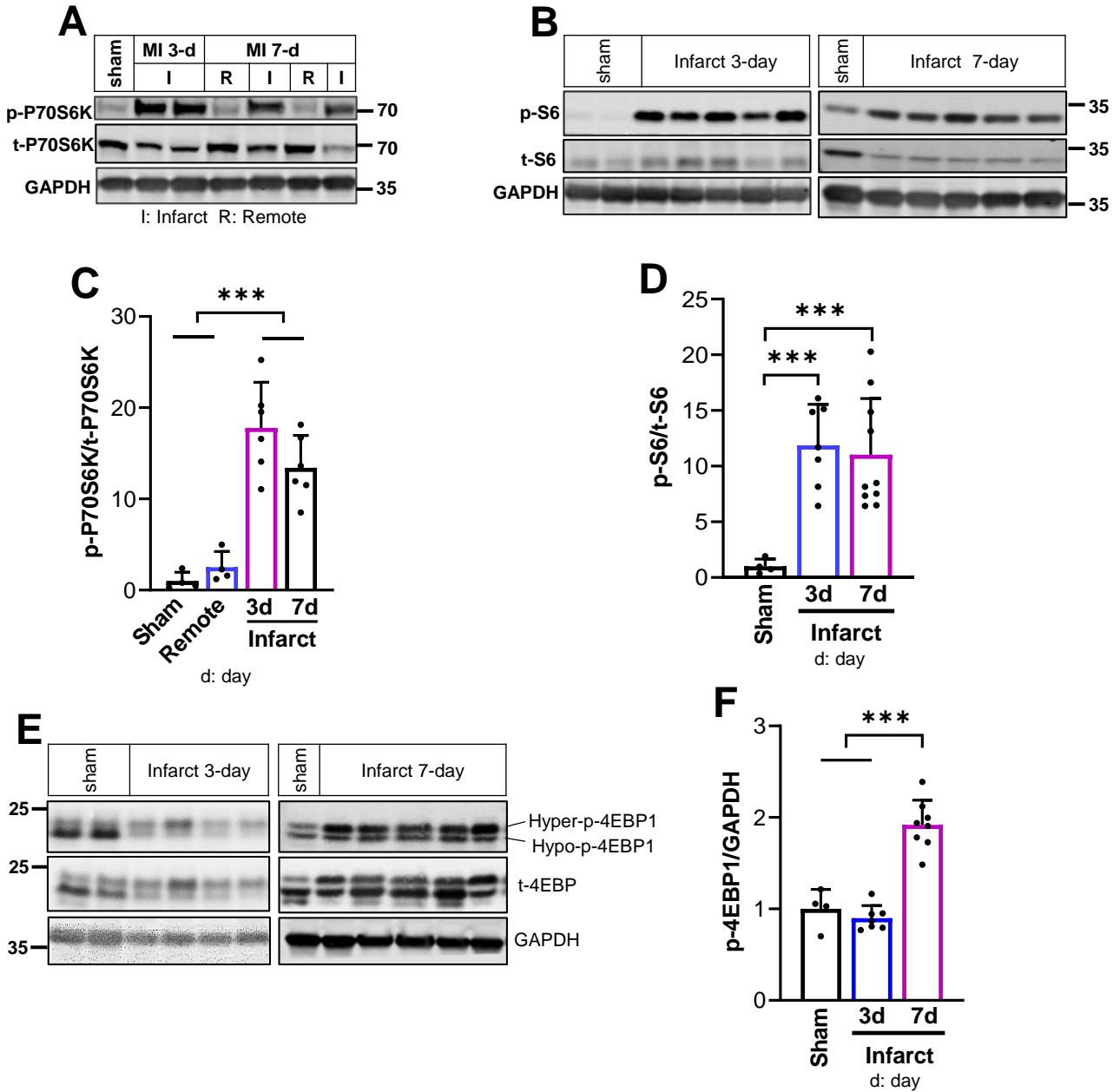
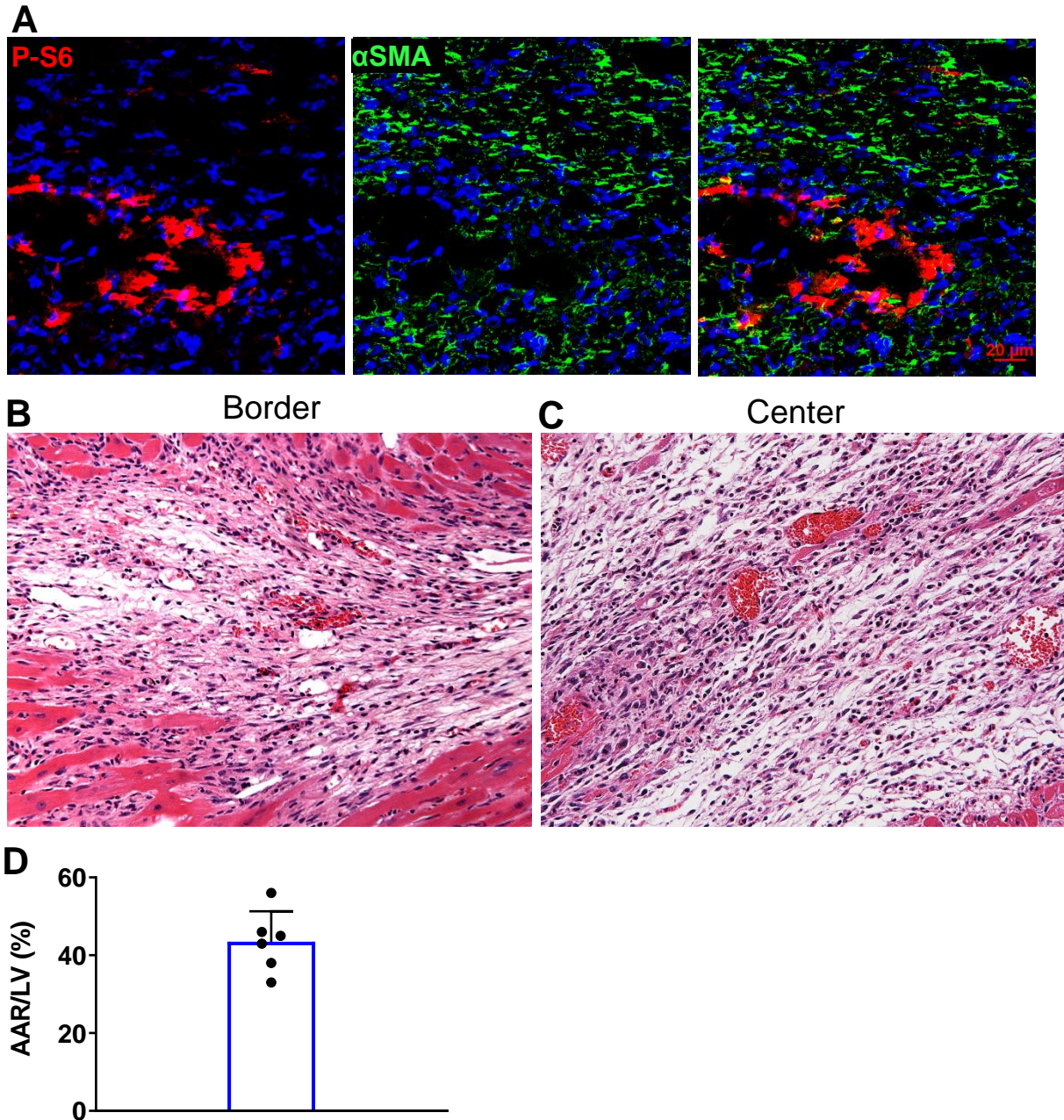


S-Figure 1. Robust activation of mTORC1 in the ischemic myocardium



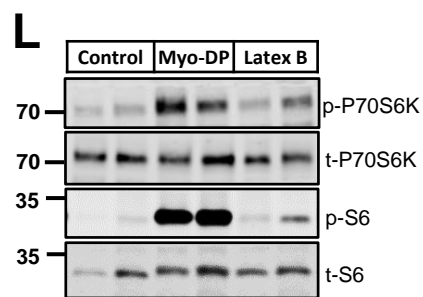
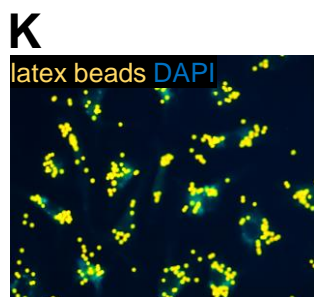
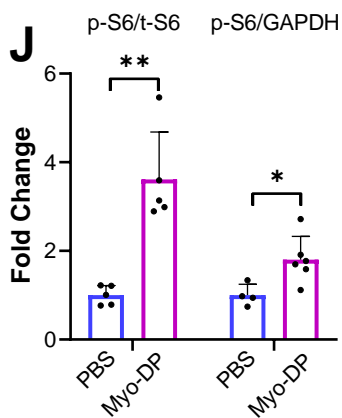
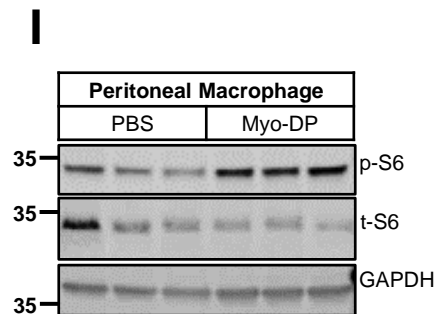
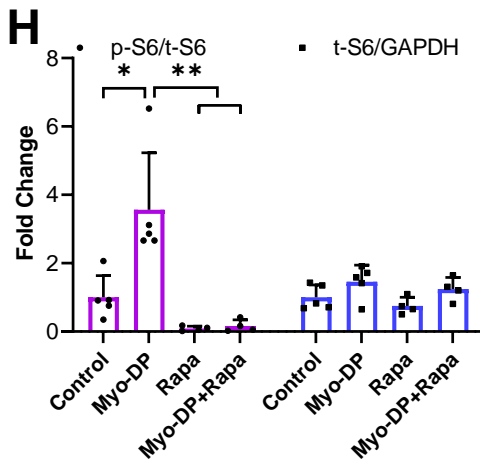
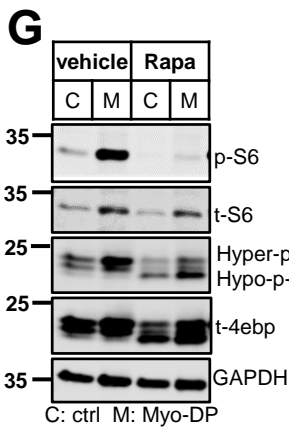
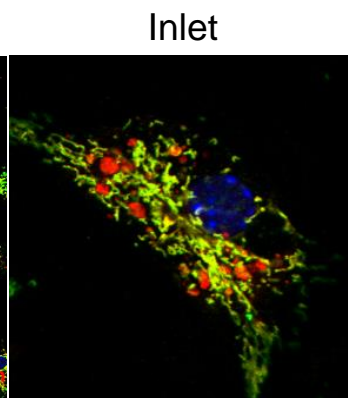
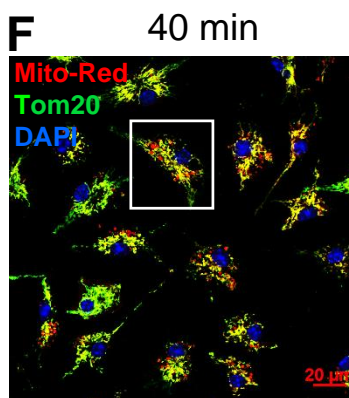
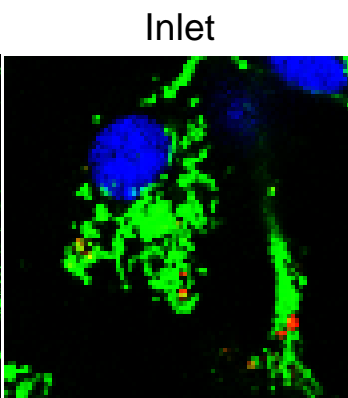
