

## SUPPLEMENTARY INFORMATION

### **Temporal variability in quantitative human gut microbiome profiles and implications for clinical research.**

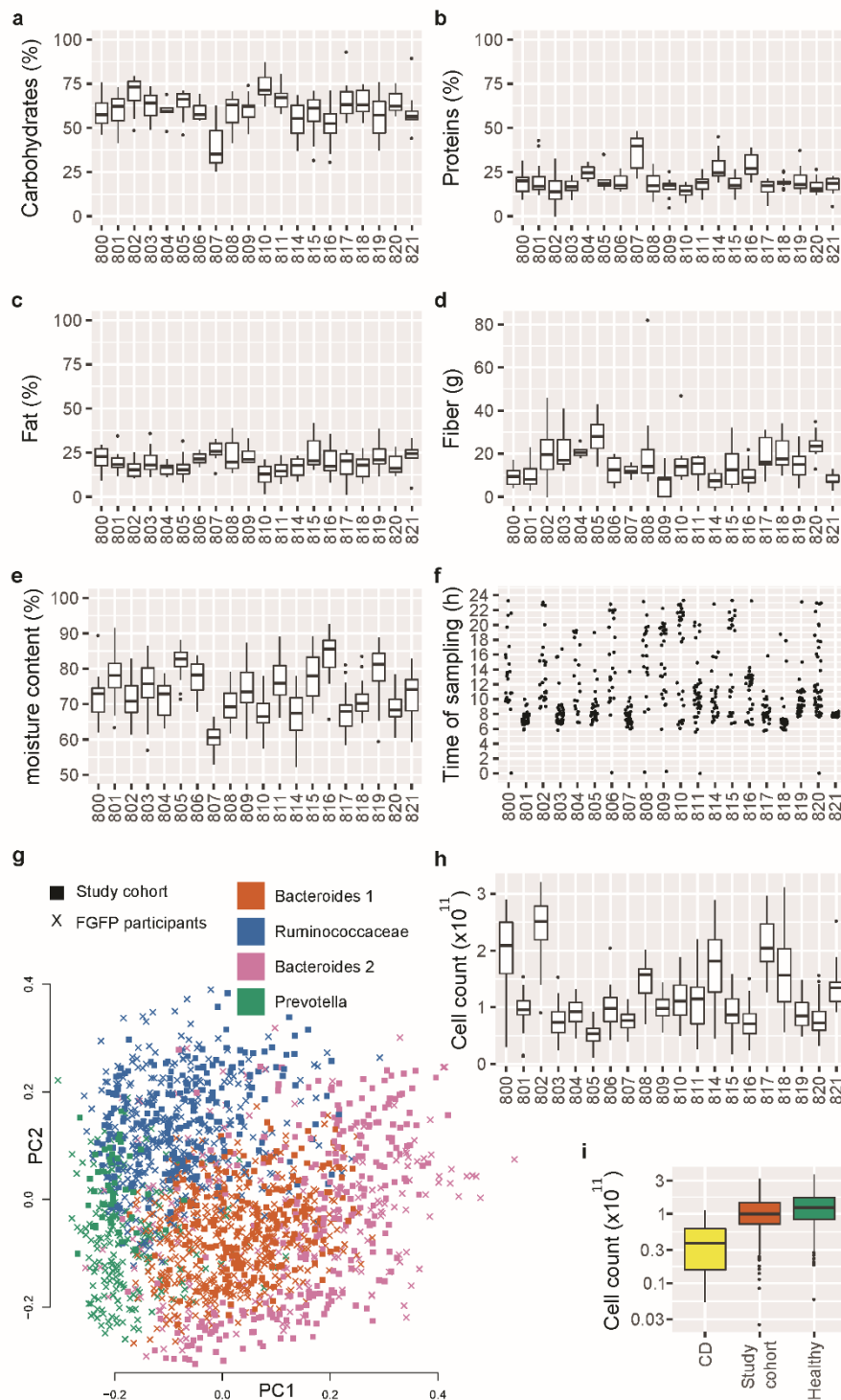
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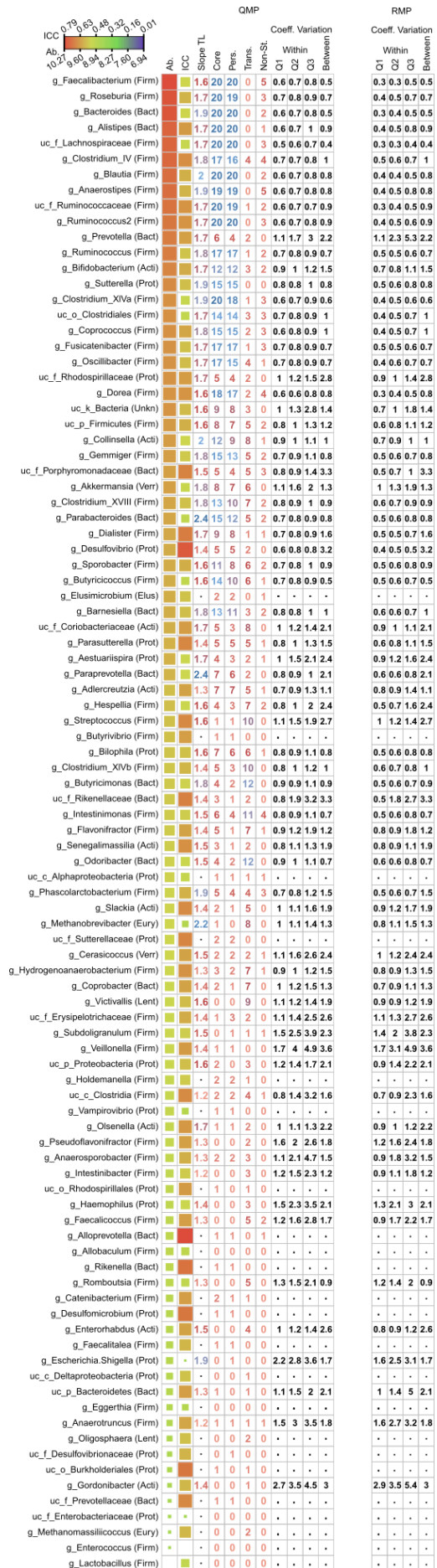
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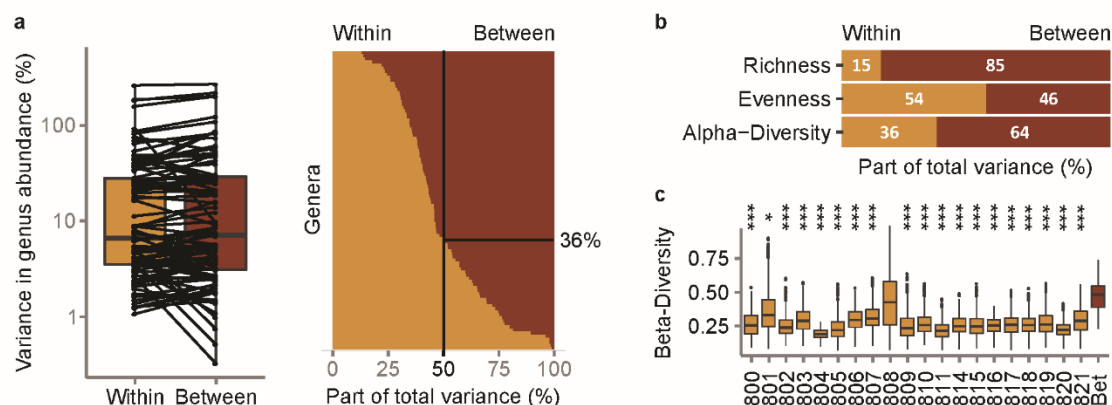
**Supplementary Figure 1: Overview of study cohort characteristics and variation in metadata.** Box plots of percentage of total weight of food products consumed per day for **a**. Carbohydrates (ICC: 0.21), **b**. Proteins (ICC: 0.40), and **c**. Fat (ICC: 0.21), for each participant. **d**. Total weight of fiber consumed per day (ICC: 0.32), for each participant. **e**. Box plots of stool moisture content (%) for each participant (ICC:0.51). **f**. Time of stool sampling according to a 24h-scale, for each participant. **g**. Principal Coordinate Analyses of RMPs of all study cohort samples (squares) and 1104 FGFP participants (crosses),

