

Supplementary Materials for

Evolutionary epigenomic analyses in mammalian early embryos reveal species-specific innovations and conserved principles of imprinting

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Published 24 November 2021, *Sci. Adv.* 7, eabi6178 (2021)
DOI: 10.1126/sciadv.abi6178

The PDF file includes:

Figs. S1 to S17
Legends for tables S1 to S4
References

Other Supplementary Material for this manuscript includes the following:

Tables S1 to S4

Figure S1

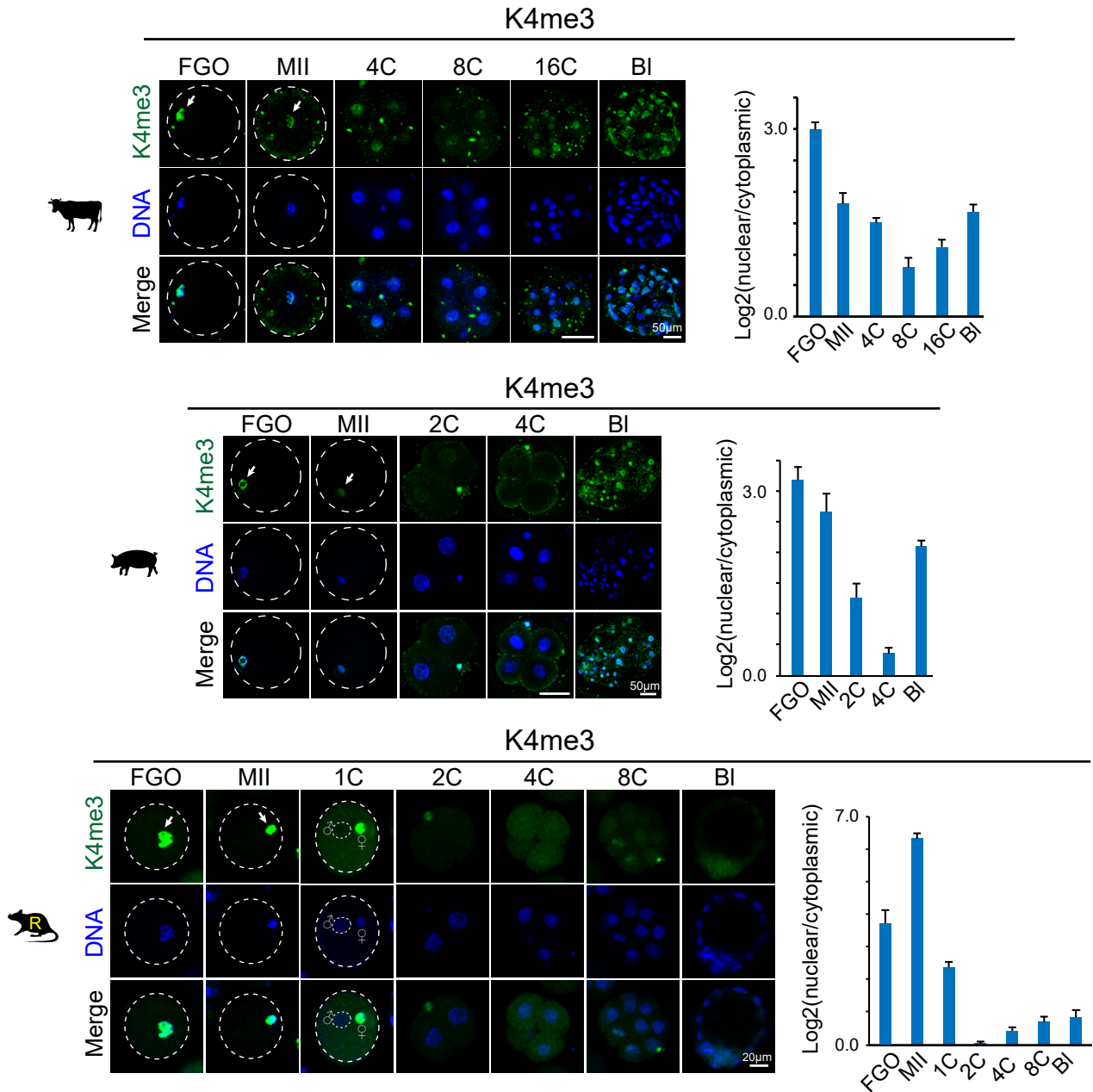


Figure S1. The dynamics of H3K4me3 in mammalian oocytes and preimplantation embryos. Immunofluorescent staining results showing the dynamics of H3K4me3 (K4me3) from FGO to blastocyst in bovine, porcine, and rat. The oocytes are circled by white dashed lines. White arrows indicate the nuclei and chromatin for FGOs and MII oocytes, respectively. In rat 1-cell, both female and male pronucleus (small white dashed lines) are shown. For bovine and porcine samples, bar=50 µm; for rat samples, bar=20 µm. The relative levels of H3K4me3 (log2 transformed nuclear/cytoplasmic fluorescence intensity, mean ± SEM) at each stage of the three species are shown on the right. FGO, full-grown oocyte; MII, metaphase II-stage oocyte; 1C, 1-cell embryo; 2C, late 2-cell embryo; 4C, 4-cell embryo; 8C, 8-cell embryo; 16C, 16-cell embryo; BI, blastocyst.

Figure S2

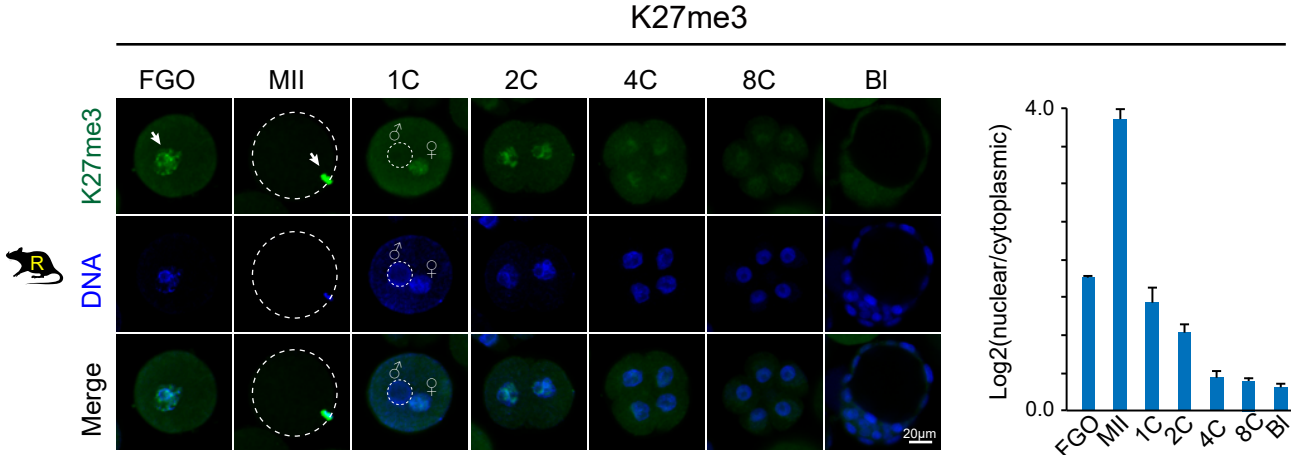
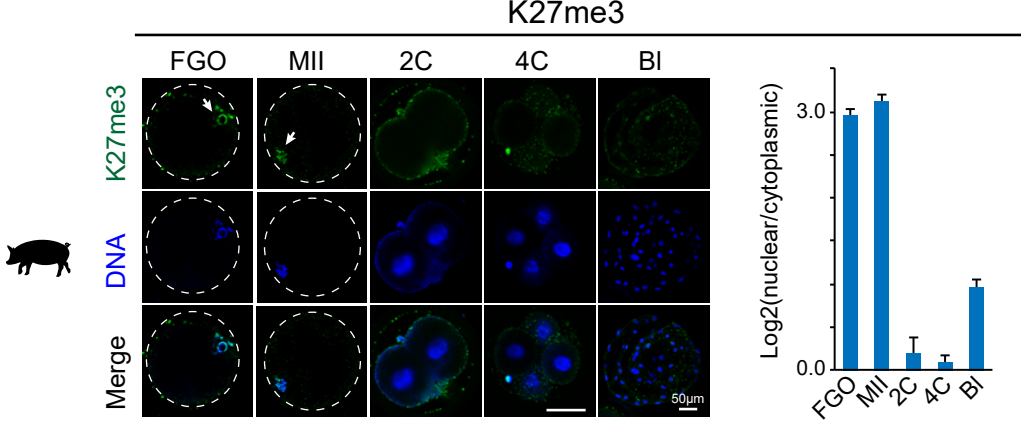
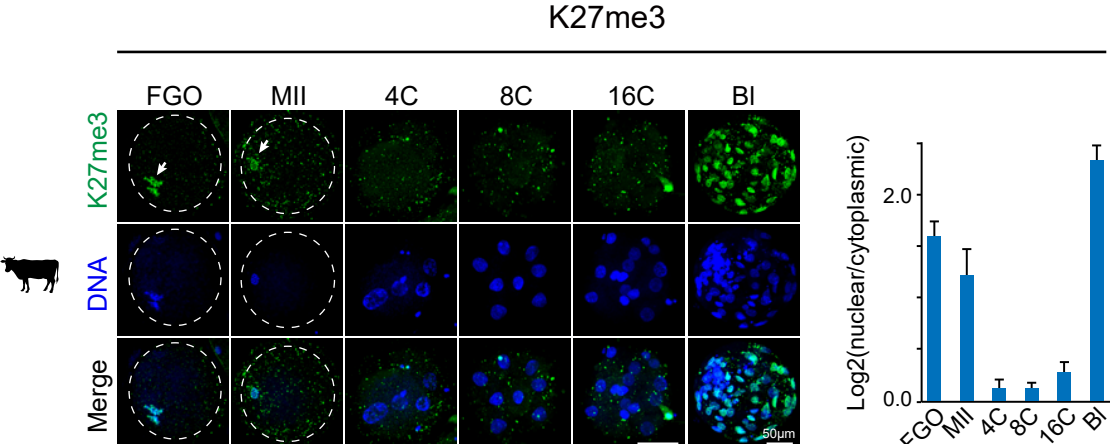
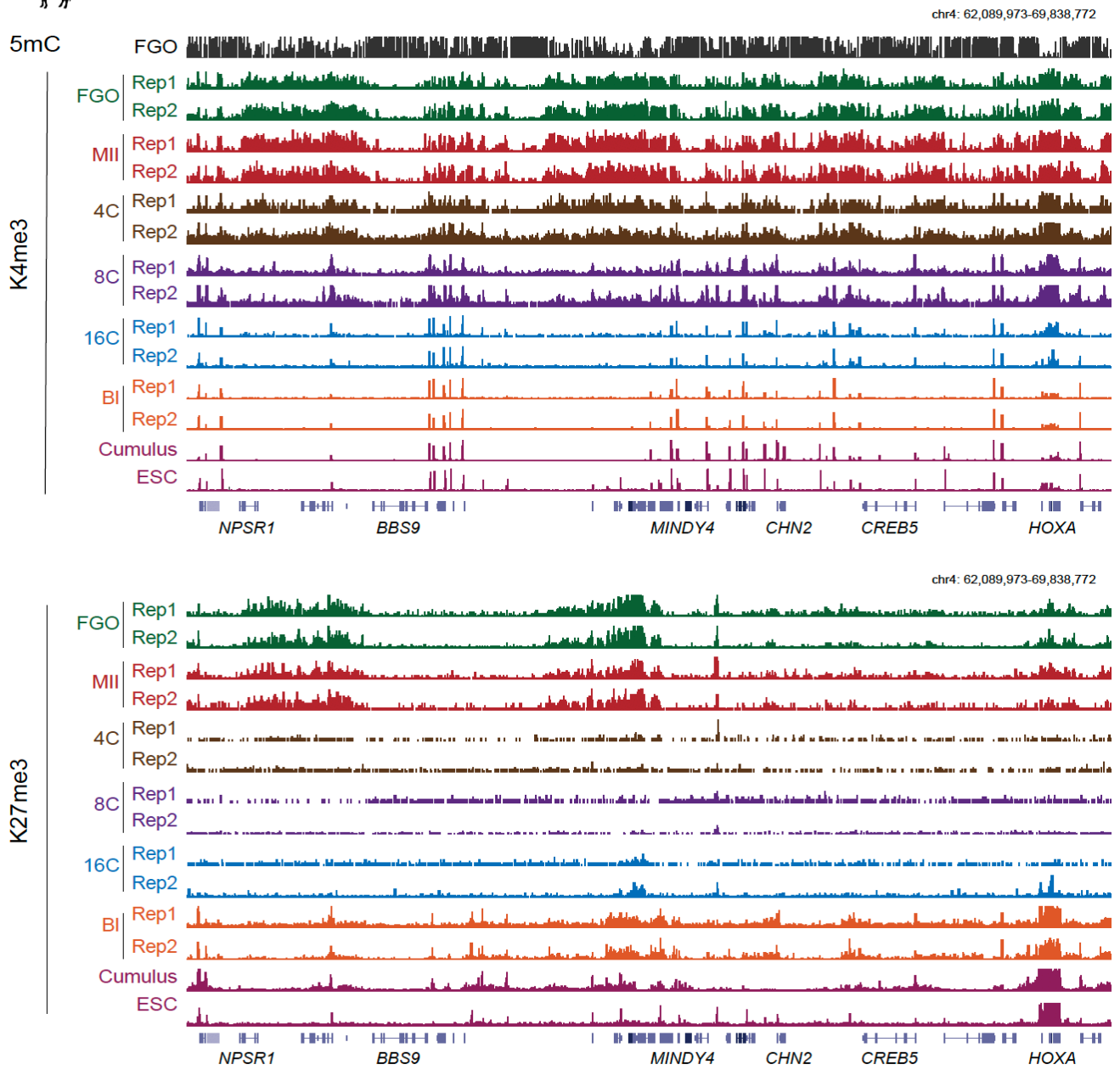


Figure S2. The dynamics of H3K27me3 in mammalian oocytes and preimplantation embryos. Immunofluorescent staining results showing the dynamics of H3K27me3 (K27me3) from FGO to blastocyst in bovine, porcine, and rat. The oocytes are circled by white dashed lines. White arrows indicate the nuclei or chromatins of FGOs or MII oocytes, respectively. In rat 1-cell, both female and male pronucleus (small white dashed lines) are shown. For bovine and porcine samples, bar=50 μm ; for rat samples, bar=20 μm . The relative levels of H3K27me3 (\log_2 transformed nuclear/cytoplasmic fluorescence intensity) at each stage of the three species are shown on the right. Data are shown as mean \pm SEM. FGO, full-grown oocyte; MII, metaphase II-stage oocyte; 1C, 1-cell embryo; 2C, late 2-cell embryo; 4C, 4-cell embryo; 8C, 8-cell embryo; 16C, 16-cell embryo; B1, blastocyst.

