

SUPPLEMENTARY INFORMATION

TABLES

Supplementary Table 1

Strain or plasmid	Relevant characteristics	Source or Reference
<i>Escherichia coli</i> strains		
CC118 λ pir	F- araD139 Δ (ara, leu)7697 Δ lacX74 phoA Δ 20 galE galK thi rpsE rpoB argE _{am} recA1 λ pir	(Herrero et al., 1990)
DH5 α λ pir	F- supE44 Δ lacU169 ϕ 80d Δ lacZ Δ M15 recA1 endA1 hsdR17 thi-1 gyrA96 relA1 λ pir	(Platt et al., 2000)
DH5 α MCR	F- mcrA Δ (mrr-hsdRMS-mcrBC) ϕ 80d Δ lacZ Δ M15 Δ (lacZYA-argF)U169 deoR recA1 endA1 phoA supE44 λ - thi-1 gyrA96 relA1	(Grant et al., 1990)
S17-1 λ pir	F- recA thi pro r _K ⁻ m _K ⁺ RP4::2-Tc::MuKm Tn7 λ pir	(Herrero et al., 1990)
TOP10	F- mcrA Δ (mrr-hsdRMS-mcrBC) ϕ 80d Δ lacZ Δ M15 Δ lacX74 recA1 araD139 Δ (ara-leu)7697 galU galK rpsL(Str ^R) endA1 nupG λ -	Invitrogen
EcN	<i>E. coli</i> Nissle 1917 wild-type	Ardeypharm, Germany
JZL95 (znuA)	EcN Δ znuA(-82 to +1000)	This study
JZL109 (znuA)	EcN Δ znuA(-82 to +1000)::Kan	This study
JZL100 (znuA zupT)	EcN Δ znuA(-82 to +1000) Δ zupT(-42 to +774)::Kan	This study
JB90 (ybtX)	EcN Δ ybtX(-108 to +1281)::Cm	This study
JB92 (znuA ybtX)	EcN Δ znuA(-82 to +1000)::Kan Δ ybtX(-108 to +1281)::Cm	This study
JB76 (znuA zupT ybtX)	EcN Δ znuA(-82 to +1000) Δ zupT(-42 to +774)::Kan Δ ybtX(-108 to +1281)::Cm	This study
HZE116 (irp2)	EcN Δ irp2(+1 to +6108)::Tet	This study
HZE112 (znuA zupT irp2)	EcN Δ znuA(-82 to +1000) Δ zupT(-42 to +774)::Kan Δ irp2(+1 to +6108)::Tet	This study
<i>Salmonella enterica</i> serovar Typhimurium strains		

