

Supplementary Information for

**The *E. coli* two-component signal sensor BarA binds protonated acetate via a conserved hydrophobic binding pocket**

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**This PDF file includes:**

Figure S1

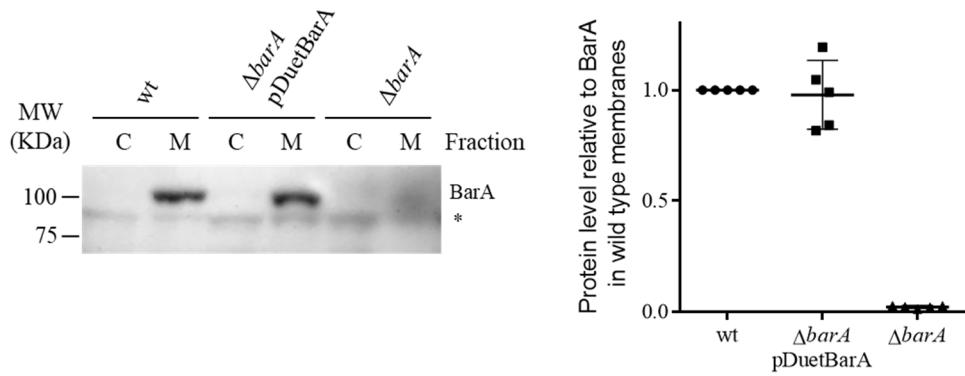
Figure S2

Table S1

Table S2

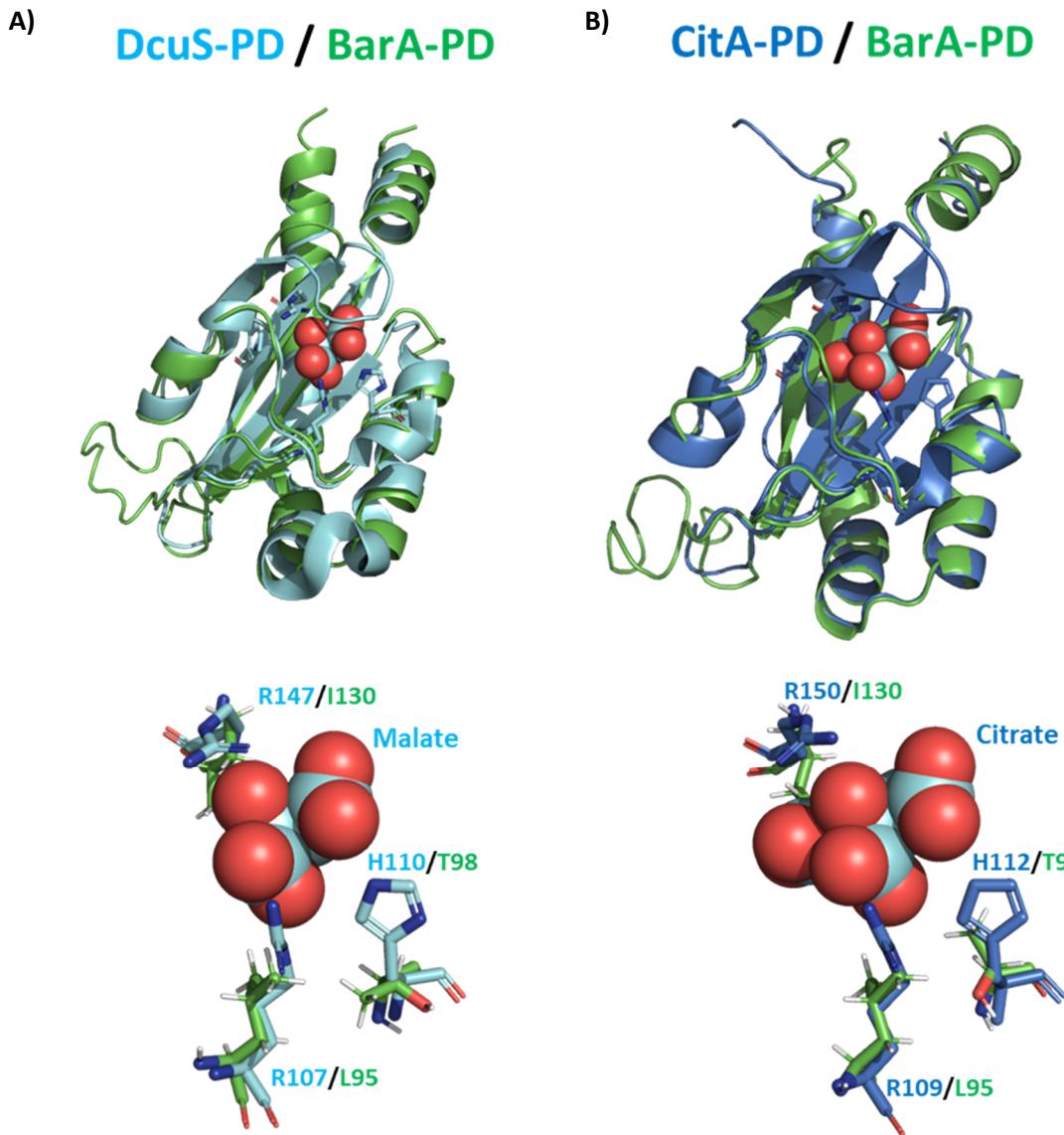
Table S3

**Figure S1**



**Fig. S1. Complementation of a *barA* mutant strain with plasmid pDuetBarA results in nearly wild-type BarA levels.** BarA protein (102.5 kDa) levels in cytosolic (C) and membrane (M) fractions of strains KSB837 (WT), IFC5035 ( $barA^-$ ) carrying the pDuetBarA plasmid, and IFC5035 ( $barA^-$ ), as determined by Western blot analyses using BarA polyclonal antibodies. A quantification from five independent blots is shown on the right, indicating mean  $\pm$  SD, with individual values indicated (wt, filled circles; IFC5035 ( $\Delta barA$ ) carrying pDuetBarA, filled squares; and IFC5035 ( $\Delta barA$ ), triangles). \* indicates a nonspecific band present in all stains and fractions.

**Figure S2**



**Fig. S2. Structural models of BarA-PD based on the DcuS and CitA sensor domains.** (A) (Upper panel) Cartoon of the structural model of BarA-PD generated by using the program I-Tasser and the crystal structure of the DcuS sensor domain as the template (PDB: 3BY8). The BarA-PD model (green) and the template (light blue), which includes the bound malate molecule, are shown overlapped. (Lower panel) The relative positions of the DcuS amino acid residues needed for malate binding (light blue) to the corresponding BarA amino acid residues (green), in the generated model, are depicted. (B) (Upper panel) Cartoon of the structural model of BarA-PD generated by using the program I-Tasser and the crystal structure of the CitA sensor domain as the template (PDB: 1P0Z). The BarA-PD model (green) and the template (blue), which includes a bound citrate molecule, are shown overlapped. (Lower panel) The relative positions of the CitA amino acid residues needed for citrate binding (light blue) to the corresponding BarA amino acid residues (green), in the generated model, are depicted.

