <i>C. ryanae</i> infection also had a significant age-related distribution
$(\chi^2 = 18.020, P = 0.001)$ . In contrast, <i>C. bovis</i> was detected only in two calves aged 10 days
and 35 days, and there was no statistical significance in the age-related distribution ( $P = 0.590$ ).

# Distribution of Cryptosporidium spp. and C. parvum subtypes

All 72 C. parvum-positive samples were successfully amplified and subtyped by

sequence analysis of the *gp60* gene. A total of 11 different subtypes belonging to the family IIa were identified (Table 3). Subtype family IId was not detected. The distinction of each subtype within the IIa was in the number of trinucleotide region of TCA and TGA repeats (i.e., had one copy of sequence ACATCA immediately after the trinucleotide repeats). As shown in Table 3, in pre-weaned Korean native calves, the most frequently detected subtype was IIaA18G3R1 (72.2%), followed by IIaA17G3R1 (5.6%), and then IIaA15G2R1 (4.2%) and IIaA19G4R1 (4.2%). Other subtypes, namely, IIaA14G1R1 (1.4%), IIaA14G3R1 (1.4%), IIaA15G1R1 (1.4%), IIaA16G4R1 (2.8%), IIaA17G4R1 (2.8%), IIaA19G1R1 (1.4%), and IIaA19G3R1 (2.8%) were also identified. There was not much of a correlation between calf age and a specific subtype. IIaA19G4R1 was observed only in calves aged 1–10 days, whereas IIaA17G3R1 was found exclusively in calves aged 11–20 days. There were more various subtypes found in calves aged 1–10 days (Table 3). The most predominant subtype, IIaA18G3R1, was seen in all ages. This is the first report on the various zoonotic subtypes circulating in the KOR.

Based on the 18S rRNA gene, 14 (12 *C. ryanae* and 2 *C. bovis*) sequences were obtained and compared with the published literature. Twelve sequences of *C. ryanae* showed 95.1%–100% similarity with each other. The *C. ryanae* sequences shared 95.7%–100% identity with those found in Austria, China, India, Thailand, and Japan. Two sequences of *C. bovis* shared 94.1% similarity. These sequences demonstrated 95.5%–96.2% identity with those identified previously in the KOR and had 91.9%–96.2% homology with those from Austria, USA, Japan, and China. Interestingly, differences in nucleotides between *C. ryanae* and *C. bovis* were observed. As shown in Fig. 1, the nucleotides in the six positions, i.e., 440, 460, 464–466, and 470 were different between the two species.

#### **Discussion**

Cryptosporidium, along with rotavirus, has been well recognized as the main pathogen causing diarrhea in neonatal calves worldwide [29]. Our findings established the prevalence of Cryptosporidium spp. in pre-weaned diarrheic calves according to age, and the presence of various zoonotic subtypes of C. parvum in the KOR were identified. In the present study, the overall prevalence of Cryptosporidium spp. was found to be 18.7%, which is higher than that reported previously in the KOR [18, 24, 30]. These variations could be explained by the age of the animals, time of sample collection, and the differences in geographical location. However, the percentage of Cryptosporidium spp.-positive samples found in our study was lower than that reported in other countries such as Germany (88.9%), Japan (83.8%), China (38.4%), Italy (38.8%), Colombia (26.6%), Argentina (22.5%), and Estonia (22.6%) [25, 31-36].

In this study, the presence of three Cryptosporidium spp. in pre-weaned Korean native calves was ascertained:  $C.\ bovis$ ,  $C.\ parvum$ , and  $C.\ ryanae$ . Of them,  $C.\ parvum$  was the most predominant species in the KOR. This finding agrees with the results observed in several other countries [7, 25, 32, 35, 37, 38]. Most studies have proven that  $C.\ parvum$  mainly infects calves up to 1 month of age [32, 39-42]. The results of the present study demonstrated that  $C.\ parvum$  was detected only in calves aged  $\leq 20$  days, and the infection rate was the highest in calves aged 11–20 days. This observation is consistent with a previous study performed by our group [18]. According to our findings,  $C.\ parvum$  was not detected in calves over 21 days of age. It is speculated that calves in this age group are less susceptible to  $C.\ parvum$  infection owing to the partial development of the immune system with increasing age, which reduces the effects compared to the neonatal calves. Moreover,  $C.\ parvum$  is known to cause watery diarrhea [23, 29]. In this study, the number of animals with watery feces was small; hence, the association

with diarrhea was not evaluated. Although we were not able to compare the occurrence of C. parvum with the diarrhea status, C. parvum was found to be the causative agent of diarrhea in young calves. Our results suggest that C. parvum infection is attributed to the significant agerelated distribution (P = 0.000). Consequently, C. parvum was strongly associated with diarrhea in calves aged  $\leq 20$  days.

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

230

Cryptosporidium ryanae was the second most frequently detected species in preweaned Korean native calves. In general, C. ryanae is often found in post-weaned calves [15]. The results revealed that C. ryanae was detected in all age groups with a low prevalence and that its occurrence increased with age. Especially, the infection rate of C. ryanae was the highest in calves aged  $\geq 31$  days. The prevalence of *C. ryanae* found in this study was similar to that of a previous study performed in the KOR [24]. Our observation confirmed that C. ryanae has an age-associated distribution, similar to C. parvum. A recent study has reported that *C. ryanae* was common in pre-weaned as well as post-weaned calves and that the infection was associated with the occurrence of moderate diarrhea in pre-weaned calves [23]. In contrast, other studies have shown that C. ryanae was not associated with diarrhea [26, 38, 43]. So far, the pathogenicity of C. ryanae is controversial. A previous study conducted in the KOR demonstrated that although it is not a single infection, the prevalence of C. ryanae was significantly high in hemorrhagic diarrhea [24]. We could not arrive at a conclusion regarding the correlation with diarrhea since the number of C. ryanae-positive samples from diarrheic calves was small. Hence, C. ryanae infection may cause diarrhea in calves  $\geq 21$  days of age and should be considered as a causative agent of diarrhea in this age group. Further studies are necessary to clarify the pathogenicity of *C. ryanae* in pre-weaned calves.

