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### Supplemental information

### Effective chimeric antigen receptor

#### T cells against SARS-CoV-2

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## **Supplemental Figures**

SFigure 1: viSNE profiles of anti-Flag and EGFP expression on CR3022-8a-28Z T cells, related to Figure 1B. Anti-Flag staining shows the surface expression of the Flag-tagged CARs in islands i, ii and iii (following expansion and prior to incubation with 293-ACE2 cells), while GFP shows the expression of EGFP by the pHIV bicistronic lentiviral construct containing IRES-EGFP (n=4).

SFigure 2: CD4 and CD8 CAR-Ts generated within the lentiviral transduced human peripheral Tcell population, related to Figure 1B. Flow cytometric staining with anti-CD4 and anti-CD8 shows similar proportions of CD8+ and CD4+ cells amongst the CR3022-28Z, CR3022-8a-28Z, CR3022-CH3-28Z or CR3022-IgG4-28Z CAR-Ts.

SFigure 3: Flow cytometric profile of human ACE2 expression on 293 cells, related to Figure 1C. 293 cells were transduced to stably express the hACE2 receptor. Light blue: isotype control; pink: anti-h ACE2 receptor.

SFigure 4: Changes in the expression of CD69 in subsets of CAR-Ts following incubation with 293-ACE2 or 293-ACE2-RBD cells, related to Figure 1E. From Figure 1e, the viSNE is subdivided into Islands i ii and and iii based on the surface expression of the activation marker CD69. Fig. 1e, right panels shows the changes in expression for island i. SFigure 4 shows that change in the percent representation amongst CAR-Ts for islands ii and and iii (Panel A); MFI changes in expression (Panel B).

SFigure 5: Response of transduced Jurkat T-cells expressing CARs, related to Figure 1. Induction of CD69 on Jurkat cells expressing different CARs in response to Vero cells loaded with RBD peptide. Data are shown as mean  $\pm$  SD (n=3). \* represents p < 0.05. \*\* represents p < 0.01. \*\*\*\* represents p < 0.0001. (compared with CAR-Jurkat only group by one-way ANOVA, n=3).

### Supplemental Figures

SFigure 6: Examples of the cytolytic response of SARS-CoV-2 CAR-Ts with different donor to the RBD and S1 peptides, related to Figure 2. The killing of 293-ACE2 target cells that had been precoated with either the RBD (upper panels: donors B and C) or S1 peptide (lower panels: donors A and E). Data are shown as mean  $\pm$  SD (n=3). \* represents p < 0.05. \*\* represents p < 0.01. \*\*\* represents p < 0.001 (compared with CR3014-28Z by two-way ANOVA).

SFigure 7: CAR-Ts do not specifically block SARS-CoV-2 pseudotyped viral particle entry into cells, related to Figure 7. Recombinant pseudotyped viral particles containing SARS-CoV-2 spike protein were used to mimic SARS-CoV-2 cell infection and cell entry. The SARS-CoV-2 pseudo-virus particles encode DsRed in the viral genome. The DsRed gene will be strongly expressed after the SARS-CoV-2 pseudo-virus entry into ACE2-expressing cells. It shows a trend in the blockade of pseudotyped viral particle entry by both CR3014 and CR3022 at different ratios.

### **SMovies**

SMovie 1: Panels of time-lapse movies of co-culturing of CAR-Ts with 293-ACE2 cells, related to Figure 3. CR3022-8a-28Z CAR-Ts were co-cultured with the 293-ACE2 cells and tracked for 20 hours. EGFP+ CAR-T cells failed to form clusters with 293-ACE2.

SMovie 2: Panels of time-lapse movies of co-culturing of CAR-Ts with RBD peptide loaded 293-ACE2 cells, related to Figure 3. CR3022-8a-28Z CAR-Ts were co-cultured with 293-ACE2 that had been pre-incubated with RBD peptide and tracked for 20 hours. EGFP+ CAR-T cells were observed to form clusters with 293-ACE2+RBD cells.

SMovie 3: Panels of time-lapse movies of co-culturing of CAR-Ts with S1 peptide loaded 293-ACE2 cells, related to Figure 3. CR3022-8a-28Z CAR-Ts were co-cultured with 293-ACE2 that had been pre-incubated with S1 peptide and tracked for 20 hours. EGFP+ CAR-T cells were observed to form clusters with 293-ACE2+S1 cells.



CR3022-8a-28Z



CR3022-8a-28Z



CR3022-CH3-28Z

CR3022-IgG4-28Z

1.95

23.3

10<sup>5</sup>

10<sup>4</sup>















Effecter/Target ratio

### 293-ACE2 with soluble S1 (Donor A)



# Neutralization of SARS-CoV-2 spike pseudotyped VSV

