

SUPPLEMENTARY MATERIAL

High-resolution phenotypic analysis of the mouse hematopoietic hierarchy using spectral cytometry – from stem cell subsets to early progenitor compartments

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Supplementary Figures

Supplementary Figure 1. *Validation of unmixing accuracy in 14-color panel.*

Supplementary Figure 2. *Validation of the gating strategy using fluorescence-minus-one (FMO) controls.*

Supplementary Figure 3. *Side-by-side comparison of the panel on spectral and “conventional” polychromatic flow cytometers.*

Supplementary Figure 4. *Changes in hematopoietic stem and progenitor cells at different ages.*

Supplementary Figure 5. *Validation of unmixing accuracy between GFP and FITC.*

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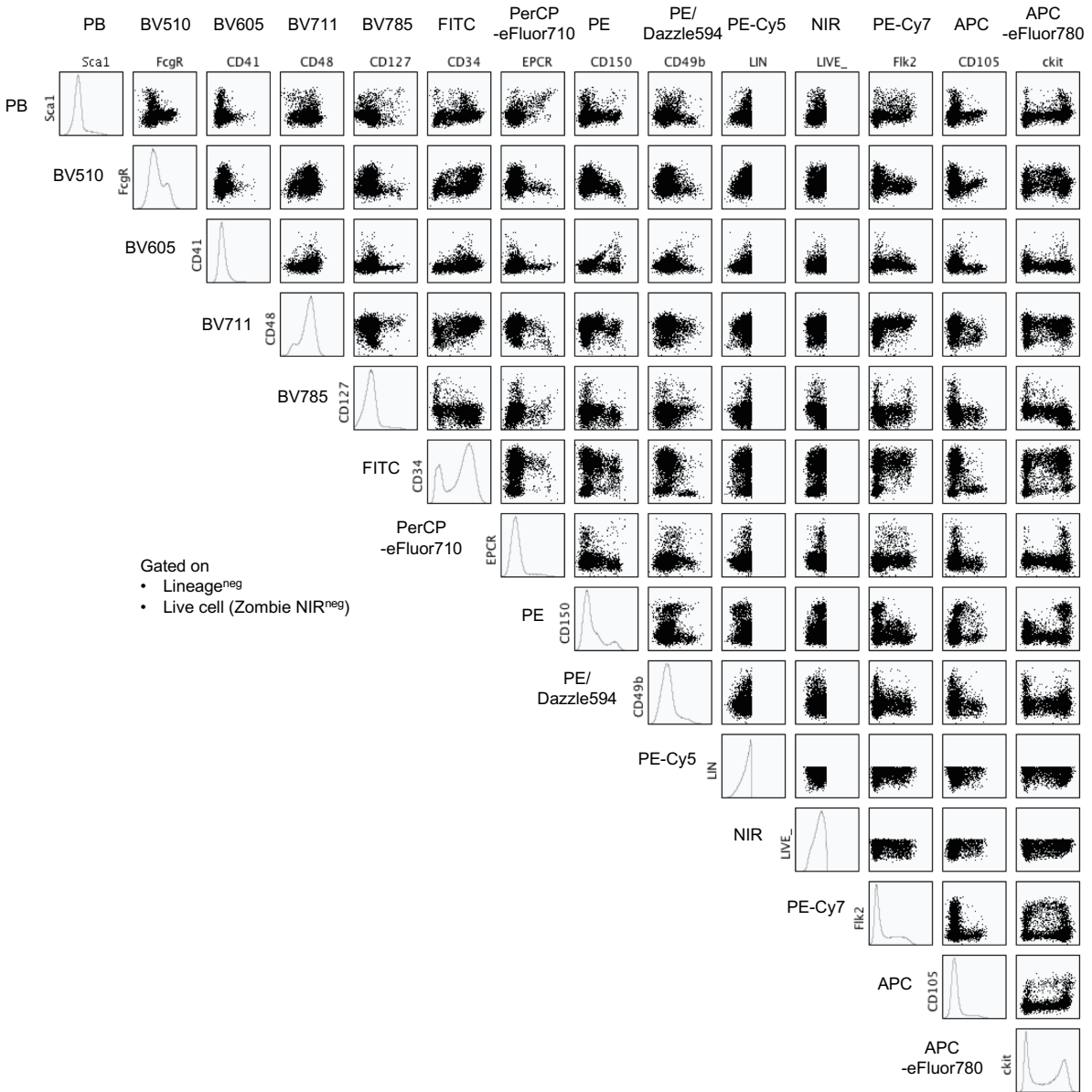
Supplementary Table 1. *Reagents used for the 14-color panel and reference controls.*