We found that the prevalence of *C. bovis* was the lowest in pre-weaned Korean native

calves. This observation is contradictory to the results reported by several studies in which C. bovis was the dominant species in pre-weaned calves [20, 44-47]. In this study, C. bovis was detected only in two calves aged 10 and 35 days. Several studies have stated that C. bovis is common in 2–3-week-old calves [41, 43]. However, our result signified that C. bovis was not detected in this age (Table 1). Cai et al. mentioned that C. bovis usually appears after weaning and that the infection can last weeks or months and contribute to the small increase in Cryptosporidium infection rates soon after weaning [26]. This observation may also explain the low prevalence of *C. bovis* in the present study. To date, information on the prevalence and clinical signs of C. bovis infection in both pre-weaned and post-weaned calves is very limited in the KOR. C. bovis could have probably been considered to be less important than C. parvum and therefore overlooked as an etiological agent of diarrhea in calves. Moreover, the results revealed that infection by C. bovis, unlike the two other species, was not age-related. Most importantly, the involvement of *C. bovis* in diarrhea remains unclear. Unlike *C. ryanae*, many studies have suggested that C. bovis was associated with diarrhea [23, 26, 38, 48]. However, infection by C. bovis/C. ryanae may lead to clinical signs owing to the presence of C. parvum [32]. Therefore, the prevalence and pathogenicity of C. bovis in pre-weaned and post-weaned calves must be investigated through large-scale epidemiological surveys.

231

232

233

234

235

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

*C. parvum* IIa family is common in humans as well as calves and is considered potentially zoonotic. To date, three *C. parvum* subtypes have been detected in calves in the KOR [18, 24], whereas one subtype (IIaA16G3R1) was not found in this study. In addition to the two subtypes (IIaA15G2R1 and IIaA18G3R1) described above, nine other subtypes (IIaA14G1R1, IIaA14G3R1, IIaA15G1R1, IIaA16G4R1, IIaA17G3R1, IIaA17G4R1, IIaA19G1R1, IIaA19G3R1, and IIaA19G4R1) that have not previously been detected in the

KOR were identified for the first time, showing the presence of high genetic diversity. Among them, IIaA18G3R1 was most commonly found in pre-weaned Korean native calves with diarrhea. This result is inconsistent with that of a previous study in which IIaA15G2R1 was shown as the predominant subtype [18]. This difference could be attributed to the fact that in the previous study, both normal and diarrheic feces were used and that IIaA15G2R1 was detected regardless of diarrhea [18]. Other variations are due to the differences in the season of sampling, regions, the number of samples, and herd management. IIaA15G2R1 has been known as the most prevalent *C. parvum* subtype infecting humans and cattle in many countries [7, 33, 49-53] and has also been detected in calves without diarrhea [18, 32, 54]. There seems to be no relationship between the subtype and diarrhea. In the present study, IIaA15G2R1 was detected only in three calves with diarrhea and the third frequent subtype along with IIaA19G4R1.

Here, IIaA18G3R1 was the dominant subtype that accounted for 72.2% of *C. parvum*infected pre-weaned Korean native calves and was the frequent cause of human
cryptosporidiosis, besides being reported in calves and foals [55-60]. The second common
subtype in the KOR, IIaA17G3R1, has been found in calves and humans in several countries
[61-65]. IIaA19G4R1 was the third frequent subtype identified in the pre-weaned Korean
native calves and was also detected in small ruminants and fish as well as humans and calves
[55, 64, 66-68]. Interestingly, all sequences belonging to the IIaA19G4R1 subtype were
identical to those reported from other countries previously. These subtypes are considered to
be the most common ones in calves in the KOR.

The other seven subtypes were also identified in pre-weaned Korean native calves with diarrhea, but their prevalence was relatively low. Subtypes IIaA14G1R1, IIaA14G3R1, and

IIaA15G1R1 were each detected in one calf. IIaA14G1R1 was identified in calves, goat kids, and humans [7, 12, 17, 19, 25, 33, 51, 52]. IIaA14G3R1 was found in humans, calf, lambs, and fresh molluscan shellfish [19, 25, 69, 70]. IIaA15G1R1 has been reported in humans [28, 51, 52, 71, 72] as well as in cattle and goat kids [22, 73-75]. Subtypes IIaA16G4R1 and IIaA17G4R1 were each found in two calves in the current study. Unlike the other subtypes, IIaA16G4R1 has so far been noted only in neonatal calf with diarrhea [76], which is consistent with our findings. Subtype IIaA16G4R1 has not yet been detected in humans; however, it cannot be excluded the possibility that this may represent a significant health risk. IIaA17G4R1 subtype has been identified in humans, cattle, and goats [31, 33, 59, 70, 76, 77] and has further been detected in diarrheic calves [31]. Finally, subtypes IIaA19G1R1 and IIaA19G3R1 have each been identified in one calf. IIaA19G1R1 has been reported in humans, cattle, and sheep [35, 52, 63, 78-80]. IIaA19G3R1 has been identified in humans, cattle, and deer [60, 81-84]. This study is the first to report the presence of various subtypes in pre-weaned calves in the KOR.

To detect *C. bovis* and *C. ryanae*, 18S rRNA and heat-shock protein 70 genes are generally used [15]. According to sequence analysis of the 18S rRNA gene, *C. bovis* and *C. ryanae* showed ≥99% identity, and it is not always possible to differentiate between them by PCR [85, 86]. However, in this study, we used only the 18S rRNA gene. Even without phylogenetic analysis, the difference between the two species could be confirmed by sequencing analysis. At the six nucleotide positions of 440, 460, 464–466, and 470, *C. bovis* had C, T, A, T, C, and A, while *C. ryanae* had T, C, G, C, T, and G. These positions are representative markers that distinguish *C. ryanae* from *C. bovis*. Our results suggest that these two species can be discerned using the 18S rRNA gene.