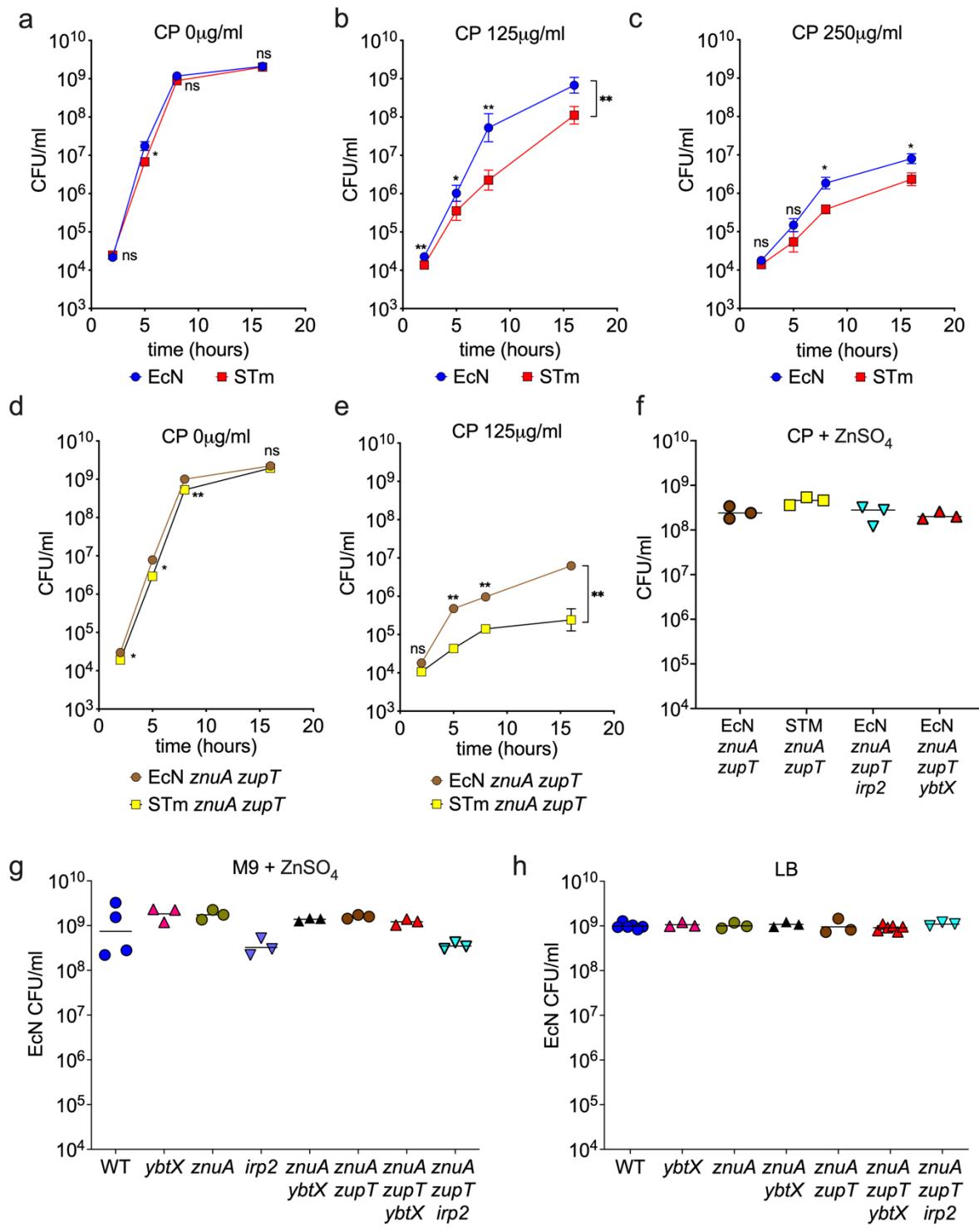
IR715	ATCC 14028s Nal ^R	(Stojiljkovic et al., 1995)
JZL3	IR715 $\Delta znuA::Cm$	(Liu et al., 2012)
APJ4	IR715 $\Delta znuA::Cm \Delta zupT::KSAC$	(Cerasi et al., 2013)
MSC74	IR715 $\Delta iroN::tetRA$	(Sassone-Corsi et al., 2016)
Plasmids		
pBS34	pBluescriptII KS+ ::[XbaI][PstI]KSAC[PstI][XbaI], Carb ^R Kan ^R	(Raffatellu et al., 2009)
pCRBlunt II-TOPO	TA-Cloning Vector, Kan ^R	Invitrogen
pGP704	<i>oriR6K mobRP4</i> , Carb ^R	(Miller and Mekalanos, 1988)
pHP45Ω	Strep ^R Carb ^R	(Prentki and Krisch, 1984)
pKD3	Carb ^R Cm ^R	(Datsenko and Wanner, 2000)
pKD4	Carb ^R Kan ^R	(Datsenko and Wanner, 2000)
pJK611	pKD46:: <i>sacB</i>	Kelly T. Hughes
pJB10	pGP704:: <i>ybtX</i> FR1-Cm-FR2, Cm ^R , Carb ^R	This study
pNM3	pCRBlunt II-TOPO:: <i>znuA</i> FR1+FR2, Kan ^R	This study
pNM4	pRDH10:: <i>znuA</i> FR1-FR2, Cm ^R	This study
pNM4::KSAC	pRDH10:: <i>znuA</i> FR1-KSAC-FR2, Cm ^R Kan ^R	This study
pRDH10	<i>oriR6K mobRP4 sacRB</i> , Cm ^R Tet ^R	(Kingsley et al., 1999)
pSW172	<i>oriR101 repA101ts</i> , Carb ^R	(Lopez et al., 2012)
pHZE107	pGP704:: <i>irp2</i> FR1-Tet-FR2, Tet ^R Carb ^R	This study