Figure S3

A



B

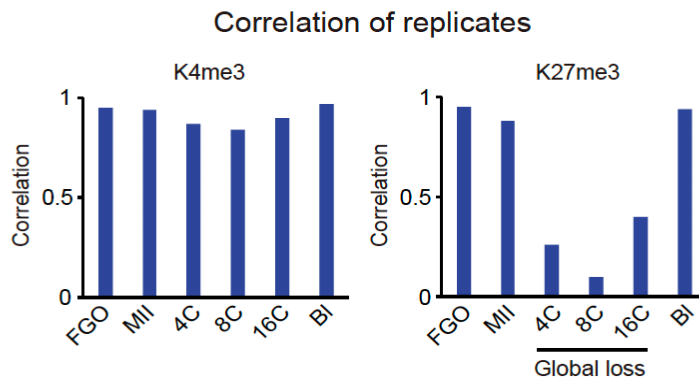
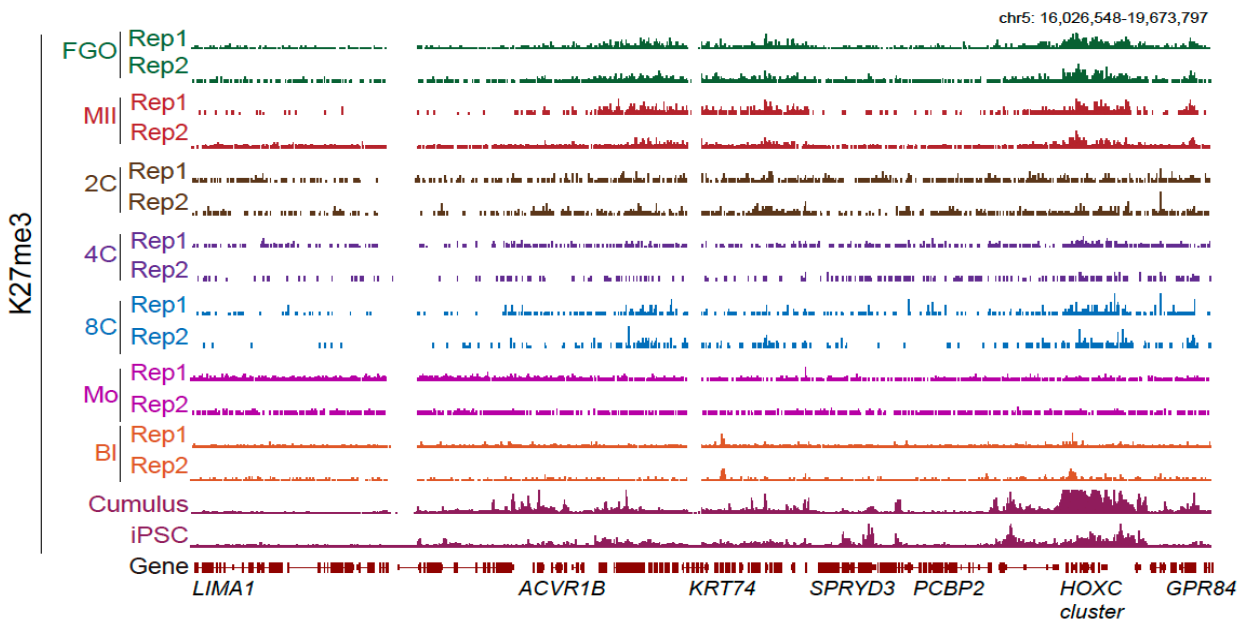
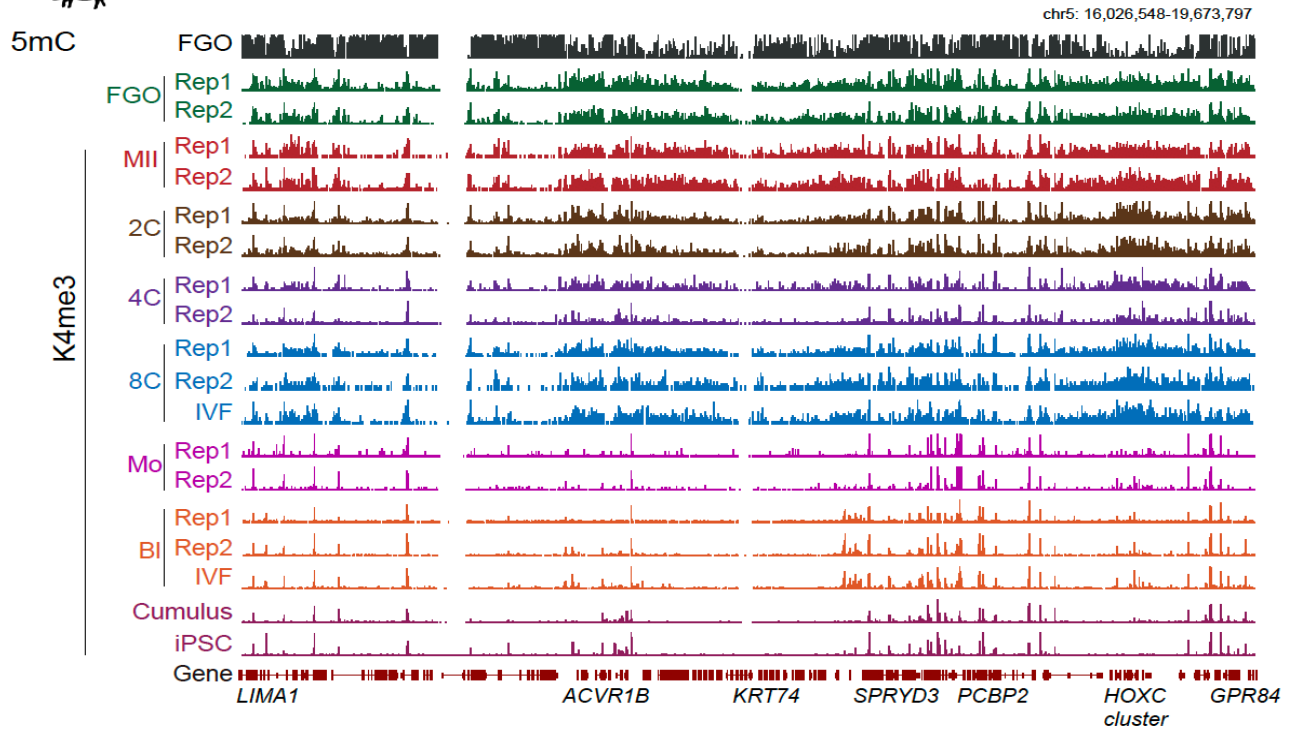


Figure S3. Global view of DNA methylation in FGOs and histone modifications in oocytes and early embryos in bovine. (A) UCSC genome browser views showing DNA methylation (5mC) in FGOs, two replicates of H3K4me3 (K4me3) and H3K27me3 (K27me3) in FGO, MII, 4C, 8C, 16C and Bl. Cumulus cell and ESC data (69) are also shown for comparison. (B) Barplots showing the Pearson correlation between replicates for H3K4me3 and H3K27me3 data in bovine. FGO, full-grown oocyte; MII, metaphase II-stage oocyte; 4C, 4-cell embryo; 8C, 8-cell embryo; 16C, 16-cell embryo; Bl, blastocyst. Note the stages showing lower correlation are those when H3K27me3 undergoes global loss (indicated).

Figure S4

A



B



Correlation of replicates

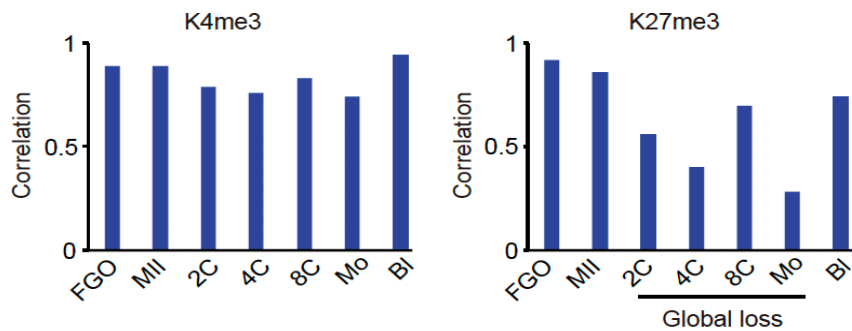
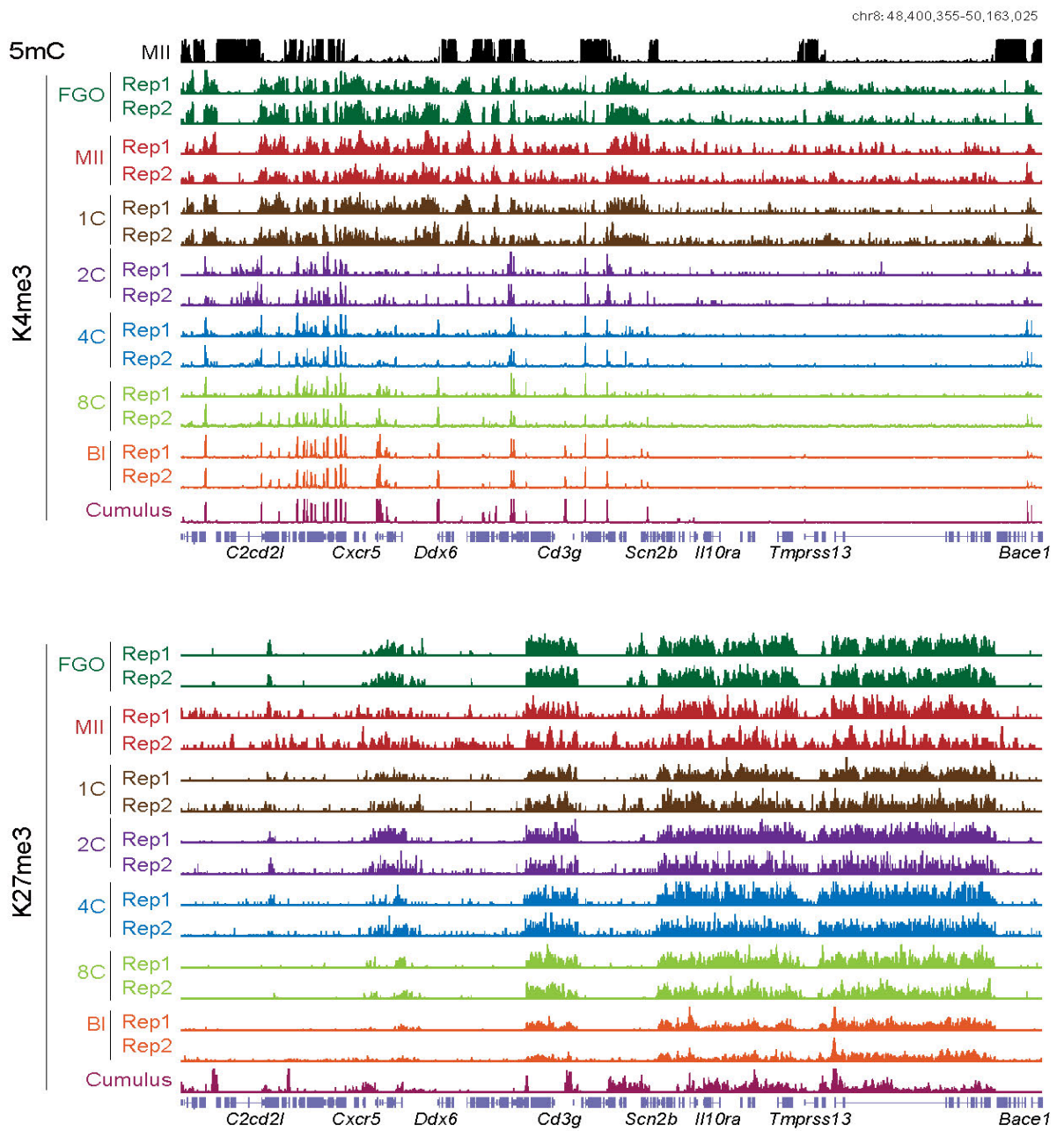


Figure S4. Global view of DNA methylation in FGOs and histone modifications in oocytes and early embryos in porcine. (A) UCSC genome browser views showing DNA methylation (5mC) in FGOs, two replicates of H3K4me3 (K4me3) and H3K27me3 (K27me3) in FGO, MII, 2C, 4C, 8C, Mo and Bl. Cumulus cell and iPSC data (70) are also shown for comparison. H3K4me3 in *in vitro* fertilized (IVF) 8C and blastocyst are also shown. (B) Barplots showing the Pearson correlation between replicates for H3K4me3 and H3K27me3 data in porcine. FGO, full-grown oocyte; MII, metaphase II-stage oocyte; 2C, late 2-cell embryo; 4C, 4-cell embryo; 8C, 8-cell embryo; Mo, morula, Bl, blastocyst. Note the stages showing lower correlation are those when H3K27me3 undergoes global loss (indicated).

