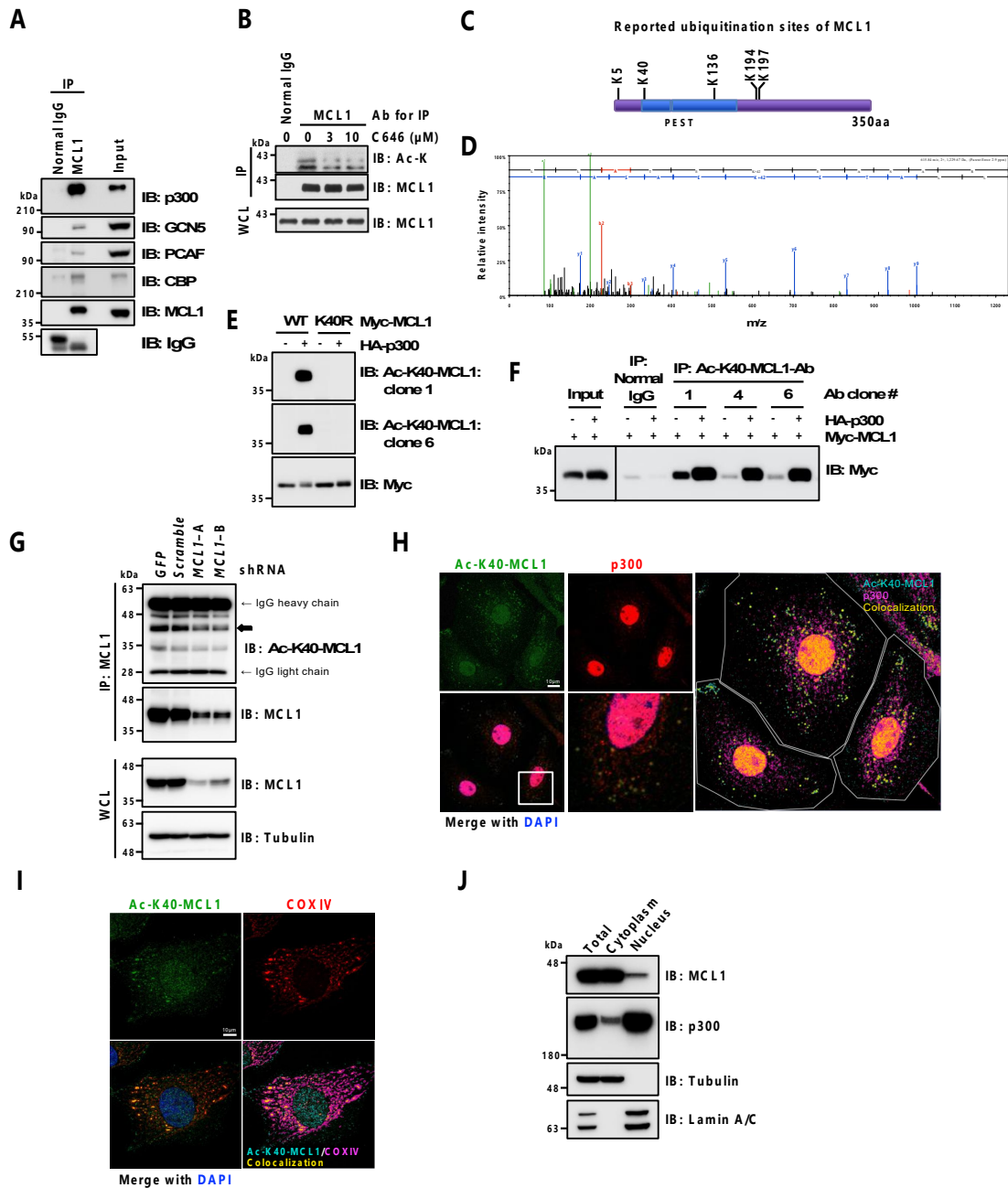


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**Supplemental information**

**Interplay between protein acetylation  
and ubiquitination controls MCL1 protein stability**

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**Figure S1. p300 binds and acetylates MCL1 at the lysine 40 (K40) residue, related to Figure 1.**

**A**, Evaluation of endogenous interaction between MCL1 and the indicated lysine acetyltransferases. Immunoblot (IB) analysis of whole-cell lysates (WCL) (input: 60  $\mu$ g of cell lysates) and anti-MCL1 immunoprecipitates (IP) derived from HeLa cells.