#### Conclusion

Our results confirm the presence of three *Cryptosporidium* spp. in pre-weaned calves with diarrhea: *C. bovis*, *C. parvum*, and *C. ryanae*. *C. parvum* was found to be the dominant species in young calves in the KOR. The occurrence of *C. ryanae* and *C. parvum*, but not *C. bovis*, in pre-weaned Korean native calves was significantly related to age; the prevalence of *C. parvum* decreased with age, whereas that of *C. ryanae* increased with age. The most frequently detected subtype in calves with diarrhea was IIaA18G3R1, which was responsible for zoonotic transmission. This is the first report to identify nine potentially zoonotic subtypes belonging to the family IIa, which have not previously been reported in cattle in the KOR. This study establishes the high genetic diversity of *C. parvum* in diarrheic calves and the widespread distribution of zoonotic *C. parvum* in the KOR. Therefore, the results emphasize that young calves may be a potential source of infection and may serve as an important zoonotic reservoir for human cryptosporidiosis.

#### Acknowledgements

Not applicable

#### **Author Contributions**

- Conceptualization: Kyoung-Seong Choi, Jinho Park
- **Data curation:** Kyoung-Seong Choi, Jinho Park
- Formal analysis: Dong-Hun Jang, Hyung-Chul Cho, Seung-Uk Shin, Eun-Mi Kim<sup>1</sup>, Yu-Jin
- 322 Park, Sunwoo Hwang,

Methodology: Dong-Hun Jang, Hyung-Chul Cho, Seung-Uk Shin, Eun-Mi Kim, Yu-Jin Park, 323 Sunwoo Hwang 324 325 Funding acquisition: Kyoung-Seong Choi Writing - original draft: Kyoung-Seong Choi, 326 327 328 **Data Availability Statement** All data generated or analyzed during this study are included in the article. The 329 nucleotide sequence obtained in the present study has been deposited in the GenBank database 330 under the accession numbers MZ736314-MZ736399. 331 332 **Funding** 333 This research was supported by the Korea Institute of Planning and Evaluation for 334 Technology in Food, Agriculture, and Forestry (IPET) (Grant No. 321016-01-1-HD020). The 335 funders had no role in study design, data collection and analysis, decision to publish, or 336 preparation of the manuscript. 337 338 **Competing interests** 339 340 The authors have declared that they no competing interests exit.

#### References

- 343 1. Fayer R (2004) *Cryptosporidium*: a water-borne zoonotic parasite. Vet Parasitol 126:
- 37-56. https://doi.org/10.1016/j.vetpar.2004.09.004 PMID: 15567578
- 345 2. Xiao L, Ryan UM (2004) Cryptosporidiosis: an update in molecular epidemiology.
- 346 Curr Opin Infect Dis 17:483-490. https://doi.org/10.1097/00001432-200410000-
- 347 00014 PMID: 15353969
- 348 3. McLauchlin J, Amar C, Pedraza-Diaz S, Nichols GL (2000) Molecular
- epidemiological analysis of *Cryptosporidium* spp. in the United Kingdom: results of
- genotyping *Cryptosporidium* spp. in 1,705 fecal samples from humans and 105 fecal
- samples from livestock animals. J Clin Microbiol 38:3984-3990.
- https://doi.org/10.1128/JCM.38.11.3984-3990.2000 PMID: 11060056
- 353 4. Xiao L (2010) Molecular epidemiology of cryptosporidiosis: an update. Exp Parasitol
- 354 124:80-89. https://doi.org/10.1016/j.exppara.2009.03.018 PMID: 19358845
- Wells B, Shaw H, Hotchkiss E, Gilray J, Ayton R, Green J, et al. (2015) Prevalence,
- species identification and genotyping *Cryptosporidium* from livestock and deer in a
- catchment in the Cairngorms with a history of a contaminated public water supply.
- Parasit Vectors 8:66. https://doi.org/10.1186/s13071-015-0684-x PMID: 25650114
- Feng Y, Ryan UM, Xiao L (2018) Genetic diversity and population structure of
- 360 *Cryptosporidium*. Trends Parasitol 34:997-1011.
- 361 https://doi.org/10.1016/j.pt.2018.07.009 PMID: 30108020
- 362 7. Lichtmannsperger K, Harl J, Freudenthaler K, Hinney B, Wittek T, Joachim A (2020)
- 363 Cryptosporidium parvum, Cryptosporidium ryanae, and Cryptosporidium bovis in
- samples from calves in Austria. Parasitol Res 119:4291-4295.

- 365 https://doi.org/10.1007/s00436-020-06928-5 PMID: 33057813
- 366 8. Dessi G, Tamponi C, Varcasia A, Sanna G, Pipia AP, Carta S, et al. (2020)
- 367 *Cryptosporidium* infections in sheep farms from Italy. Parasitol Res 119:4211-4218.
- 368 https://doi.org/10.1007/s00436-020-06947-2 PMID: 33140165
- 369 9. Thomson S, Hamilton CA, Hope JC, Katzer F, Mabbott NA, Morrison LJ, et al. (2017)
- Bovine cryptosporidiosis: impact, host-parasite interaction and control strategies. Vet
- 371 Res 48:42. https://doi.org/10.1186/s13567-017-0447-0 PMID: 28800747
- 372 10. Caffarena RD, Meireles MV, Carrasco-Letelier L, Picasso-Risso C, Santana BN, Riet-
- Correa F, et al. (2020) Dairy calves in Uruguay are reservoirs of zoonotic subtypes of
- 374 *Cryptosporidium parvum* and pose a potential risk of surface water contamination.
- Front Vet Sci 7:562. https://doi.org/10.3389/fvets.2020.00562 PMID: 32974408
- 376 11. Aberg M, Emanuelson U, Troell K, Bjorkman C (2020). A single-cohort study of
- 377 Cryptosporidium bovis and Cryptosporidium ryanae in dairy cattle from birth to
- 378 calving. Vet Parasitol Reg Stud Reports 20:100400. https://doi.org/doi:
- 379 10.1016/j.vprsr.2020.100400 PMID: 32448548
- 380 12. Yildirim A, Adanir R, Inci A, Yukari BA, Duzlu O, Onder Z, et al. (2020) Prevalence
- and genotyping of bovine *Cryptosporidium* species in the Mediterranean and Central
- Anatolia region of Turkey. Comp Immunol Microbiol Infect Dis 69:101425.
- 383 https://doi.org/10.1016/j.cimid.2020.101425 PMID: 31978845
- 13. Liang N, Wu Y, Sun M, Chang Y, Lin X, Yu L, et al. (2019) Molecular epidemiology
- of Cryptosporidium spp. in dairy cattle in Guangdong Province, South China.
- Parasitology146:28-32. https://doi.org/10.1017/S0031182018001129 PMID:
- 387 29986775