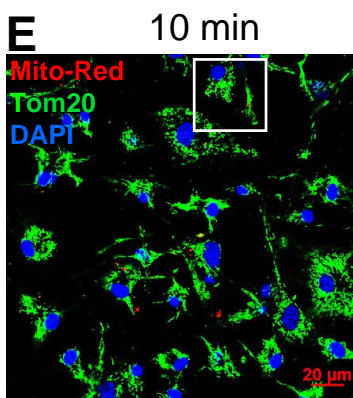
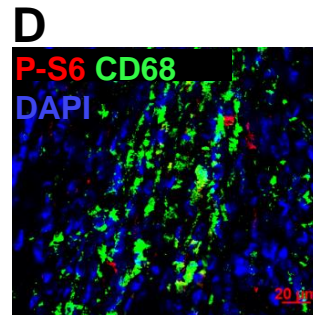
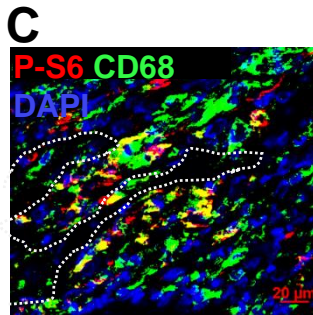
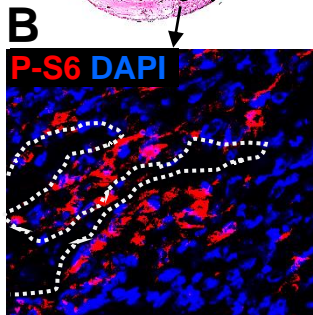
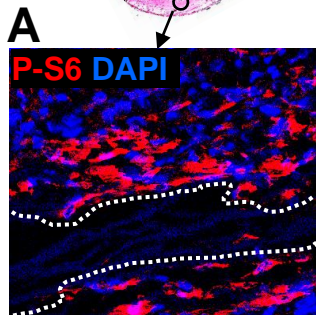
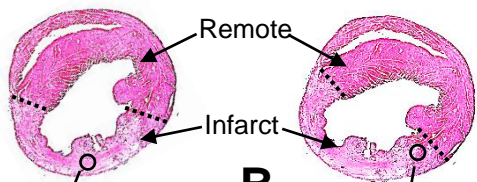
S-Figure 1. C57BL/6J mice were subjected to sham procedure or LAD ligation to generate myocardial infarction. Heart tissue was separated into remote and infarct region under a dissection scope. Immunoblot analyses were performed using samples from cohorts as indicated. Individual lane on all immunoblots represents different experimental animals. **A.