Supplementary Table 2

Name	Sequence (5' - 3')	Source
<i>znuA</i> FR1-Fw	CACATT <u>GGATCC</u> GTTGTCAGCACACGAG	This study

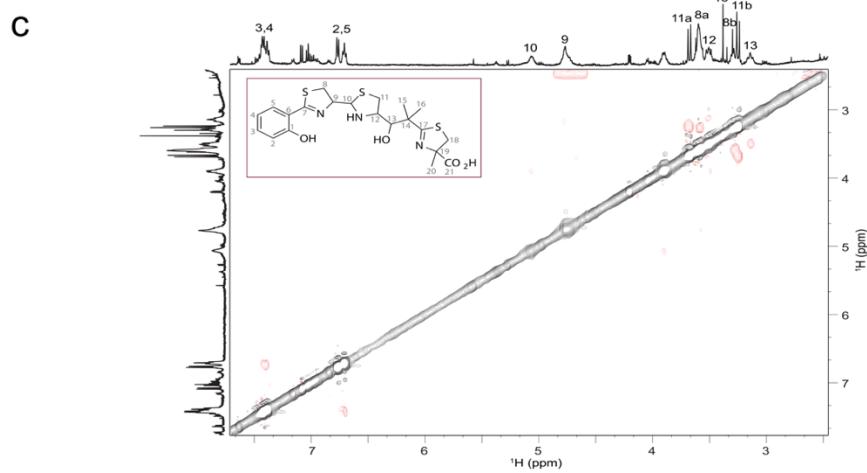
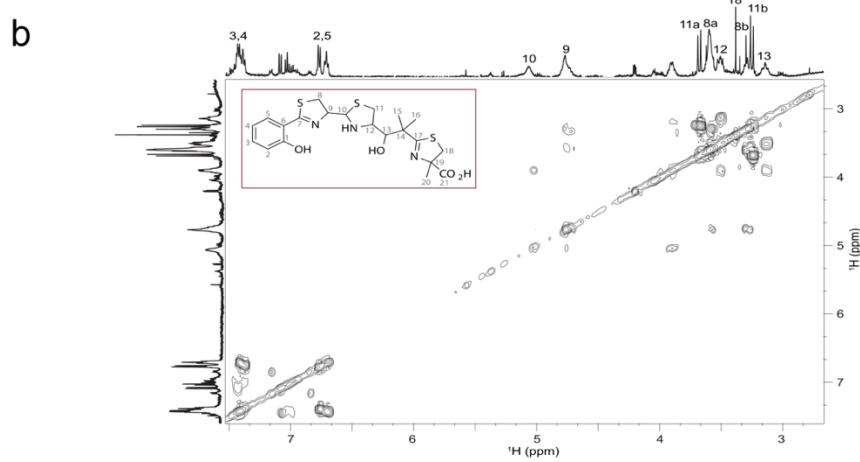
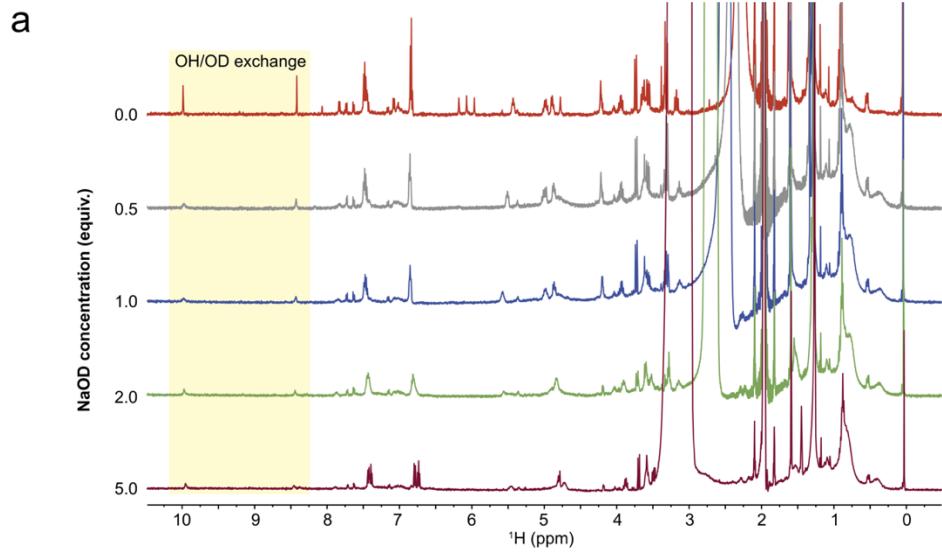
<i>znuA</i> FR1-Rv	TATGCCT <u>CTAGACAAGTCTGGTTCCCTGG</u>	This study
<i>znuA</i> FR2-Fw	TATGCCT <u>CTAGACACACCGCGTTATGTTGG</u>	This study
<i>znuA</i> FR2-Rv	CACATT <u>GGATCCCGGA</u> ACTGTTGCCGTC	This study
<i>zupT</i> -Fw	GTAAGAACCCGGATAACAATGATGATGATCATCA GTTATTGTGTAGGCTGGAGCTGCTTC	This study
<i>zupT</i> -Rv	TGTTGCCTTAGCAATGGGAAACATCTGTCATTAT CGTCTATGGGAATTAGCCATGGTCC	This study
LB_ybtX_fwd	gccacactgcagatctgcaggCAGACCCCAGATGCTGAAC	This study
LB_ybtX_rev	gtcacaggtaTCCCGCGAATAAACACAGAG	This study
CmR_fwd	tattcgccgaTACCTGTGACGGAAGATCACTTCGC	This study
CmR_rev	cagggtactcCTTACGCCCGCCCTGCC	This study
RB_ybtX_fwd	ggggcgtaagGAGTACCCCTGCTCAACCACCTGTCCC TC	This study
RB_ybtX_rev	aattcccggagagctcgatAGCCGGCGCTGCTGGCG	This study
ybtX_pres_FW	AAAGAGGGTGAAGTCGACAC	This study
ybtX_pres_RV	GTCTATGCAGTCCTACCCG	This study
ybtX_LB_FW	CTGACGGAACATAAACGAGC	This study
ybtX_RB_RV	TACAGGTGGTGGTATTCC	This study
Gib_out_LB_FW	CAATTGCATAATCGCGTTCA	This study
Gib-out-RB-RV	AGGGGATGATGGTATTGCGC	This study
ybtX_RED_FW	CACTGATTTCATCCCTTACCTCTGTGTTATT GC GGAGTGTAGGCTGGAGCTGCTTC	This study
ybtX_LB_FW	CTGGTGATGGAAGAGGGACAGGTGGTTGAGCAG GGTACTCCATATGAATATCCTCCTTA	This study