Figure S5

A



B



Correlation of replicates

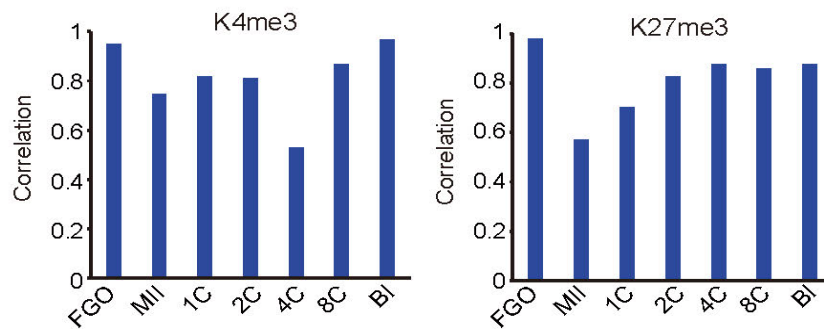


Figure S5. Global view of DNA methylation in MII oocytes and histone modifications in oocytes and early embryos in rat. (A) UCSC genome browser views showing DNA methylation (5mC) in MII oocytes (26), two replicates of H3K4me3 (K4me3) and H3K27me3 (K27me3) in FGO, MII, 1C, 2C, 4C, 8C and Bl. Cumulus cell data are also shown for comparison. (B) Barplots showing the Pearson correlation between replicates for H3K4me3 and H3K27me3 data in rat. FGO, full-grown oocyte; MII, metaphase II-stage oocyte; 1C, 1-cell embryo; 2C, late 2-cell embryo; 4C, 4-cell embryo; 8C, 8-cell embryo; Bl, blastocyst.

Figure S6

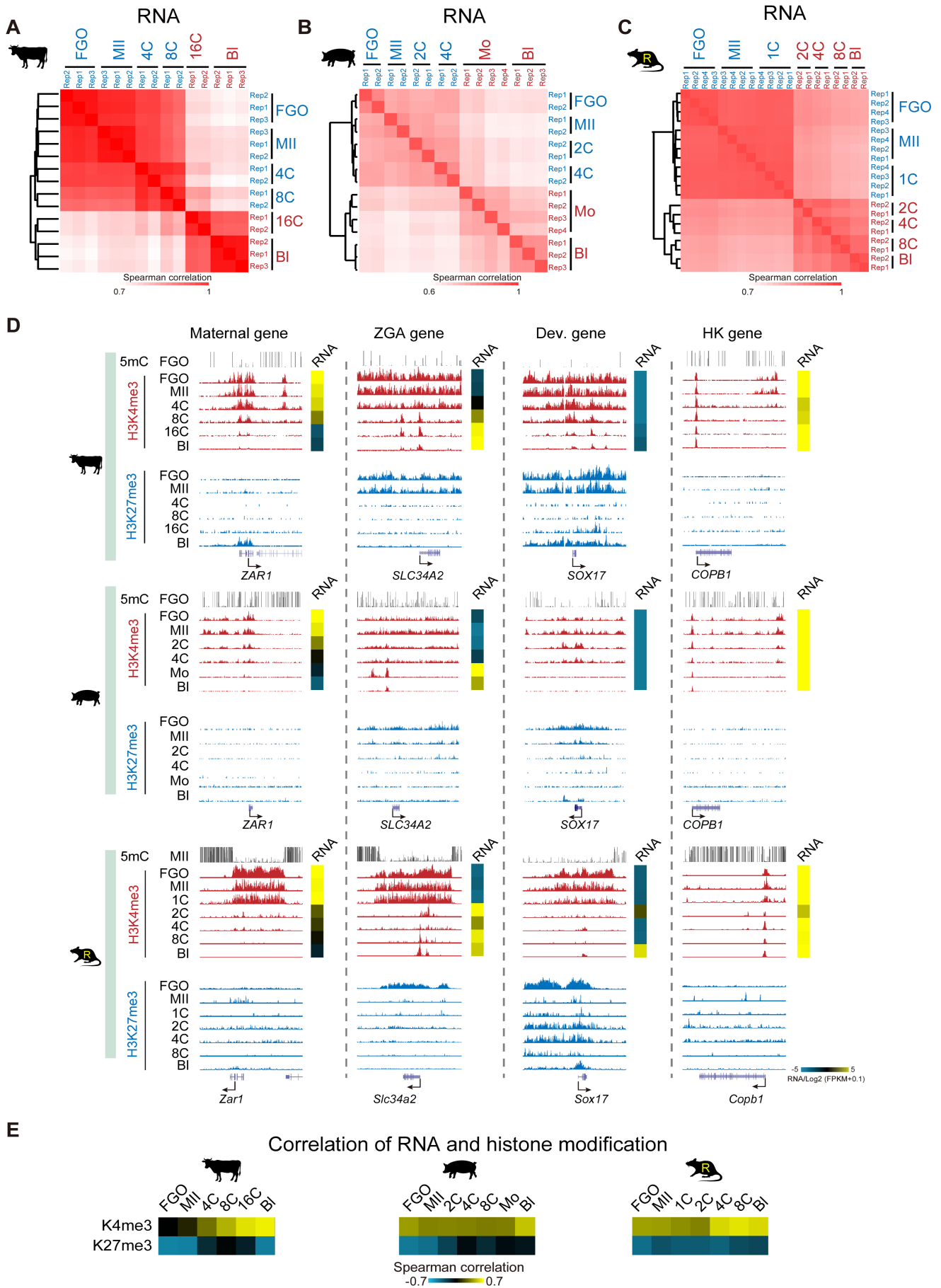


Figure S6. Global view of RNA-seq data in mammalian oocytes and early embryos. (A) Heatmap showing the pairwise Spearman correlation between replicates of the RNA-seq data among all development stages in bovine. The cluster on the left shows the relationship of all development stages. Blue, the oocyte and pre-ZGA stages; red, the post-ZGA stages. (B and C) Similar analysis as a for porcine (B) and rat (C). (D) Snapshots from UCSC genome browser showing the distribution of DNA methylation (5mC), H3K4me3 (K4me3) and H3K27me3 (K27me3) signals at representative gene loci in the oocytes and pre-implantation embryos of bovine, porcine and rat. Gene expression is also shown by heatmaps (\log_2 transformed FPKM+0.1). ZGA, zygotic genome activation; Dev., developmental; HK, housekeeping gene. (E) Heatmaps showing the Spearman correlation between promoter H3K4me3 (K4me3) or H3K27me3 (K27me3) and gene expression across all genes. FGO, full-grown oocyte; MII, metaphase II-stage oocyte; 1C, 1-cell embryo; 2C, late 2-cell embryo; 4C, 4-cell embryo; 8C, 8-cell embryo; 16C, 16-cell embryo; Mo, morula; Bl, blastocyst; ICM, inner cell mass.