coloured according to enterotype, namely Ruminococaceae (R, blue), Bacteroides 1 (B1, orange), Bacteroides 2 (B2, Pink), Prevotella (P, green). **h.** Box plots of microbial load, in cell counts per gram of faeces, for each participant (ICC:0.61). **i.** Microbial load, in cell counts per gram of faeces, of all samples (Study Cohort) in comparison with previously sampled cohorts of Crohn's Disease patients (CD) and healthy volunteers (Healthy)<sup>11</sup>. The body of the box plots within these figures represents the first and third quartiles of the distribution, and the median line. The whiskers extend from the quartiles to the last data point within 1.5× IQR, with outliers beyond. Figures and summary measures are based on all data (N=20, n=713) (Supplementary Table S1 & S2).



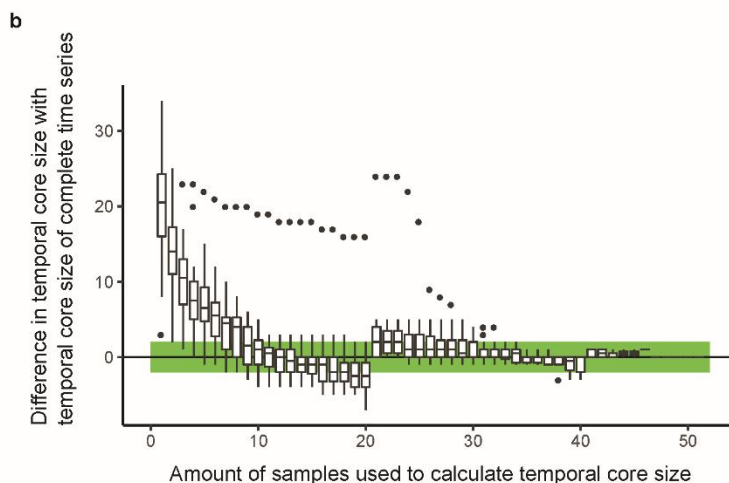
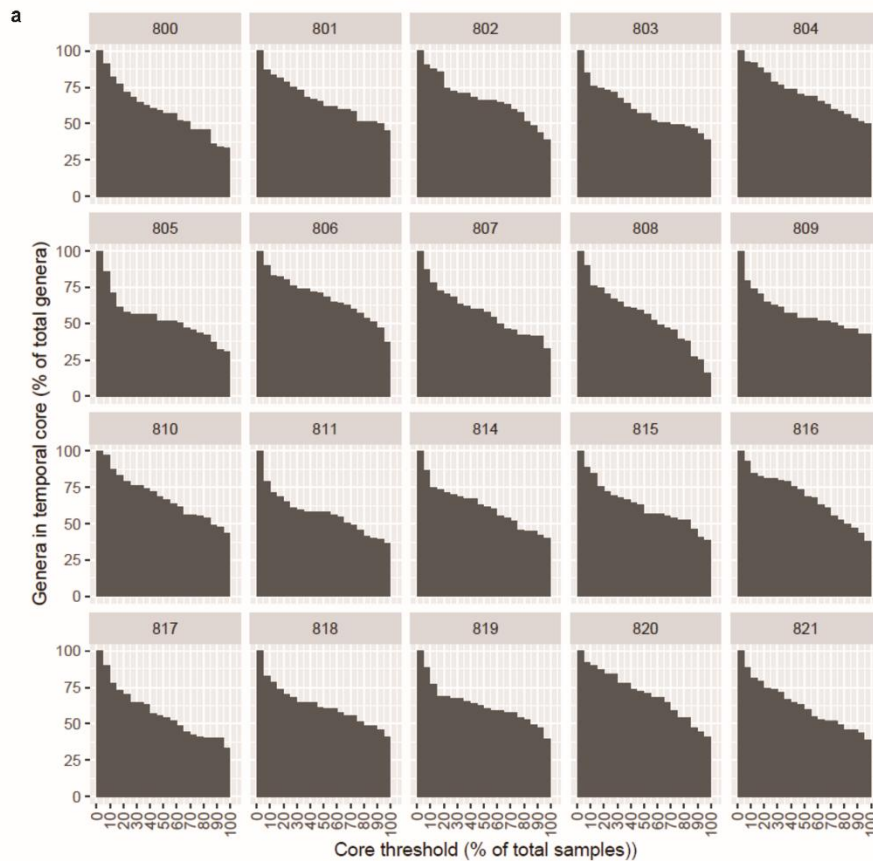
### Supplementary Figure 2. Variation and core measurements for all genera.