Gib-out-RB-RV	AGGGGATGATGGTATTGCGC	This study
ybtX_RED_FW	CACTGATTTCATCCCTTACCTCTCTGTGTTATTC GCGGAGTGTAGGCTGGAGCTGCTTC	This study
ybtX_LB_FW	CTGGTGATGGAAGAGGGACAGGTGGTGAGCAG GGTACTCCATATGAATATCCTCCTTA	This study
TetR-F	TTAAGACCCACTTCACATTAAAG	This study
TetA-R	CTAACGCAC TTGTCT CCTGTTAC	This study
pGP704-sall US irp2 F	acctgcagatctgcaggcgacTCAGTCTGGTGCTGGATG	This study
TetR-US irp2 R	AATGTGAAAGTGGGTCTTAATCTTCCTCCTGATG GCACG	This study
TetA-DS irp2 F	AACAGGAGACAAGTGCTTAGCGCGAAGCAA GATTTCC	This study
pGP704-sacl DS-irp2 R	tgcgaattcccgggagagctcTGGCAATATA TG	This study
irp2up-TetRA F	ATGCTTTCGGTAAGACGTGCCATCAGGAGGAAG ATTAAGACCCACTTCACATTAAAG	This study
irp2dn-TetRA R	GATGGCGTCCGGGAAAATCAGTTGCTTCGC GCTAACGCAC TTGTCTCCTGTTAC	This study
irp2-inF	ACAACGCTTCCTCGTACAATG	This study
Irp2-inR	TCTGTTCGAGCACCTGTTGC	This study
Irp2-upF	TTTGGCGTAACTCCTTCGAC	This study
Irp2-dnR	TTCATAAAGTCAATGGAACG	This study



