

## **Supplementary Material**

### **Supplementary Figure Legends**

**Supplementary Figure 1.** Mean cell densities of immune cell subpopulations (A) and their co-expression with immune checkpoints (B) in ER-positive and ER-negative tumors

**Supplementary Figure 2.** Heatmaps of clusters of immune cell subpopulations derived from multiplex IHC using unsupervised hierarchical clustering in stroma (A) and tumor (B) compartments

**Supplementary Figure 3.** CUTseq experimental replicates' concordance

**Supplementary Figure 4.** Percentage (A) and size (B) of the altered genome (amplified or deleted) in ER-positive and ER-negative tumors. In the boxplots, each box extends from the 25th to the 75th percentile, the midline represents the median, and the whiskers extend from  $-1.5 \times \text{IQR}$  to  $+1.5 \times \text{IQR}$  from the closest quartile, where IQR is the inter-quartile range

### **Supplementary Tables**

**Supplementary Table 1.** Cell densities (cells/mm<sup>2</sup>) of the different immune cell subsets as derived from the multiplex fluorescent IHC method

**Supplementary Table 2.** Correlation matrix values for the different immune cell subsets

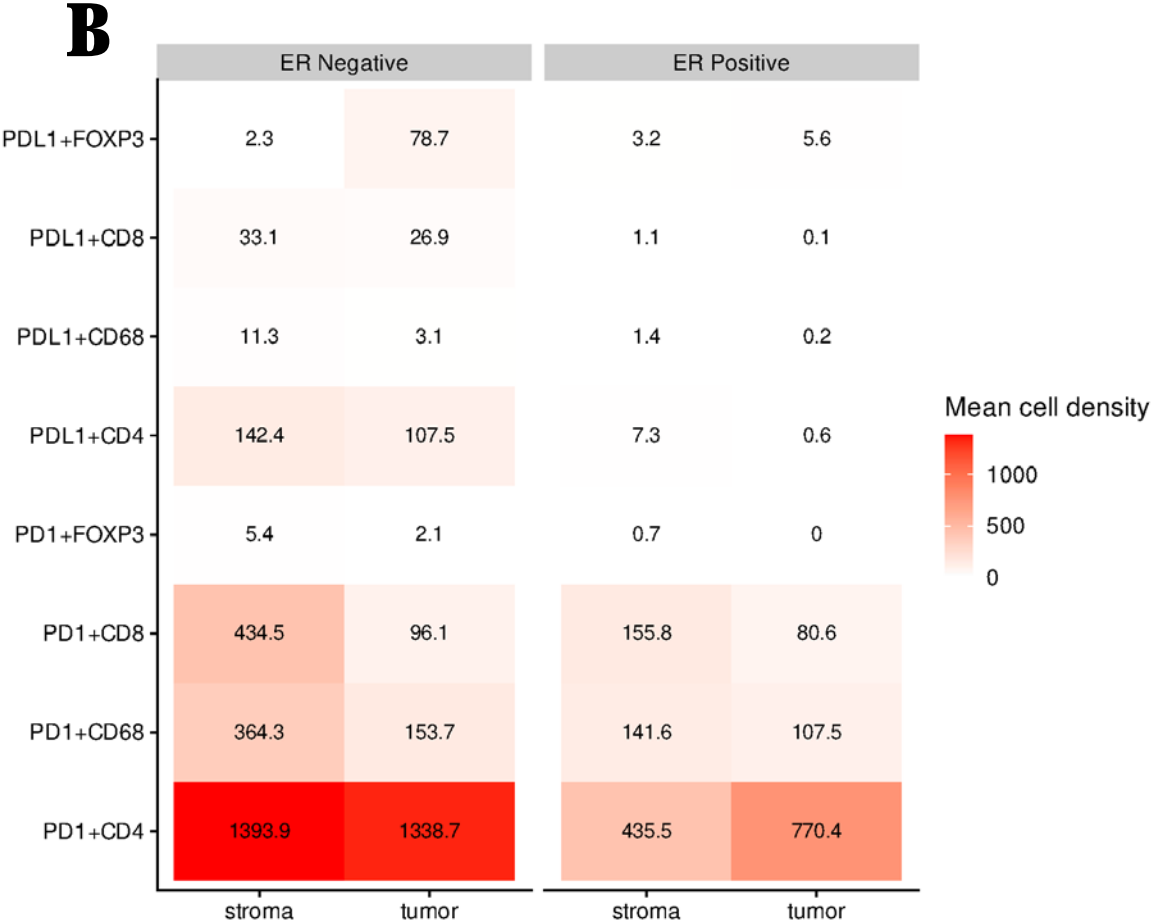
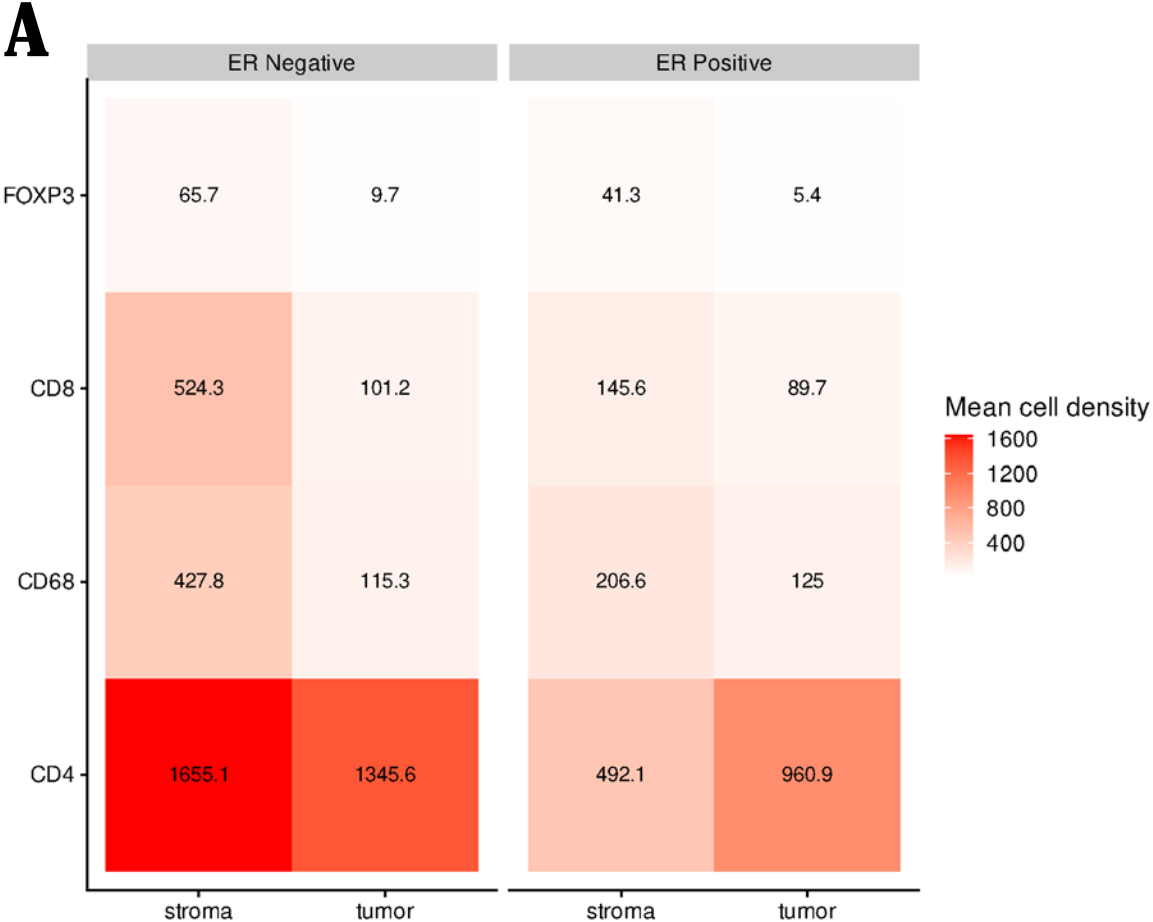
**Supplementary Table 3.** Patient characteristics per manual TILs group in the SBG-2004-1 study cohort