Figure S7

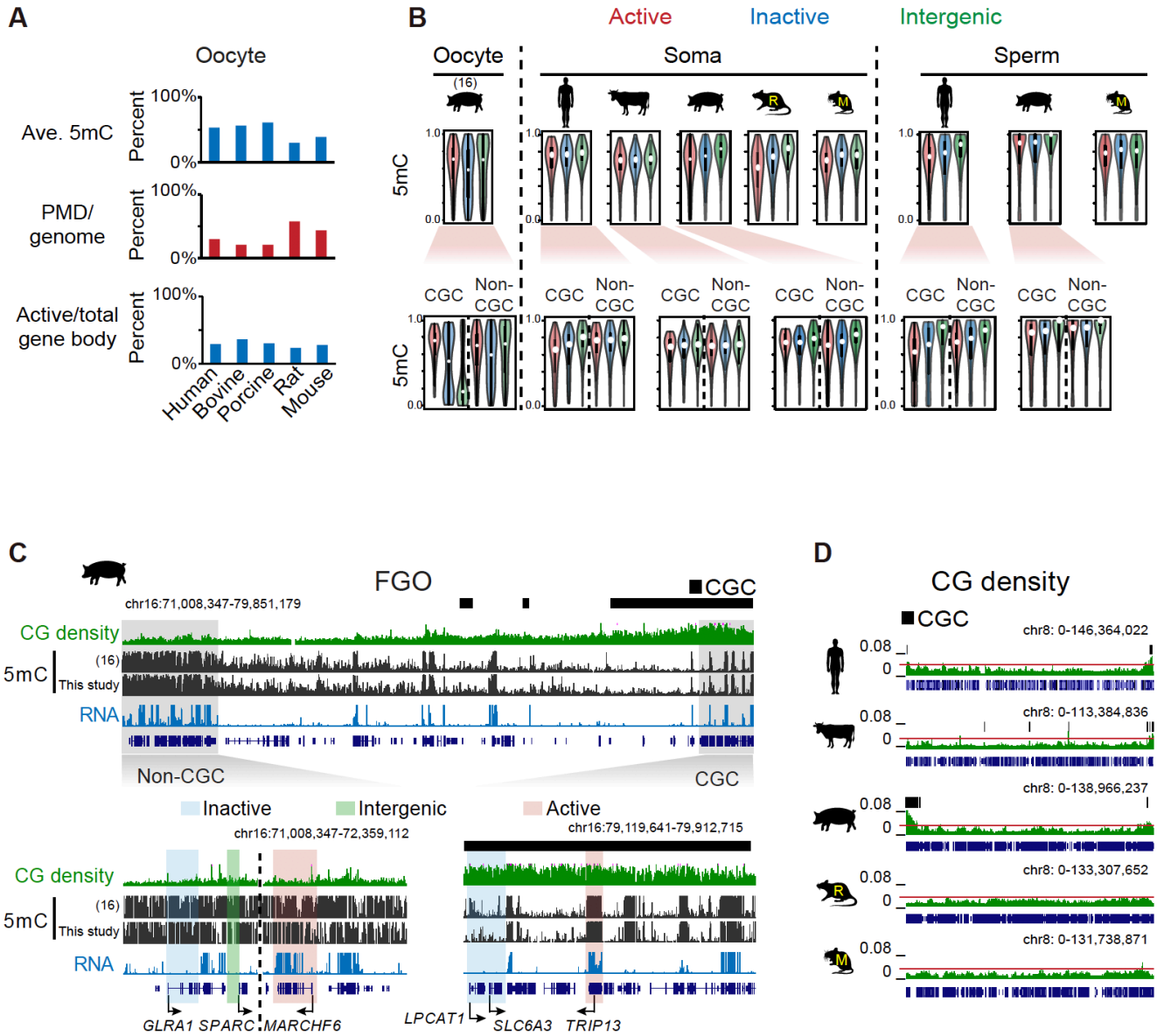
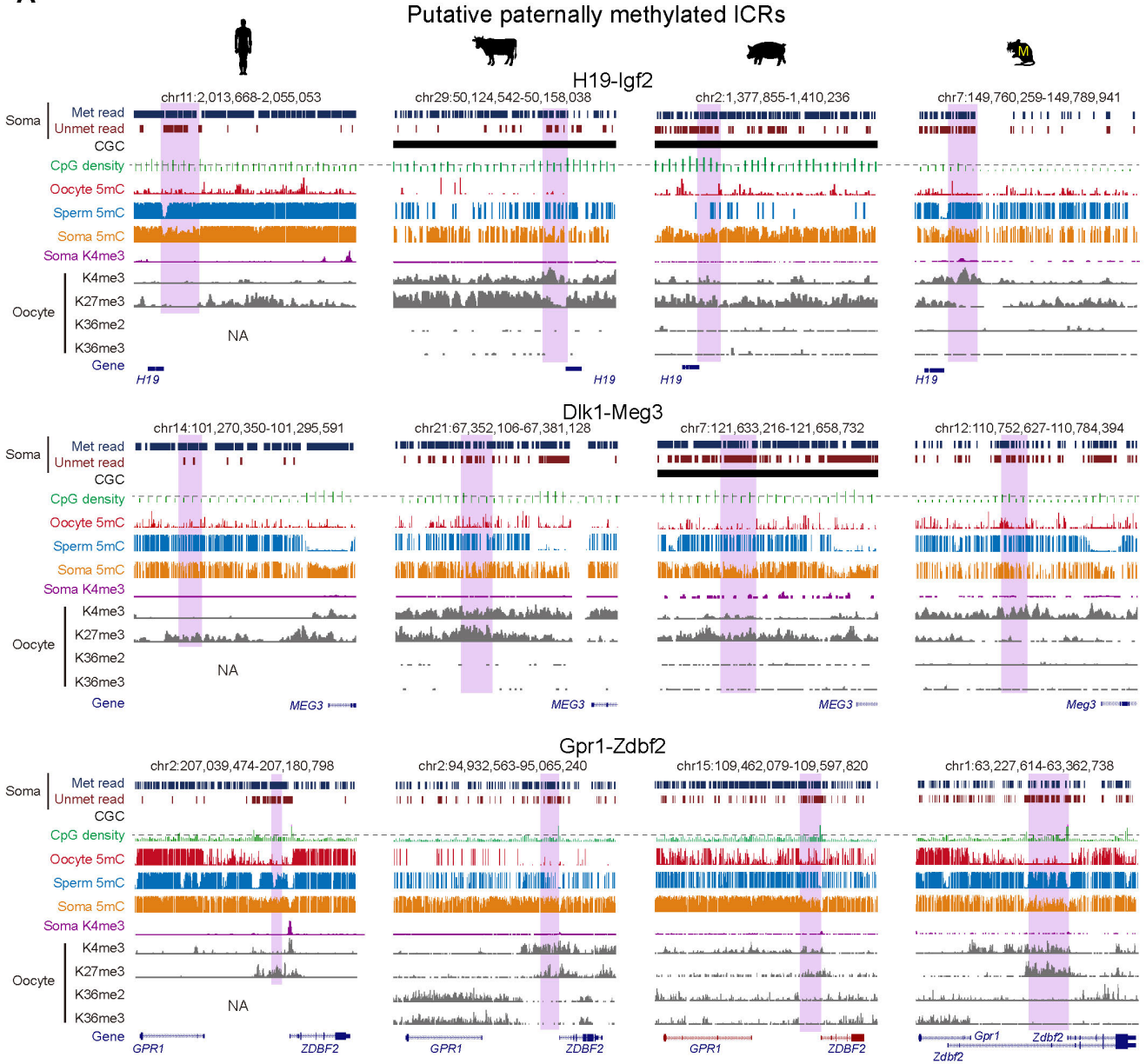
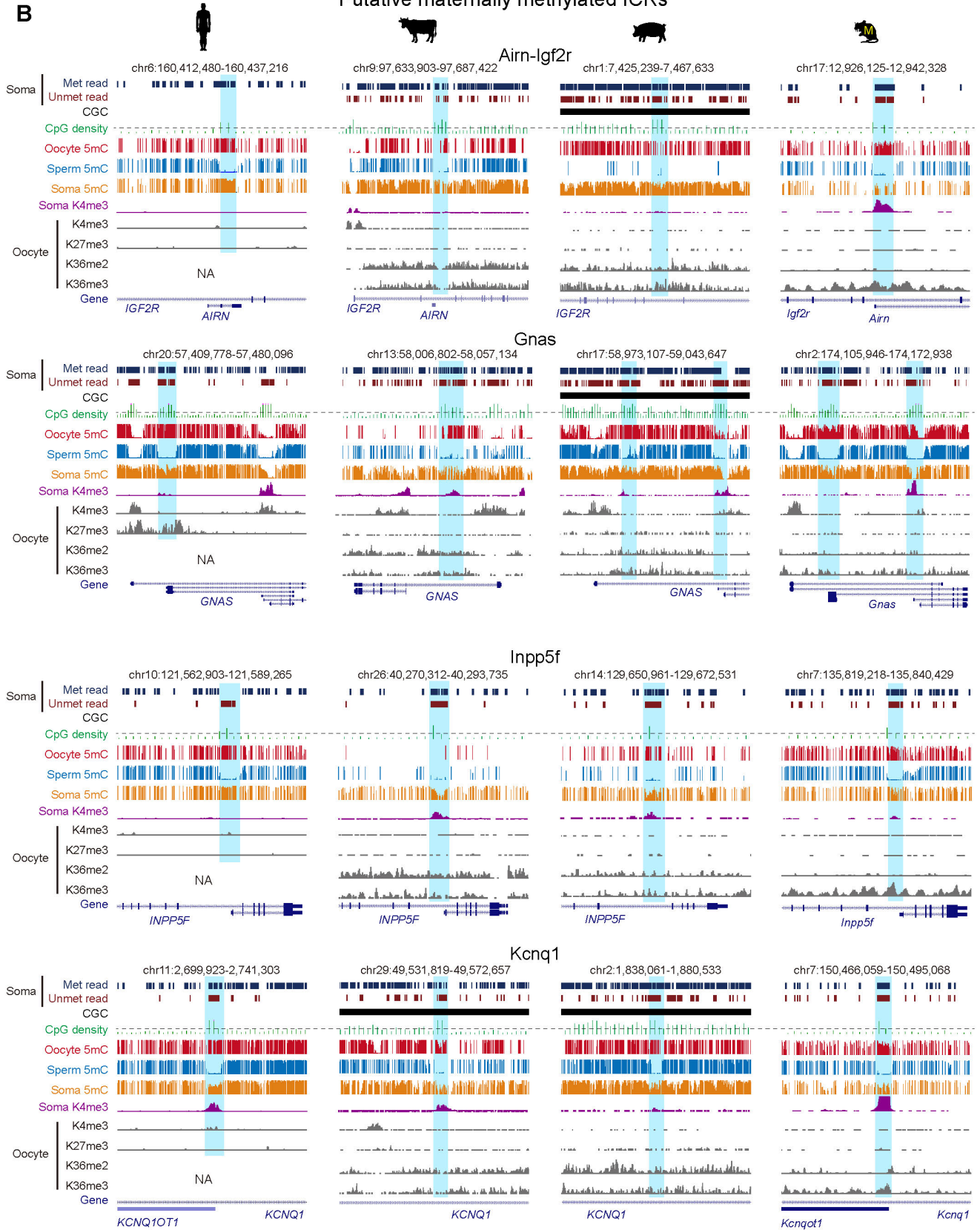


Figure S7. DNA methylation and CpG continents (CGCs) in mammalian oocytes. (A) Barplots showing the global DNA methylation (5mC) levels (top), PMD coverages (middle) and the percentages of active/total gene body (bottom) in the oocytes of human, bovine, porcine, rat and mouse. (B) Top, violin plots showing the DNA methylation levels in active gene bodies (red), inactive gene bodies (blue), and intergenic regions (green) in porcine oocytes, somatic tissues (Soma) in the five species, and sperm in human, porcine and mouse. Bottom, similar analyses were conducted for CGC and non-CGC regions in corresponding tissues/cells (top) in human, bovine and porcine. Data resources used in this analysis: porcine MII oocytes (16); human liver (73); bovine lung (74); rat left ventricle (75); mouse cerebellum (76); human sperm (24); porcine sperm (16); mouse sperm (81). (C) The UCSC genome browser views showing DNA methylation states in CGC (right) and non-CGC (left) regions in porcine FGOs. Active gene bodies, inactive gene bodies and intergenic regions are shaded. Data from (16) are included as a positive control. (D) The UCSC genome browser views showing the CG density across the entire chromosome (chr8) among the five species. CGCs are shown. The red line indicates the cutoff (0.03) for CGCs.

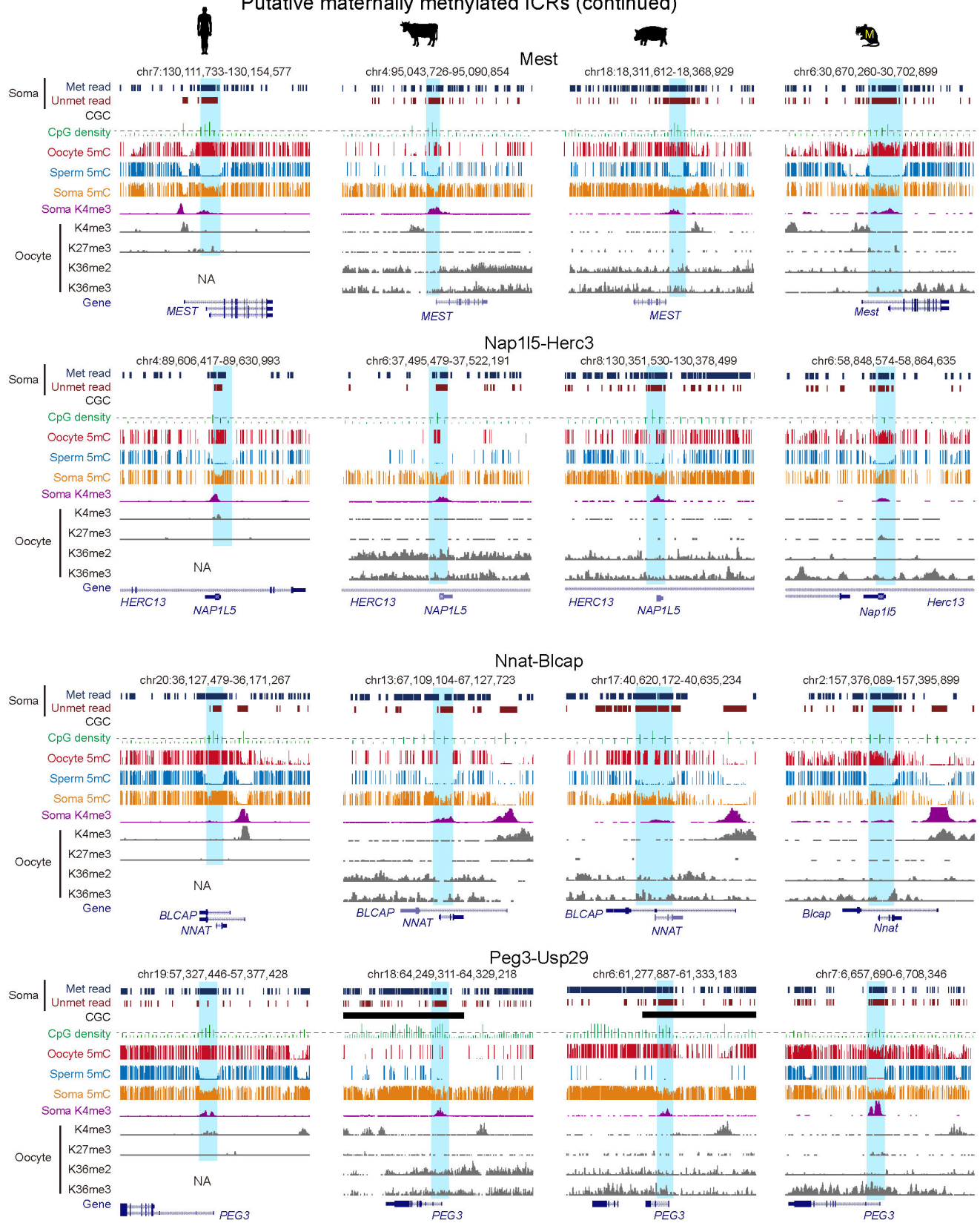
Figure S8

A

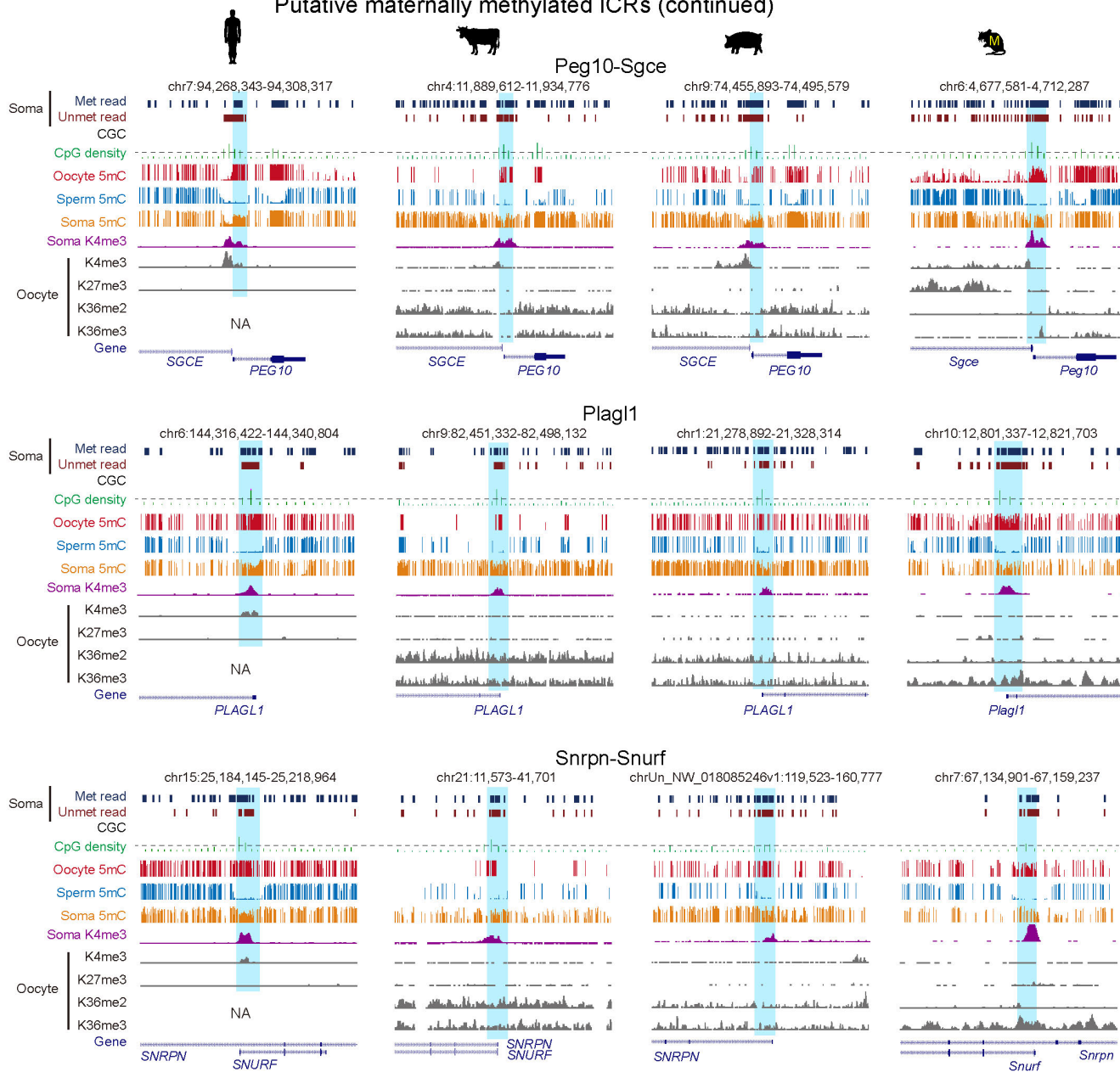


B**Putative maternally methylated ICRs**

Putative maternally methylated ICRs (continued)