For all genera above the abundance threshold: (Ab.) Log mean abundance of the genus over all samples in which it was present; (ICC) Intraclass correlation coefficient of the genus; (Slope TL) Slope of Taylor's law for the genus calculated over all individuals ( $1.6 \pm 0.25$  [1.2-2.4]); (Core) The number of participants for which the genus belongs to the core, defined as present in more than 95% of the samples; (Pers.) The number of participants for which the genus can be considered persistent, defined as present in more than 20% of the samples with at least 90% of these observations being consecutive; (Trans.) The number of participants for which the genus can be considered transient, defined as present in more than 60% of the samples while less than 75% of these observations are consecutive; (Non-St.) The number of participants in which the genus shows non-stationary dynamics, defined as not satisfying a unit root process in which abundances vary around an equilibrium level (Augmented Dicky Fuller test,  $p > 0.05$ ). (Coef. Variation) Coefficient of variation in genus abundance within and between individuals. The coefficient of variation is defined as the ratio of the standard deviation to the mean and is a standardized measure of dispersion. Within-subject variation is summarized as the first, second and third quartile (Q1, Q2, Q3, respectively) of all values for the within-subject coefficient of variation, calculated for each time series (N=20). Data are based on either quantitative (QMP) or relative (RMP) microbiome profiling.



**Supplementary Figure 3. Within- compared to between-subject variance for relative abundance profiles.** **a.** Within- versus between-subject variance in relative genus abundance (Wilcoxon test, two-sided,  $n=98$ ,  $P= 0.007154$ ) (left) and within-subject and between-subject variance as part of the total variance in genus abundance (right) for all genera over the abundance threshold. **b.** Within- and between-subject variance as part of the total variance in alpha diversity measures, for observed richness, Pielou evenness, and Shannon diversity index based on RMPs. **c.** Beta-diversity, as assessed through Bray Curtis dissimilarity, between samples of the same individual (800-821), and between the first sample of each individual (Bet) based on RMPs (N=20,  $n=709$ ). Significance of the differences within and between individuals were tested through an ANOVA on multivariate homogeneity of group dispersions.

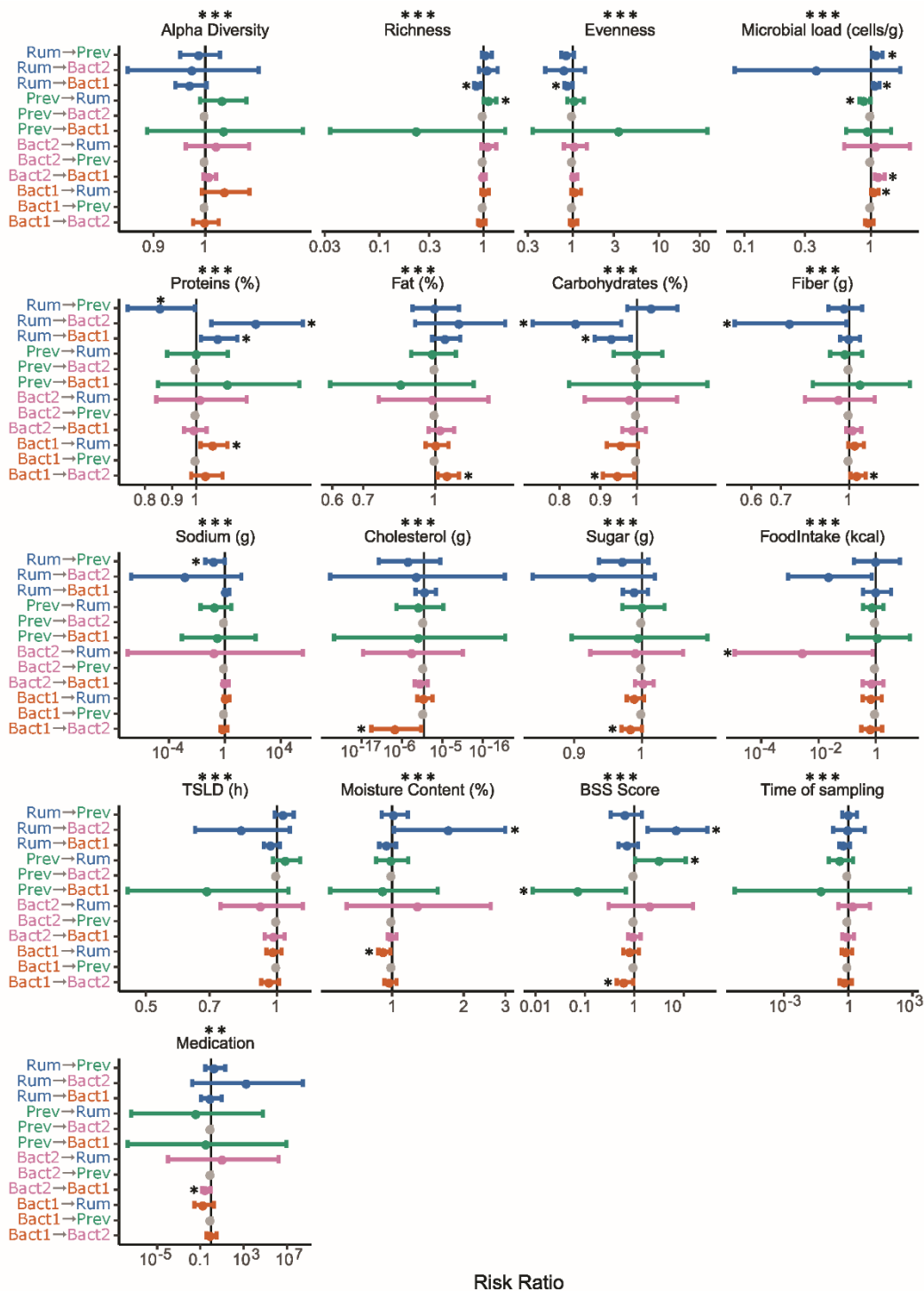
The body of the box plots represents the first and third quartiles of the distribution, and the median line. Whiskers extend from the quartiles to the last data point within  $1.5 \times \text{IQR}$ , with outliers beyond. Significance levels: \*\*\*: 0.001, \*\*: 0.01, \*: 0.05.



