

Additional file 1

CD4+ effector T cells accelerate Alzheimer's disease in mice

Jatin Machhi^{*1}, Pravin Yeapuri¹, Yaman Lu¹, Emma Foster², Rupesh Chikhale³, Jonathan Herskovitz⁴, Krista L. Namminga¹, Katherine E. Olson¹, Mai Mohamed Abdelmoaty^{5,6}, Ju Gao¹, Rolen M. Quadros^{1,7}, Tomomi Kiyota⁸, Liang Jingjing¹, Bhavesh D. Kevadiya¹, Xinglong Wang¹, Yutong Liu^{1,9}, Larisa Y Poluektova¹, Channabasavaiah B. Gurumurthy^{1,7}, R Lee Mosley¹, Howard E Gendelman^{1,5*}

¹*Department of Pharmacology and Experimental Neuroscience, University of Nebraska Medical Center, Omaha, NE 68198, USA*

²*Department of Biological Sciences, Northern Kentucky University, Highland Heights, KY 41099, USA*

³*University College London School of Pharmacy, Bloomsbury, London WC1E 6DE, UK*

⁴*Department of Pathology and Microbiology, College of Medicine, University of Nebraska Medical Center, Omaha, NE 68198, USA*

⁵*Department of Pharmaceutical Sciences, College of Pharmacy, University of Nebraska Medical Center, NE 68198, USA*

⁶*Therapeutic Chemistry Department, Pharmaceutical and Drug Industries Research Division, National Research Centre, Giza, Egypt*

⁷*Mouse Genome Engineering Core Facility, Vice Chancellor for Research Office, University of Nebraska Medical Center, Omaha, Nebraska, USA*

⁸*Department of Safety Assessment, Genentech Inc., South San Francisco, CA 94080, USA*

⁹*Department of Radiology, University of Nebraska Medical Center, Omaha, NE 68198, USA*

Corresponding authors:

Howard E. Gendelman, Department of Pharmacology and Experimental Neuroscience, University of Nebraska Medical Center, Omaha, NE 6898-5880; phone 402-559-8920; fax 402-559-3744.

Email hegendel@unmc.edu (for review and correspondence)

Jatin Machhi, Department of Pharmacology and Experimental Neuroscience, University of Nebraska Medical Center, Omaha, NE 6898-5880; phone 402-559-2779.

Email jatin.machhi@unmc.edu (shared correspondence).