**Table S1.** Bacterial strains and plasmids used in this study

| Strain or plasmid  | Relevant characteristics  | Source or reference |
|--------------------|---|---------------------|
| <b>Strain</b>      |   |                     |
| KSB837             | CF7789 λΦ( <i>csrB'-lacZ</i> )  | (7)                 |
| IFC5035            | KSB837 Δ <i>barA</i> ::Kan <sup>r</sup>   | This work           |
| IFC5036            | KSB837 Δ <i>barA</i>  | This work           |
| IFC5038            | KSB837 <i>barA</i> <sup>S99A</sup> ::Cam <sup>r</sup>   | This work           |
| IFC5039            | KSB837 <i>barA</i> <sup>H102A</sup> ::Cam <sup>r</sup>  | This work           |
| IFC5040            | KSB837 <i>barA</i> <sup>R124A</sup> ::Cam <sup>r</sup>  | This work           |
| IFC5041            | KSB837 <i>barA</i> <sup>I130A</sup> ::Cam <sup>r</sup>  | This work           |
| IFC5042            | KSB837 <i>barA</i> <sup>R132A</sup> ::Cam <sup>r</sup>  | This work           |
| KSB-ackA-ptA       | KSB837 <i>ackA</i> ::Tet <sup>r</sup> ::ptA   | (15)                |
| <b>Plasmid</b>     |   |                     |
| pEXT22             | Low copy number vector, Kan <sup>r</sup>  | (56)                |
| pEXT22(-NdeI)      | pEXT22 derivative, carries 2-bp insertion as a result of end filling and ligation at the NdeI site  | This work           |
| pEXT22-barA        | <i>barA</i> under native promoter in pEXT22(-NdeI)  | This work           |
| pEXT22-barAcyt     | <i>barA</i> <sup>198-918</sup> under native promoter in pEXT22(-NdeI)   | This work           |
| pEXT22-barAPDarcB  | <i>barA</i> <sup>1-31</sup> - <i>arcB</i> <sup>43-57</sup> - <i>barA</i> <sup>176-918</sup> under <i>barA</i> native promoter in pEXT22(-NdeI)  | This work           |
| pEXT22-barAPDgacS  | <i>barA</i> <sup>1-34</sup> - <i>gacS</i> <sup>23-154</sup> - <i>barA</i> <sup>176-918</sup> under <i>barA</i> native promoter in pEXT22(-NdeI) | This work           |
| pUC18              | Cloning vector, Amp <sup>r</sup>  | (55)                |
| pUC18-barA         | <i>barA</i> <sup>WT</sup> in pUC18  | This work           |
| pUC18-Cam          | Cloning vector, Amp <sup>r</sup> Cam <sup>r</sup>   | This work           |
| pBarAS99ACam       | <i>barA</i> <sup>S99A</sup> ::Cam <sup>r</sup> in pUC18   | This work           |
| pBarAH102ACam      | <i>barA</i> <sup>H102A</sup> ::Cam <sup>r</sup> in pUC18  | This work           |
| pBarAR124ACam      | <i>barA</i> <sup>R124A</sup> ::Cam <sup>r</sup> in pUC18  | This work           |
| pBarAI130ACam      | <i>barA</i> <sup>I130A</sup> ::Cam <sup>r</sup> in pUC18  | This work           |
| pBarAR132ACam      | <i>barA</i> <sup>R132A</sup> ::Cam <sup>r</sup> in pUC18  | This work           |
| pKD3               | Template plasmid for <i>cat</i> cassette amplification flanked by FRT sequences, Amp <sup>R</sup> , Cam <sup>r</sup>                            | (53)                |
| pKD4               | Template plasmid for <i>kan</i> cassette amplification flanked by FRT sequences, Amp <sup>R</sup> , Kan <sup>r</sup>                            | (53)                |
| pACYCDuet-1        | T7 promoter/ <i>lac</i> operator expression vector, Cam <sup>r</sup>  | Novagen             |
| pACYCDuet-1(-Xhol) | pACYCDuet-1 derivative, carries 4-bp insertion as a result of end filling and ligation at the Xhol site   | This work           |
| pDuetBarA          | <i>barA</i> under the control of the T7 promoter in pACYCDuet-1(-Xhol)  | This work           |
| pDuetBarAXho       | <i>barA</i> with an inserted Xhol restriction site by silent mutation ( <i>barA</i> <sup>Xhol</sup> ) in pACYCDuet-1(-Xhol)                     | This work           |

