

# Supplementary Materials for

# Deubiquitination of proteasome subunits by OTULIN regulates type I IFN production

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Published 19 November 2021, *Sci. Adv.* 7, eabi6794 (2021) DOI: 10.1126/sciadv.abi6794

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Figs. S1 to S4





IFIT3

MDA5

Rig-I

β-actin

ISG15 (LE) ISG15 (SE) β-actin

### Fig. S1. Excessive inflammation signatures in Otulipenia patients.

(A) NanoString analysis of NF- $\kappa$ B signaling in whole blood samples from two patients and four healthy controls, and a type I interferonopathy patient control (PC) with DNASE2 deficiency.

(B) Gene expression levels of IFN-I pathway in PBMCs. The PBMCs from P3 and two healthy controls were stimulated with poly(I:C) (20  $\mu$ g/mL) or LPS (1  $\mu$ g/mL) for 22 hours before RNA extraction.

(C) Expression levels of *OAS1*, *IFI44L* and *RSAD2* in patients' fibroblasts. Fibroblasts from P1 and P2 were treated with 0.5  $\mu$ M Baricitinib for the indicated amount of time.

(**D**) Gene expression levels of IFN pathway in P1's and P2's fibroblasts. Cells were treated with 0.5  $\mu$ M Baricitinib or 10  $\mu$ g/mL Enbrel for 24 hours.

(E, F) Western blots of the inhibition level of type I IFN signaling (E) and ISG15 conjugates (F) in P1's and P2's fibroblasts. Cells were treated with 0.5  $\mu$ M Baricitinib or 10  $\mu$ g/mL Enbrel for 4 or 24 hours. LE, long exposure; SE, short exposure.



WТ ко

1.5

-1

-0.5

0

0.5

1

1.5

Fig. S2. The elevated type I IFN signals in OTULIN deficient cells are NF-KB-independent.

(A) The transcription level of NF-KB, ISRE (Interferon Stimulation Response Element) and IFN-

 $\beta$  in wild type (WT) and OTULIN KO HEK293T cells measured by luciferase assay.

(B) Western blots of the activation levels of type I IFN signaling in wild type (WT) and OTULIN KO 293T cell lines.

(C) ELISA analysis of IP10 level in the supernatants of wild type and OTULIN KO 293T cell lines.

(**D**) Quantitative real-time PCR analysis of expression levels of genes in IFN-I and NF- $\kappa$ B signaling pathways in wild type and OTULIN KO 293T cell lines.

(E) RNA sequencing analysis of the expression levels of genes in the IFN-I pathway (left) and NFκB pathway (right) in wild type and OTULIN KO 293T cells.

(F) Gene enrichment analysis, using GO database, of differentially expressed genes by RNA sequencing in wild type and OTULIN KO 293T cells reveals enrichment of antiviral-related pathways.

(G) Gene expression levels of IFN pathway in OTULIN KO HEK293T cells complemented with different amounts of wild type OTULIN.

(H) Luciferase assay of the transcription level of NF- $\kappa$ B, ISRE and IFN- $\beta$  in OTULIN KO HEK293T cells treated with a NF- $\kappa$ B inhibitor. OTULIN\_KO2 cells were treated with 10  $\mu$ M I $\kappa$ B $\alpha$  ubiquitination inhibitor GS-143 for 8 hours.

(I) Luciferase assay of the transcription level of NF- $\kappa$ B, ISRE and IFN- $\beta$  in both OTULIN KO cell lines after inhibition of the NF- $\kappa$ B by knockdown of p65 using 3 different siRNAs.

(J) The expression levels of genes in IFN and NF-kB signaling pathways. p65 in HEK293T

OTULIN KO cells was knocked down by 3 different siRNAs.

Data in panel A, C, D are shown as mean  $\pm$  SEM from at least three repeated biological analyses.

*P* values were determined by unpaired two-tailed *t*-test. \*\*P < 0.01, \*\*\*P < 0.001, \*\*\*P < 0.0001.



#### Fig. S3. OTULIN deficiency causes proteasome dysfunction.

(A) Mass spectrometry analysis identified the interactions between OTULIN or HOIP and various proteasome subunits. Numbers represent peptides captured by mass spectrometry.

(B) The K48 ubiquitination levels in wild type (WT) and OTULIN KO 293T cell lines.

**(C)** Proteasome activities in empty vector (EV), OTULIN wild type (WT), the catalytic-dead variant Cys129Ala (C129A) and the disease-associated variant Leu272Pro (L272P) overexpressed OTULIN KO 293T cells.

**(D)** Native gel analysis of the proteasome assembly in OTULIN WT and KO (KO1, KO2) 293T cell lines. The assembly of different parts of the proteasome was illustrated using antibodies of different subunits correspondingly.

(E) Western blots analysis of linear ubiquitination levels on purified proteasome from OTULIN KO 293T cells.

(F) Glycerol gradient centrifugation of lysed wild type and OTULIN KO 293T cells (KO1, KO2), and OTULIN complemented OTULIN KO 293T cells (KO2+OTULIN). The fractions were analyzed using proteasome chymotrypsin-like activity (upper panel) and immunoblotting with indicated antibodies (lower panel).

(G) Increased linear ubiquitination of proteasome fraction in OTULIN KO 293T cells as indicated by glycerol gradient centrifugation followed by proteasome chymotrypsin-like activity assay (upper panel) and immunoblotting (lower panel).

Data in panel B is shown as mean  $\pm$  SEM from four repeated technical replicates. *P* values were determined by unpaired two-tailed *t*-test. \**P* < 0.1, \*\*\**P* < 0.001, \*\*\*\**P* < 0.0001.



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GAPDH

PSMA7

PSMC6 PSMA7 PSMC6

Flag GAPDH Fig. S4. Linear ubiquitination on proteasome subunits could be removed by OTULIN deubiquitinase activity.

(A, B) Detection of OTULIN-mediated deubiquitination of the proteasome subunits using immunoprecipitation (IP). LUBAC complex consists of the catalytic subunit HOIP and two accessory proteins: HOIL1 and Sharpin. WCL, whole cell lysate. Ub-WT: wild type Ub plasmid.
(C) Immunoprecipitation confirmed the interactions between OTULIN and proteasome subunits. HEK293T WT cells were transfected with OTULIN and flag tagged proteasome subunits before lysed.

(**D**) The interactions between OTULIN and endogenous proteasome subunits PSMA7 and PSMC6 were indicated by immunoprecipitation in HEK293T cells.