**B,** Treatment with the p300/CBP inhibitor C646 reduces the MCL1 acetylation. IB analysis of WCL and anti-MCL1 IP derived from HeLa cells. The cells were treated overnight with C646 at the indicated concentration before harvesting.

**C,** A schematic diagram of reported ubiquitination sites in MCL1.

**D,** The LC-MS/MS spectrum for peptide LLATEkEASAR shows the location of acetylation at Lys40 in the human MCL1 protein via the +42 Da shift from the y5 to y6 fragment ions.

**E,** IB analysis of WCL derived from 293T cells transfected with the Myc-MCL1 constructs together with HA-p300 as indicated. Cells were harvested at 48 h after transfection.

**F,** IB analysis of anti-acetylated (Ac)-K40-MCL1-IP derived from 293T cells transfected with Myc-MCL1 and HA-p300 constructs as indicated. Cells were harvested at 48 h after transfection for IP.

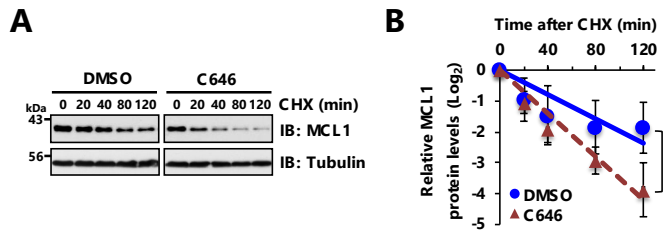
**G,** IB analysis of WCL and anti-MCL1-IP derived from SKBR3 cells stably expressing the lentiviral shRNA specific for *GFP*, *Scramble*, or *MCL1* (using two independent shRNAs).

**H,** Endogenous acetylated-K40 (Ac-K40) MCL1 colocalizes with the p300 acetyltransferase in the nucleus and cytoplasm of BT-20 breast cancer cells. Immunofluorescence (IF) images of Ac-K40-MCL1 (green) and p300 (red) with DAPI staining (blue) in BT-20 cells. Scale bar, 10  $\mu$ m.

**I,** Endogenous Ac-K40-MCL1 is present in mitochondria of BT-20 breast cancer cells. Immunofluorescence (IF) images of Ac-K40-MCL1 (green) and COX IV (red), a mitochondrial marker protein, with DAPI staining (blue) in BT-20 cells. Scale bar, 10  $\mu$ m.

**J,** IB analysis of WCL, cytoplasmic and nuclear extracts derived from HeLa cells.

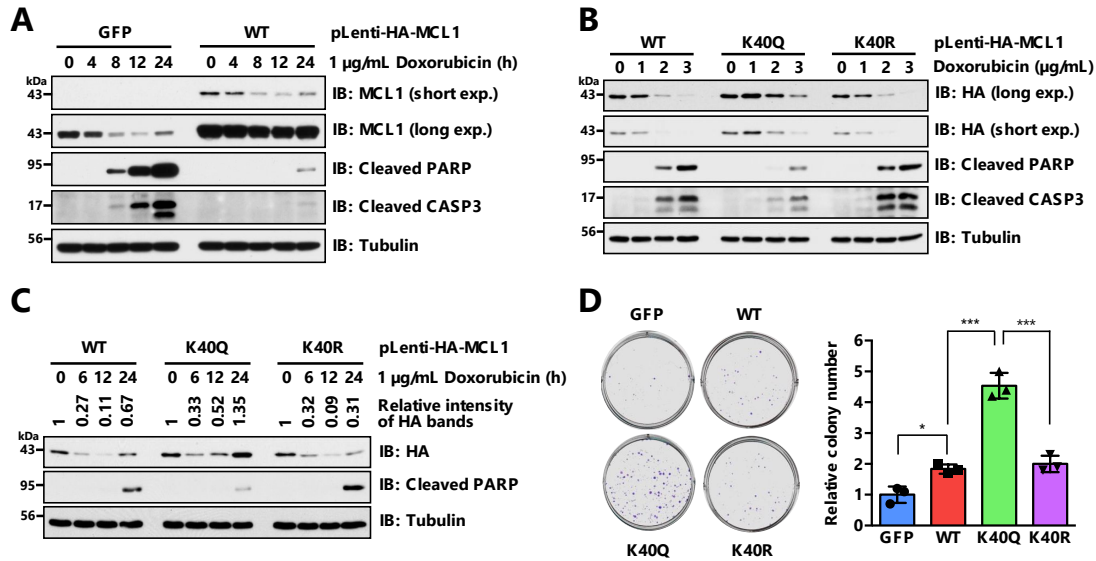
Data (A), (B), (E)-(G), and (J) are representative of at least two independent experiments.