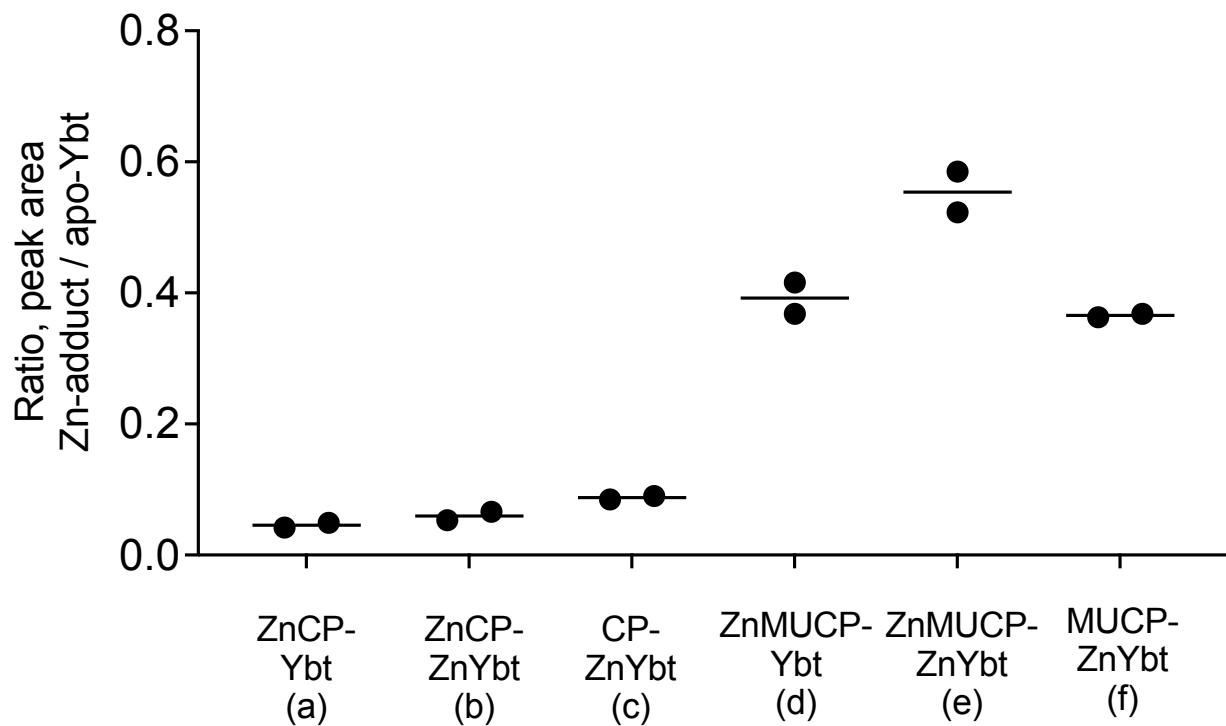
Supplementary Figure 1. Wild-type and mutant strains grow to similar abundance in rich media or in zinc-limiting media supplemented with zinc.

(a-f) EcN and STm strains were grown in modified LB medium containing 0, 125, 250 or 150 µg/ml CP and 5 µM ZnSO₄ for 16 h static incubation at 37 °C. For **a-e** samples were taken 2 h, 5 h, 8 h and 16 h, and for **f** 16 h after inoculation. Two-sided one sample *t* test was used on log transformed data to accept or reject null hypothesis (theoretical mean = 0). For **a-c**, EcN n = 4 biologically independent replicates, STm n = 3 biologically independent replicates. For **d-f**, n = 3 biologically independent replicates. **(g)** EcN strains were grown in M9 medium supplemented with 5 µM ZnSO₄ for 20 h shaking incubation at 37 °C. Different batches of calprotectin were used for **a-e** compared to **f** and main Figure 1. **(h)** EcN strains were grown in LB medium for 20 h shaking incubation at 37 °C. **(f-h)** Each symbol represents an independent biological replicate. **(a-e)** Data are presented as geometric mean values +/- geometric SD. * *P* value ≤0.05; ** *P* value ≤0.01; ns = not significant. Exact *P* values are reported in Supplementary Data 2. Source data are provided as a Source Data file.