**Supplementary Table 4.** Variables derived from the digital TILs evaluation

**Supplementary Table 5.** List of antibodies used for the multiplex fluorescent IHC method

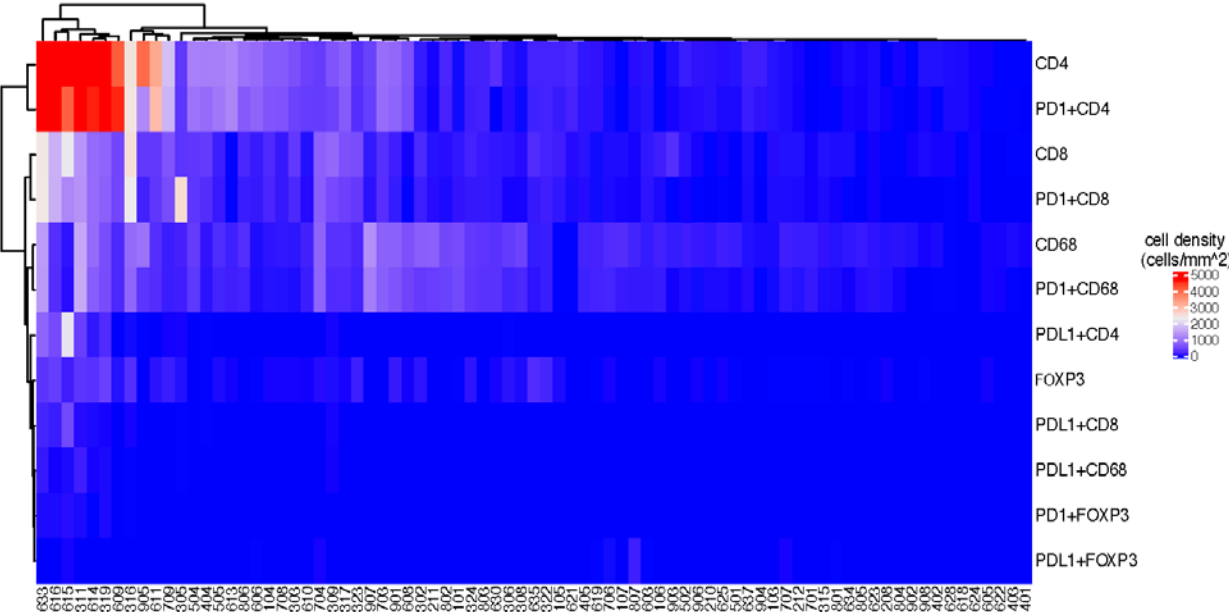
**Supplementary Data 1.** Segmented Somatic Copy Number Alterations profiles by CUTseq