- Fayer R, Santin M, Xiao L (2005) Cryptosporidium bovis n. sp. (Apicomplexa:
- 389 Cryptosporidiidae) in cattle (Bos taurus). J Parasitol 91:624-629.
- 390 https://doi.org/10.1645/GE-3435 PMID: 16108557
- 391 15. Fayer R, Santin M, Trout JM (2008) Cryptosporidium ryanae n. sp. (Apicomplexa:
- 392 Cryptosporidiidae) in cattle (Bos taurus). Vet Parasitol 156:191-198.
- 393 https://doi.org/10.1016/j.vetpar.2008.05.024 PMID: 18583057
- 394 16. Peng MM, Wilson ML, Holland RE, Meshnick SR, Lal AA, Xiao L (2003) Genetic
- diversity of *Cryptosporidium* spp. in cattle in Michigan: implications for understanding
- the transmission dynamics. Parasitol Res 90:175-180. https://doi.org/10.1007/s00436-
- 397 003-0834-5 PMID: 12783304
- 398 17. Khan A, Shaik JS, Grigg ME (2018) Genomics and molecular epidemiology of
- 399 Cryptosporidium species. Acta Trop 184:1-14.
- 400 https://doi.org/10.1016/j.actatropica.2017.10.023 PMID: 29111140
- 401 18. Lee YJ, Ryu JH, Shin SU, Choi KS (2019) Prevalence and molecular characterization
- of *Cryptosporidium* and *Giardia* in pre-weaned native calves in the Republic of Korea.
- 403 Parasitol Res 118:3509-3517. https://doi.org/10.1007/s00436-019-06482-9 PMID:
- 404 31624910
- 405 19. Kabir MHB, Ceylan O, Ceylan C, Shehata AA, Bando H, Essa MI, et al. (2020)
- Molecular detection of genotypes and subtypes of *Cryptosporidium* infection in
- diarrheic calves, lambs, and goat kids from Turkey. Parasitol Int 79:102163.
- 408 https://doi.org/10.1016/j.parint.2020.102163 PMID: 32589940
- 409 20. Silverlas C, Naslund K, Bjorkman C, Mattsson JG (2010) Molecular characterisation
- of *Cryptosporidium* isolates from Swedish dairy cattle in relation to age, diarrhoea and

- region. Vet Parasitol 169:289-295. https://doi.org/10.1016/j.vetpar.2010.01.003 PMID:
- 412 20138705
- 413 21. Muhid A, Robertson I, Ng J, Ryan U (2011) Prevalence of and management factors
- 414 contributing to *Cryptosporidium sp.* infection in pre-weaned and post-weaned calves
- in Johor, Malaysia. Exp Parasitol 127:534-538.
- 416 https://doi.org/10.1016/j.exppara.2010.10.015 PMID: 21050848
- Taylan-Ozkan A, Yasa-Duru S, Usluca S, Lysen C, Ye J, Roellig DM, et al. (2016)
- 418 Cryptosporidium species and Cryptosporidium parvum subtypes in dairy calves and
- goat kids reared under traditional farming systems in Turkey. Exp Parasitol 170:16-20.
- 420 https://doi.org/10.1016/j.exppara.2016.06.014 PMID: 27373430
- 421 23. Li N, Wang R, Cai M, Jiang W, Feng Y, Xiao L (2019) Outbreak of cryptosporidiosis
- due to *Cryptosporidium parvum* subtype IIdA19G1 in neonatal calves on a dairy farm
- in China. Int J Parasitol 49:569-577. https://doi.org/10.1016/j.ijpara.2019.02.006
- 424 PMID: 31071320
- Lee SH, VanBik D, Kim HY, Lee YR, Kim JW, Chae M, et al. (2016) Multilocus typing
- of Cryptosporidium spp. in young calves with diarrhea in Korea. Vet Parasitol 229:81-
- 427 89. https://doi.org/10.1016/j.vetpar.2016.09.019 PMID: 27809984
- 428 25. Kabir MHB, Itoh M, Shehata AA, Bando H, Fukuda Y, Murakoshi F, et al. (2020)
- Distribution of *Cryptosporidium* species isolated from diarrhoeic calves in Japan.
- 430 Parasitol Int 78:102153. https://doi.org/10.1016/j.parint.2020.102153 PMID:
- 431 32504804
- 432 26. Cai M, Guo Y, Pan B, Li N, Wang X, Tang C, et al. (2017) Longitudinal monitoring of
- 433 *Cryptosporidium* species in pre-weaned dairy calves on five farms in Shanghai, China.