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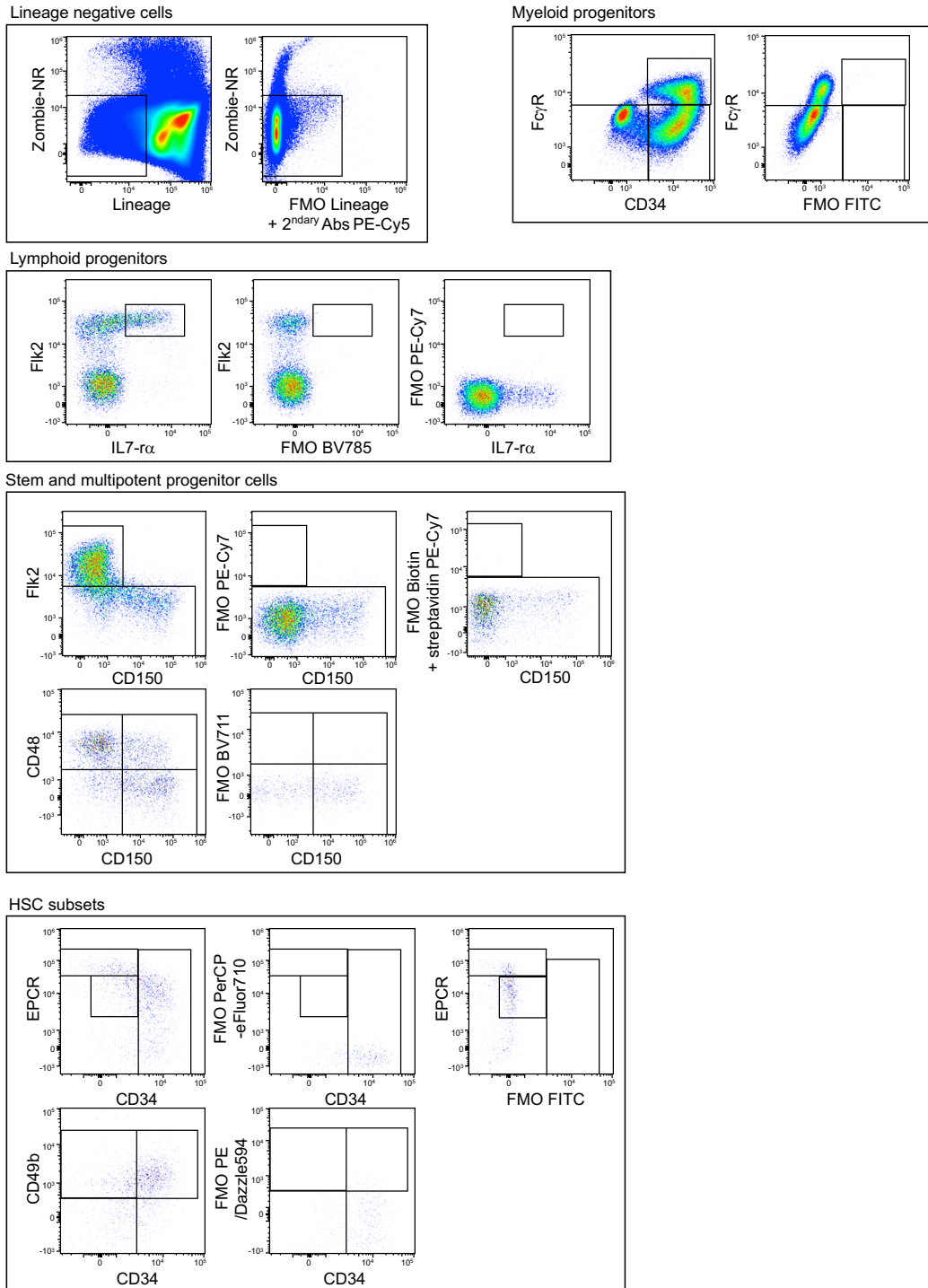
Supplementary Table 4. *Mean percentage (frequency of parent) and standard deviation (SD) for all gates presented in Figure 3.*