# Supplementary Figure 1

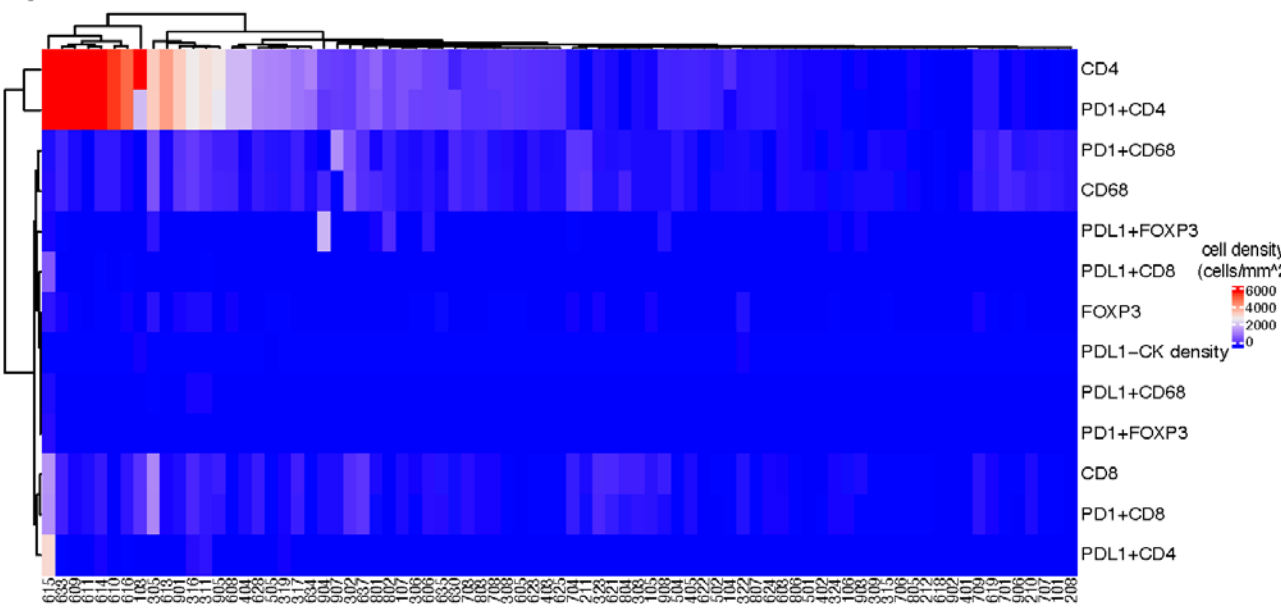


# Supplementary Figure 2

## A) Stroma compartment

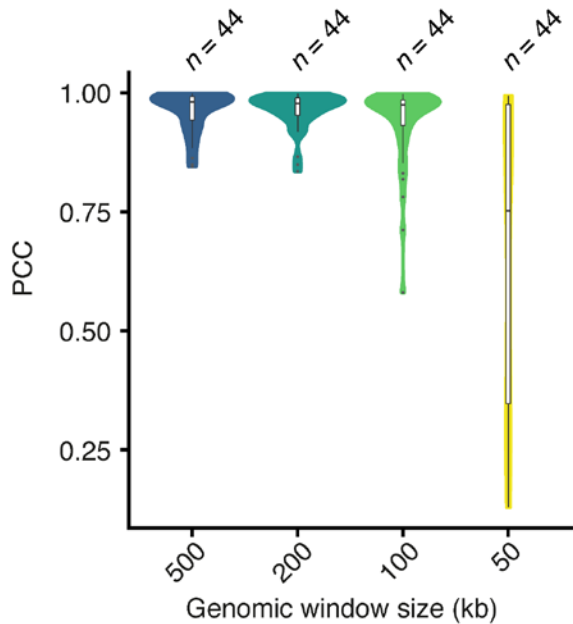


## B) Tumor compartment

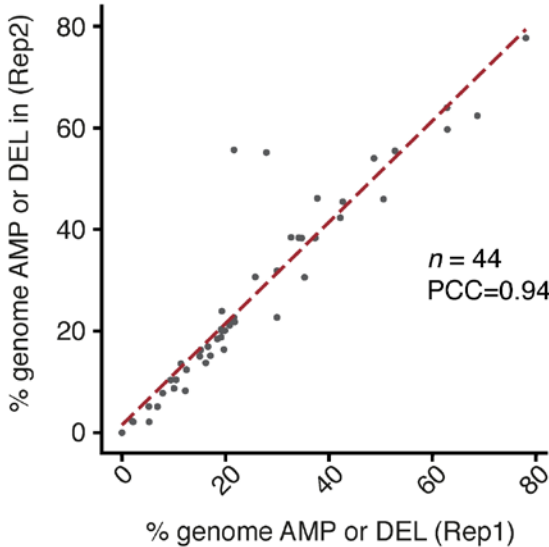


# Supplementary Figure 3

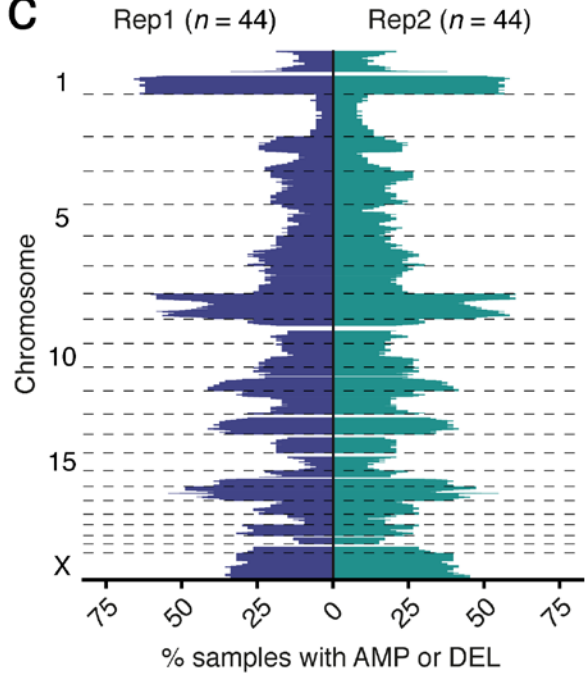
**a**



**b**

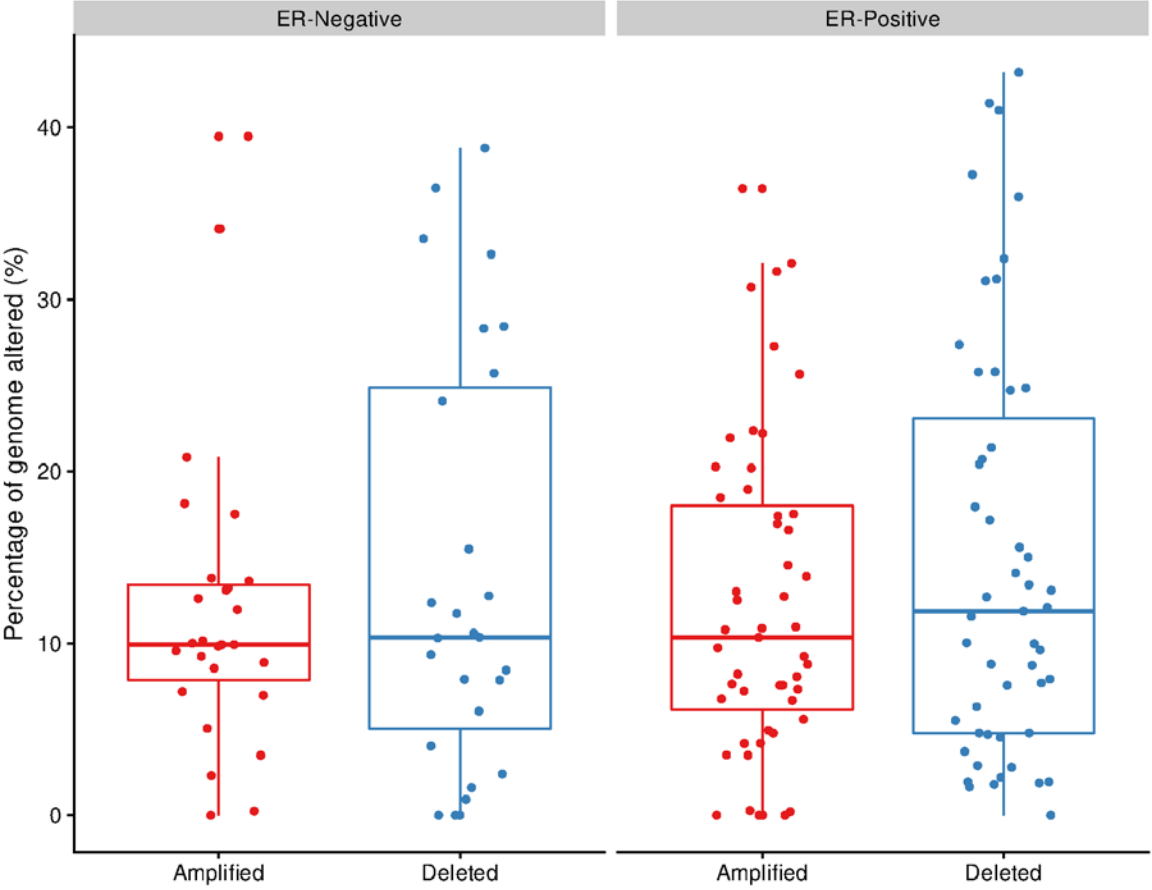


**c**

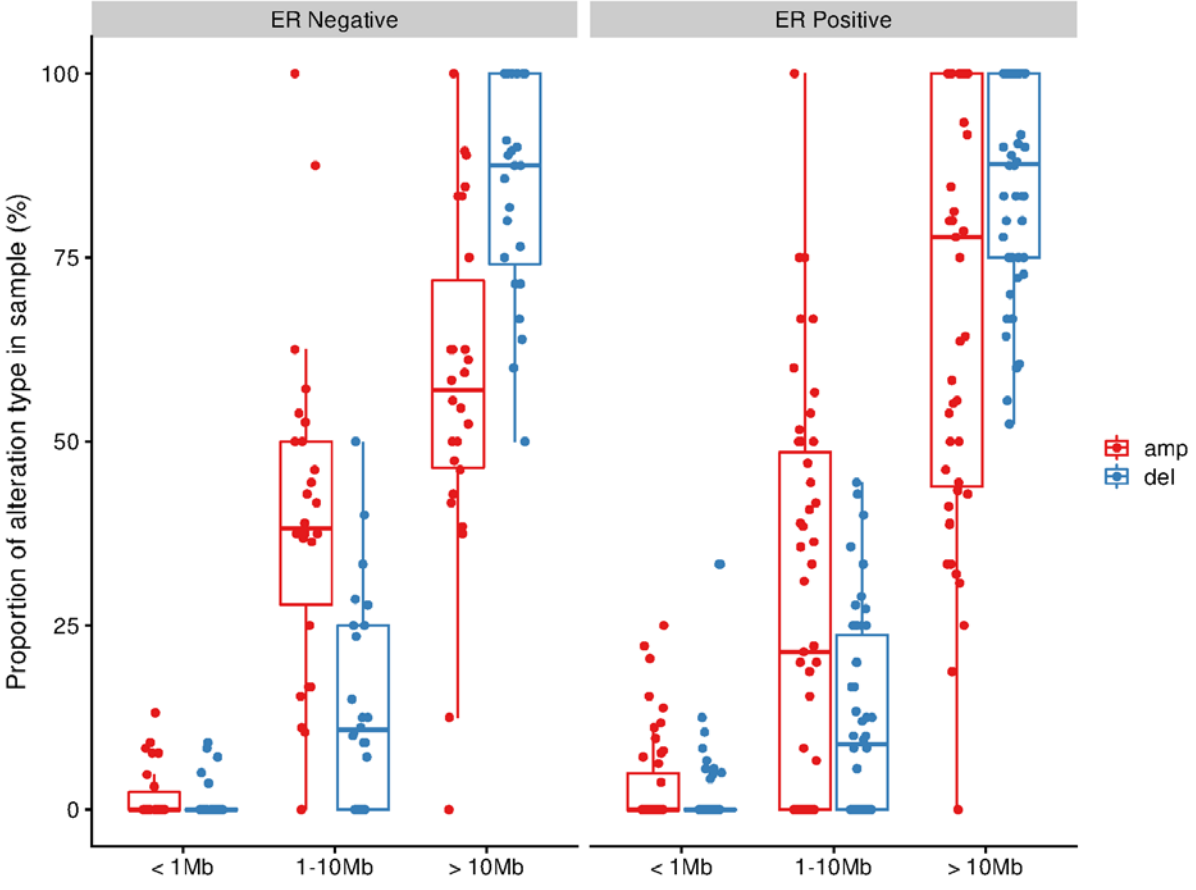


# Supplementary Figure 4

## A



## B



**Supplementary Table 1.** Cell densities (cells/mm<sup>2</sup>) of the different immune cell subsets as derived from the multiplex fluorescent IHC method (n=79)

<b>Cell type</b>	<b>Median</b>	<b>Mean</b>	<b>SD</b>	<b>p-value<sup>§</sup></b>