**Figure S2. p300-mediated MCL1 acetylation leads to MCL1 stabilization in part through decreasing MCL1 ubiquitination, related to Figure 2.**

**A**, Treatment with the p300/CBP inhibitor C646 shortens MCL1 protein half-life. IB analysis of WCL derived from PF382 cells. Cells were pretreated with C646 (10  $\mu$ M) overnight and then treated with 100  $\mu$ g/mL cycloheximide (CHX) for the periods indicated before harvesting.

**B**, Quantification of the MCL1 band intensities of IB replicates of (A). Data are presented as the mean  $\pm$  SD; n = 3 independent experiments, \* p < 0.05.

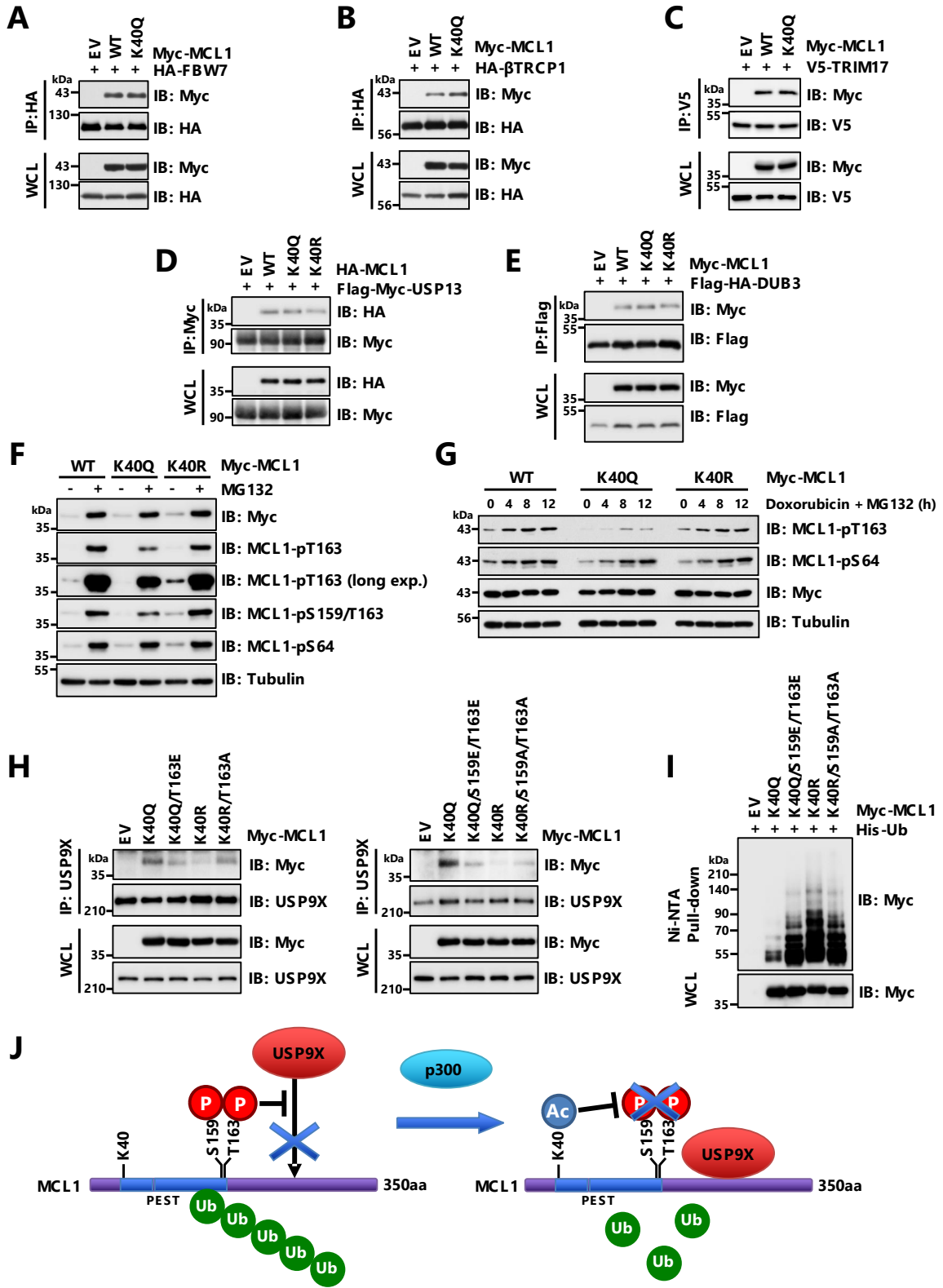


**Figure S3. Acetylation-mimetic MCL1 K40Q displays enhanced anti-apoptotic function and oncogenicity, related to Figure 3.**

**A-C**, Acetylation-mimetic MCL1 K40Q exerts enhanced anti-apoptotic function. IB analysis of WCL derived from HeLa cells stably expressing GFP (as a control) or HA-MCL1 (WT, K40Q, or K40R). Cells were treated with doxorubicin for the time period indicated at 1 µg/mL (**A** and **C**) or at the indicated concentration for 24 h (**B**).