**Supplementary Figure 4. a. Percentage of genera belonging to the temporal core, according to different core thresholds, for each individual.** With a core threshold of 90%, about 44% [27-52%] of a person's genera are included. With less stringent temporal core thresholds more genera satisfy the conditions (e.g. about 73% [57-86%] of a person's genera are detected more than 20% of the time (the threshold for the persistent category),

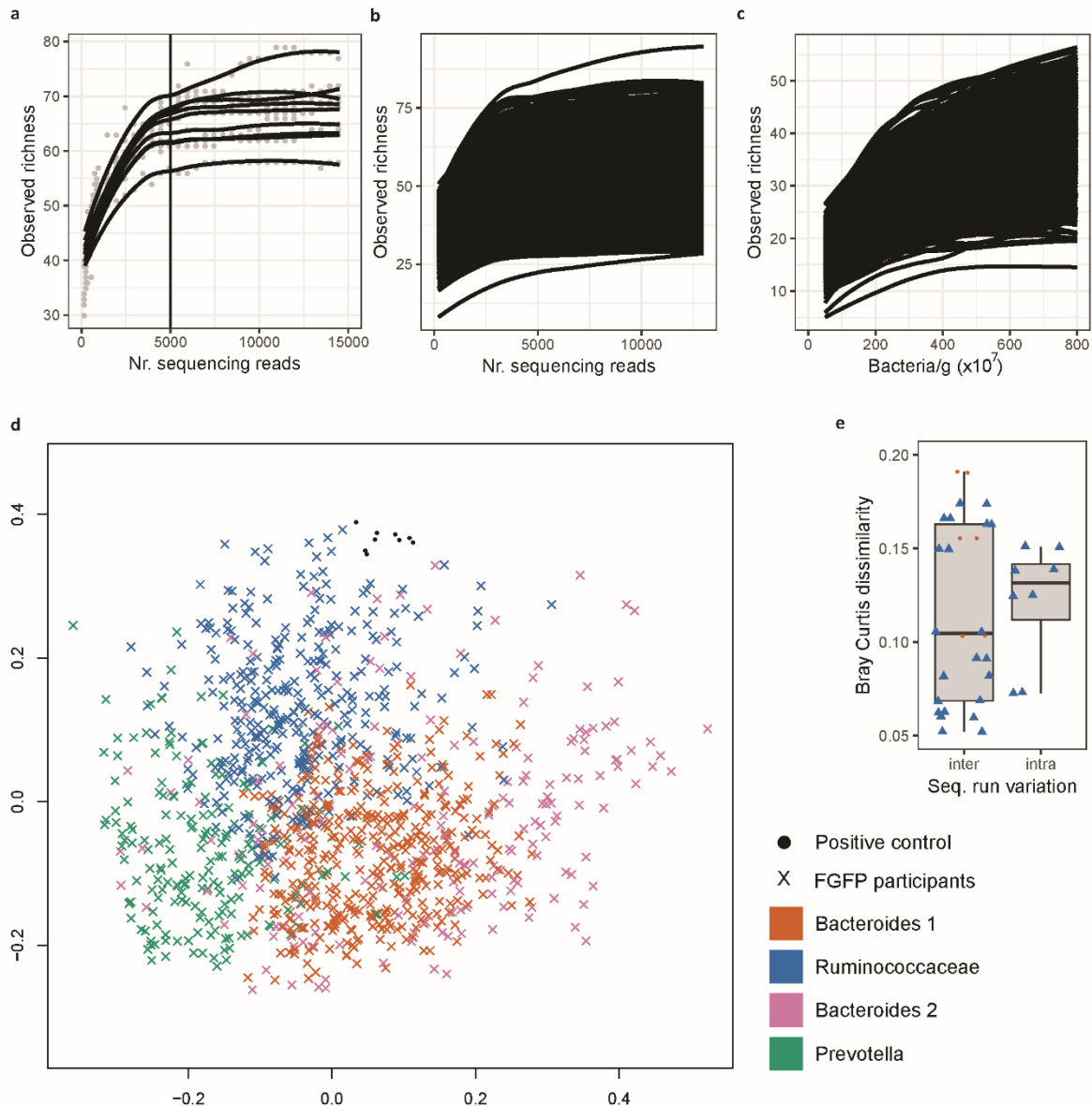
and about 58% [48-69%] of a person's genera are detected more than 60% of the time (the threshold for the transient category). **b. Temporal core size falls within 90% of the all-samples estimate from 11 samples onwards for all non-perturbed time series.** For increasing lengths of time series (x-axis) we calculated the temporal core size (with a 95% threshold, which explains the stepwise increase after 20 and 40 samples,  $N=20$ ,  $n=694$ ) and compared it with the temporal core size obtained if all samples were to be included (y-axis). The time series of participant 808, which included an infection event, is responsible for the major outliers.