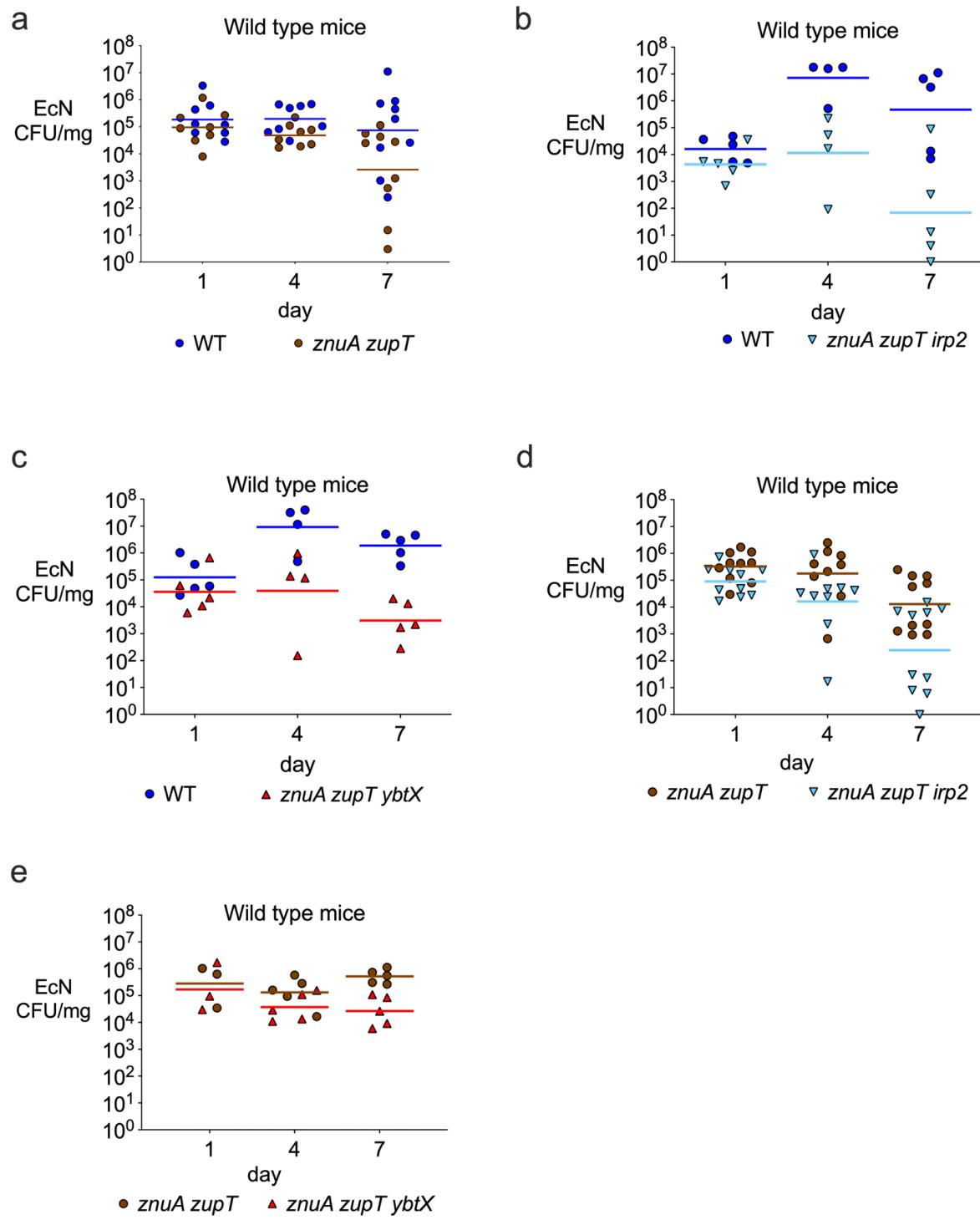


Supplementary Figure 2. Verification of NMR resonance assignments of Ybt by 2D NMR, and lack of significant changes in Ybt resonances upon the addition of base.