Supplementary Table 5. *Number of hematopoietic stem and progenitor cells at different mouse ages.*



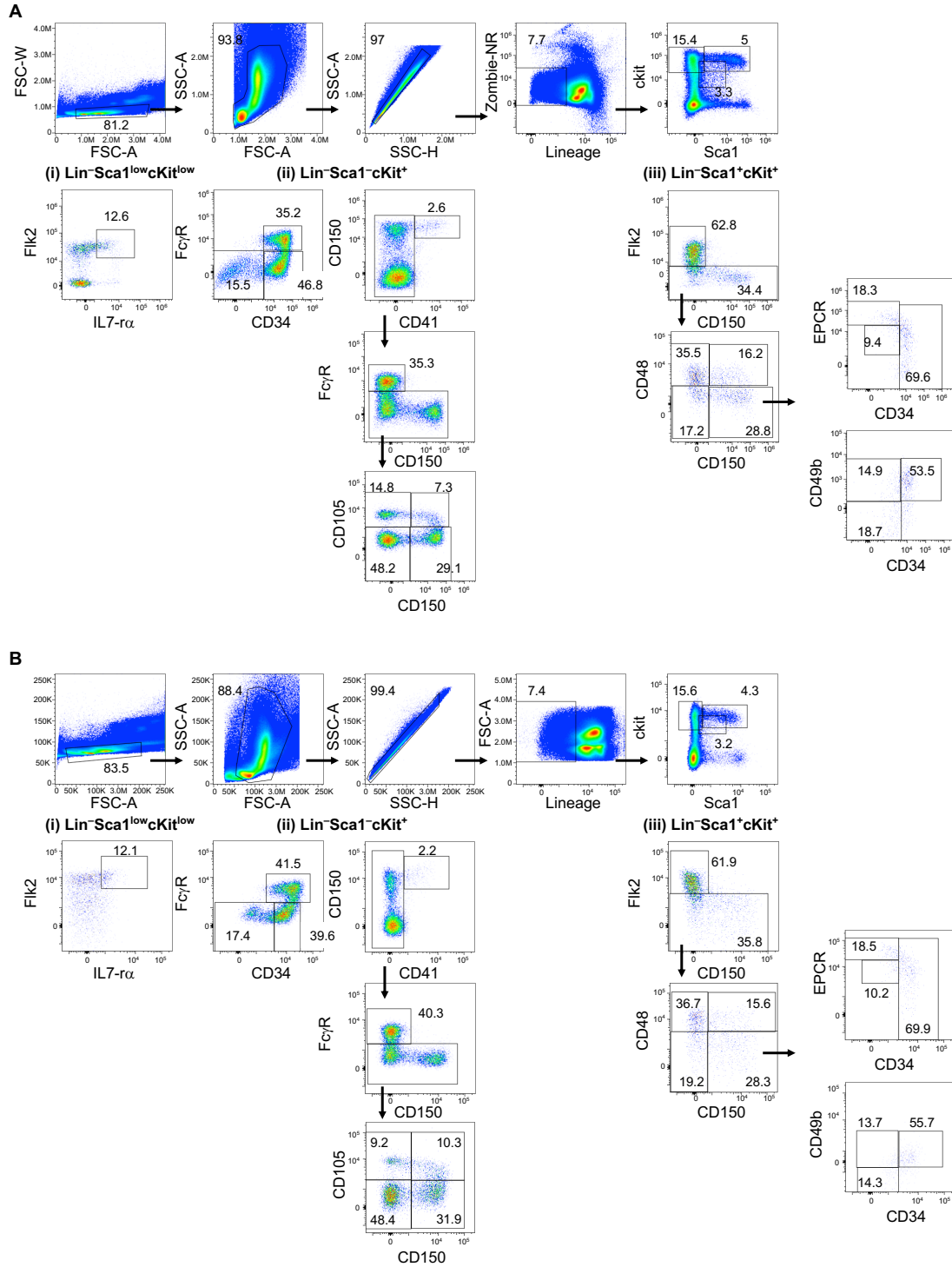
Supplementary Figure 1. Validation of unmixing accuracy in 14-color panel.