- 434 Vet Parasitol 241:14-19. https://doi.org/10.1016/j.vetpar.2017.05.005 PMID:
- 435 28579024
- 436 27. Cheun HI, Choi TK, Chung GT, Cho SH, Lee YH, Kimata I, et al. (2007) Genotypic
- characterization of *cryptosporidium* oocysts isolated from healthy people in three
- different counties of Korea. J Vet Med Sci 69:1099-1101.
- 439 https://doi.org/10.1292/jvms.69.1099 PMID: 17984603
- 440 28. Sulaiman IM, Hira PR, Zhou L, Al-Ali FM, Al-Shelahi FA, Shweiki HM, et al. (2005)
- 441 Unique endemicity of cryptosporidiosis in children in Kuwait. J Clin Microbiol
- 43:2805-2809. https://doi.org/10.1128/JCM.43.6.2805-2809.2005 PMID: 15956401
- 443 29. Meganck V, Hoflack G, Piepers S, Opsomer G (2015) Evaluation of a protocol to
- reduce the incidence of neonatal calf diarrhoea on dairy herds. Prev Vet Med 118:64-
- 70. https://doi.org/10.1016/j.prevetmed.2014.11.007 PMID: 25475689
- 446 30. Lee SH, Kim HY, Choi EW, Kim D (2019) Causative agents and epidemiology of
- diarrhea in Korean native calves. J Vet Sci 20:e64.
- https://doi.org/10.4142/jvs.2019.20.e64 PMID: 31775191
- 449 31. Avendano C, Ramo A, Vergara-Castiblanco C, Sanchez-Acedo C, Quilez J (2018)
- Genetic uniqueness of *Cryptosporidium parvum* from dairy calves in Colombia.
- 451 Parasitol Res 117:1317-1323. https://doi.org/10.1007/s00436-018-5818-6 PMID:
- 452 29484550
- 453 32. Diaz P, Varcasia A, Pipia AP, Tamponi C, Sanna G, Prieto A, et al. (2018) Molecular
- characterisation and risk factor analysis of *Cryptosporidium* spp. in calves from Italy.
- 455 Parasitol Res 117:3081-3090. https://doi.org/10.1007/s00436-018-6000-x PMID:
- 456 30008134

- 457 33. Holzhausen I, Lendner M, Gohring F, Steinhofel I, Daugschies A (2019) Distribution
- of *Cryptosporidium parvum gp60* subtypes in calf herds of Saxony, Germany. Parasitol
- Res 118:1549-1558. https://doi.org/10.1007/s00436-019-06266-1 PMID: 30790038
- 460 34. Lombardelli JA, Tomazic ML, Schnittger L, Tiranti KI (2019) Prevalence of
- 461 Cryptosporidium parvum in dairy calves and gp60 subtyping of diarrheic calves in
- 462 Central Argentina. Parasitol Res 118:2079-2086. https://doi.org/10.1007/s00436-019-
- 463 06366-y PMID: 31187226
- 35. Santoro A, Dorbek-Kolin E, Jeremejeva J, Tummeleht L, Orro T, Jokelainen P, et al.
- 465 (2019) Molecular epidemiology of *Cryptosporidium* spp. in calves in Estonia: high
- prevalence of *Cryptosporidium parvum* shedding and 10 subtypes identified.
- 467 Parasitology 146:261-267. https://doi.org/10.1017/S0031182018001348 PMID:
- 468 30086806
- 469 36. Wu Y, Zhang K, Zhang Y, Jing B, Chen Y, Xu C, et al. (2020) Genetic diversity of
- 470 *Cryptosporidium parvum* in neonatal dairy calves in Xinjiang, China. Pathogens 9:
- 471 692. https://doi.org/10.3390/pathogens9090692 PMID: 32842484
- 472 37. Kaupke A, Rzezutka A (2015) Emergence of novel subtypes of *Cryptosporidium*
- parvum in calves in Poland. Parasitol Res 114:4709-4716.
- 474 https://doi.org/10.1007/s00436-015-4719-1 PMID: 26358098
- 475 38. Qi M, Zhang K, Huang M, Wang S, Xu C, Wang T, et al. (2020) Longitudinal detection
- of *Cryptosporidium* spp. in 1-10-week-old dairy calves on a farm in Xinjiang, China.
- 477 Parasitol Res 119:3839-3844. https://doi.org/10.1007/s00436-020-06904-z PMID:
- 478 32996049
- 39. Santin M, Trout JM, Xiao L, Zhou L, Greiner E, Fayer R (2004) Prevalence and age-

- related variation of *Cryptosporidium* species and genotypes in dairy calves. Vet
- Parasitol 122:103-117. https://doi.org/10.1016/j.vetpar.2004.03.020 PMID: 15177715
- 482 40. Santin M (2013) Clinical and subclinical infections with *Cryptosporidium* in animals.
- 483 N Z Vet J 61:1-10. https://doi.org/10.1080/00480169.2012.731681 PMID: 23134088
- 484 41. Feng Y, Xiao L (2017) Molecular epidemiology of cryptosporidiosis in China. Front
- 485 Microbiol 8:1701. https://doi.org/10.3389/fmicb.2017.01701 PMID: 28932217
- 486 42. Tao W, Li Y, Yang H, Song M, Lu Y, Li W (2018) Widespread occurrence of zoonotic
- 487 Cryptosporidium species and subtypes in dairy cattle from Northeast China: public
- health concerns. J Parasitol 104:10-17. https://doi.org/10.1645/17-140 PMID:
- 489 29088547
- 490 43. Wang R, Zhao G, Gong Y, Zhang L (2017) Advances and perspectives on the
- 491 epidemiology of bovine *Cryptosporidium* in China in the past 30 years. Front
- 492 Microbiol 8:1823. https://doi.org/10.3389/fmicb.2017.01823 PMID: 28979256
- 493 44. Bjorkman C, Lindstrom L, Oweson C, Ahola H, Troell K, Axen C (2015)
- 494 *Cryptosporidium* infections in suckler herd beef calves. Parasitology142:1108-1114.
- 495 https://doi.org/10.1017/S0031182015000426 PMID: 2589955
- 496 45. Feng Y, Ortega Y, He G, Das P, Xu M, Zhang X, et al. (2007) Wide geographic
- distribution of *Cryptosporidium bovis* and the deer-like genotype in bovines. Vet
- 498 Parasitol 144:1-9. https://doi.org/10.1016/j.vetpar.2006.10.001 PMID: 17097231
- 499 46. Seppa-Lassila L, Orro T, Lassen B, Lasonen R, Autio T, Pelkonen S, et al. (2015)
- Intestinal pathogens, diarrhoea and acute phase proteins in naturally infected dairy
- 501 calves. Comp Immunol Microbiol Infect Dis 41:10-16.
- 502 https://doi.org/10.1016/j.cimid.2015.05.004 PMID: 26264522