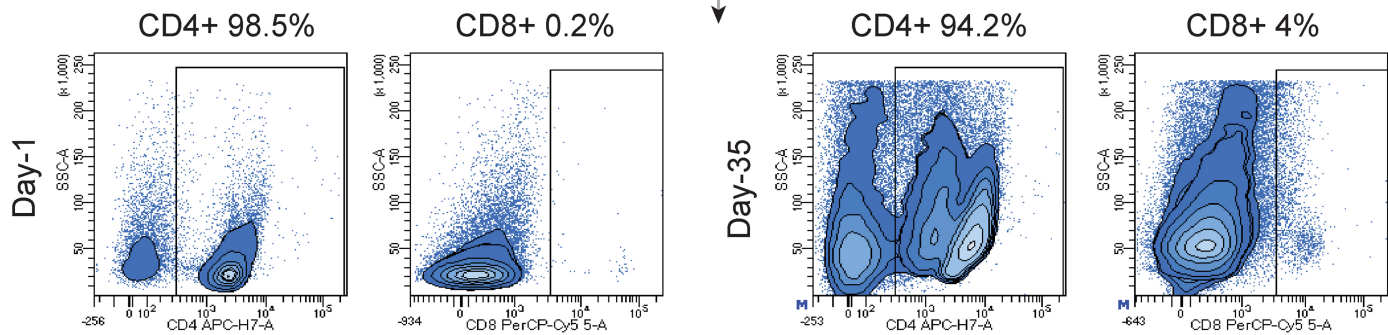
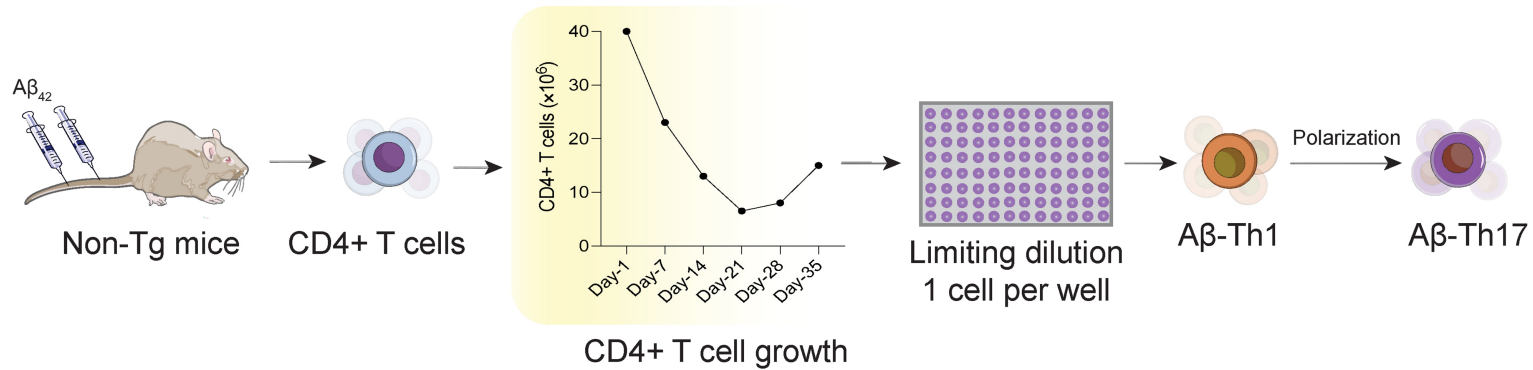
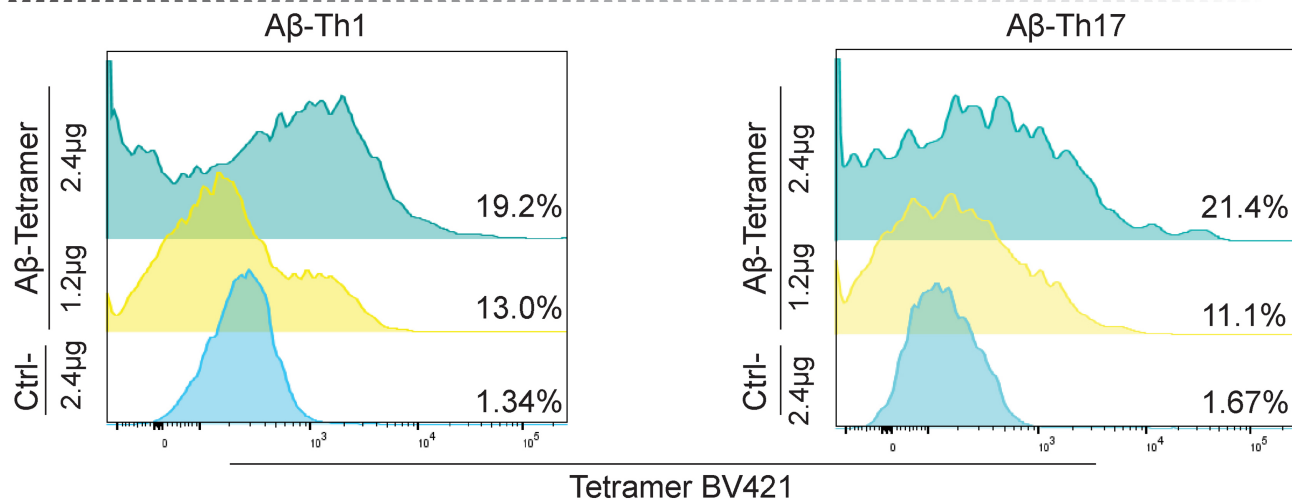
a**b**

Fig. S1. Development of monoclonal A β -Th1 and A β -Th17 cells. (a) Workflow showing development of antigen-specific T cells. Non-Tg mice immunized with A β_{1-42} followed by isolation of CD4⁺ T cells from spleen and lymph nodes. In vitro culture of CD4⁺ T cells in the presence of feeder cells and A β_{1-42} for enrichment of antigen-rich population. Growth pattern of CD4⁺ T cells over in vitro incubation where cells growth declined for a several weeks followed by increased cell number without compromising CD4⁺ T cell purity (shown as % of CD3⁺ cells). Limiting dilution culture to seed as low as 1 cell per well in presence of feeder cells, A β_{1-42} , and IL2 to obtain A β -Th1 cell clone. A β -Th1 cells polarized into A β -Th17 cells using conditional culture media in presence of feeder cells and A β_{1-42} . (b) Staining of three-month propagated A β -Th1 and A β -Th17 cells with two concentrations of MHCII-IA^b-KLVFFAEDVGSNKGA (A β T cell epitope) tetramer after incubation at 37 °C for 3 hr. Control tetramer MHCII-IA^b-PVSKMRMATPLLMQA (Ctrl) was used to determine non-specific binding of T cells.