** Immunoblot analysis of p-P70S6K, t-P70S6K, and GAPDH from samples obtained 3-day or 7-day after MI. **B.** Immunoblot analysis of p-S6, t-S6, and GAPDH using samples from sham or infarct tissues at the time points as indicated. **C.** Summarized data from different experiments performed as in A. n=4 to 6 hearts. **D.** Summarized data of different experiments performed as in B. n= 4 to 10 hearts. **E.** Protein samples obtained from sham or infarct tissue at different time points were analyzed for p-4EBP1. The hyper- and hypo-phosphorylated 4EBP1 were indicated. **F.** Quantification of the p-4EBP1 (combined both hyper and hypo-p-4EBP1) after normalization to GAPDH. n= 4 to 8 hearts. *** p<0.001

S-Figure 2. Lack of mTORC1 activation in myofibroblasts



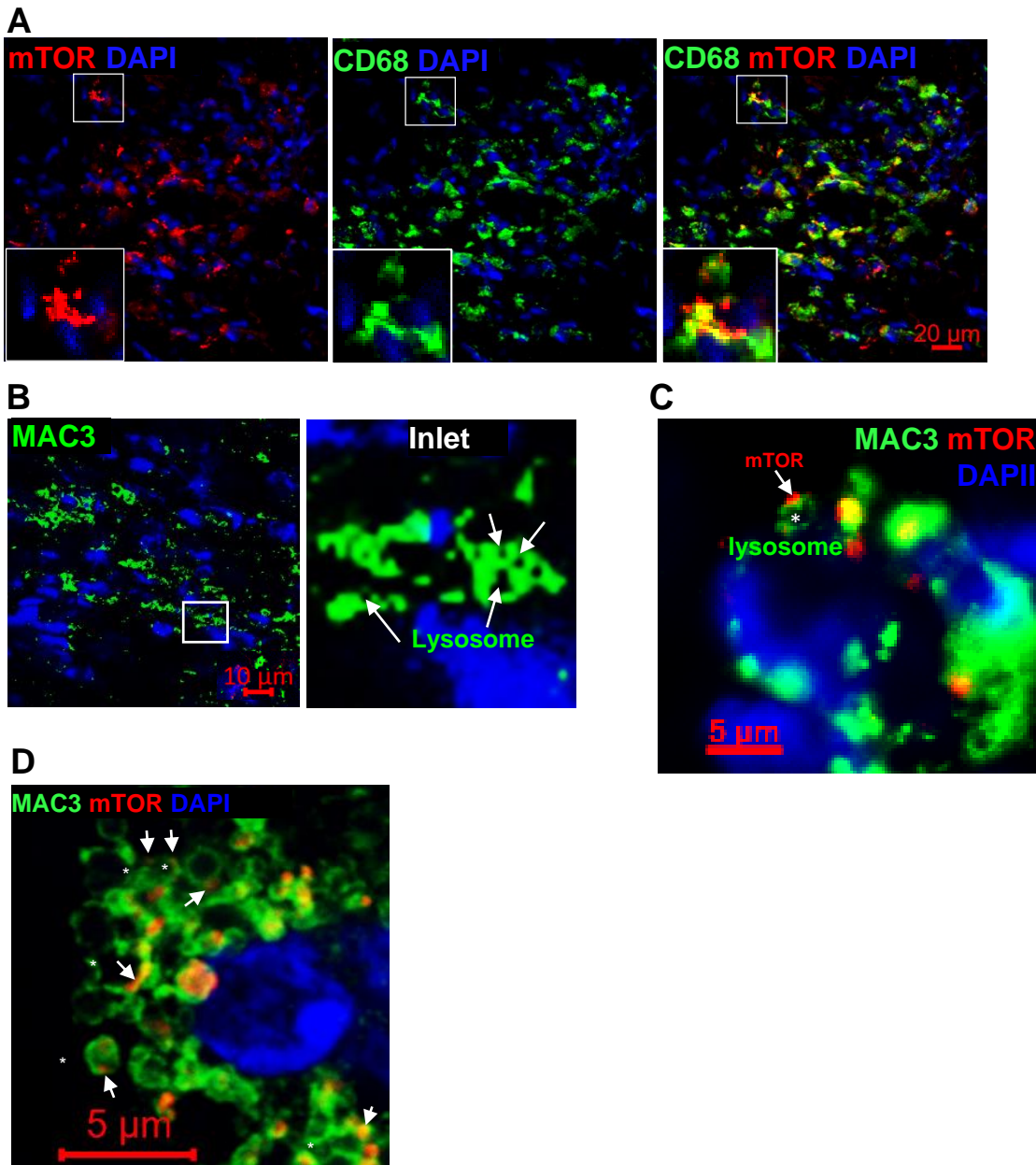
S-Figure 2. **A.** Myocardial infarction was generated by LAD ligation. One week after the MI procedure, the heart was perfusion fixed and prepared for cryosection. The tissue sections were labeled with p-S6 and α SMA antibodies. **B and C.** Representative images of HE staining performed on MI hearts. Hearts were harvested 7-day after the MI procedure and fixed with 4%PFA for 48 hours at 4°C. HE staining were performed on cross-sections obtained from paraffin-embedded tissue blocks. Images were acquired from both infarct border and Infarct center. **D.** Estimation of area at risk (AAR). Following occlusion of a coronary artery, The heart was perfused with 2% Evans blue. Unstained area (AAR) to total LV area were quantified using ImageJ. n=6 hearts.