|                |   |           |
|----------------|---|-----------|
| pDuetBarAL52P  | $\text{barA}^{\text{Xhol L52P}}$ in pACYCDuet-1(-Xhol)  | This work |
| pDuetBarAV83D  | $\text{barA}^{\text{Xhol V83D}}$ in pACYCDuet-1(-Xhol)  | This work |
| pDuetBarAV88D  | $\text{barA}^{\text{Xhol V88D}}$ in pACYCDuet-1(-Xhol)  | This work |
| pDuetBarAL95F  | $\text{barA}^{\text{Xhol L95F}}$ in pACYCDuet-1(-Xhol)  | This work |
| pDuetBarAT98A  | $\text{barA}^{\text{Xhol T98A}}$ in pACYCDuet-1(-Xhol)  | This work |
| pDuetBarAT98H  | $\text{barA}^{\text{Xhol T98H}}$ in pACYCDuet-1(-Xhol)  | This work |
| pDuetBarAI136N | $\text{barA}^{\text{Xhol I136N}}$ in pACYCDuet-1(-Xhol) | This work |

**Table S2.** Oligonucleotides used in this study

| Primer name       | Primer sequence (5'-3')   | Use  |
|-------------------|---|--|
| BarA100up-Fw      | GACGCGTCGTGCGTC   | Used to amplify the <i>barA</i> promoter region  |
| BA5R              | TGCTCTAGAAGATCTGATCATATGGAGTTCCGTTATGGG                         | Used to amplify the <i>barA</i> promoter region  |
| barA-Ndel-Fw      | CAACATATGACCAACTACAGCCTGCGC                                     | Used to amplify the full-length <i>barA</i> gene and the megaprimer to generate <i>barA</i> punctual mutants |
| BAfullR-Hind      | CCCAAGCTTGCCGATTGCTACTGACAAGA                                   | Used to amplify the full-length <i>barA</i> gene   |
| BarA5'(198-x)     | CCCGGATCCCATATGCGCGATGTAACCGGTCCG                               | Used to amplify the coding sequence of the cytosolic BarA variant  |
| ChGS-perip-Fw     | CTGAGTATCTTTCGTCGTGCATCGCTATTGGCCGAGAT<br>GCACGGC               | Used to construct the BarA <sup>PDGacS</sup> chimeric HK   |
| ChGS-perip-Rv     | CATCATCACGCTGAAATAAGATCTCGCGTAGCCCTGC<br>AGCAG                  | Used to construct the BarA <sup>PDGacS</sup> chimeric HK   |
| GS-perip-Rv       | GCGGTAGCCCTGCAGCAG  | Used to construct the BarA <sup>PDGacS</sup> chimeric HK   |
| GS-perip-Fw       | TTGGCCGAGATGCACGGC  | Used to construct the BarA <sup>PDGacS</sup> chimeric HK   |
| Arc-Bar-peripl-Fw | GTCGAAAGCATTGATGTTATTCGTGAGATCTTATTCCAG<br>CGTG                 | Used to construct the BarA <sup>PDArcB</sup> chimeric HK   |
| Arc-Bar-peripl-Rv | CTGACCATGCAGCACCATGGTTACGACGAAAAAGATACTC<br>AGC                 | Used to construct the BarA <sup>PDArcB</sup> chimeric HK   |
| barAdel-Fw        | ATTTAACAGTGTGACCTAATTGTCCTAACGGAACTCCG<br>TGTAGGCTGGAGCTGCTTC   | Used to delete the <i>barA</i> gene  |
| barAdel-Rv        | CATAAACACAGGCACCTTGTACCAATCTGAAACCAGCGTA<br>TGAATATCCTCCTAGTTCC | Used to delete the <i>barA</i> gene and to integrate <i>barA</i> variants into the chromosome                |
| pKDEco2-Rv        | CGGAATTCATGAATATCCTCCTAGTTTC                                    | Used to amplify the <i>cat</i> gene from   |