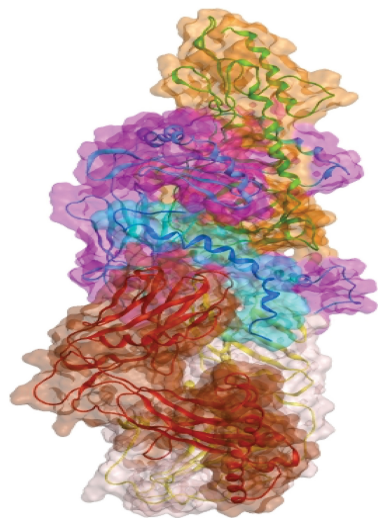
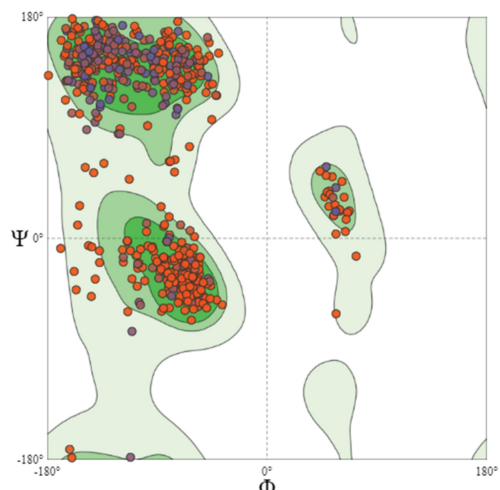
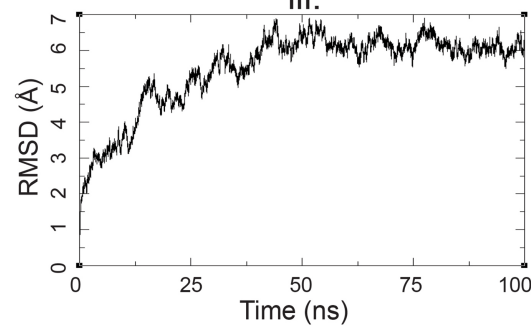
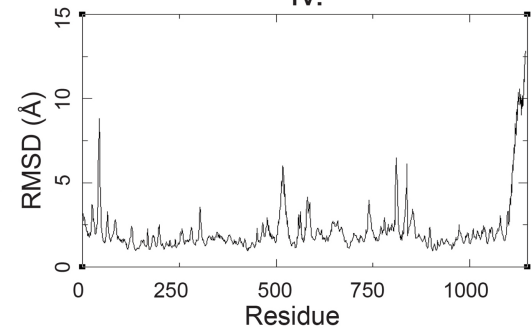
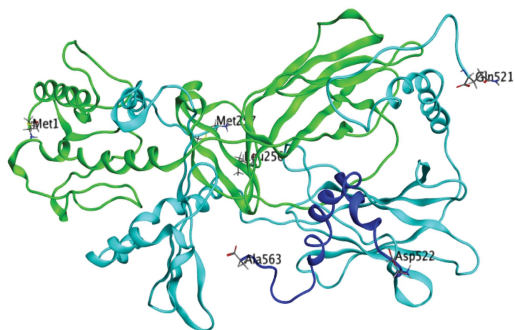
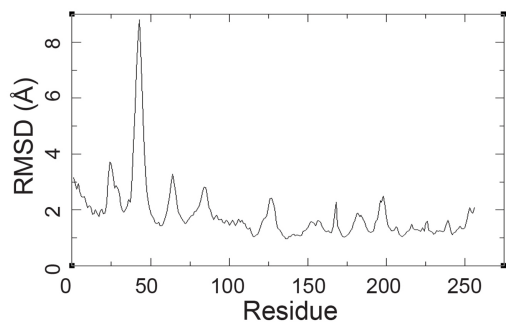
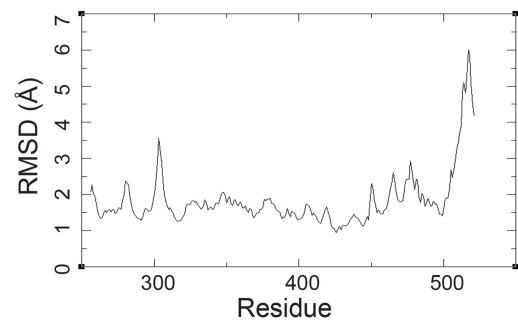
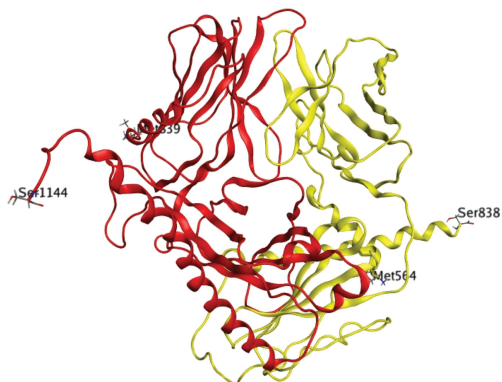