NxN plot displays every parameter versus every other parameter to assess unmixing accuracy of the fluorochromes in the 14-color panel. Plots were gated on Live Lin^{neg} BM cells from an 8-week-old wild-type mouse with gating exclusion of off-scale antibody aggregates.



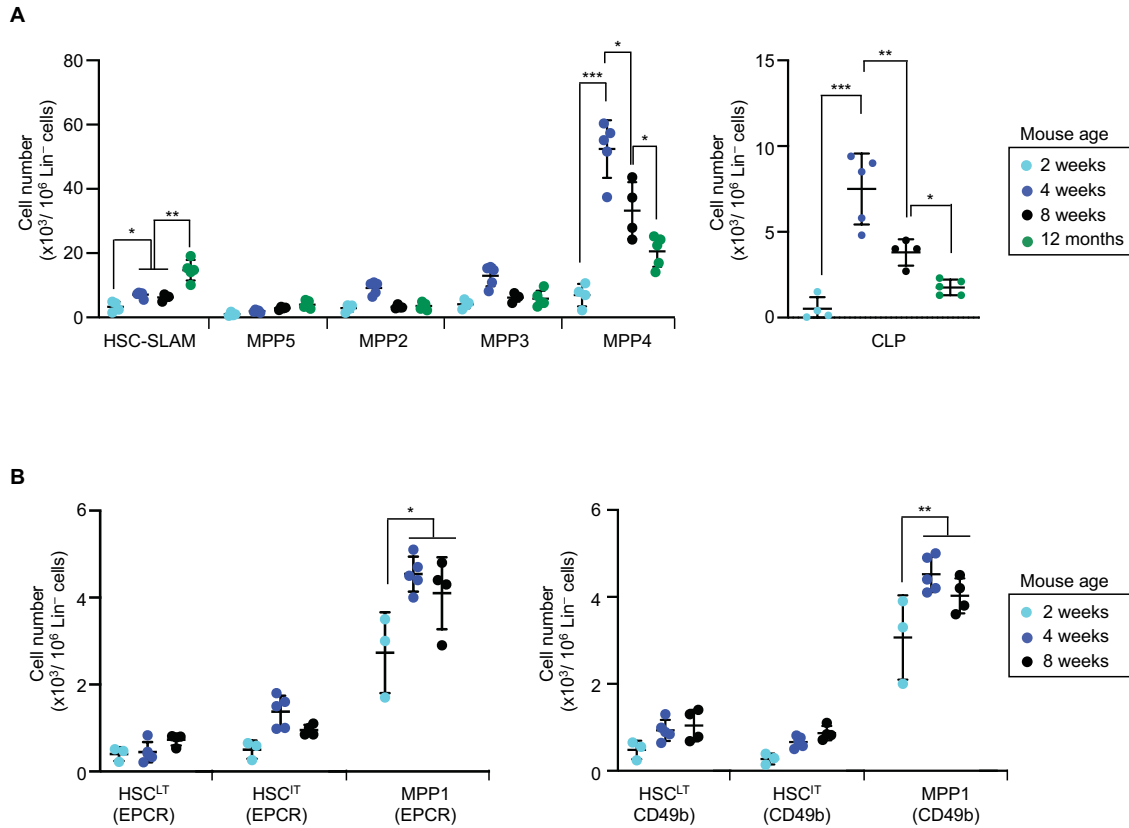
Supplementary Figure 2. Validation of the gating strategy using fluorescence-minus-one (FMO) controls.