(a) ^1H NMR 1D spectra of Ybt dissolved in CD_3CN in the presence of 5.0 equiv. of ZnCl_2 (top trace, 0 equiv. of NaOD , red trace) and increasing amounts of NaOD (0.5 equiv., gray trace), (1.0 equiv., blue trace), (2.0 equiv., green trace), and (5.0 equiv., brown trace). There are not changes in the spectrum other than the loss of OH signal intensity upon addition of NaOD (highlighted in yellow), which is attributed to increased solvent exchange. The starting spectrum of Ybt in the presence of 5.0 equiv. ZnCl_2 shows hydrolysis products (signal at 6.2 ppm) in addition to bound and unbound Ybt isomers. (b) 2D gradient-enhanced ^1H - ^1H COSY showing aliphatic and aromatic region of spectrum for Ybt dissolved in CD_3CN . (c) 2D gradient-enhanced ^1H - ^1H ROESY showing aliphatic and aromatic region of spectrum for Ybt dissolved in CD_3CN . All spectra were acquired at 500 MHz.

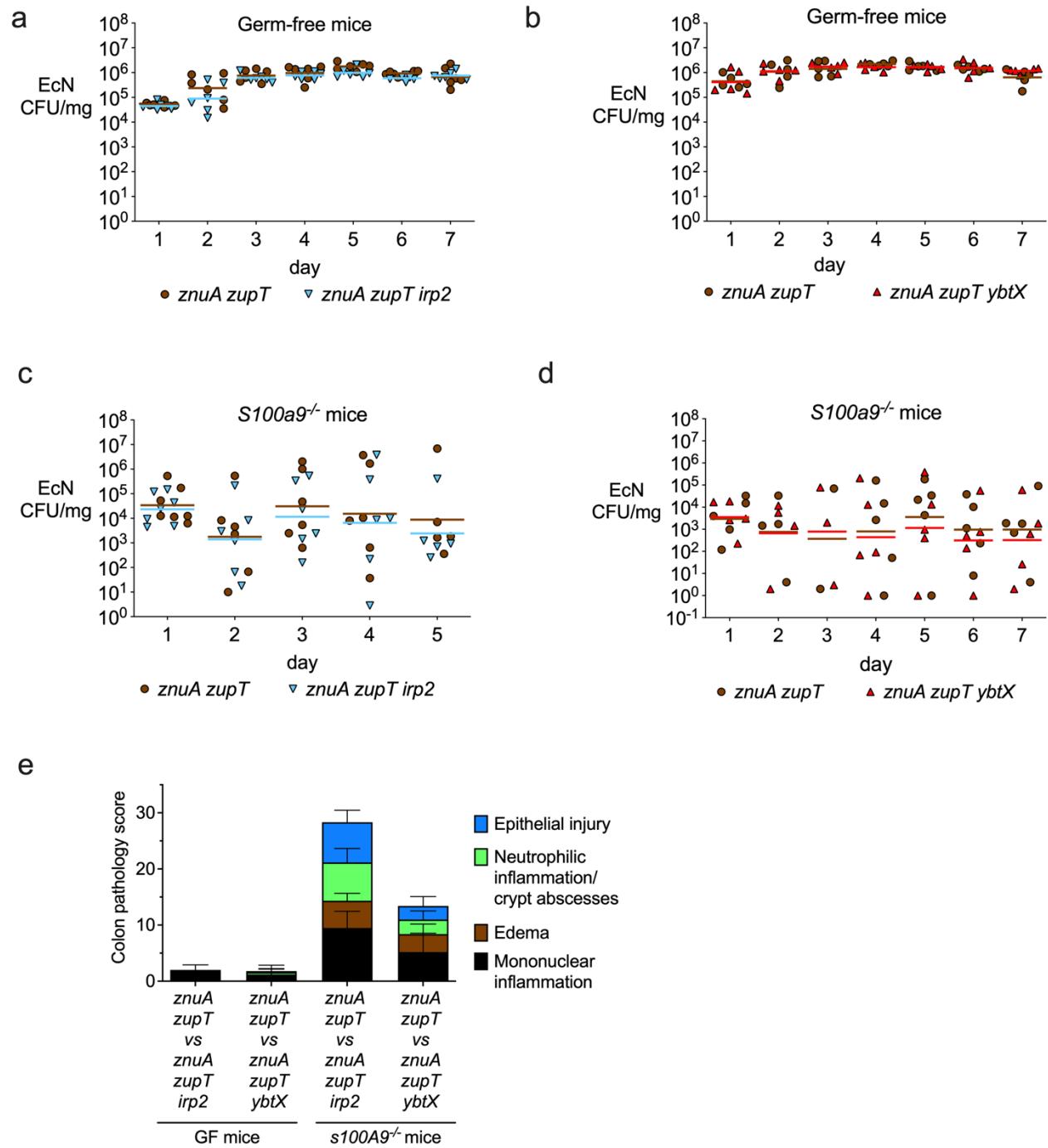


Supplementary Figure 3. Direct-injection mass spectrometry competition experiments address whether calprotectin (CP) and yersiniabactin (Ybt) exchange zinc. Competition was carried out between (a) zinc-CP and apo-Ybt, (b) zinc-CP and zinc-Ybt, (c) CP and zinc-Ybt. Control experiments were carried out with mutant-CP (mutCP), which is unable to bind zinc, in place of CP in the following (d) zinc-mutCP and Ybt, (e) zinc-mutCP and zinc-Ybt, and (f) mutCP and zinc-Ybt. Zinc-CP = pre-incubated 0.8 eq Zn and CP-zinc binding site, zinc-Ybt = preincubated with 1eq Zn, zinc-mutCP = preincubated 0.8 eq of Zn with 1 eq mutated CP. Source data are provided as a Source Data file.



Supplementary Figure 4. EcN fecal CFU from mice in Figure 4.