Putative maternally methylated ICRs (continued)



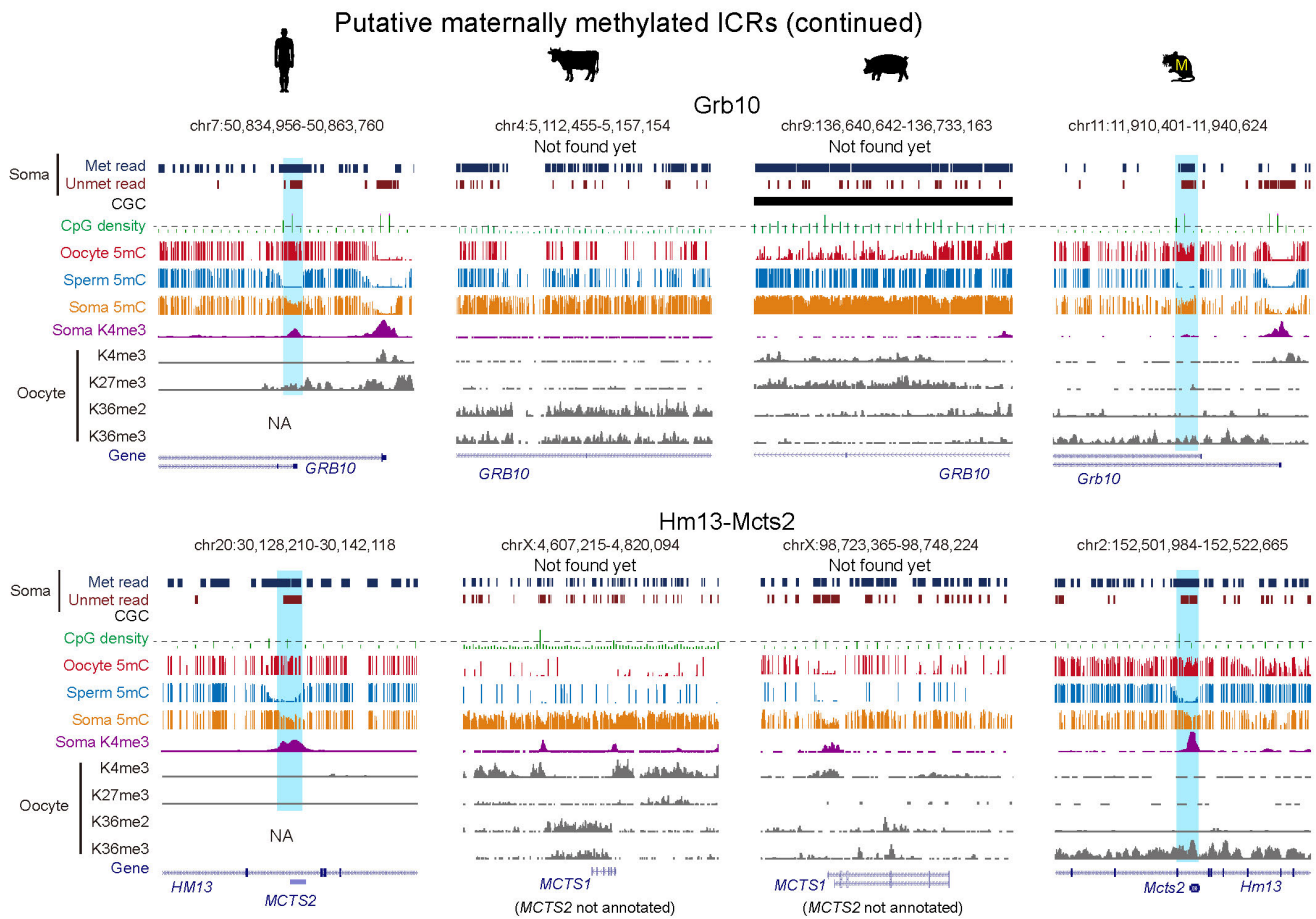
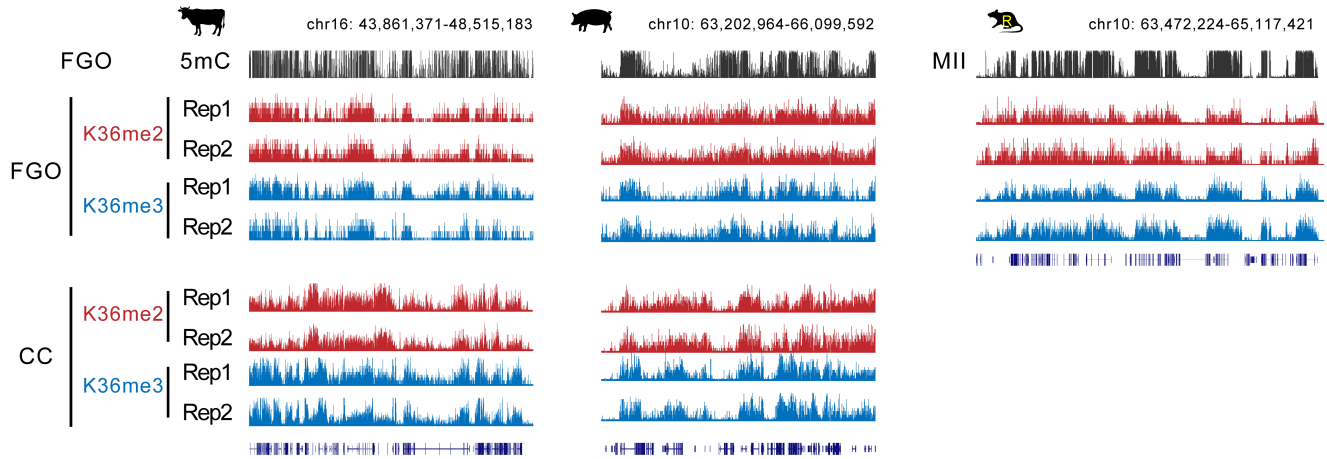


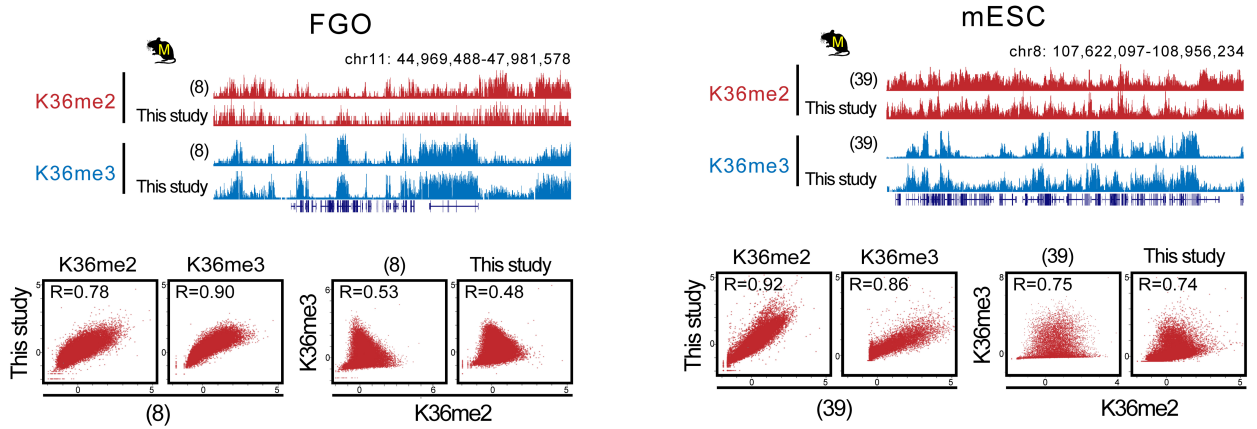
Figure S8. The putative paternal and maternal ICRs and their relationship with CGCs in bovine and porcine. The UCSC genome browser view showing the putative paternal (A, shaded in purple) and maternal (B, shaded in light blue) ICRs in bovine and porcine. Only germline ICRs conserved in mouse and human were considered. ICR positions for human and mouse were obtained from previous studies and related data were independently processed in this study (24, 35). The *GRB10* and *HM13-MCTS2* ICRs in bovine and porcine were not found yet. Only *MCTS1*, but not *MCTS2*, is annotated and is shown in bovine and porcine. CGC, CpG density (1 kb bin), DNA methylation (5mC) in oocyte, sperm and somatic tissues (Soma) (human: liver (73); bovine: lung (74); porcine: muscle (78); mouse: liver (76)), H3K4me3 (K4me3), H3K27me3 (K27me3), H3K36me2 (K36me2) and H3K36me3 (K36me3) in oocyte around these ICRs are shown. The dashed lines for CG density indicate the cutoff of 0.03 (100 kb bin) used for calling CGCs. Met read, reads with all CpGs (at least 3) methylated; Unmet read, reads with all CpGs (at least 3) unmethylated (human: liver; bovine: lung; porcine: muscle; mouse: liver).

Figure S9

A



B



C

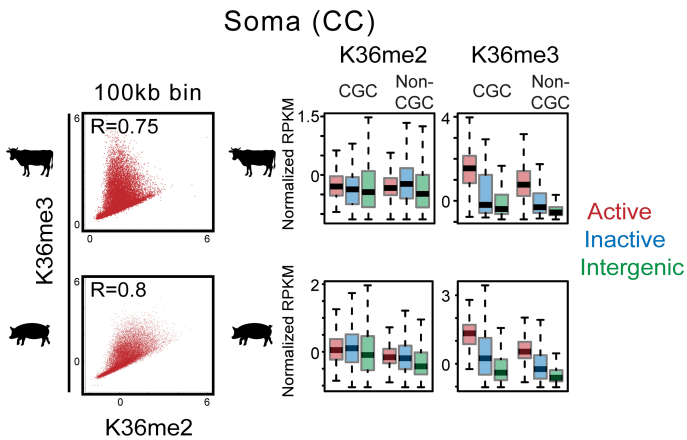
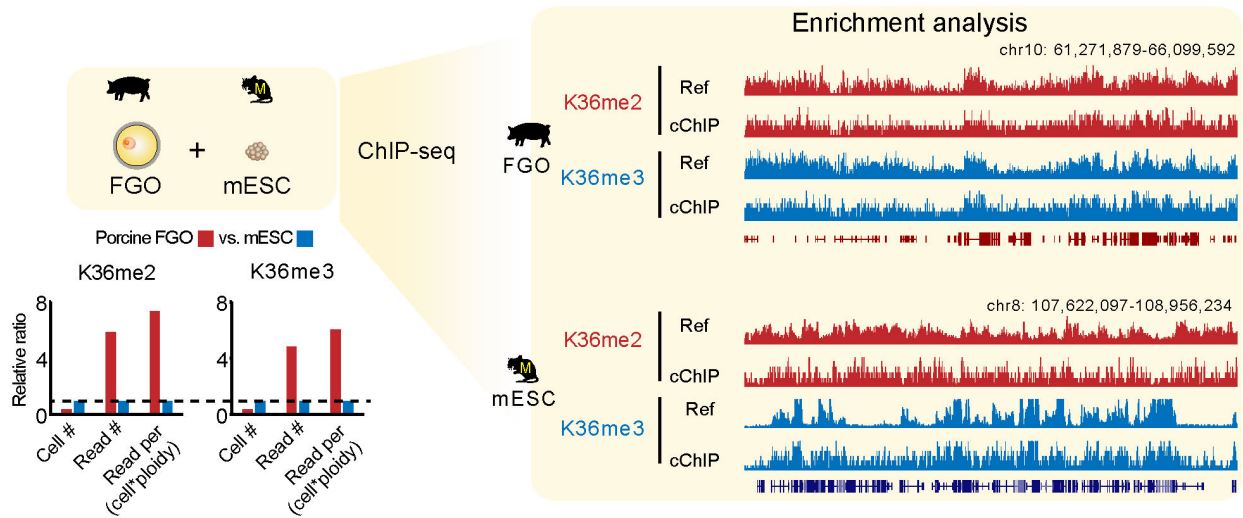


Figure S9. H3K36me2 and H3K36me3 in the oocytes and somatic cumulus cells (CCs) across mammals. (A) UCSC genome browser views showing DNA methylation (5mC) in oocytes, two replicates of H3K36me2 (K36me2) and H3K36me3 (K36me3) in FGOs of bovine, porcine and rat. H3K36me2 and H3K36me3 in cumulus cells (CCs) for bovine and porcine are also shown. (B) The UCSC genome browser views (top) and scatterplots (bottom) showing the comparison of H3K36me2 and H3K36me3 patterns in mouse FGOs in this study and a previous study (8), or in mESCs in this study and a previous study (39). Scatterplots comparing H3K36me2 and H3K36me3 from the same study are also shown. Data from previous studies were downloaded and processed independently in this study. (C) Scatterplots comparing H3K36me2 and H3K36me3 in CCs for bovine and porcine (left). Boxplots showing the enrichment of H3K36me2 and H3K36me3 in active gene bodies (red), inactive gene bodies (blue), and intergenic regions (green) in CGCs and non-CGCs in CCs for bovine and porcine.