Background staining and negative/positive boundaries were determined using FMO controls for the definition of (i) lineage negative cells, (ii) myeloid progenitors, (iii) lymphoid progenitors, (iv) stem and multipotent progenitor cells and (v) HSC subsets.

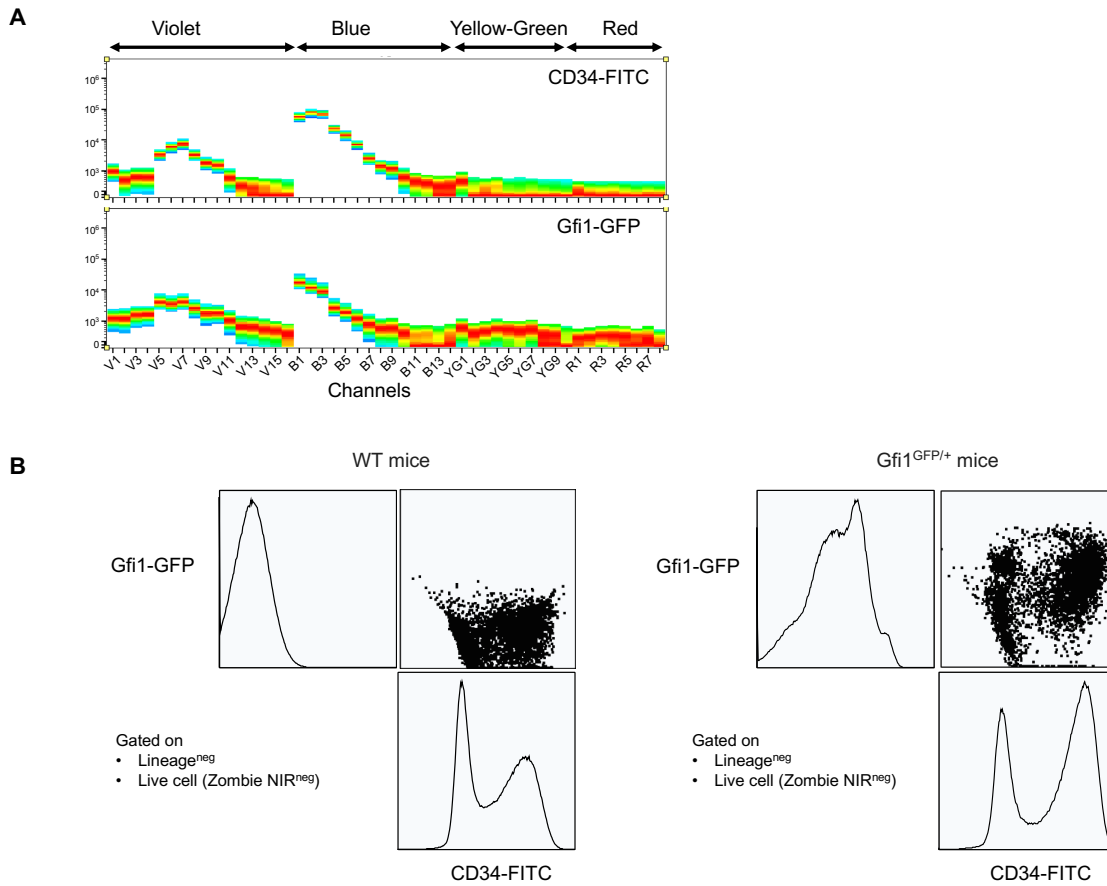


Supplementary Figure 3. Side-by-side comparison of the panel on spectral and “conventional” polychromatic flow cytometer.

Representative flow cytometry plots of the 14-fluorescent parameter flow analysis of 8-week-old murine bone marrow (BM) cells on spectral flow cytometer (**A**) and “conventional” polychromatic flow cytometer (**B**). Figure is representative of 3 independent experiments performed side-by-side on the two cytometers.