<b><i>CD4+ cells</i></b>				
intra-tumoral	271	1104	1967	0.4541
stromal	141.8	909	1860	
<b><i>CD8+ cells</i></b>				
intra-tumoral	23.87	91.46	228.2	<0.0001 ****
stromal	81.3	278.8	510.9	
<b><i>CD68+ cells</i></b>				
intra-tumoral	59.32	114.2	136.1	<0.0001 ****
stromal	146.9	278.3	329.1	
<b><i>FOXP3+ cells</i></b>				
intra-tumoral	0	6.9	16.84	<0.0001 ****
stromal	0	49.56	95.97	
<b><i>PD-1+CD4+ cells</i></b>				
intra-tumoral	285.8	976.4	1606	0.0432 *
stromal	86.87	779.6	1649	
<b><i>PD-1+CD8+ cells</i></b>				
intra-tumoral	22.28	83.8	222.7	<0.0001 ****
stromal	40.27	254.8	522.2	
<b><i>PD-1+CD68+ cells</i></b>				
intra-tumoral	52.44	117	197.1	<0.0001 ****
stromal	112.6	217.5	302.1	
<b><i>PD-1+FOXP3+ cells</i></b>				
intra-tumoral	0	0.75	6.2	0.0195
stromal	0	2.3	8.8	*
<b><i>PD-L1+CD4+ cells</i></b>				
intra-tumoral	0	38.52	320.5	0.0080 **
stromal	0	55.25	269.1	
<b><i>PD-L1+CD8+ cells</i></b>				
intra-tumoral	0	9.58	83.94	0.0840
stromal	0	12.45	63.07	
<b><i>PD-L1+CD68+ cells</i></b>				
intra-tumoral	0	1.25	5.63	0.0923
stromal	0	4.87	21.37	
<b><i>PD-L1+FOXP3+ cells</i></b>				
intra-tumoral	0	31.55	212.3	0.2221
stromal	0	2.88	16.22	