Figure S10

A



B

Porcine FGO vs. mESC

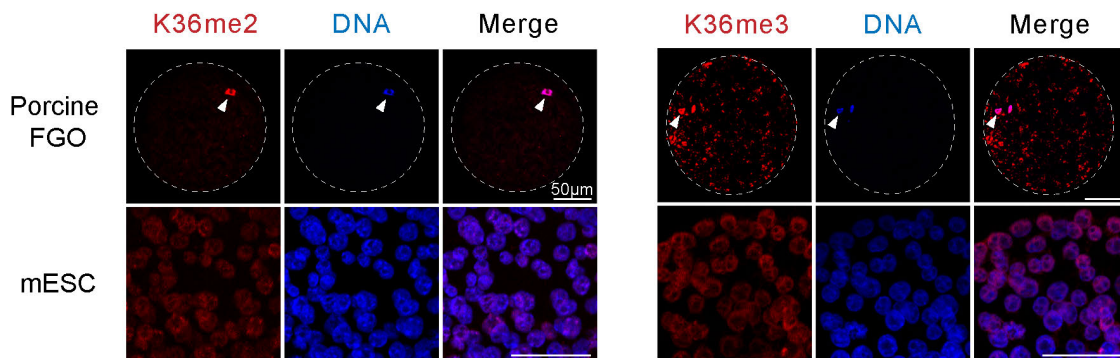


Figure S10. Validation of H3K36me2 and H3K36me3 in porcine oocytes. (A) Competitive and calibrated ChIP-seq (cChIP) result of H3K36me2 (K36me2) and H3K36me3 (K36me3) for porcine FGOs, with mESCs as references. A total of 100 FGOs (2n, 4C) and 250 mESCs (2n, 2C) were mixed and subjected to STAR ChIP-seq for H3K36me2 and H3K36me3. Sequencing reads were mapped separately to mouse and porcine genomes. Barplots showing the comparison of cell numbers, read numbers, read per (cell number*ploidy) between porcine FGOs and mESCs (set as 1). UCSC genome browser views (right) showing the comparison of H3K36me2 and H3K36me3 patterns in porcine FGOs and mESCs obtained from pure samples (Ref) or mixed samples (cChIP). (B) Immunofluorescent staining showing the abundance of H3K36me2 and H3K36me3 in porcine FGOs and mESCs. The oocytes are circled by white dashed lines. White arrows indicate the nuclei of porcine FGOs. Bar=50 µm.

Figure S11

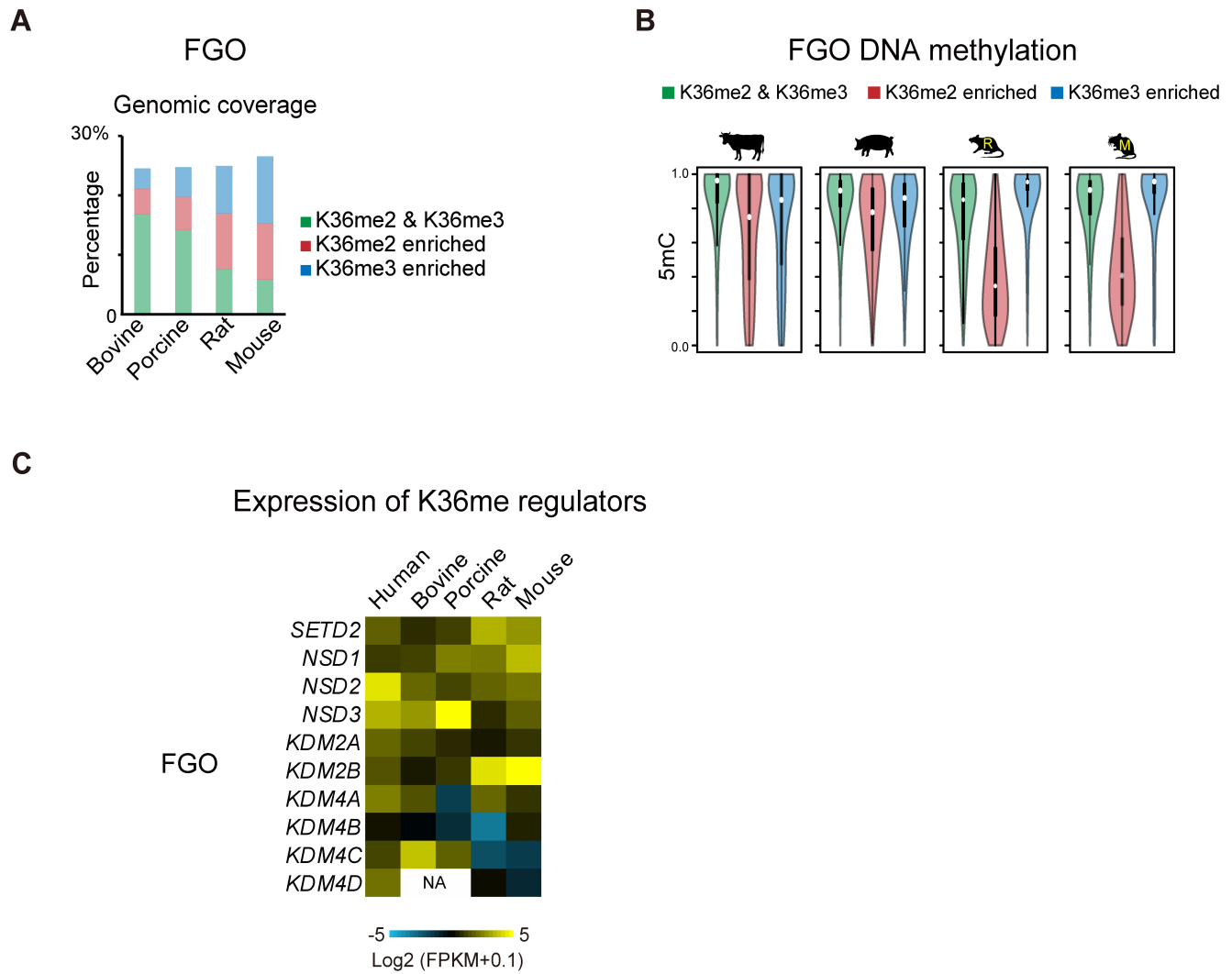


Figure S11. H3K36me2- and H3K36me3-correlated DNA methylation in the oocytes across mammals. (A) Barplot showing the percentages of the genome enriched by H3K36me2 (red), H3K36me3 (blue) or both H3K36me2 and H3K36me3 (green) in the oocytes of different species. (B) Violin plots showing the DNA methylation levels at regions enriched by H3K36me2 (K36me2 enriched, red), H3K36me3 (K36me3 enriched, blue) or both H3K36me2 and H3K36me3 (K36me2 & K36me3, green) in the oocytes across mammals. (C) Heatmap showing the expression of H3K36me2/3 regulators in the oocytes across the five species.

Figure S12

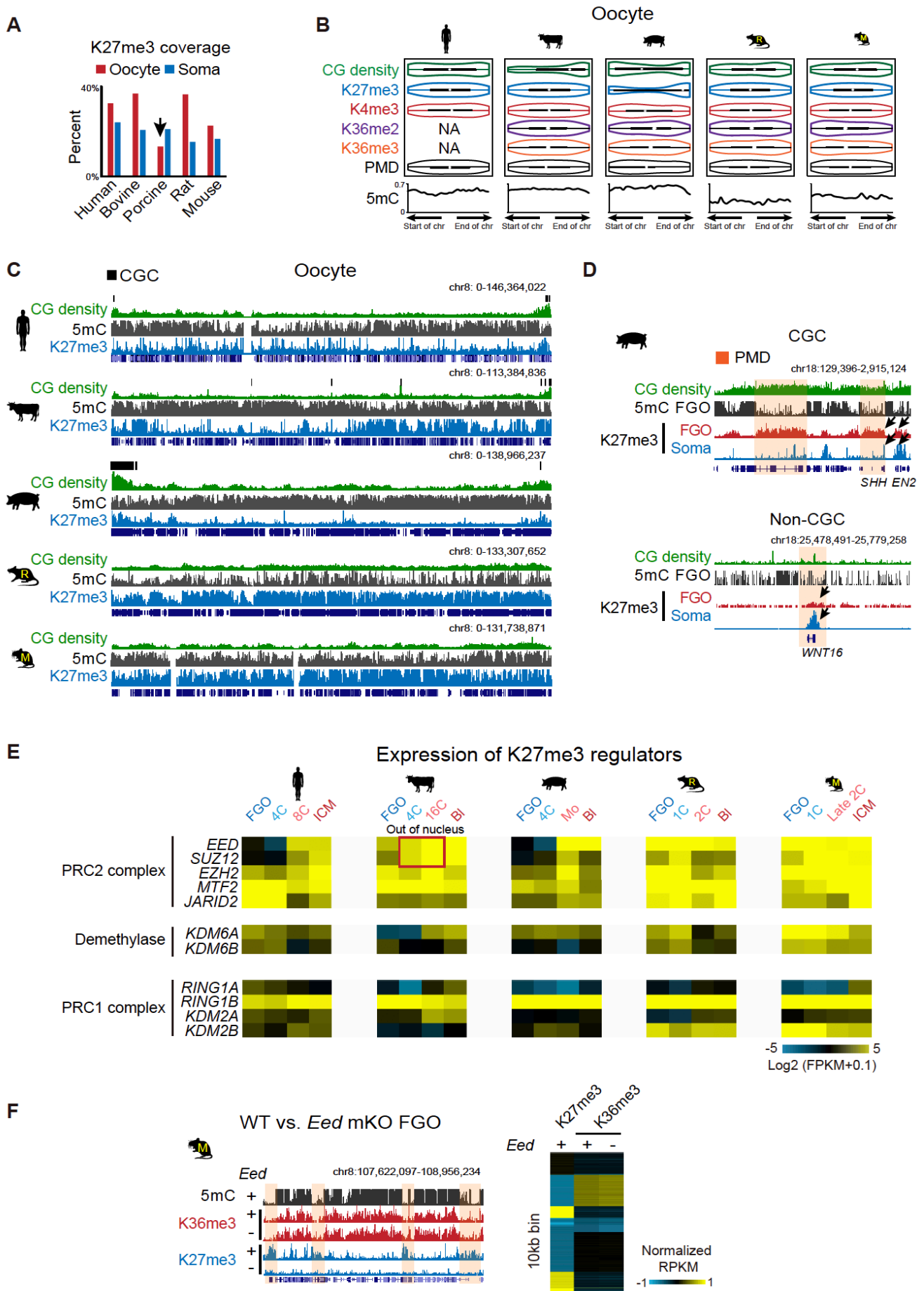


Figure S12. H3K27me3 and its relationship with other epigenetic marks in mammalian oocytes. (A) Bar chart showing the percentages of the genome covered by H3K27me3 (K27me3) in the oocytes among different species. The somatic tissues (Soma) in each species are also similarly analyzed as controls. Arrow indicates the relatively lower genome coverage of H3K27me3 in porcine FGOs. (B) Violin plots showing the distribution of CpG density, H3K27me3, H3K4me3 (K4me3), H3K36me2 (K36me2), H3K36me3 (K36me3), PMDs and DNA methylation (5mC) along all chromosomes in the oocytes of all five species. (C) UCSC browser views showing H3K27me3 on the whole chromosome 8 in the oocytes of all five species. The DNA methylation and CG density are also shown. (D) UCSC browser views showing H3K27me3 in CGCs (left) and non-CGCs (right) in porcine FGOs and somatic cells (Soma). DNA methylation and CG density are also shown. (E) Heatmaps showing the expression of H3K27me3 regulators in the oocyte and during the preimplantation development of the five species. Note *EED* and *SUZ12* are expressed but are excluded from nucleus before the morula stage in bovine (30) (red box). (F) UCSC browser views (top) and heatmap (bottom, 10kb bin) showing the H3K36me3 in wild type (+) and maternal *Eed* knockout (-) mouse FGOs. H3K27me3 in wild type FGOs is also shown.