|                 |  |  |
|-----------------|--|--|
|                 |  | plasmid pKD3   |
| pKD-Kpn-Hind-Fw | GGGGTACCAAGCTGTGAGGCTGGAGCTGCTTC                                 | Used to amplify the <i>cat</i> gene from plasmid pKD3  |
| barAmut-ins-Fw  | ATTTAACAGTGTGACCTTAATTGTCCCATAACGGAACCTCCA<br>TGACCAACTACAGCCTGC | Used to integrate <i>barA</i> variants into the chromosome   |
| BarA-S99A-Rv    | GATCAAGATGAAAATTGGCGGTGACAAAGAGTCGG                              | Used to generate <i>barA</i> punctual mutant by megaprimer amplification   |
| BarA-H102A-Rv   | CATTGATGAGGGATCAAGAGCAAATTGGAGGTGACAAA<br>GAG                    | Used to generate <i>barA</i> punctual mutant by megaprimer amplification   |
| BarA-R124A-Rv   | GATCATAATATGCCGTAGCAGTGACAGTGAGCTGGCG                            | Used to generate <i>barA</i> punctual mutant by megaprimer amplification   |
| BarA-I130A-Rv   | GAAATAATCGCGTGCAGGAGGGCCATAATATGCCGTAC<br>G                      | Used to generate <i>barA</i> punctual mutant by megaprimer amplification   |
| BarA-R132A-Rv   | CTCAGAAATAATCGCGTGGCGAGGATCATAATATGCC                            | Used to generate <i>barA</i> punctual mutant by megaprimer amplification   |
| barAf1Ncol      | CAACCATGGCGACCAACTACAGCCTGCGCGCAC                                | Used to amplify the full-length <i>barA</i> gene and the <i>barA</i> periplasmic region  |
| BarAXholTM2-Fw  | GCAGCAATATAAGAGATCTTATCTGAGCGTGATGATG<br>CTG                     | Used to create a Xhol restriction site in the BarA sequence by a silent mutation   |
| BarAXholTM2-Rv  | CAGCATCATCACGCTCGAGATAAGATCTTTATATTGCT<br>GC                     | Used to create a Xhol restriction site in the BarA sequence by a silent mutation and to amplify the <i>barA</i> periplasmic region |



Table S3. NCBI reference sequence accessions of BarA orthologs used in phylogenetic analysis.

| Accession      | Organism                                    | Class               | Order             | Family                      |
|----------------|---|---------------------|-------------------|-----------------------------|
| WP_197471059.1 | <i>Azotobacter vinelandii</i>               | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_186415863.1 | <i>Pseudomonas stutzeri</i>                 | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_022965696.1 | <i>Pseudomonas caeni</i>                    | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_053111169.1 | <i>Oblitimonas alkaliphila</i>              | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_034024066.1 | <i>Pseudomonas aeruginosa</i>               | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_011062714.1 | <i>Pseudomonas protegens</i>                | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_071871354.1 | <i>Pseudomonas hussainii</i>                | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_022960986.1 | <i>Pseudomonas pelagia</i>                  | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_093397665.1 | <i>Pseudomonas xinjiangensis</i>            | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_083727290.1 | <i>Pseudomonas pachastrella</i>             | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_092288012.1 | <i>Pseudomonas sabulinigri</i>              | Gammaproteobacteria | Pseudomonadales   | <i>Pseudomonadaceae</i>     |
| WP_072324365.1 | <i>Marinospirillum alkaliphilum</i>         | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| WP_091961386.1 | <i>Marinospirillum celere</i>               | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| WP_068997271.1 | <i>Terasakiispira papahanaumokuakeensis</i> | Gammaproteobacteria | Oceanospirillales | <i>incertae sedis</i>       |
| WP_027849455.1 | <i>Marinospirillum minutulum</i>            | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| WP_051789246.1 | <i>Endozoicomonas montiporae</i>            | Gammaproteobacteria | Oceanospirillales | <i>Endozoicomonadaceae</i>  |
| WP_020583285.1 | <i>Endozoicomonas elysicola</i>             | Gammaproteobacteria | Oceanospirillales | <i>Endozoicomonadaceae</i>  |
| WP_177166874.1 | <i>Pseudospirillum japonicum</i>            | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| WP_036567047.1 | <i>Oceanospirillum beijerinckii</i>         | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| WP_077242806.1 | <i>Oceanospirillum linum</i>                | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| WP_027706946.1 | <i>Zooshikella ganghwensis</i>              | Gammaproteobacteria | Oceanospirillales | <i>Hahellaceae</i>          |
| WP_011395717.1 | <i>Hahella chejuensis</i>                   | Gammaproteobacteria | Oceanospirillales | <i>Hahellaceae</i>          |
| WP_020406163.1 | <i>Hahella ganghwensis</i>                  | Gammaproteobacteria | Oceanospirillales | <i>Hahellaceae</i>          |
| WP_015486099.1 | <i>Thalassolituus oleivorans</i>            | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| WP_156879095.1 | <i>Oceanobacter kriegii</i>                 | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| TPD55691.1     | <i>Oleibacter marinus</i>                   | Gammaproteobacteria | Oceanospirillales | <i>Oceanospirillaceae</i>   |
| WP_044617057.1 | <i>Gynuella sunshinyii</i>                  | Gammaproteobacteria | Oceanospirillales | <i>Saccharospirillaceae</i> |