**D**, Acetylation-mimetic MCL1 K40Q enhances colony formation potential of HeLa cells. Colony-formation assay was conducted using HeLa cells presented in (**A-C**). These cells were pretreated overnight with doxorubicin (0.02 µg/mL) before plating for the assays. Data are presented as the mean ± SD; n = 3 biological replicates, \* p < 0.05, \*\*\* p < 0.001.

Data (A)-(C) are representative of at least two independent experiments.



**Figure S4. MCL1 acetylation promotes its interaction with USP9X, resulting in MCL1 deubiquitination and stabilization, related to Figure 4.**

**A-E**, IB analysis of WCL and anti-HA, anti-V5, anti-Myc, or anti-Flag IP derived from 293T cells transfected with the indicated constructs. At 36 h after transfection, cells were treated overnight with MG132 (10  $\mu$ M) before harvesting for IP.

**F**, The acetylation-mimetic K40Q mutation impairs MCL1 phosphorylation at S159/T163. IB analysis of WCL derived from CRISPR/Cas9-mediated *MCL1*-knockout HeLa cells transfected with the indicated Myc-MCL1 constructs. At 36 h after transfection, cells were treated with MG132 (10  $\mu$ M) for 12 h before harvesting.

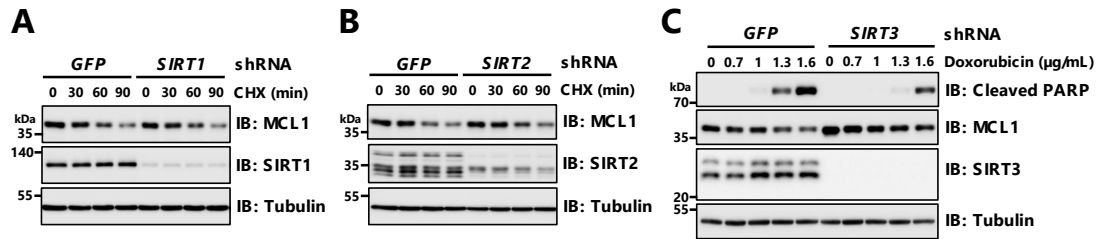
**G**, IB analysis of WCL derived from HeLa cells transfected with the indicated Myc-MCL1 constructs. At 36 h after transfection, cells were co-treated with doxorubicin (1  $\mu$ g/mL) and MG132 (15  $\mu$ M) for the periods indicated before harvesting.