Supplementary Figure 4. Changes in hematopoietic stem and progenitor cells at different ages. (A) Absolute numbers \pm SD of HSPC populations per 10^6 live Lin^- cells in the BM of 2-, 4-, 8- and 52-week-old mice. (B) Absolute numbers \pm SD of HSC subsets, defined by EPCR (left panel) and CD49b (right panel) expression, per 10^6 live Lin^- cells in the BM of 2-, 4-, and 8-week-old mice.



Supplementary Figure 5. Validation of unmixing accuracy between GFP and FITC.
(A) Spectral histograms of CD34-FITC and Gfi1-GFP detected on the Aurora flow cytometer. **(B)** NxN plot of Lin^{neg}Live BM cells from 8-week-old wild-type and Gfi1-GFP reporter mice that assesses unmixing accuracy between FITC and GFP in the 14-fluorescent parameter panel.

SUPPLEMENTARY TABLE 1:

Reagents used for the 14-color panel and reference controls.

	Name	Alternative name	Clone	Fluorochrome	Source	Catalog number	Dilution	Reference Control Dilution
#1	Ter119	Ly-76	TER119	Purified rat Ab	BioLegend	116202	1/400	
	Mac1	CD11b	M1/70	Purified rat Ab	BioLegend	101202	1/800	
	Gr1	Ly-6C	RB6-8C5	Purified rat Ab	BioLegend	108402	1/800	
	B220	CD45R	RA-3-6B2	Purified rat Ab	BioLegend	103202	1/400	1/50
	CD5		53-7.3	Purified rat Ab	BioLegend	100602	1/800	
	CD3		17A2	Purified rat Ab	BioLegend	100202	1/200	
	CD4		GK1.5	Purified rat Ab	BioLegend	100402	1/800	
	CD8		53-6.7	Purified rat Ab	BioLegend	100702	1/800	
	Goat Anti-Rat		F(ab') ₂ -IgG	PE-Cy5	Invitrogen	A10691	1/400	1/50
#2	cKit	CD117	2B8	APC-eFluor780	eBioscience	47-1171-82	1/400	1/50
#3	Sca1	Ly-6a/e	D7	Pacific Blue	Biolegend	108120	1/400	1/50
#4	Flk2	CD135	A2F10	Biotin	eBioscience	13-1351-85	1/400	1/50
	Streptavidin			PE-Cy7	BioLegend	405206	1/400	1/50
#5	CD48	SLAMF2	HM48-1	BV711	Biolegend	103439	1/400*	1/50
#6	CD150	SLAM	TC15-12F12.2	PE	Biolegend	115904	1/400	1/50
#7	CD34	Mucosialin	RAM34	FITC	eBioscience	11-0341-85	1/25	1/50
#8	CD127	IL-7Ra	A7R34	BV785	Biolegend	135037	1/100	1/50
#9	FcγR	CD16/32	93	BV510	Biolegend	101333	1/400	1/50
#10	CD49b	ItgαII	DX5	PE/Dazzle594	Biolegend	108923	1/200	1/50
#11	EPCR	CD201	1560	PerCP-eFluor710	eBioscience	46-2012-80	1/200	1/50
#12	CD105	Endoglin	MJ7/18	APC	Biolegend	120413	1/400*	1/50
#13	CD41	ItgαIIb	MWReg30	BV605	Biolegend	133921	1/400	1/50
#14	LIVE/DEAD			Zombie NIR	BioLegend	423105	1/200	100μL/test
	UltraComp ebeads				Invitrogen	01-2222-42		~50μL/test

* Antibody used at a concentration 1/100 for LSRFortessa “conventional” flow cytometer

SUPPLEMENTARY TABLE 2
LSRFortessa instrument configuration.

