

Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- | | | |
|-------------------------------------|-------------------------------------|--|
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | The statistical test(s) used AND whether they are one- or two-sided
<i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i> |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | A description of all covariates tested |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
<i>Give P values as exact values whenever suitable.</i> |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> | For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> | Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated |

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

Fiji - used for analysis of fluorescence microscopy images
Olympus FluoView - used for batch Z projection of confocal images
ImageLab 6.0 - used for quantification of immunoblot data
SerialEM - used to collect cryo-EM data
GraphPad Prism 8 - data recording, analysis, and plot generation

Data analysis

cisTEM - used to process cryo-EM data. Version 2.0.0 (available on github)
Coot 0.9.3 - as integrated into CCPEM used for manual model building (open source)
Phenix 1.19.1-4122 used for model refinement (open source)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Structural models have been deposited in the PDB under the accession codes 7LS2 (class I) and 7LS1 (class II). Cryo-EM maps have been deposited to the EMDB under the accession codes EMD-23501 (class I) and EMD-23500 (class II). Single-cell RNA-seq data generated by Usoskin et al. are accessible through Gene

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	Sample sizes for cell imaging experiments was similar to that of prior publications (e.g. DOI 10.1523/JNEUROSCI.4155-08.2008, DOI 10.1016/j.neulet.2013.09.048). Sample sizes for immunoblot and biochemical experiments was similar to that of prior publications (e.g. DOI 10.1038/s41467-021-21637-y, DOI 10.1038/s41467-020-15412-8, DOI 10.1371/journal.pbio.3000780) For single particle cryo-EM, we collected a dataset encompassing 2,995 movies that yielded a dataset of 193,796 ribosome coordinates. A high-resolution reconstruction of a ribosome requires ~5000 images of a ribosome (particles). We collected sufficient data to determine the high-resolution structures of each eEF2-containing class (23,297 and 11,878, for classes 1 and 2, respectively).
Data exclusions	No data were excluded from analyses
Replication	We define technical replicates as those performed in parallel from the same original biological sample, and biological replicates as those performed using separate biological samples. For confocal imaging experiments, several individual cells were measured from at least two biological replicates to achieve an appropriate n. Biological replicates for immunoblots from total cell lysates and ribosome isolations were conducted in parallel. All polysome profile experiments were performed with three biological replicates. Overlaid representative polysome profiles were conducted in parallel. Reported data for confocal imaging and polysome profiles were derived from successfully replicated experiments.
Randomization	Randomization was not relevant to this study, as no in vivo or human studies were conducted
Blinding	No data in this study derived from subjective assessments by the experimenter. As such, blinding was not relevant to this study

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

- | n/a | Involved in the study |
|-------------------------------------|---|
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Antibodies |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Eukaryotic cell lines |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology and archaeology |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Animals and other organisms |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Human research participants |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Clinical data |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Dual use research of concern |

Methods

- | n/a | Involved in the study |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Flow cytometry |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |

Antibodies

Antibodies used	Anti-Rck, Rb polyclonal (1:1000) MBL PD009 Lot 032 Anti-Rck, Mm monoclonal (1:500) SCBT 376433 Lot J0317 Anti-Peripherin, Ck polyclonal (1:1000) Novus NBP1-0543 Anti-eEF2K Rb polyclonal (1:400) Invitrogen PA5-22175 Lot UH2831084A Phalloidin-TRITC (1:200) Sigma P1951 Anti-p-eEF2 (T56), Rb polyclonal (1:1000) CST 2331S Lot 9 Anti-eEF2, Rb polyclonal (1:1000) CST 2332S Lot 7 Anti-SERBP1, Rb polyclonal Bethyl (1:1000) A303-938A Lot1 Anti-RPL5, Rb polyclonal Bethyl (1:1000) A303-933A Anti-RPS6, Rb monoclonal (1:1000) CST 2217s Lot 10
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Anti-ATF-4, Rb monoclonal (1:1000) CST 118155 Lot 4
 Anti-GAPDH, Rb polyclonal (1:10,000) Proteintech 10494
 Anti-PELO, Mm polyclonal (12µg per 30µl protein A/G beads) SCBT 393418 Lot B0315
 Gt-anti-Rb-Cy5 polyclonal (1:1000) Invitrogen A10523 Lot 1675037
 Gt-anti-Ck-AlexaFluor488 (1:1000) polyclonal Invitrogen A11039 Lot 1869581
 Gt-anti-Mm-Cy5 polyclonal (1:1000) Invitrogen A10524 Lot 1675775
 Gt-anti-Rb-peroxidase (1:10,000) Invitrogen 32460 Lot QG220833

Validation

Anti-Rck, Rb readily detects RCK in fixed mouse cells (e.g. DOI 10.1523/JNEUROSCI.0104-08.2008, DOI 10.1038/s41598-018-30805-y)
 Anti-Rck, Mm readily detects RCK in fixed human cells (e.g. DOI 10.1038/s41467-019-08548-9)
 Anti-Peripherin, Ck indicated by manufacturer for immunofluorescence detection of peripherin; specifically labels peripheral neurons in heterogenous primary cultures (e.g. DOI: 10.1111/bph.15646)
 Anti-eEF2K, Rb indicated by manufacturer for immunofluorescence detection of eEF2K in fixed cells
 Phalloidin-TRITC readily detects actin cytoskeleton of eukaryotic cells (e.g. DOI 10.1038/s41467-017-02449-5)
 Anti-p-eEF2 (T56), Rb readily detects phospho-eEF2 (Thr56) in immunoblot (e.g. DOI 10.3389/fnmol.2019.00097)
 Anti-eEF2, Rb readily detects eEF2 in immunoblot (e.g. DOI 10.3389/fnmol.2019.00097)
 Anti-SERBP1, Rb readily detects SERBP1 on immunoblots (e.g. DOI 10.1093/nar/gkaa1189)
 Anti-RPL5, Rb readily detects RPL5 on immunoblots (e.g. DOI 10.1158/0008-5472.CAN-18-2718)
 Anti-RPS6, Rb readily detects RPS6 on immunoblots (e.g. DOI 10.1038/s41467-019-11227-4)
 Anti-ATF-4, Rb readily detects ATF-4 on immunoblots (e.g. DOI 10.1038/s41467-019-08908-5)
 Anti-GAPDH, Rb readily detects GAPDH on immunoblots of mouse heart, brain, and skin tissue per manufacturer
 Anti-PELO, Mm readily binds PELO on immunoblots from mouse brain and thymus, and rat brain per manufacturer

Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)	U2-OS cells (ATCC), F11 (Sigma)
Authentication	These cell lines were directly ordered from ATCC or Sigma, but not tested
Mycoplasma contamination	Cell lines were tested for Negative for Mycoplasma by PCR and found to be negative
Commonly misidentified lines (See ICLAC register)	No commonly misidentified cell lines were used in this study

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals	Mouse (<i>Mus musculus</i>), Swiss-Webster, male, 4-6 weeks old Mouse, eEF2K KO, male, 4-6 weeks old
Wild animals	No wild animals were used in this study
Field-collected samples	No field-collected samples were used in this study
Ethics oversight	Institutional Animal Care and Use Committee of the University of Texas at Dallas protocol # 18-08

Note that full information on the approval of the study protocol must also be provided in the manuscript.