- 503 47. Rieux A, Paraud C, Pors I, Chartier C (2013) Molecular characterization of
- 504 *Cryposporidium* isolates from pre-weaned calves in Western France in relation to age.
- Vet Parasitol 197:7-12. https://doi.org/10.1016/j.vetpar.2013.05.001 PMID: 23735428
- 506 48. Xu Z, Li N, Guo Y, Feng Y, Xiao L (2020) Comparative genomic analysis of three
- intestinal species reveals reductions in secreted pathogenesis determinants in bovine-
- specific and non-pathogenic *Cryptosporidium* species. Microb Genom 6:e000379.
- 509 https://doi.org/ 10.1099/mgen.0.000379 PMID: 32416746
- 510 49. Feng Y, Torres E, Li N, Wang L, Bowman D, Xiao L (2013) Population genetic
- 511 characterisation of dominant *Cryptosporidium parvum* subtype IIaA15G2R1. Int J
- Parasitol 43:1141-1147. https://doi.org/10.1016/j.ijpara.2013.09.002 PMID: 24126186
- 513 50. Ichikawa-Seki M, Aita J, Masatani T, Suzuki M, Nitta Y, Tamayose G, et al. (2015)
- Molecular characterization of *Cryptosporidium parvum* from two different Japanese
- prefectures, Okinawa and Hokkaido. Parasitol Int 64:161-166. https://doi.org/
- 516 10.1016/j.parint.2014.11.007 PMID: 25481361
- 517 51. Ramo A, Quilez J, Vergara-Castiblanco C, Monteagudo L, Del Cacho E, Clavel A
- 518 (2015) Multilocus typing and population structure of *Cryptosporidium* from children
- 519 in Zaragoza, Spain. Infect Genet Evol 31:190-197.
- 520 https://doi.org/10.1016/j.meegid.2015.01.023 PMID: 25660036
- 521 52. Soba B, Logar J (2008) Genetic classification of Cryptosporidium isolates from
- humans and calves in Slovenia. Parasitology 135:1263-1270. https://doi.org/
- 523 10.1017/S0031182008004800 PMID: 18664309
- 524 53. Valenzuela O, Gonzalez-Diaz M, Garibay-Escobar A, Burgara-Estrella A, Cano M,
- Durazo M, et al. (2014) Molecular characterization of *Cryptosporidium* spp. in

https://doi.org/10.1371/journal.pone.0096128 PMID: 24755606 527 54. Trotz-Williams LA, Martin DS, Gatei W, Cama V, Peregrine AS, Martin SW, et al. 528 (2006) Genotype and subtype analyses of *Cryptosporidium* isolates from dairy calves 529 and humans in Ontario. Parasitol Res 99:346-352. https://doi.org/10.1007/s00436-006-530 0157-4 PMID: 16565813 531 Al Mawly J, Grinberg A, Velathanthiri N, French N (2015) Cross sectional study of 532 55. prevalence, genetic diversity and zoonotic potential of Cryptosporidium parvum 533 cycling in New Zealand dairy farms. **Parasit** Vectors 8:240. 534 https://doi.org/10.1186/s13071-015-0855-9 PMID: 25896433 535 536 56. Grinberg A, Learmonth J, Kwan E, Pomroy W, Lopez Villalobos N, Gibson I, et al. (2008) Genetic diversity and zoonotic potential of Cryptosporidium parvum causing 537 foal diarrhea. J Clin Microbiol 46:2396-2398. https://doi.org/10.1128/JCM.00936-08 538 PMID: 18508944 539 57. Inacio SV, Widmer G, de Brito RL, Zucatto AS, de Aquino MC, Oliveira BC, et al. 540 (2017) First description of Cryptosporidium hominis gp60 genotype IkA20G1 and 541 Cryptosporidium parvum gp60 genotypes IIaA18G3R1 and IIaA15G2R1 in foals in 542 Brazil. Vet Parasitol 233:48-51. *gp60* https://doi.org/10.1016/j.vetpar.2016.11.021 543 544 PMID: 28043388 Ng JS, Pingault N, Gibbs R, Koehler A, Ryan U (2010) Molecular characterisation of 545 58. Cryptosporidium outbreaks in Western and South Australia. Exp Parasitol 125:325-546 547 328. https://doi.org/10.1016/j.exppara.2010.02.012 PMID: 20219461

Mexico.

**PLoS** 

One

9(4):e96128.

526

548

59.

children

from

Waldron LS, Power ML (2011) Fluorescence analysis detects gp60 subtype diversity

- in Cryptosporidium infections. Infect Genet Evol 11:1388-1395. https://doi.org/
- 550 10.1016/j.meegid.2011.05.008 PMID: 21609784
- 551 60. Zintl A, Proctor AF, Read C, Dewaal T, Shanaghy N, Fanning S, et al. (2009) The
- prevalence of *Cryptosporidium* species and subtypes in human faecal samples in
- 553 Ireland. Epidemiol Infect 137:270-277. https://doi.org/10.1017/S0950268808000769
- 554 PMID: 18474128
- 555 61. Del Chierico F, Onori M, Di Bella S, Bordi E, Petrosillo N, Menichella D, et al. (2011)
- Cases of cryptosporidiosis co-infections in AIDS patients: a correlation between
- clinical presentation and *gp60* subgenotype lineages from aged formalin-fixed stool
- samples. Ann Trop Med Parasitol 105:339-349. https://doi.org/
- 559 10.1179/1364859411Y.0000000025 PMID: 21929875
- 560 62. Glaberman S, Moore JE, Lowery CJ, Chalmers RM, Sulaiman I, Elwin K, et al. (2002)
- Three drinking-water-associated cryptosporidiosis outbreaks, Northern Ireland. Emerg
- Infect Dis 8:631-633. https://doi.org/10.3201/eid0806.010368 PMID: 12023922
- 563 63. Mammeri M, Chevillot A, Chenafi I, Thomas M, Julien C, Vallee I, et al. (2019)
- Molecular characterization of *Cryptosporidium* isolates from diarrheal dairy calves in
- France. Vet Parasitol Reg Stud Reports 18:100323.
- 566 https://doi.org/10.1016/j.vprsr.2019.100323 PMID: 31796198
- 567 64. Thompson HP, Dooley JS, Kenny J, McCoy M, Lowery CJ, Moore JE, et al. (2007)
- Genotypes and subtypes of *Cryptosporidium* spp. in neonatal calves in Northern
- Ireland. Parasitol Res 100:619-624. https://doi.org/10.1007/s00436-006-0305-x PMID:
- 570 17031699
- 571 65. Waldron LS, Ferrari BC, Power ML (2009) Glycoprotein 60 diversity in C. hominis