Figure S13

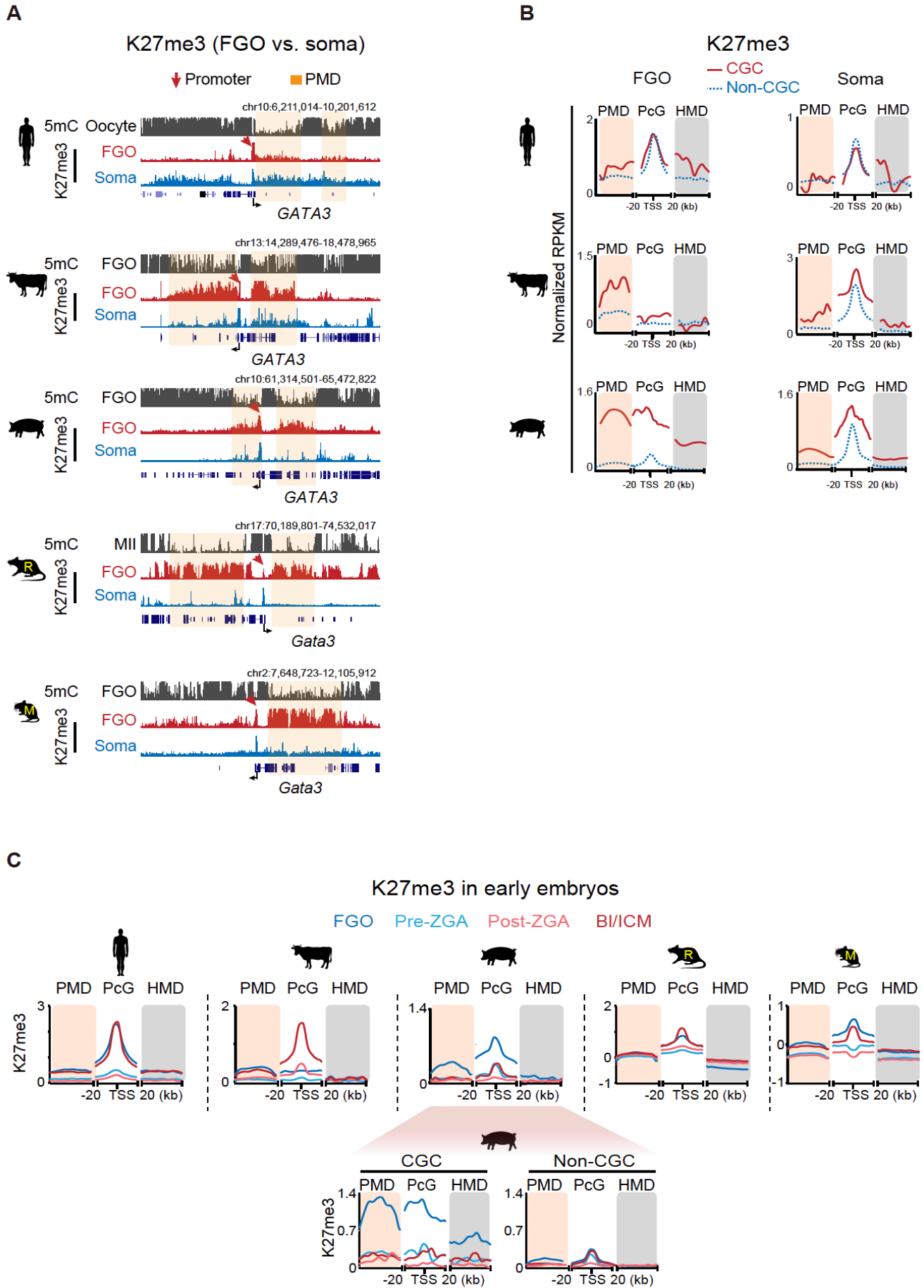
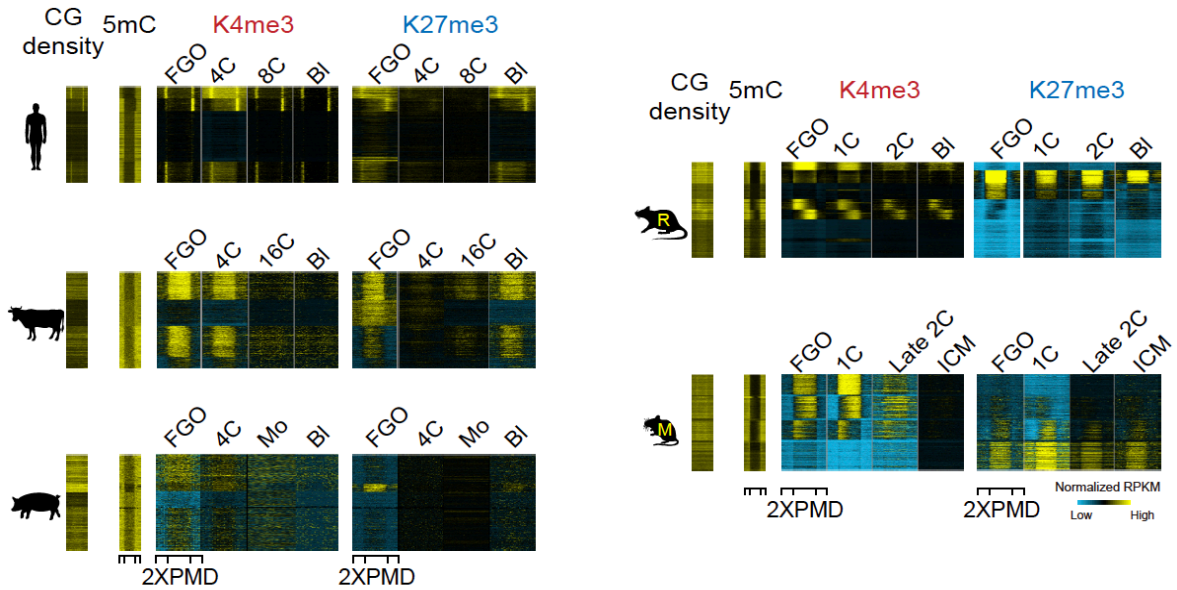


Figure S13. H3K27me3 in mammalian oocytes and preimplantation embryos. (A) UCSC genome browser views showing H3K27me3 (K27me3) enrichment around the *Gata3* loci in the FGOs of all five species. DNA methylation in oocytes and H3K27me3 in somatic tissues (Soma) are also shown. (B) Metaplots showing H3K27me3 enrichment in PMDs, Polycomb group target gene (PcG) promoters (TSS \pm 20kb) and HMDs in CGC (red) and non-CGC (blue) regions in FGOs (left) and somatic tissues (Soma, right) for human, bovine and porcine. (C) Metaplots showing the enrichment of H3K27me3 in PMDs, PcG promoters and HMDs in FGOs and early embryos among the five species (top). CGCs and non-CGCs for porcine are also separately analyzed (bottom). FGO, full-grown oocyte; Bl, blastocyst; ICM, inner cell mass; PMD, partially methylated domain; PcG, Polycomb group gene, HMD, highly methylated domain. CGC, CpG content.

Figure S14

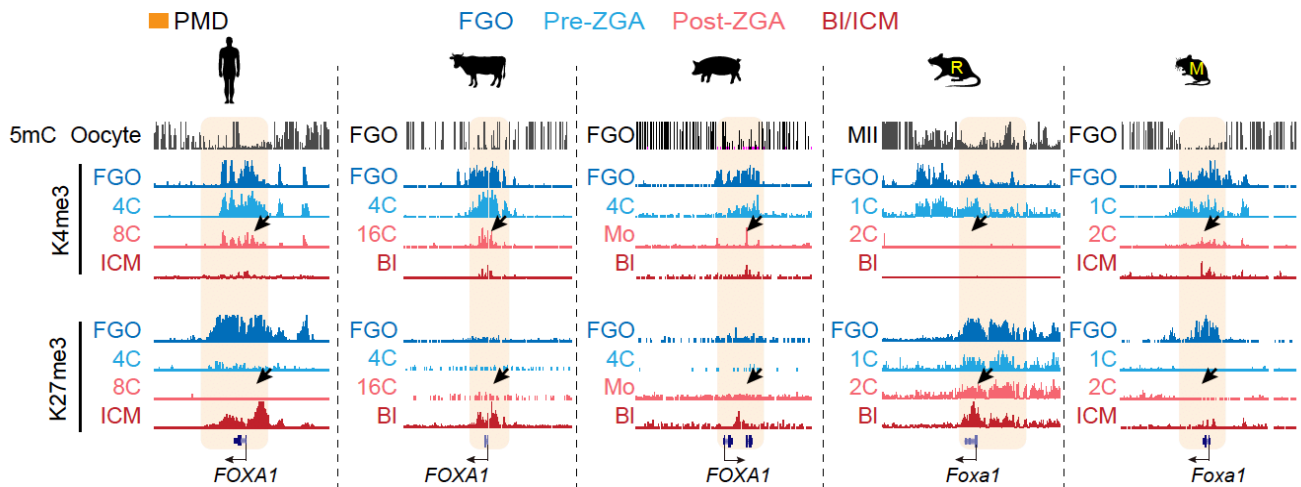
A

K4me3 and K27me3 in PMD



B

Developmental genes



C

Developmental genes

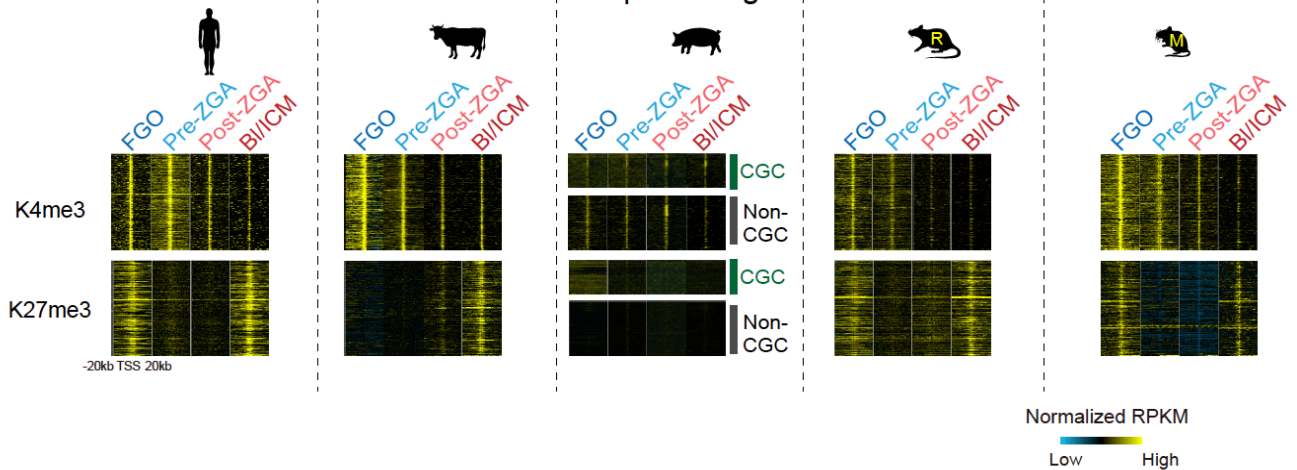


Figure S14. Dynamics of bivalency at PMD and developmental gene promoters during mammalian preimplantation development. (A) Heatmaps showing the enrichment of H3K4me3 and H3K27me7 in the PMDs of mammalian oocytes and their dynamics during mammalian early development. CG density and DNA methylation in PMDs are also shown. FGO, full-grown oocyte; PMD, partially methylated domain; HMD, highly methylated domain; 1C, 1-cell embryo; 2C, late 2-cell embryo; 4C, 4-cell embryo; 8C, 8-cell embryo; 16C, 16-cell embryo; Mo, morula; Bl, Blastocyst; ICM, inner cell mass. (B) UCSC genome browser views showing the dynamics of H3K4me3 (K4me3) and H3K27me3 (K27me3) marks at a representative bivalent gene *Foxa1* during preimplantation development of the five species. The arrows indicate the promoter regions at ZGA. The PMD regions are shaded. (C) Heatmaps showing the enrichment of H3K4me3 and H3K27me3 marks at bivalent genes from FGO to blastocyst in the five species. Developmental genes are identified in the pluripotent stem cells of each species (Materials and Methods).