The body of the box plots represents the first and third quartiles of the distribution, and the median line. Whiskers extend from the quartiles to the last data point within  $1.5 \times \text{IQR}$ , with outliers beyond. Significance levels: \*\*\*: 0.001, \*\*: 0.01, \*: 0.05.



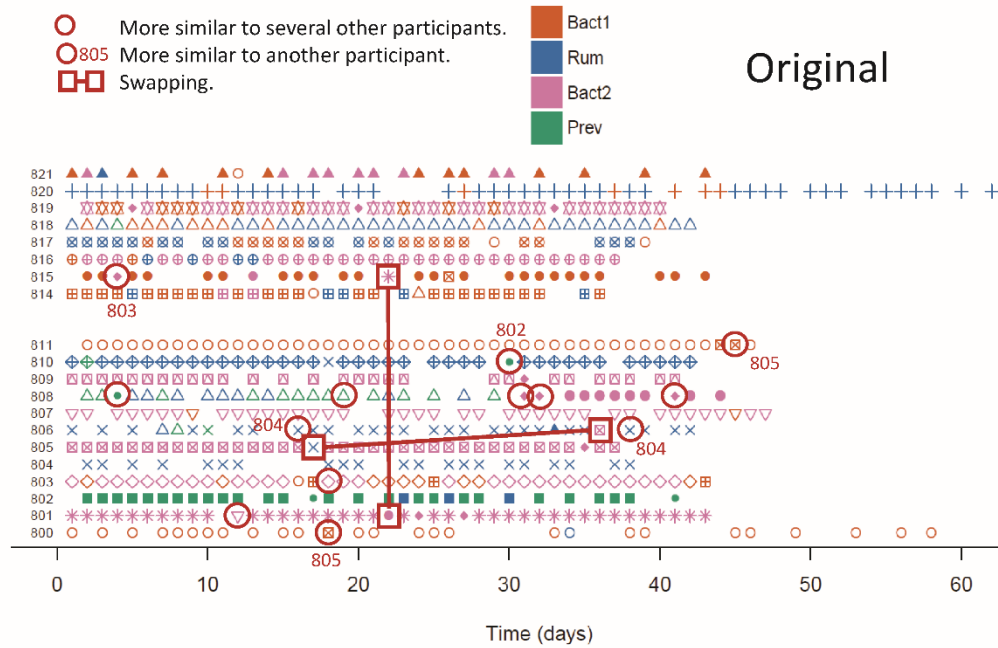
**Supplementary Figure 5. Enterotype switches are linked to several investigated parameters.** The relative risk of an enterotype switch with each unit increase in the parameter (risk ratio, x-axis), set out for each possible enterotype switch (y-axis). Switches are color coded according to data availability: no data (grey), and subsequently to direction of the switch: away from the Ruminococaceae (R, blue), Bacteroides 1 (B1, orange), Bacteroides 2 (B2, Pink), Prevotella (P, green) enterotype (N=20, n=709). Error bar dots indicate the median value with whiskers extending to the 95% confidence interval. Significance levels: \*\*\*: 0.001, \*\*: 0.01, \*: 0.05.