Laser wave length (nm)	Laser power (mW)	Dichroic filters (nm)	Bandpass filters (nm)	Fluorochromes
355		505LP	525/50	-
		450LP	450/50	-
405	50	750LP	780/60	BV785
		690LP	710/50	BV711
		635LP	670/30	-
		600LP	610/20	BV605
		505LP	525/50	BV510
		-	450/50	Pacific Blue
488	50	685LP	695/40	PerCP-eFluor710
		505LP	530/30	FITC
561	50	750LP	780/60	PE-Cy7
		685LP	710/50	-
		635LP	660/20	PE-Cy5 / PI
		600LP	610/20	PE/Dazzle594
		-	586/15	PE
640	40	750LP	780/60	APC-eFluor780
		690LP	710/50	-
		-	670/14	APC

SUPPLEMENTARY TABLE 3:

Mean percentage (frequency of parent) and standard deviation (SD) for all gates presented in Figure 2A (n=7).

	Lin⁻	LK	LS^{low}K^{low}	LSK	LSK Flk2⁻
Mean	7.0	14.6	2.2	4.4	34.5
SD	2.2	6.0	0.8	1.8	6.8

Lymphoid and myeloid progenitors

	CLP	CMP	GMP	MEP
Mean	10.8	57.1	23.9	17
SD	5.0	6.2	3.9	4.0

Myeloid progenitors

	Pre-GM	GMP	Pre MegE	MKP	CFU-E	Pre CFU-E
Mean	43.3	22.8	33.0	4.18	12.6	10.7
SD	3.9	4.4	3.3	0.6	5.0	2.3

Stem and multipotent progenitor cells

	HSC-SLAM	MPP5	MPP2	MPP3	MPP4
Mean	32.1	14.2	19.7	32.2	62.9
SD	4.3	2.8	3.6	4.9	6.8

HSC subsets

	HSC^{LT} (EPCR)	HSC^{IT} (EPCR)	MPP1 EPCR	HSC^{LT} (CD49b)	HSC^{IT} (CD49b)	MPP1 (CD49b)
Mean	15.2	17.0	60.1	12.1	20.7	56.0
SD	5.8	1.1	6.9	1.6	6.8	4.2

SUPPLEMENTARY TABLE 4:

Mean percentage (frequency of parent) and standard deviation (SD) for all gates presented in Figure 3.

2-week-old mice (n=4)

Stem and multipotent progenitor cells

	HSC-SLAM	MPP5	MPP2	MPP3	MPP4
Mean	27	7.34	27.5	37.5	33.1
SD	3.9	1.8	2.4	3.9	19.1

HSC Subsets

	HSC ^{LT} (EPCR)	HSC ^{IT} (EPCR)	MPP1 EPCR
Mean	9.61	8.15	79.9
SD	4.8	1.1	4.5

	HSCL ^T (CD49b)	HSC ^{IT} (CD49b)	MPP1 (CD49b)
Mean	8.96	11.7	69.5
SD	2.9	3.7	3.1

	CD34 ⁻ CD41 ⁺	CD34 ⁻ CD41 ⁻	CD34 ⁺ CD41 ⁻
Mean	0.22	12	85.3
SD	0.2	4.9	3.6

	CD34 ⁻ CD105 ⁺	CD34 ⁻ CD105 ⁻	CD34 ⁺ CD105 ⁻
Mean	3.52	16.6	71.1
SD	1.7	3.1	7.0

4-week-old mice (n=6)

Stem and multipotent progenitor cells

	HSC-SLAM	MPP5	MPP2	MPP3	MPP4
Mean	32.1	14.2	19.7	32.2	62.9
SD	4.3	2.8	3.6	4.9	6.8

HSC Subsets

	HSC ^{LT} (EPCR)	HSC ^{IT} (EPCR)	MPP1 EPCR
Mean	15.2	17	60.1
SD	5.8	1.1	6.9

	HSCL ^T (CD49b)	HSC ^{IT} (CD49b)	MPP1 (CD49b)
Mean	12.1	20.7	56
SD	1.6	6.8	4.2

	CD34 ⁻ CD41 ⁺	CD34 ⁻ CD41 ⁻	CD34 ⁺ CD41 ⁻
Mean	1.3	12.9	81.4
SD	2.3	4.8	11.4

	CD34 ⁻ CD105 ⁺	CD34 ⁻ CD105 ⁻	CD34 ⁺ CD105 ⁻
Mean	6.4	13	65.6
SD	1.42	6.72	5.4

8-week-old mice (n=7)