**H**, Phosphorylation-mimetic S159E/T163E mutations impair the interaction of MCL1 with USP9X regardless of the acetylation-mimetic K40Q mutation. IB analysis of WCL and anti-USP9X IP derived from CRISPR/Cas9-mediated *MCL1*-knockout HCT116 cells transfected with the indicated Myc-MCL1 constructs. At 36 h after transfection, cells were treated overnight with MG132 (10  $\mu$ M) before harvesting for IP.

**I**, Additive phosphorylation-mimetic S159E/T163E mutations restore the ubiquitination of acetylation-mimetic MCL1 K40Q. 293T cells were transfected with the indicated Myc-MCL1 and His-ubiquitin (His-Ub) constructs. At 36 h after transfection, cells were treated overnight with MG132 (10  $\mu$ M) before harvesting. His-Ub-conjugated proteins were captured with Ni-NTA agarose beads and subjected to IB analysis.

**J**, A schematic diagram of the crosstalk between phosphorylation at S159/T163 and acetylation at K40 of the MCL1 protein in regulating MCL1 interaction with USP9X.

Data (A)-(I) are representative of at least two independent experiments.



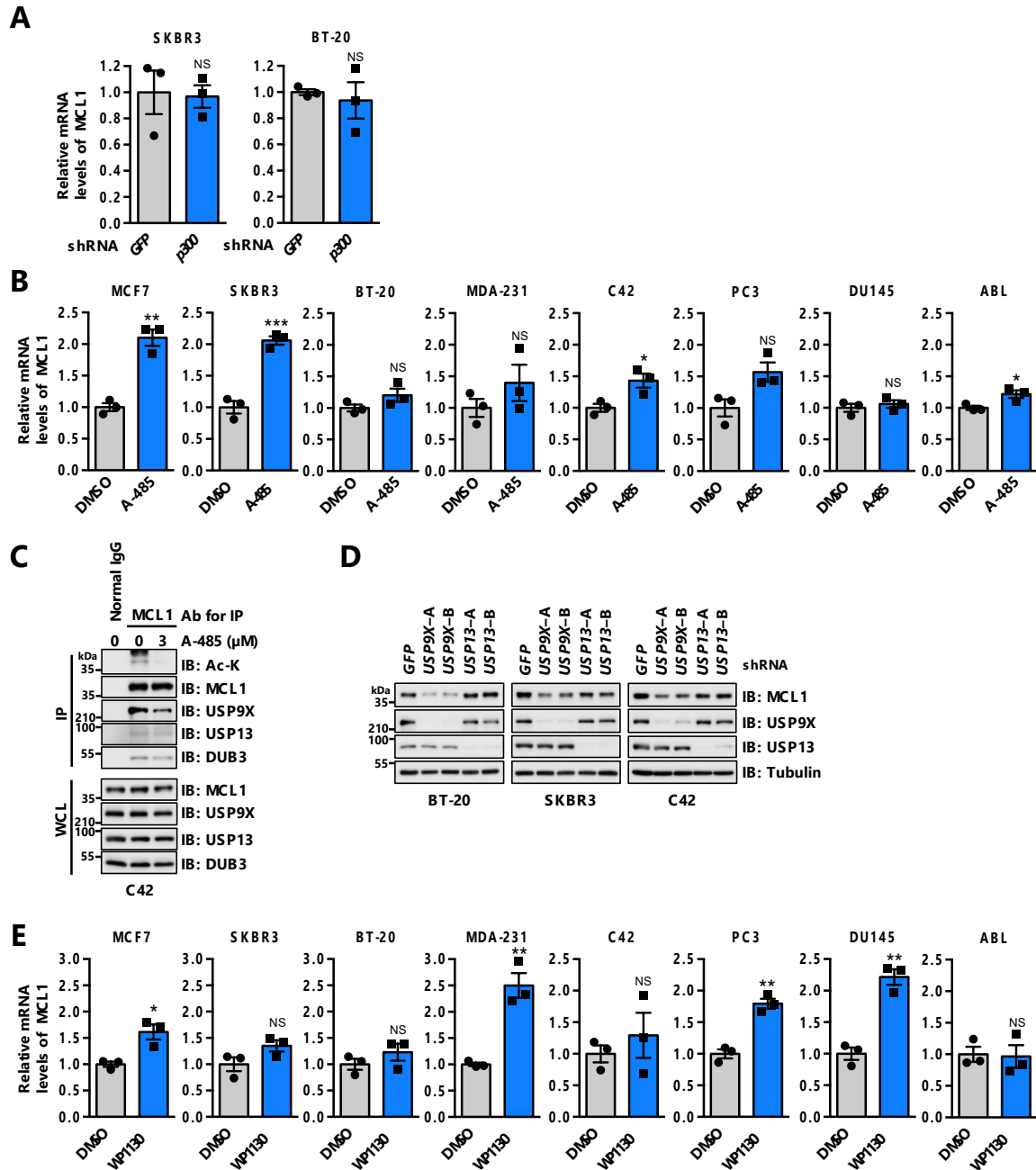
**Figure S5. SIRT3 negatively regulates MCL1 protein stability through deacetylation of MCL1, related to Figure 5.**