S-Figure 3. Digesting tissue debris activates mTORC1



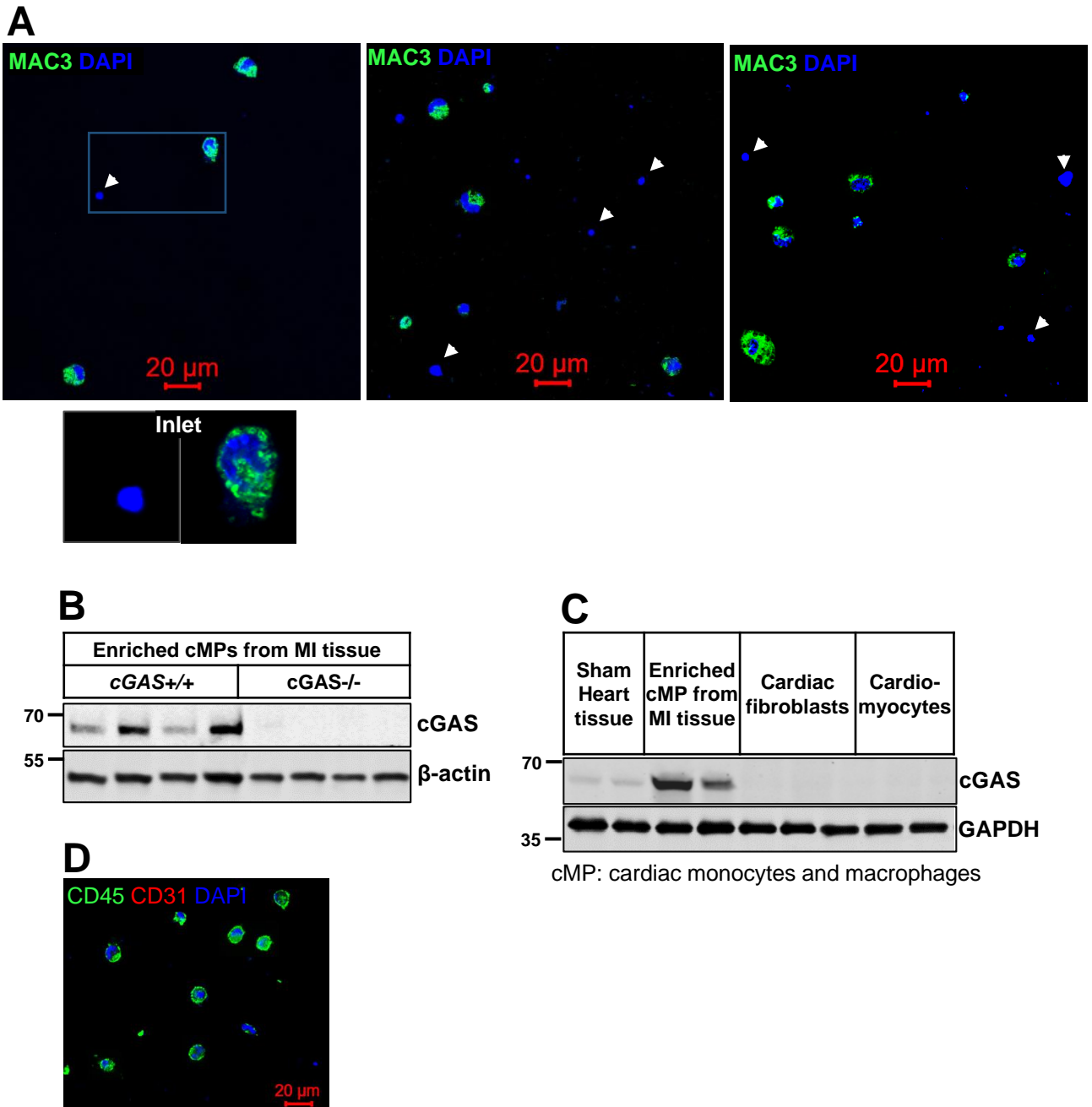
S-Figure 3. A and B. Cryosections from hearts 7-day after LAD ligation were labeled with p-S6 and CD68 antibodies. The residue necrotic tissue section was demarcated with the dotted line. The spatial location of views obtained for **A** (infarct center) and **B** (infarct border) were indicated on the HE stained cross sections. **C.** Co-labeling of CD68 and p-S6 to confirm cMPs was the main cell type with positive p-S6. **D.** Co-labeling of CD68 and p-S6 in regions where the necrotic tissue had been replaced with provisional matrix. **E and F.** Phagocytotic assay with myocardial derived particles (Myo-DP). Myo-DPs were labeled with MitoTracker Red at 100 nM for 30 minutes. The labeled particles were washed and incubated with BMDM for the 10 minutes (**E**) and 40 minutes (**F**). Endogenous mitochondria was labeled with Tom20 (green). Engulfed particles was shown in red. **G.** Macrophages (BMDM) were treated with control or Myo-DP for 6 hours in the absence or presence of 20 nM rapamycin. The levels of p-S6, t-S6, p-4EBP and t-4EBP were detected by immunoblot. **H.** Quantified data for p-S6 level normalized to t-S6 and t-S6 level normalized to GAPDH. n=4 to 5 experiments. **I.** Peritoneal macrophages were treated with PBS or Myo-DP for 15 hours. Protein samples were subjected to immunoblot analysis for p-S6, t-S6, and GAPDH. **J.** Summarized data from different experiments performed as in **I.** n=4 to 6 experiments. **K.** Macrophages (BMDM) were incubated with latex beads for 6 hours. **L.** Protein samples prepared from macrophages treated with either beads or Myo-DP were subjected to SDS-PAGE. n=2 experiments for each group. Membranes were probed with p-P70S6, t-P70S6, p-S6, t-S6, and GAPDH. Individual lane on all immunoblots represented different experiments. * p<0.05 ** p<0.01 .