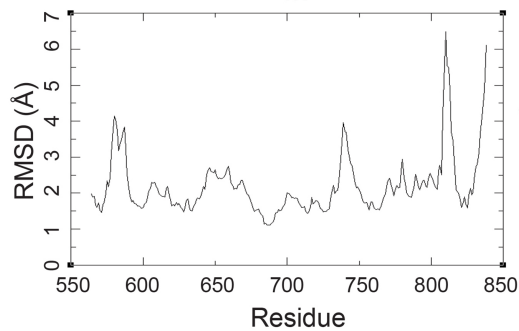
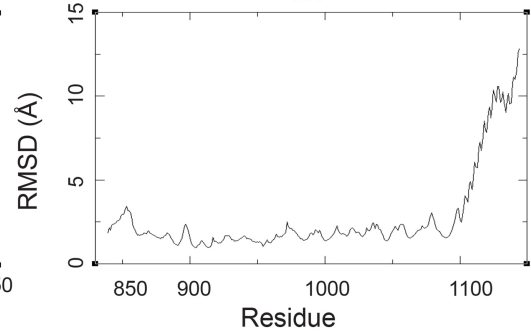
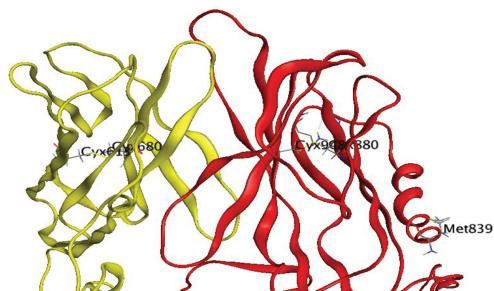
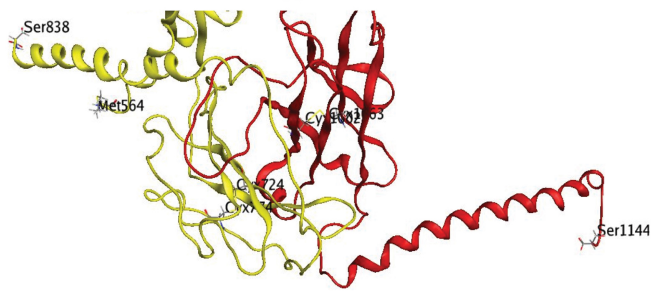
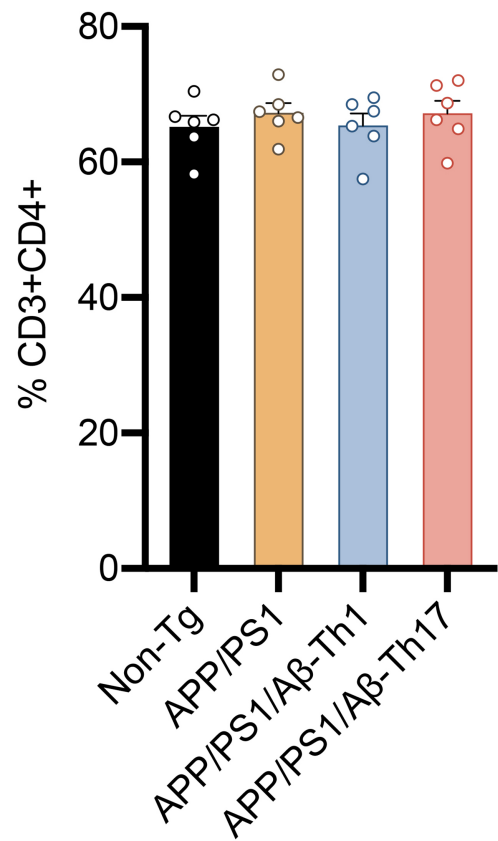
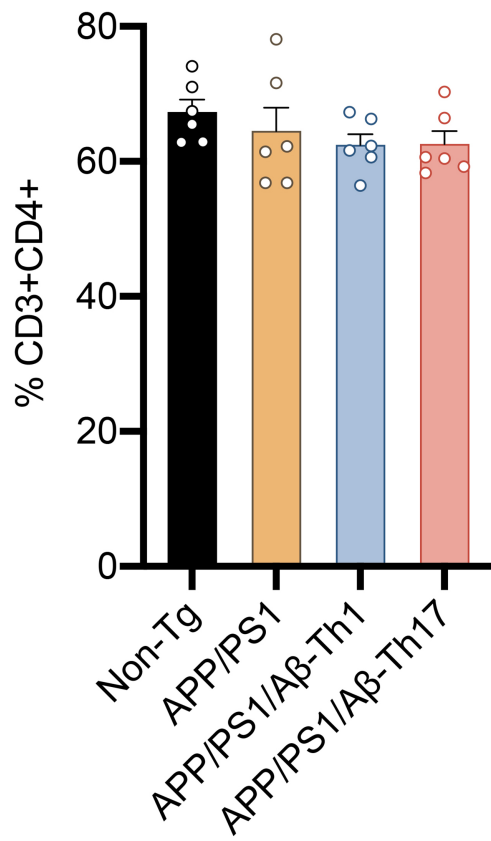
a**ii.****iii.****iv.****b****i.****ii.****iii.****c****i.****ii.****iii.****iv.****v.**