- and C. parvum causing human cryptosporidiosis in NSW, Australia. Exp Parasitol
- 573 122:124-127. https://doi.org/10.1016/j.exppara.2009.02.006 PMID: 19233175
- 574 66. Koinari M, Karl S, Ng-Hublin J, Lymbery AJ, Ryan UM (2013) Identification of novel
- and zoonotic *Cryptosporidium* species in fish from Papua New Guinea. Vet Parasitol
- 576 198:1-9. https://doi.org/10.1016/j.vetpar.2013.08.031 PMID: 24064001
- 577 67. Koinari M, Lymbery AJ, Ryan UM (2014) *Cryptosporidium* species in sheep and goats
- from Papua New Guinea. Exp Parasitol 141:134-137.
- 579 https://doi.org/10.1016/j.exppara.2014.03.021 PMID: 24703974
- 580 68. Shrestha RD, Grinberg A, Dukkipati VS, Pleydell EJ, Prattley DJ, French NP (2014)
- Infections with multiple *Cryptosporidium* species and new genetic variants in young
- dairy calves on a farm located within a drinking water catchment area in New Zealand.
- Vet Parasitol 202:287-291. https://doi.org/10.1016/j.vetpar.2014.03.034 PMID:
- 584 24780161
- 585 69. Giangaspero A, Papini R, Marangi M, Koehler AV, Gasser RB (2014)
- 586 Cryptosporidium parvum genotype IIa and Giardia duodenalis assemblage A in
- 587 Mytilus galloprovincialis on sale at local food markets. Int J Food Microbiol 171:62-
- 588 67. https://doi.org/10.1016/j.ijfoodmicro.2013.11.022 PMID: 24334090
- 589 70. Waldron LS, Dimeski B, Beggs PJ, Ferrari BC, Power ML (2011) Molecular
- epidemiology, spatiotemporal analysis, and ecology of sporadic human
- cryptosporidiosis in Australia. Appl Environ Microbiol 77:7757-7765. https://doi.org/
- 592 10.1128/AEM.00615-11 PMID: 21908628
- 593 71. Deshpande AP, Jones BL, Connelly L, Pollock KG, Brownlie S, Alexander CL (2015)
- Molecular characterization of *Cryptosporidium parvum* isolates from human

- cryptosporidiosis cases in Scotland. Parasitology 142:318-325. https://doi.org/
- 596 10.1017/S0031182014001346 PMID: 25244937
- 597 72. Osman M, Benamrouz S, Guyot K, El Safadi D, Mallat H, Dabboussi F, et al. (2018)
- Molecular epidemiology of *Cryptosporidium* spp. in North Lebanon. J Infect Dev
- 599 Ctries 12:34S. https://doi.org/10.3855/jidc.10014 PMID: 31805009
- 600 73. Mahfouz ME, Mira N, Amer S (2014) Prevalence and genotyping of *Cryptosporidium*
- spp. in farm animals in Egypt. J Vet Med Sci 76:1569-1575.
- 602 https://doi.org/10.1292/jvms.14-0272 PMID: 25649937
- 603 74. Mi R, Wang X, Huang Y, Zhou P, Liu Y, Chen Y, et al. (2014) Prevalence and molecular
- characterization of *Cryptosporidium* in goats across four provincial level areas in
- 605 China. PLoS One 9:e111164. https://doi.org/10.1371/journal.pone.0111164 PMID:
- 606 25343501
- 75. Naguib D, El-Gohary AH, Mohamed AA, Roellig DM, Arafat N, Xiao L (2018) Age
- patterns of *Cryptosporidium* species and *Giardia duodenalis* in dairy calves in Egypt.
- Parasitol Int 67:736-741. https://doi.org/10.1016/j.parint.2018.07.012 PMID:
- 610 30055334
- 611 76. Mercado R, Pena S, Ozaki LS, Fredes F, Godoy J (2015) Multiple Cryptosporidium
- 612 parvum subtypes detected in a unique isolate of a Chilean neonatal calf with diarrhea.
- Parasitol Res 114:1985-19888. https://doi.org/10.1007/s00436-015-4364-8 PMID:
- 614 25673079
- 615 77. Al-Habsi K, Yang R, Williams A, Miller D, Ryan U, Jacobson C (2017) Zoonotic
- 616 Cryptosporidium and Giardia shedding by captured rangeland goats. Vet Parasitol Reg
- Stud Reports 7:32-35. https://doi.org/10.1016/j.vprsr.2016.11.006 PMID: 31014653

618 78. Chalmers RM, Robinson G, Elwin K, Elson R (2019) Analysis of the *Cryptosporidium* spp. and gp60 subtypes linked to human outbreaks of cryptosporidiosis in England and 619 620 Wales, 2009 to 2017. Parasit Vectors 12:95. https://doi.org/10.1186/s13071-019-3354-6 PMID: 30867023 621 Connelly L, Craig BH, Jones B, Alexander CL (2013) Genetic diversity of 622 79. Cryptosporidium spp. within a remote population of soay sheep on St. Kilda Islands, 623 Scotland. Environ Microbiol 79:2240-2246. 624 Appl 625 https://doi.org/10.1128/AEM.02823-12 PMID: 23354707 Del Coco VF, Cordoba MA, Bilbao G, de Almeida Castro AP, Basualdo JA, Fayer R, 80. 626 et al. (2014) Cryptosporidium parvum gp60 subtypes in dairy cattle from Buenos Aires, 627 628 Argentina. Res Vet Sci 96:311-314. https://doi.org/10.1016/j.rvsc.2013.12.010 PMID: 24480390 629 81. Abeywardena H, Jex AR, Nolan MJ, Haydon SR, Stevens MA, McAnulty RW, et al. 630 (2012) Genetic characterisation of Cryptosporidium and Giardia from dairy calves: 631 discovery of species/genotypes consistent with those found in humans. Infect Genet 632 633 Evol 12:1984-1993. https://doi.org/10.1016/j.meegid.2012.08.004 PMID: 22981927 82. Nolan MJ, Jex AR, Koehler AV, Haydon SR, Stevens MA, Gasser RB (2013) 634 Molecular-based investigation of Cryptosporidium and Giardia from animals in water 635 catchments in Southeastern Australia. Water Res 47:1726-1740. https://doi.org/ 636 10.1016/j.watres.2012.12.027 PMID: 23357792 637 O'Brien E, McInnes L, Ryan U (2008) Cryptosporidium gp60 genotypes from humans 638 83. 639 and domesticated animals in Australia, North America and Europe. Exp Parasitol