S-Figure 4. mTOR in cMPs localizes to the lysosomes membrane



S-Figure 4. **A.** Tissue sections of the heart obtained 1 week after LAD ligation were labeled with mTOR and CD68 antibodies. **B.** Cryosections of the heart were stained with a lysosomal marker MAC3. Inlet demonstrates the circular lysosomal structure. **C.** Tissue sections from MI heart 7-day after LAD ligation was co-labeled with mTOR and MAC3. The arrow indicates the mTOR on the membrane of lysosomes (indicated with *). **D.** cMP isolated from infarct tissue was labeled with MAC3 and mTOR. Arrows indicate the mTOR localized to the membrane of lysosomes (circular structure, some indicated with *). For **A** and **B**, cryosections from 3 hearts were examined. For **D**, cMPs from at least three infarcts were examined.

S-Figure 5. Characterization of cMPs enriched from infarcting myocardium



S-Figure 5. **A.** Condensed and fragmented nuclei without cytosol from multiple fields were indicated with arrows. **B.** Protein samples prepared from cells enriched with cMPs from infarct tissue of cGAS^{+/+} or cGAS^{-/-} mice were subjected to immunoblot analysis to detect cGAS. **C.** Protein samples from sham myocardium, cells enriched with cMPs from infarct tissue, mouse cardiomyocytes, and fibroblasts were probed for cGAS. **D.** Cells isolated from infarct tissue were labeled with CD45 and CD31. For **B** and **C**, each lane represents 1 heart. For **D**, the cells were pooled from MI tissue obtained from two hearts.