Fig. S2. Modelling and explicit solvent molecular dynamics simulations. (a) (i) TCR-pMHC complex with surface interactions (post MD production); (ii) Ramachandran plot for the optimized and energetically stabilized model; (iii) RMSD plot for the TCR-pMHC complex for 100 ns MD simulation; (iv) RMSF for complete TCR-pMHC complex over the period of 100 ns MD simulation. (b) (i) pMHC complex from the modelled complex; RMSF for the (ii) MHC α chain and (iii) MHC β chain. (c) (i) TCR complex; (ii) RMSF of TCR α chain and (iii) TCR β chain from the complex. (iv) Close-up of the Fab (fragment antigen binding) and (v) the Fc (fragment crystallizable) regions.

Blood



Spleen



Lymph nodes

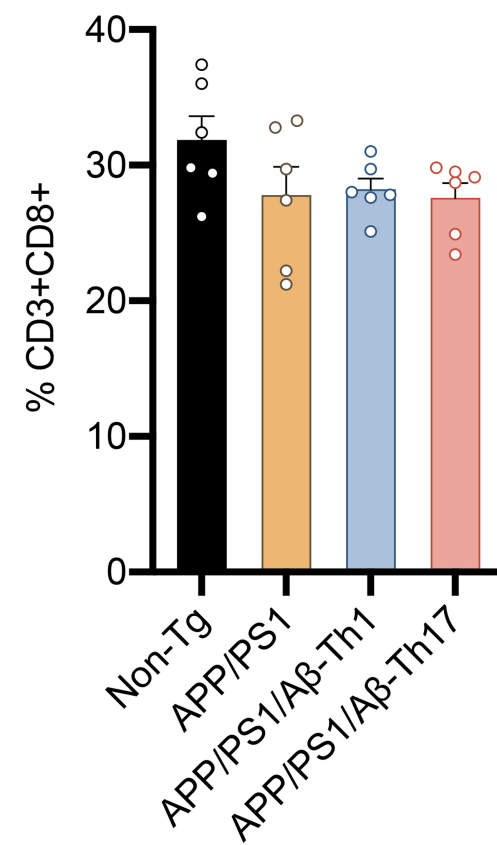
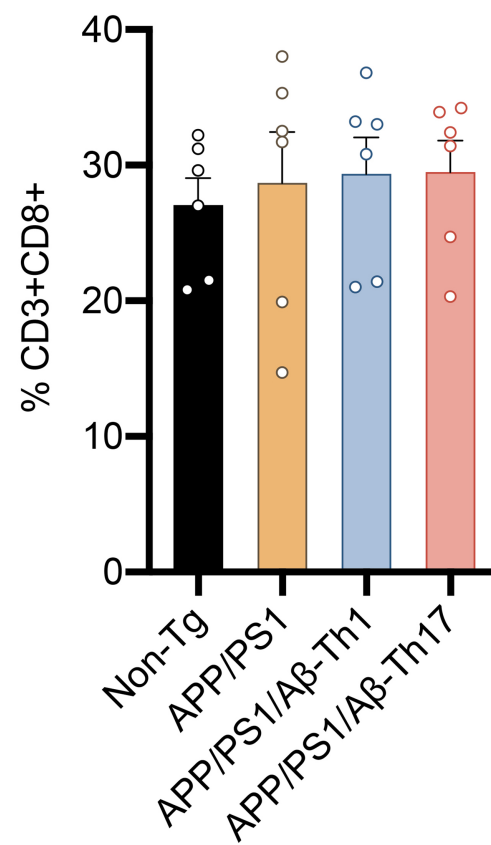
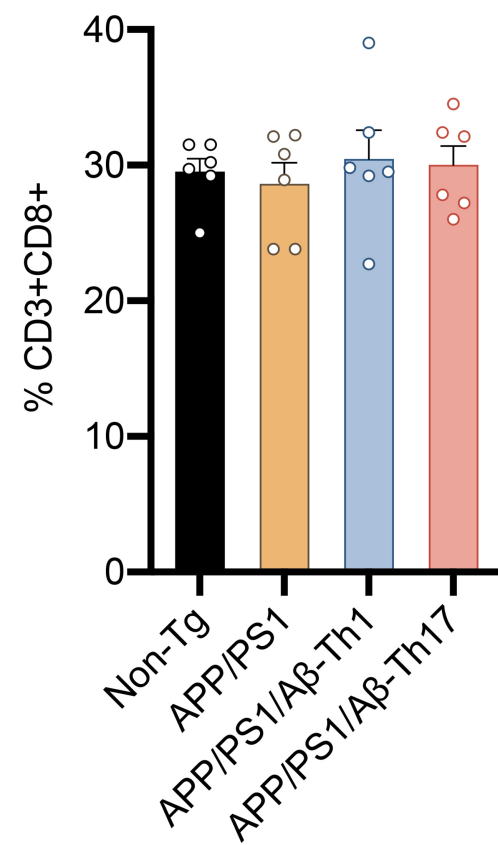
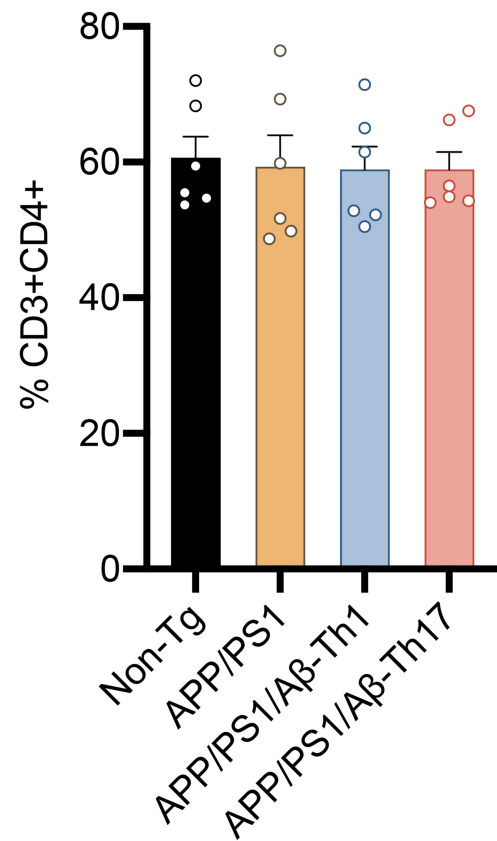
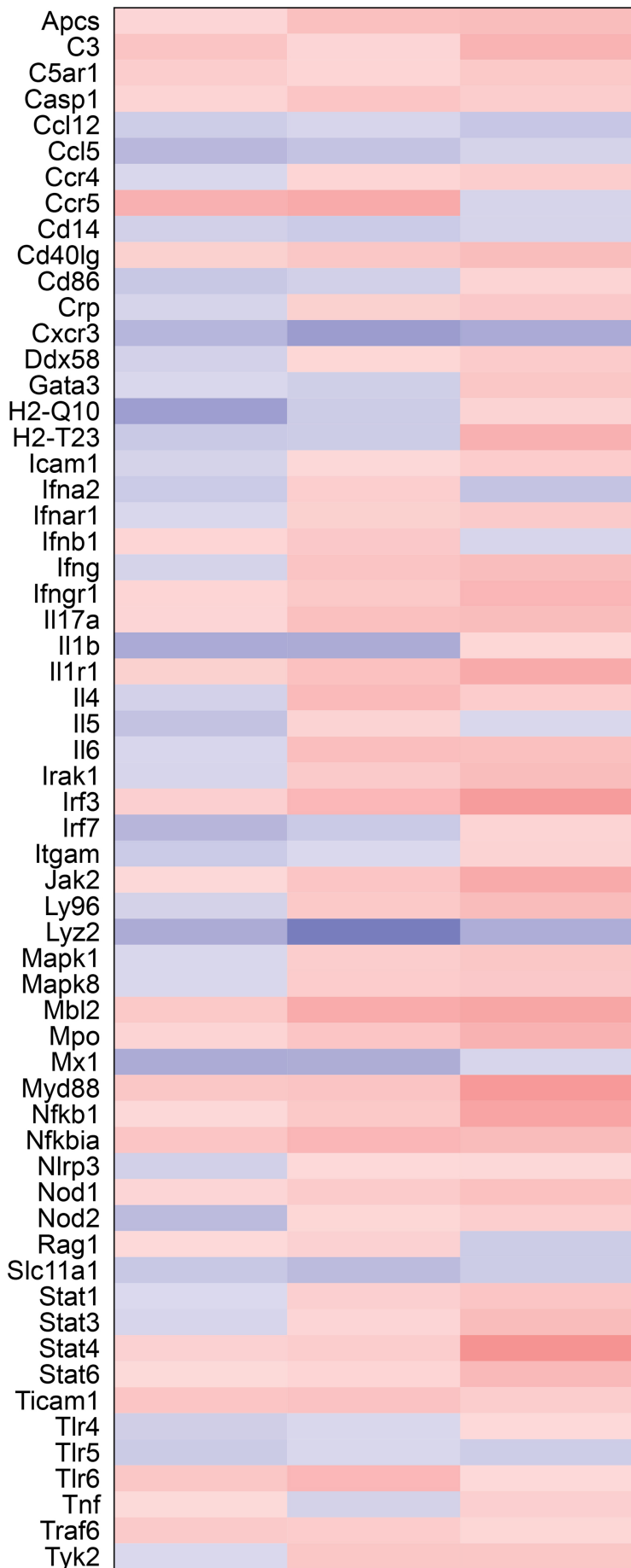