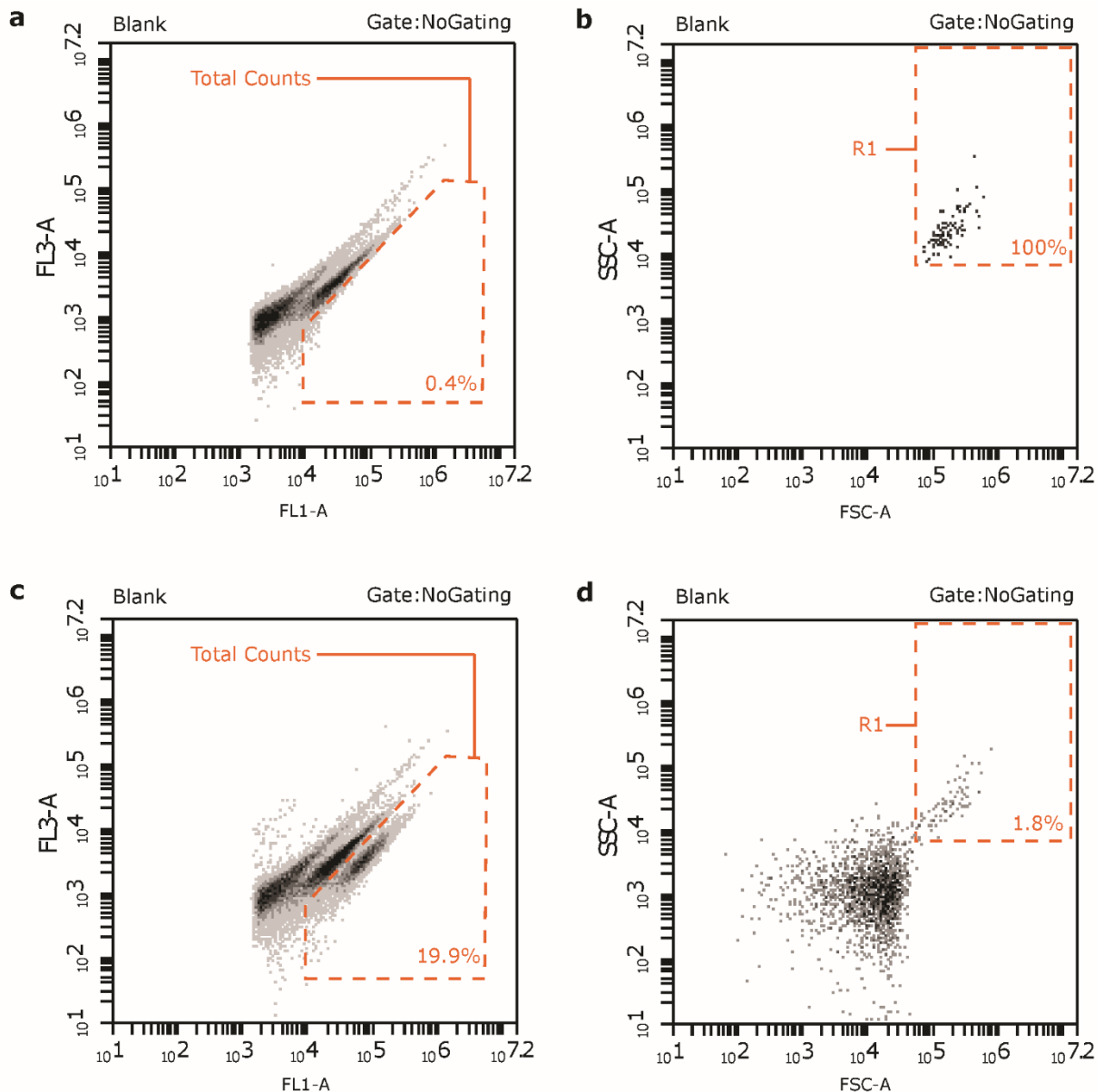


**Supplementary Figure 6. Quality control of sequencing data.** Richness could be considered stable in **a.** controls (n=9) and **b.** RMPs (N=20, n=709), from 5000 reads, and in **c.** QMPs (N=20, n=694), from  $4 \times 10^9$  bacterial cells per gram of faeces, which is well below the applied subsampling thresholds. **d.** PCoA of the positive control samples (black points) together with the RMPs of 1103 FGFP participants (crosses), coloured according to enterotype, namely Ruminococcaceae (R, blue), Bacteroides 1 (B1, orange), Bacteroides 2 (B2, Pink), Prevotella (P, green). **e.** Bray Curtis Dissimilarity between positive control samples, derived from 2 aliquots of DNA from the same high-richness stool sample (orange dots, blue triangles), between (inter) and within (intra) sequencing runs (n=9).

The body of the box plots represents the first and third quartiles of the distribution, and the median line. Whiskers extend from the quartiles to the last data point within  $1.5 \times \text{IQR}$ , with outliers beyond. Significance levels: \*\*\*: 0.001, \*\*: 0.01, \*: 0.05.



**Supplementary Figure 7 Graphical presentation of the mislabelling analyses and corrections.** Two swapping events were identified based on comparisons between BC dissimilarities of a sample with those of same or different participants, and DMM-clustering. For each participant (800-821), samples are shown according to sampling moment with an indication of enterotype, namely, Ruminococcaceae (blue), Bacteroides 1 (orange), Bacteroides 2 (Pink), and Prevotella (green), and the assigned DMM sub-cluster (20 shapes). In our original data, 15 samples showed more similarity with samples from one (circles with participant number) or several other participants (circle). Swapping events (connected squares) were identified based on according mismatches in BC-dissimilarities, enterotyping, and DMM-clustering.



### Supplementary Figure 8: Gating Strategy Flow cytometry

A fixed gating and staining approach was applied. Both blank and sample solutions were stained with SYBR Green I. **a**, The FL1-A/FL3-A acquisition plot of a blank sample (0.85% w/v physiological solution) with gate boundaries indicated. A threshold value of 2,000 was applied on the FL1 channel. **b**, Secondary gating was performed on the FSC-A/SSC-A channels to further discriminate between debris or background and microbial events. **c**, **d**, FL1-A/FL3-A count acquisition of a faecal sample (**c**) with secondary gating on FSC-A/SSC-A channels on the basis of blank analyses (**d**). Total counts were defined as events registered in the FL1-A/FL3-A gating area, excluding debris or background events observed in the FSC-A/SSC-A R1 gate. The flow rate was set at 14  $\mu$ l per minute and the acquisition rate did not exceed 10,000 events per second. Each panel reflects events registered over the course of a 30-s acquisition period.

**Supplementary Table 1: Determination of lag-time.** Proposed lag time based on cross-correlation analysis between stool moisture and the P/C ratio of the previous day for each participant, slope and significance of the correlation, related information, and final lag time as used in the analyses.

Participant	Proposed lag time *	Associated slope	Significant?	Median stool moisture (%)	Median time since last defecation (h)	Time of sampling	Final lag time
800	-4	-0.263856122	yes	73.09	30	Afternoon:12_Evening:6_Morning:10_Night:1	-4
801	-1	-0.302007752	yes	78.07	24	Morning:42	-1
802	-2	-0.166510592	no	70.98	23.75	Afternoon:11_Evening:5_Morning:14	-1
803	-3	-0.067174532	no	75.9	14.5	Afternoon:4_Morning:41	-1
804	-1	-0.170934213	no	72.53	32.5	Afternoon:5_Evening:5_Morning:11	-1
805	-2	-0.408542242	yes	82.71	22.5	Afternoon:3_Evening:1_Morning:35	-2
806	-2	-0.189618863	no	77.07	22.5	Afternoon:8_Evening:9_Morning:8_Night:1	-1
807	-1	-0.195957759	no	60.23	17.5	Afternoon:1_Morning:39	-1
808	-4	-0.091100488	no	70.805	25	Afternoon:11_Evening:11_Morning:10_Night:1	-1
809	-2	-0.226582804	no	74.84	23.5	Afternoon:10_Evening:13_Morning:7_Night:1	-1
810	-5	-0.343098783	yes	67.01	22.5	Afternoon:5_Evening:19_Morning:14	-5
811	-5	-0.19020666	no	75.94	14	Afternoon:7_Evening:5_Morning:31_Night:1	-1
814	-2	-0.147461484	no	67.51	16	Afternoon:5_Evening:2_Morning:22_Night:1	-1
815	0	-0.510107276	yes	77.98	25	Afternoon:4_Evening:13_Morning:14	0
816	-2	-0.029807728	no	86.18	14	Afternoon:22_Evening:1_Morning:14	-1
817	0	-0.183358611	no	67.94	24.5	Afternoon:1_Morning:31	-1
818	-2	-0.288589038	no	70.155	24	Afternoon:1_Evening:3_Morning:37	-1
819	-3	-0.207215476	no	81.18	23	Afternoon:2_Evening:1_Morning:36	-1
820	-1	-0.249295696	yes	68.57	14	Afternoon:8_Evening:10_Morning:29_Night:1	-1
821	-2	0.037792959	no	74.12	12	Morning:45	-1