**A and B**, *SIRT1* or *SIRT2* depletion shows minimal effects on MCL1 protein half-life. IB analysis of WCL derived from HeLa cells stably expressing the lentiviral shRNA specific for *GFP*, *SIRT1*, or *SIRT2*. Cells were treated with 100 µg/mL CHX for the period indicated before harvesting.

**C**, *SIRT3* depletion desensitizes cells to doxorubicin through MCL1 stabilization. IB analysis of WCL derived from HeLa cells stably expressing the lentiviral shRNA specific for *GFP* or *SIRT3*. Cells were treated with the indicated concentrations of doxorubicin for 24 h before harvesting.

Data (A)-(C) are representative of at least two independent experiments.





**Figure S6. MCL1 acetylation promotes cancer cell survival in a p300 and USP9X dependent manner, related to Figure 6.**

**A**, Real-time RT-PCR analysis to determine the relative mRNA expression levels of MCL1 in SKBR3 and BT-20 cells presented in Figure 6B. *GAPDH* was utilized for normalization. Data are presented as the mean  $\pm$  SEM, n = 3 biological replicates, NS: not significant.

**B**, Real-time RT-PCR analysis to determine the relative mRNA expression levels of MCL1 in breast and prostate cancer cells presented in Figure 6C. These cells were treated with A-485 (3  $\mu$ M) for 48 h before harvesting. *GAPDH* was utilized for normalization. Data are presented as the mean  $\pm$  SEM, n = 3 biological replicates, \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001, NS: not significant.

**C**, Treatment with the p300/CBP inhibitor A-485 reduces MCL1 acetylation and promotes dissociation of USP9X from MCL1. IB analysis of WCL and anti-MCL1 IP derived from C42 cells treated overnight with A-485 at the indicated concentration before harvesting.

**D**, *USP9X* depletion reduces MCL1 protein levels in the indicated breast and prostate cancer cells. IB analysis of WCL derived from BT-20, SKBR3, and C42 cells stably expressing the lentiviral shRNA specific for *GFP*, *USP9X*, or *USP13*.

**E**, Real-time RT-PCR analysis to determine the relative mRNA expression levels of *MCL1* in breast and prostate cancer cells presented in Figure 6G-H. These cells were treated with WP1130 (3  $\mu$ M) for 8 h (breast cancer cells) or 12 h (prostate cancer cells) before harvesting. *GAPDH* was utilized for normalization. Data are presented as the mean  $\pm$  SEM, n = 3 biological replicates, \* p < 0.05, \*\* p < 0.01, NS: not significant.

Data (C) and (D) are representative of at least two independent experiments.