Fig. S3. Adoptive transfer of A β -Teffs did not affect T cell frequency in APP/PS1 mice. Frequency of CD3+CD4+ and CD3+CD8+ T cells in blood, spleen and lymph nodes by flow cytometric analysis for n=6 mice per group. One-way ANOVA followed by Newman–Keuls post-hoc test was used to determine statistical significance.

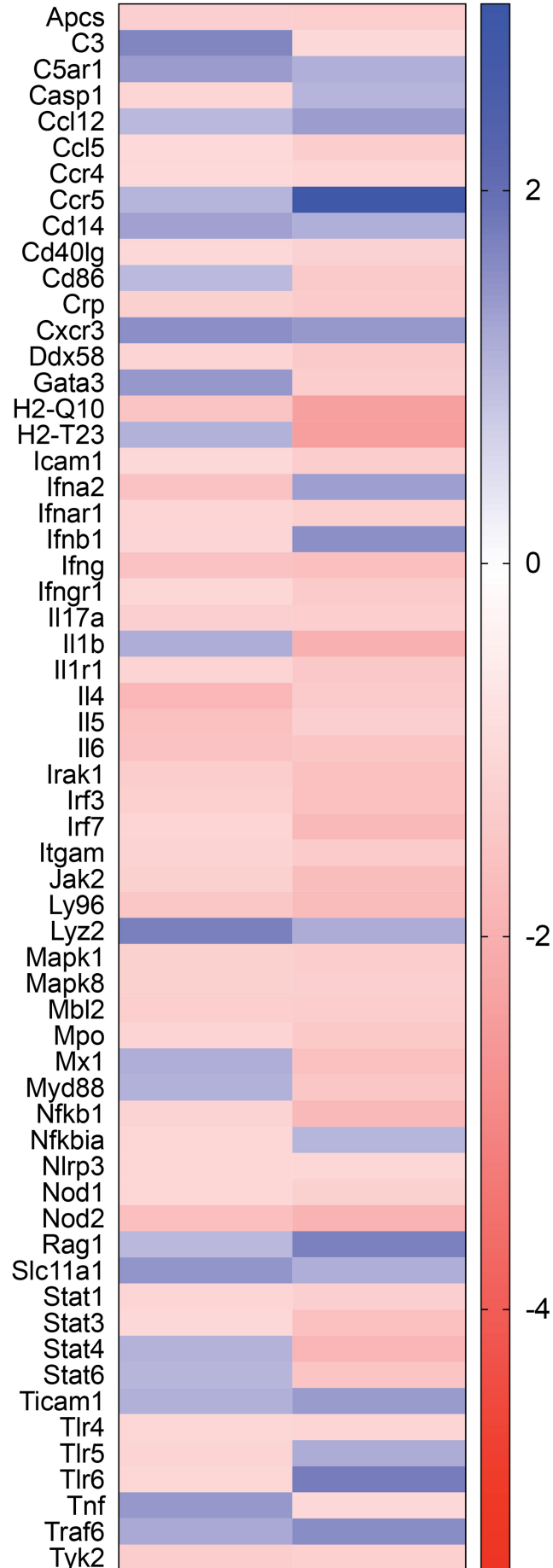
APP/PS1

APP/PS1/A β -Th1APP/PS1/A β -Th17Fold regulation
vs non-Tg

5

0

-5

APP/PS1/A β -Th1APP/PS1/A β -Th17Fold regulation
vs APP/PS1

2

0

-2

-4

Fig. S4. Transcriptomic analysis of immune genes. Gene expression of different innate and adaptive immune genes from the hippocampus of each treatment group were assessed in Qiagen RT²-PCR arrays. Genes less affected and not covered in **Fig. 5c** are summarized here. (left) Gene expression in different APP/PS1 mice compared to non-Tg mice. (right) Gene expression in APP/PS1/A β -Th1 and APP/PS1/A β -Th17 mice compared to untreated APP/PS1 mice for n=4 mice per group. Fold changes in different innate and adaptive immune genes are presented as heat maps.