Figure S15

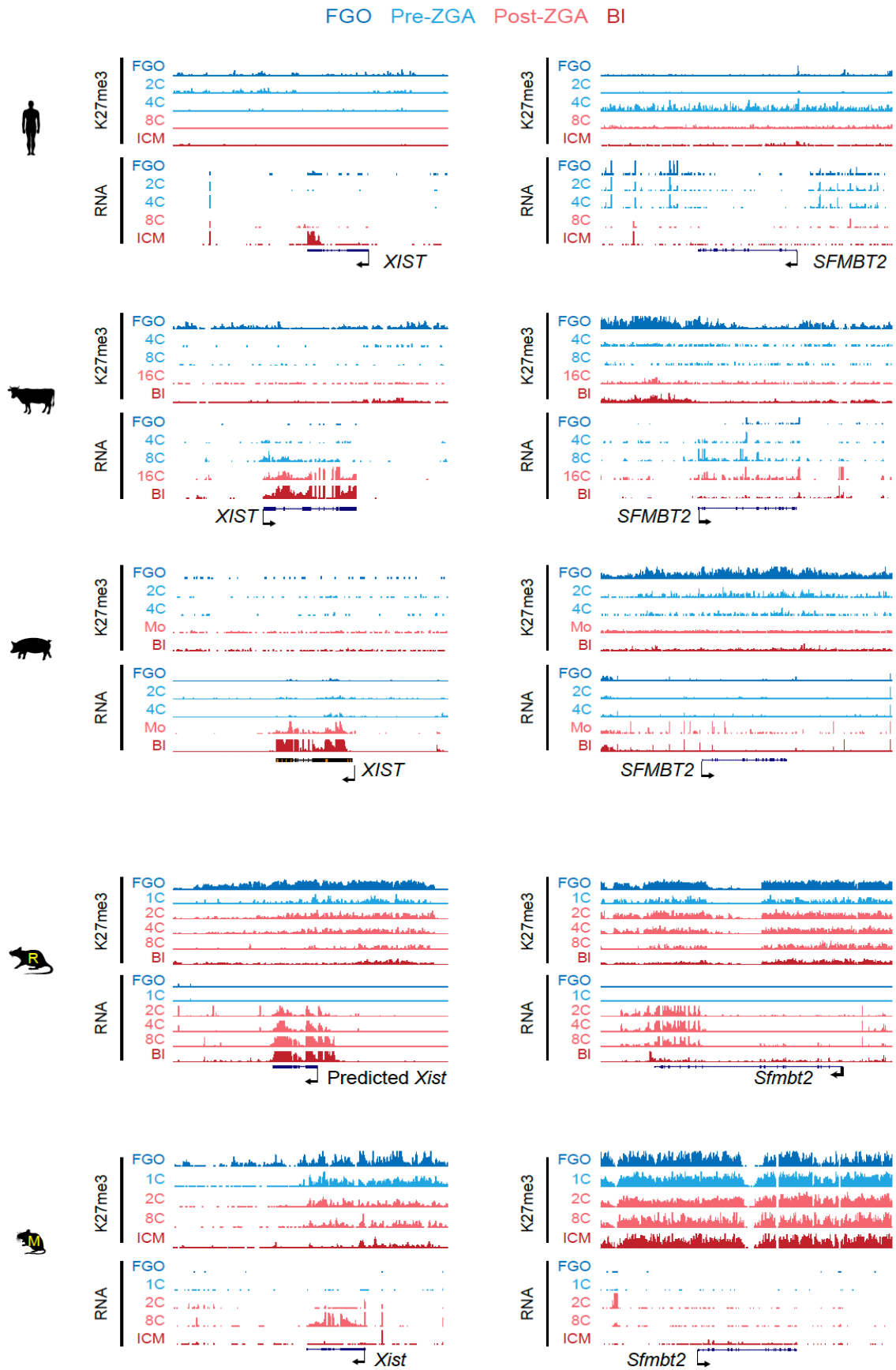


Figure S15. H3K27me3 and gene expression at the *Xist* and *Sfmbt2* loci during preimplantation development among different species. The UCSC genome browser views showing the enrichment of H3K27me3 (K27me3) and gene expression at *Xist* and *Sfmbt2* loci from FGO to blastocyst in different species. Note the absence of H3K27me3 signals at these two gene loci by ZGA in non-rodents.

Figure S16

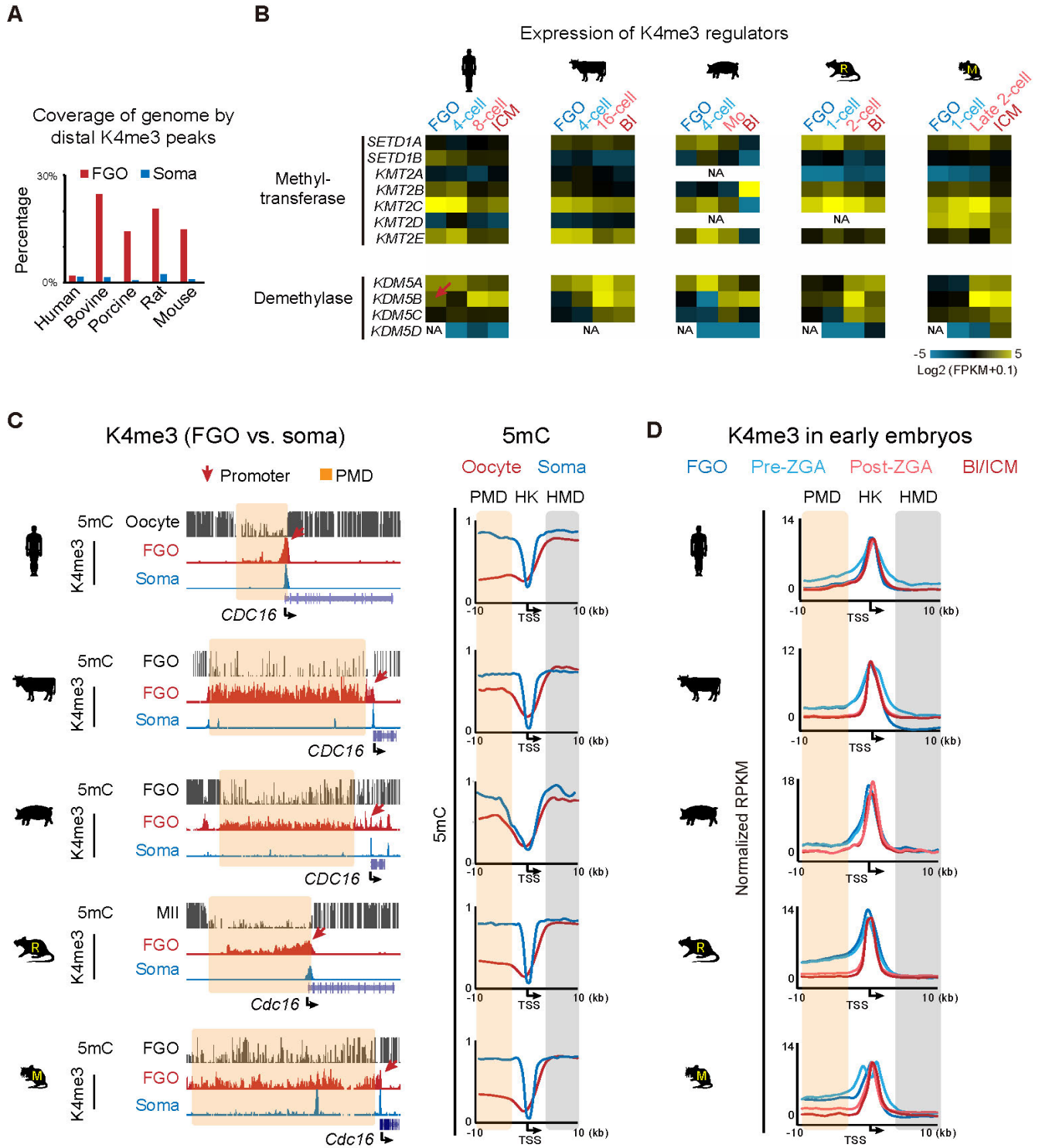
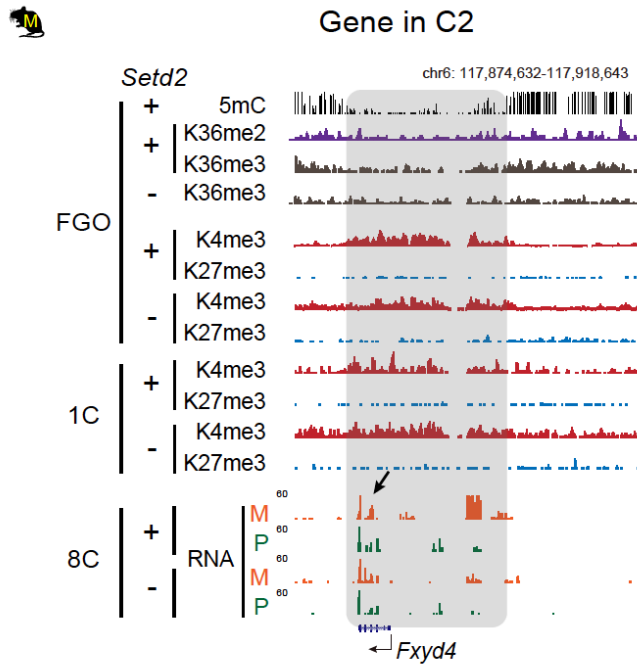


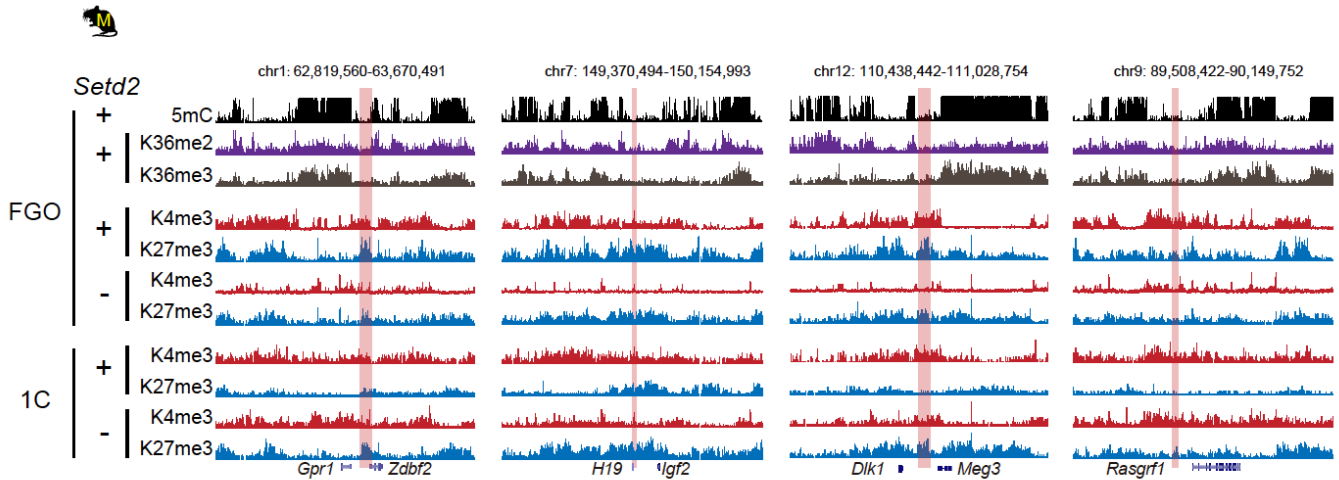
Figure S16. H3K4me3 in mammalian oocytes and preimplantation embryos. (A) Barplot showing the genome coverage of distal H3K4me3 (K4me3) peaks in oocyte genomes among different species. Analysis results for the somatic tissues (Soma) in each species are shown as controls. (B) Heatmaps showing the expression of H3K4me3 (K4me3) regulators in the oocyte and during the preimplantation development of the five species. Red arrow notes the selective expression of *KDM5B* in the oocytes of human but not other species. (C) Left, UCSC genome browser views showing the enrichment of H3K4me3 (K4me3) at the promoter (*Cdc16*) and PMD regions in FGOs of all the five species. H3K4me3 at the promoters and PMD regions are indicated by arrows and shades, respectively. Similar analyses for somatic tissues or cells are shown as controls. Right, metaplots showing the DNA methylation (5mC) at PMDs, promoters, and HMDs around HK genes that have upstream PMDs in oocytes among the five species oocytes (red line). The somatic tissue (blue line) in each species is similarly analyzed as a control. (D) Metaplots showing the enrichment of H3K4me3 in PMDs, housekeeping gene promoters (TSS \pm 2.5kb) and HMDs in FGOs and early embryos among the five species.

Figure S17

A



B



C

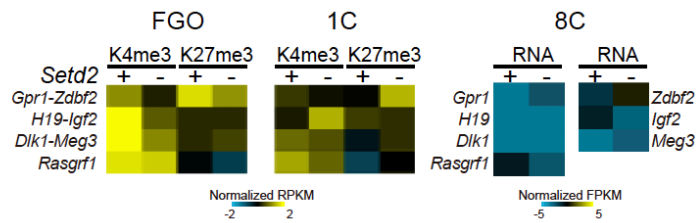


Figure S17. Exclusion of oocyte nH3K27me3 from nH3K4me3 domains in rodents. (A) The UCSC genome browser view showing the distribution of H3K36me2 (K36me2), H3K36me3 (K36me3), H3K4me3 and H3K27me3 (K27me3), and the maternal and paternal RNA read count signals at a representative gene in cluster 2 (C2, identified in Fig. 6h, where no H3K27me3 invasion is observed) in wild type (“+”) and maternal *Setd2* knockout (“-”) FGOs. PMD region is shaded. The arrow indicated *Fxyd4* gene locus. (B and C) The UCSC genome browser views (B) and heatmaps (C, left and middle) showing the distribution of H3K4me3 (K4me3) and H3K27me3 (K27me3) in the paternal ICRs (shaded) in wild type (“+”) and maternal *Setd2* knockout (“-”) mouse full-grown oocyte (FGO) and 1-cell (1C) embryo. DNA methylation (5mC), H3K36me2 (K36me2) and H3K36me3 (K36me3) in wild type FGO are shown in the snapshots. Expression of the paternally imprinted genes in wild type and *Setd2* mutant 8C (C, right) is shown. *Setd2* mutant 8C embryos are derived from enucleated WT oocytes transferred with *Setd2* mKO chromatin (9).

Table S1. (separate file)

Public data used in this study

Table S2. (separate file)

Information for datasets generated in this study

Table S3. (separate file)

Genes located in CGCs in human, bovine and porcine

Table S4. (separate file)

The positions of putative maternal and paternal ICRs in bovine, porcine and rat.

REFERENCES AND NOTES

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