<sup>§</sup>Wilcoxon sign-rank test



**Supplementary Table 3.** Patient characteristics per manual TILs group (median cut-off)

	<b>Study cohort</b>	<b>High TILs</b>	<b>Low TILs</b>	<b>P value</b>
	<b>N = 89 (%)</b>	<b>N = 43 (%)</b>	<b>N = 46 (%)</b>	
<b>Age</b> Median (min – max)	51.1 (28.7 – 64.7)	50.8 (31.0 – 64.2)	51.8 (28.7 – 64.7)	0.895*
<b>Tumor Size mm</b> Median (min – max)	24 (2 – 150)	21 (2 – 121)	27.50 (9 – 150)	0.264*
<b>Positive lymph nodes</b> Median (min – max)	6 (1 – 35)	5 (1 – 35)	7.5 (1 – 16)	0.581*
<b>Grade</b>				
1	11 (12.4)	2 (4.7)	9 (19.6)	<b>&lt;0.001**</b>
2	31 (34.8)	9 (20.9)	22 (47.8)	
3	47 (52.8)	32 (74.4)	15 (32.6)	
<b>Estrogen Receptor</b>				
Positive	57 (64.0)	18 (41.9)	39 (84.8)	<b>&lt;0.001**</b>
Negative	32 (36.0)	25 (58.1)	7 (15.2)	
<b>Progesterone Receptor</b>				
Positive	40 (44.9)	15 (34.9)	25 (54.3)	0.065**
Negative	49 (55.1)	28 (65.1)	21 (45.7)	
<b>HER2</b>				
Positive	27 (30.3)	14 (32.6)	13 (28.3)	0.203**
Negative	60 (67.4)	28 (65.1)	32 (69.6)	
Unknown	2 (2.2)	1 (2.3)	1 (2.2)	

\* Mann Whitney U test

\*\* Pearson's  $\chi^2$



**Supplementary Table 4.** Definition of variables derived from digital tumor- infiltrating lymphocytes (TILs) evaluation

<b>Variable</b>	<b>Definition</b>
<b>eTILs%</b>	100* TILs/Sum of Tumor Cells and TILs
<b>etTILs%</b> ( <i>total</i> )	100* TILs/ Sum of TILs, Tumor cells, fibroblast and others
<b>esTILs%</b> ( <i>stromal</i> )	100* TILs/Sum of TILs, fibroblast and others adjacent to tumor
<b>eaTILs</b> ( <i>cell density</i> )	Sum of TILs/Sum of areas of tumor region analyzed ( <b>mm<sup>2</sup></b> )
<b>easTILs%</b>	TILs Cell Area /Stroma Area*100
<b>Manual/visual stromal TILs</b> ( <i>sTIL</i> )	The percentage (%) of tumour stroma covered by infiltrating lymphocytes (International TILs Working Group)

**Supplementary Table 5.** List of antibodies used for the multiplex fluorescent IHC method

<b>Order</b>	<b>Antigen retrieval*</b>	<b>Marker</b>	<b>Clone</b>	<b>Host Species</b>	<b>Dilution</b>	<b>Company</b>
1.	pH9	CD68	PG-M1	Mouse	1:400	Agilent
2.	pH6	PD-1	NAT105	Mouse	1:100	Abcam
3.	pH6	CD4	4B12	Mouse	1:100	Agilent
4.	pH6	CD8a	C8/144B	Mouse	1:200	Thermo Fisher
5.	pH6	PD-L1	SP142	Rabbit	1:400	Abcam
6.	pH6	FoxP3	D6O8R	Rabbit	1:300	Cell Signaling
7.	pH6	PanCK	C-11	Mouse	1:500	Abcam
		Cytokeratin	AE1/AE3	Mouse	1:400	Dako
		E-cahderin	36/E	Mouse	1:2000	BD Biosciences
8.	-	DAPI	-	-	-	Perkin Elmer

\*Antigen retrieval performed in an automated Leica Bond RX<sup>m</sup> Research Stainer at 95 °C, 20min.

The ImmPRESS® HRP Anti-Mouse IgG (Peroxidase) (Cat. No: MP-7402-50) and Anti-Rabbit IgG (Peroxidase) Polymer Detection Kits, made in Horse (Cat No: MP-7401-50) (Vector Laboratories) were used as secondary antibodies