See SUPPLEMENTARY TABLE 3:

12-month-old mice (n=5)

Stem and multipotent progenitor cells

	HSC-SLAM	MPP5	MPP2	MPP3	MPP4
Mean	52.4	14.2	12.5	20.2	41.6
SD	6.9	2.6	2.4	5.0	7.8

HSC Subsets

	HSC ^{LT} (EPCR)	HSC ^{IT} (EPCR)	MPP1 EPCR
Mean	23.3	18.5	47.8
SD	11.0	7.7	22.6

	HSCL ^T (CD49b)	HSC ^{IT} (CD49b)	MPP1 (CD49b)
Mean	25.8	20.8	26.6
SD	8.3	21.4	4.8

	CD34 ⁻ CD41 ⁺	CD34 ⁻ CD41 ⁻	CD34 ⁺ CD41 ⁻
Mean	53.7	19.3	26.2
SD	10.9	1.4	7.8

	CD34 ⁻ CD105 ⁺	CD34 ⁻ CD105 ⁻	CD34 ⁺ CD105 ⁻
Mean	57.5	13.3	17.3
SD	2.3	7.0	3.4

SUPPLEMENTARY TABLE 5:

Mean number and standard deviation (SD) of hematopoietic stem and progenitor cells (per 10⁶ live Lin⁻ cells) presented in Figure 3.

	2-week-old (n=3)	4-week-old (n=5)	8-week-old (n=4)	12-month-old (n=5)
HSC-SLAM	3,900 ± 1,323	7,080 ± 896	6,225 ± 960	14,680 ± 3,211
MPP5	1,170 ± 565	1,940 ± 371	2,850 ± 420	3,980 ± 1,160
MPP2	3,400 ± 608	9,180 ± 1,973	3,275 ± 568	3,540 ± 1,097
MPP3	4,633 ± 907	12,980 ± 3,289	6,175 ± 1,276	5,820 ± 2,473
MPP4	6,600 ± 4,214	52,400 ± 8,978	33,250 ± 8,833	20,580 ± 4,810
CLP	187 ± 190	6,900 ± 2934	3,550 ± 1,256	1,760 ± 456
CMP	43,433 ± 8,886	99,140 ± 23,126	102,625 ± 6,342	98,860 ± 24,422
GMP	35,400 ± 11,781	53,660 ± 23,315	47,375 ± 14,602	59,780 ± 25,060
MEP	21,900 ± 10,290	49,500 ± 21,675	29,025 ± 12,928	22,300 ± 6,403
Pre-GM	19,733 ± 2,040	54,620 ± 10,621	53,550 ± 2,538	52,860 ± 17,204
GMP	34,867 ± 11,152	49,820 ± 22,119	45,750 ± 14,160	54,100 ± 25,068
Mk-Ery	22,733 ± 6,191	50,240 ± 13,220	44,600 ± 4,600	29,100 ± 5,436
Pre-CFUe	5,667 ± 1604	17,720 ± 6,926	11,150 ± 2,869	9,520 ± 3,571
CFU-E	15,100 ± 9,060	21,960 ± 11,283	13,650 ± 8,812	12,960 ± 4,193
HSC ^L T (EPCR)	400 ± 157	446 ± 223	730 ± 134	n.a.*
HSC ^I T (EPCR)	500 ± 210	1,376 ± 369	950 ± 122	n.a.*
MPP1 (EPCR)	2,733 ± 929	4,540 ± 404	4,100 ± 829	n.a.*
HSC ^L T (CD49b)	480 ± 214	932 ± 242	1,040 ± 363	n.a.*
HSC ^I T (CD49b)	273 ± 125	662 ± 129	868 ± 165	n.a.*
MPP1 (CD49b)	3,066 ± 971	4,520 ± 409	4,025 ± 403	n.a.*

* n.a.: not applicable due to the shift in marker expression at this age