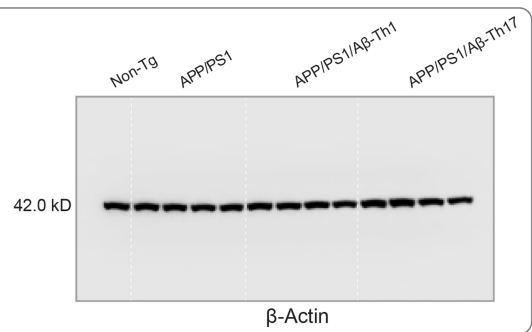
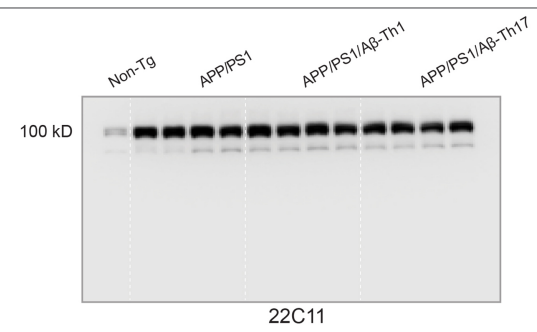
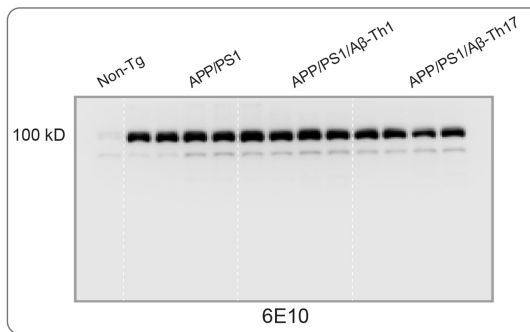
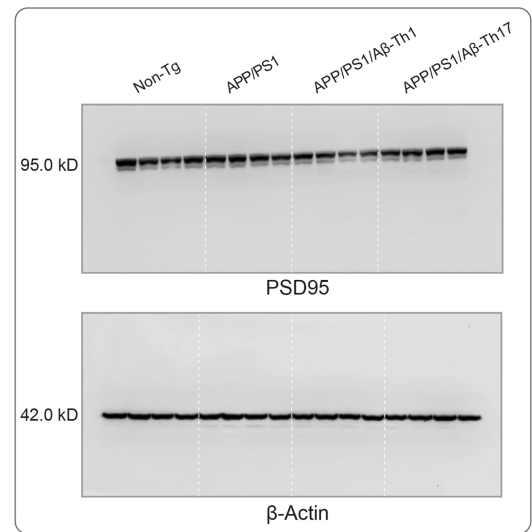
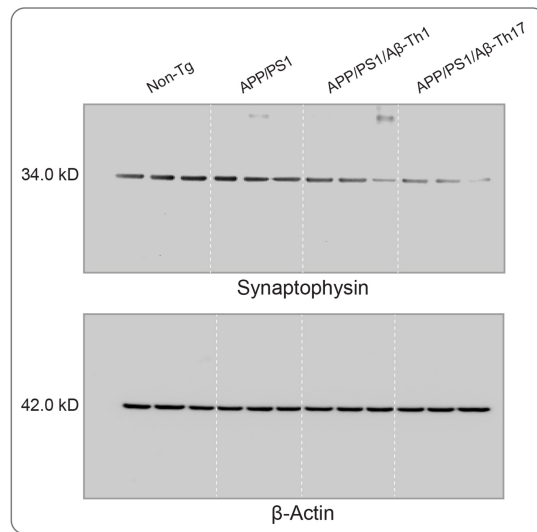
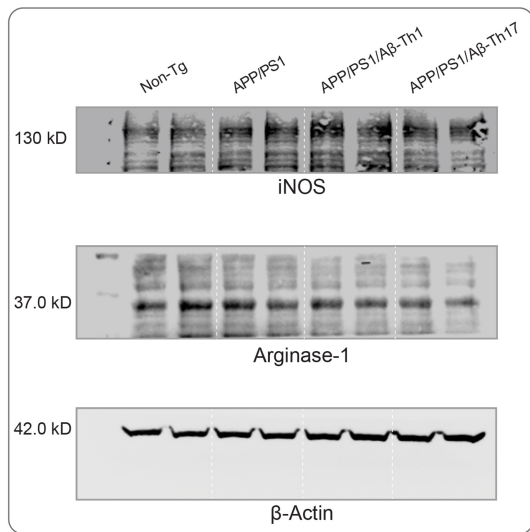


Fig. S5. Western blot images. Full blot image of iNOS, arginase-1, synaptophysin, PSD95, 6E10, 22C11 and β -actin protein expression determined from cortical tissues of each treatment group.