118:118-121. https://doi.org/10.1016/j.exppara.2007.05.012 PMID: 17618622

641	84.	Quilez J, Torres E, Chalmers RM, Robinson G, Del Cacho E, Sanchez-Acedo C (2008)
642		Cryptosporidium species and subtype analysis from dairy calves in Spain. Parasitology
643		135:1613-1620. https://doi.org/10.1017/S0031182008005088 PMID: 18980704
644	85.	Mirhashemi ME, Zintl A, Grant T, Lucy F, Mulcahy G, De Waal T (2016) Molecular
645		epidemiology of Cryptosporidium species in livestock in Ireland. Vet Parasitol 216:18-
646		22. https://doi.org/10.1016/j.vetpar.2015.12.002 PMID: 26801590
647	86.	Santin M, Zarlenga DS (2009) A multiplex polymerase chain reaction assay to
648		simultaneously distinguish Cryptosporidium species of veterinary and public health
649		concern in cattle. Vet Parasitol 166:32-37. https://doi.org/10.1016/j.vetpar.2009.07.039
650		PMID: 19713046
651		

## Figure legend

**Figure 1.** Sequence comparisons between *C. bovis* and *C. ryanae* for the partial18S rRNA gene from Korean sequences obtained in this study and reference strains. Six nucleotide differences at 440, 460, 464–466, and 470 are shown. An asterisk indicates sequences obtained in this study.

Table 1. Prevalence and distribution of *Cryptosporidium* species according to age group in pre-weaned calves

Age (days)	Complesies	No. of positive (%)	95% CI	Cryptosporidium species (No.)			
	Sample size			C. parvum	C. ryanae	C. bovis	
1–10	271	53 (19.6%)	14.8–24.3	49	3	1	
11-20	92	24 (26.1%)	17.1–35.1	23	1	0	
21–30	39	3 (7.7%)	0.0-16.1	0	3	0	
31-60	58	6 (10.3%)	2.5-18.2	0	5	1	
Total	460	86 (18.7%)	15.1-22.3	72	12	2	

Table 2. Distribution of *Cryptosporidium* species in pre-weaned Korean native calves according to age group

Age (days)	Frequency of <i>C. parvum</i> positivity (%)	$\chi^2$ ( <i>P</i> -value)	Frequency of <i>C.</i> ryanae positivity (%)	$\chi^2$ ( <i>P</i> -value)	Frequency of <i>C. bovis</i> positivity (%)	$\chi^2(P$ -value)
1–10	49/271 (18.1%)		3/271 (1.1%)		1/271 (0.4%)	
11-20	23/92 (25.0%)	25 200 (0.000)	1/92 (1.1%)	16 020 (0 001)	0	2.924 (0.410)
21-30	0	25.300 (0.000)	3/39 (7.7%)	16.020 (0.001)	0	2.824 (0.419)
31-60 (Ref.)	0		5/58 (8.6%)		1/58 (1.7%)	

1 Table 3. Distribution of *Cryptosporidium parvum* subtype according to age group

an60 subtymas —	Age gro	oups (days)	No. of
gp60 subtypes –	1-10	11-20	positive calves
IIaA14G1R1	1	0	1 (1.4%)
IIaA14G3R1	1	0	1 (1.4%)
IIaA15G1R1	1	0	1 (1.4%)
IIaA15G2R1	3	0	3 (4.2%)
IIaA16G4R1	1	1	2 (2.8%)
IIaA17G3R1	1	3	4 (5.6%)
IIaA17G4R1	1	1	2 (2.8%)
IIaA18G3R1	36	16	52 (72.2%)
IIaA19G1R1	1	0	1 (1.4%)
IIaA19G3R1	0	2	2 (2.8%)
IIaA19G4R1	3	0	3 (4.2%)
Total	46	26	72

				30 440		450		460	465	470
+ 10725200		hands 1								
* MZ736398	2.7		0.0000000		10000	TTCTGTTAAT				
* MZ736399	0.000	SECTION OF STREET	200000000000000000000000000000000000000							
KX342025	-7.63	27171277								
FJ796284										
MN540745										
MN696246				A						
MT703861										
MT835227								1		
MW767057	00707	TO THE PARTY OF								
MN918118				******					******	1757.71
MZ027077										
MT043859			and the same of th						*****	* * *
MT150693			CONTROL OF THE PARTY OF THE PAR							
MT611087		20/20/20/20 00/20		****					Maria San	120,007
			Bangladesh							
MW788440										
* MZ736386		Contract to the contract of th								
* MZ736387						G				
* MZ736388										
* MZ736389		and the second s				GA				
* MZ736390				.A A.		GA				
* MZ736391										
* MZ736392				******						1000
* MZ736393						A				
* MZ736394				1007111						
* MZ736395		11 2 4 12 5 7 5 5 1 1 1 1 1								
* MZ736396										
* MZ736397		10 100 CO				A				
JN400880										
MH458438										
MK501765										
MN540747										
		the second secon	Bangladesh							
			Bangladesh		·T			1000		
			Bangladesh		- T					
MH754181				1-1-1-1-1-1-1	- T		******		GCT.	
MT002726										
KP793013										
MT611097		A CONTRACTOR OF THE STATE OF TH								
			Ethiopia							
		1 A 1981 HOLE & A 2011 SA 2011	Thailand							
AB777177	C.	ryanae	Egypt		·T			.C	GCT.	G