(**a-e**) CFU of EcN strains in the fecal content of wild-type C57BL/6 mice treated with DSS, all inoculated with the indicated 1:1 mixtures of EcN strains. Dark blue bars represent the geometric mean of EcN wild-type in **a-c**; brown bars represent the geometric mean of the *znuA zupT* mutant in **a, d, e**; light blue bars represent the geometric mean of the *znuA zupT irp2* mutant in **b, d**; red bars represent the geometric mean of the *znuA zupT ybtX* mutant in **c, e**. Each symbol represents data from a biologically independent fecal sample from a single mouse. Source data are provided as a Source Data file.



Supplementary Figure 5. EcN fecal CFU and pathology scores from mice in Figure 5.

(**a-d**) CFU of EcN strains in the fecal content of (**a, b**) wild-type germ-free Swiss Webster mice or (**c, d**) C57BL/6 *S100a9^{-/-}* mice, all inoculated with the indicated 1:1 mixtures of EcN strains. Brown bars represent the geometric mean of the *znuA* *zupT* mutant in each experiment; light blue bars represent the geometric mean of the *znuA* *zupT* *irp2* mutant in **a, c**; red bars represent the geometric mean of the *znuA* *zupT* *ybtX* mutant in **b, d**. Each symbol represents data from a biologically independent fecal sample from a single mouse. (**e**) Pathology scores of colonic tissue from germ-free mice in panel **a, b** (day 7) or of colonic tissue from *S100a9^{-/-}* DSS-treated mice in panel **c** (day 5) and panel **d** (day 7). Germ-free EcN *znuA* *zupT* vs *znuA* *zupT* *irp2*, n = 6; germ-free EcN *znuA* *zupT* vs *znuA* *zupT* *ybtX*, n = 5; *S100a9^{-/-}* mice EcN *znuA* *zupT* vs *znuA* *zupT* *irp2*, n = 6; *S100a9^{-/-}* mice EcN *znuA* *zupT* vs *znuA* *zupT* *ybtX*, n = 5; all colon samples are biologically independent. Data are presented as mean values +/- SD. Source data are provided as a Source Data file.

Supplementary References

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