**Supplementary Table 2: Taylor law relationships per genus.** For each genus, the linearity of the relationship between log(variance) and log(mean) over time was tested using function lm, calculating slopes, p-values and adjusted R<sup>2</sup>.

	slope	pvalue	qvalue	adjR2	excluded
<i>g_Acetanaerobacterium</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Acetitomaculum</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Acidaminococcus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Actinomyces</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Adlercreutzia</i>	1.318982	4.32E-10	8.53E-10	0.925856	No
<i>g_Aestuariaispira</i>	1.719915	6.36E-08	8.59E-08	0.930594	No
<i>g_Akkermansia</i>	1.808826	4.02E-09	6.86E-09	0.915801	No
<i>g_Alistipes</i>	1.719098	1.67E-11	5.80E-11	0.919539	No
<i>g_Allisonella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Allobaculum</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Alloprevotella</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Anaerofilum</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Anaerofustis</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Anaerorhabdus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Anaerosporobacter</i>	1.272728	4.39E-12	1.89E-11	0.968111	No
<i>g_Anaerostipes</i>	1.926801	4.15E-09	6.89E-09	0.852016	No
<i>g_Anaerotruncus</i>	1.177491	0	0	0.988168	No
<i>g_Arcanobacterium</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Bacteroides</i>	1.897164	3.68E-10	7.46E-10	0.886742	No
<i>g_Barnesiella</i>	1.773776	6.85E-09	1.02E-08	0.90917	No
<i>g_Bifidobacterium</i>	1.744071	8.44E-15	2.05E-13	0.977337	No
<i>g_Bilophila</i>	1.585377	2.79E-10	5.98E-10	0.930058	No
<i>g_Blautia</i>	2.006126	7.69E-11	2.21E-10	0.904727	No
<i>g_Butyricoccus</i>	1.628223	4.58E-09	7.28E-09	0.850388	No
<i>g_Butyricimonas</i>	1.806271	6.80E-09	1.02E-08	0.893146	No
<i>g_Butyrivibrio</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Campylobacter</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Catabacter</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Catenibacterium</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Cerasicoccus</i>	1.470821	6.10E-07	7.68E-07	0.937922	No
<i>g_Christensenella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Citrobacter</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Cloacibacillus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Clostridium_III</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Clostridium_IV</i>	1.773913	7.11E-15	2.05E-13	0.96593	No
<i>g_Clostridium_sensu_stricto</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Clostridium_XIVa</i>	1.941787	4.17E-07	5.34E-07	0.754381	No
<i>g_Clostridium_XIVb</i>	1.440485	7.76E-13	4.72E-12	0.951729	No
<i>g_Clostridium_XVIII</i>	1.778503	1.15E-10	2.89E-10	0.900429	No
<i>g_Collinsella</i>	2.038511	7.35E-14	1.07E-12	0.963396	No
<i>g_Comamonas</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Coprobacillus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%

<i>g_Coprobacter</i>	1.408486	3.03E-09	5.40E-09	0.903993	No
<i>g_Coproccoccus</i>	1.817	3.13E-12	1.43E-11	0.952627	No
<i>g_Corynebacterium</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Defluviitalea</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Desulfomicrobium</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Desulfovibrio</i>	1.379804	3.69E-05	3.85E-05	0.943116	No
<i>g_Dialister</i>	1.740359	3.65E-05	3.85E-05	0.847092	No
<i>g_Dorea</i>	1.592714	4.34E-11	1.38E-10	0.910568	No
<i>g_Eggerthella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Eggerthia</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Eisenbergiella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Elusimicrobium</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Enterobacter</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Enterococcus</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Enterorhabdus</i>	1.529756	0.002209	0.002209	0.960201	No
<i>g_Escherichia.Shigella</i>	1.923327	4.32E-09	7.01E-09	0.867438	No
<i>g_Ethanoligenens</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Eubacterium</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Ezakiella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Faecalibacterium</i>	1.586017	3.70E-08	5.10E-08	0.8117	No
<i>g_Faecalicoccus</i>	1.292338	3.65E-10	7.46E-10	0.952028	No
<i>g_Faecalitalea</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Flavonifractor</i>	1.374816	6.42E-10	1.20E-09	0.908063	No
<i>g_Frondihabitans</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Fusicatenibacter</i>	1.687615	1.57E-11	5.73E-11	0.920081	No
<i>g_Fusobacterium</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Gemella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Gemmiger</i>	1.828643	6.36E-09	9.87E-09	0.861329	No
<i>g_Gordonibacter</i>	1.36327	1.46E-06	1.75E-06	0.980529	No
<i>g_Gracilibacter</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Guggenheimella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Haemophilus</i>	1.400717	2.44E-10	5.56E-10	0.918503	No
<i>g_Hafnia</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Hespellia</i>	1.604246	2.83E-08	3.98E-08	0.834914	No
<i>g_Holdemanella</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Holdemania</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Howardella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Hungatella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Hydrogenoanaerobacterium</i>	1.34709	7.88E-11	2.21E-10	0.971302	No
<i>g_Intestinibacter</i>	1.205145	9.25E-11	2.50E-10	0.915446	No
<i>g_Intestinimonas</i>	1.522959	7.51E-07	9.30E-07	0.77939	No
<i>g_Lactobacillus</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Lactococcus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Lactonifactor</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Lautropia</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Leuconostoc</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%

<i>g_Macellibacteroides</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Mannheimia</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Methanobrevibacter</i>	2.221264	9.70E-05	9.97E-05	0.810709	No
<i>g_Methanomassiliococcus</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Methanosphaera</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Mitsuokella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Mogibacterium</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Murdochiella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Neisseria</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Odoribacter</i>	1.526895	2.22E-06	2.57E-06	0.770637	No
<i>g_Oligosphaera</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Olsenella</i>	1.708358	0.000836	0.000847	0.941462	No
<i>g_Oscillibacter</i>	1.725078	1.93E-14	3.53E-13	0.961946	No
<i>g_Oxalobacter</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Papillibacter</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Parabacteroides</i>	2.365413	3.74E-11	1.24E-10	0.923954	No
<i>g_Paraprevotella</i>	2.391589	1.74E-05	1.92E-05	0.929521	No
<i>g_Parasutterella</i>	1.430587	4.76E-13	3.52E-12	0.976771	No
<i>g_Parvibacter</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Parvimonas</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Pelospora</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Peptococcus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Peptoniphilus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Phascolarctobacterium</i>	1.871308	1.51E-05	1.70E-05	0.903855	No
<i>g_Porphyrromonas</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Prevotella</i>	1.668891	2.02E-10	4.92E-10	0.989488	No
<i>g_Proteus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Pseudobutyrvibrio</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Pseudoflavonifractor</i>	1.327879	5.36E-13	3.56E-12	0.953775	No
<i>g_Raoultella</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Rikenella</i>	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
<i>g_Romboutsia</i>	1.278416	1.11E-10	2.89E-10	0.913648	No
<i>g_Roseburia</i>	1.652482	5.24E-10	1.01E-09	0.882216	No
<i>g_Rothia</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Ruminococcus</i>	1.813903	5.84E-11	1.78E-10	0.919877	No
<i>g_Ruminococcus2</i>	1.729004	1.46E-11	5.62E-11	0.920702	No
<i>g_Senegalimassilia</i>	1.523291	2.24E-08	3.27E-08	0.981002	No
<i>g_Slackia</i>	1.405686	4.04E-09	6.86E-09	0.969562	No
<i>g_Solobacterium</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Sporobacter</i>	1.576867	2.22E-10	5.24E-10	0.944212	No
<i>g_Staphylococcus</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Streptococcus</i>	1.635934	1.24E-11	5.04E-11	0.922133	No
<i>g_Subdoligranulum</i>	1.451881	1.82E-12	9.49E-12	0.946651	No
<i>g_Succinatimonas</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Succiniclasticum</i>	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
<i>g_Sutterella</i>	1.920742	8.70E-08	1.15E-07	0.88912	No

g_Terrisporobacter	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
g_Turcibacter	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
g_Ureaplasma	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
g_Vampirovibrio	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
g_Varibaculum	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
g_Veillonella	1.426921	1.19E-12	6.71E-12	0.980074	No
g_Victivallis	1.580487	1.97E-06	2.32E-06	0.895632	No
g_Weissella	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
g_Yersinia	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_c_Alphaproteobacteria	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
uc_c_Clostridia	1.227716	3.21E-13	2.96E-12	0.983721	No
uc_c_Deltaproteobacteria	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
uc_c_Gammaproteobacteria	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_c_Opitutae	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_f_Clostridiales_Incertae_Sedis_XIII	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_f_Coriobacteriaceae	1.715084	4.82E-13	3.52E-12	0.962483	No
uc_f_Cryomorphaceae	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_f_Desulfovibrionaceae	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
uc_f_Enterobacteriaceae	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
uc_f_Erysipelotrichaceae	1.423754	2.60E-08	3.73E-08	0.940943	No
uc_f_Lachnospiraceae	1.743849	1.42E-07	1.86E-07	0.781691	No
uc_f_Peptostreptococcaceae	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_f_Porphyromonadaceae	1.488836	1.38E-06	1.68E-06	0.92567	No
uc_f_Prevotellaceae	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
uc_f_Rhodospirillaceae	1.741228	6.81E-10	1.24E-09	0.986237	No
uc_f_Rikenellaceae	1.423355	2.42E-05	2.64E-05	0.973752	No
uc_f_Ruminococcaceae	1.747086	2.64E-13	2.96E-12	0.949162	No
uc_f_Sutterellaceae	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
uc_f_Synergistaceae	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_f_Veillonellaceae	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_k_Bacteria	1.582117	3.24E-13	2.96E-12	0.964295	No
uc_o_Bacillales	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_o_Bacteroidales	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_o_Burkholderiales	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
uc_o_Clostridiales	1.650315	2.64E-10	5.84E-10	0.904405	No
uc_o_Desulfovibrionales	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_o_Lactobacillales	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_o_Rhodospirillales	NA	NA	NA	NA	Yes, present in < 5 ind with > 3 timepoints per timeseries
uc_p_Bacteroidetes	1.306841	3.07E-05	3.30E-05	0.971147	No
uc_p_Firmicutes	1.581177	2.95E-12	1.43E-11	0.969873	No
uc_p_Lentisphaerae	NA	NA	NA	NA	Yes, < abundance threshold 0.5%
uc_p_Proteobacteria	1.58872	2.81E-06	3.21E-06	0.975789	No