|                       |   |                       |                       |                                   |
|-----------------------|---|-----------------------|-----------------------|-----------------------------------|
| <b>WP_041655427.1</b> | <i>Marinobacter hydrocarbonoclasticus</i> | Gammaproteobacteria   | Alteromonadales       | <i>Alteromonadaceae</i>           |
| <b>WP_008937433.1</b> | <i>Marinobacter santoriniensis</i>        | Gammaproteobacteria   | Alteromonadales       | <i>Alteromonadaceae</i>           |
| <b>WP_091852392.1</b> | <i>Marinobacter segnicrescens</i>         | Gammaproteobacteria   | Alteromonadales       | <i>Alteromonadaceae</i>           |
| <b>WP_004583184.1</b> | <i>Marinobacter nanhaiicus</i>            | Gammaproteobacteria   | Alteromonadales       | <i>Alteromonadaceae</i>           |
| <b>WP_020680514.1</b> | <i>Marinobacterium rhizophilum</i>        | Gammaproteobacteria   | Oceanospirillales     | <i>Oceanospirillaceae</i>         |
| <b>WP_010947629.1</b> | <i>Legionella pneumophila</i>             | Gammaproteobacteria   | Legionellales         | <i>Legionellaceae</i>             |
| <b>WP_058535550.1</b> | <i>Legionella saoudiensis</i>             | Gammaproteobacteria   | Legionellales         | <i>Legionellaceae</i>             |
| <b>MBV35913.1</b>     | <i>Rickettsiales bacterium</i>            | Alphaproteobacteria   | Rickettsiales         | <i>unclassified Rickettsiales</i> |
| <b>WP_045972244.1</b> | <i>Xenorhabdus doucetiae</i>              | Gammaproteobacteria   | Enterobacterales      | <i>Morganellaceae</i>             |
| <b>VTQ57351.1</b>     | <i>Campylobacter jejuni</i>               | Epsilonproteobacteria | Campylobacterales     | <i>Campylobacteraceae</i>         |
| <b>WP_026741805.1</b> | <i>Lonsdalea quercina</i>                 | Gammaproteobacteria   | Enterobacterales      | <i>Pectobacteriaceae</i>          |
| <b>WP_029593062.1</b> | <i>Siccibacter turicensis</i>             | Gammaproteobacteria   | Enterobacterales      | <i>Enterobacteriaceae</i>         |
| <b>WP_101348641.1</b> | <i>Escherichia coli</i>                   | Gammaproteobacteria   | Enterobacterales      | <i>Enterobacteriaceae</i>         |
| <b>WP_194445129.1</b> | <i>Klebsiella pneumoniae</i>              | Gammaproteobacteria   | Enterobacterales      | <i>Enterobacteriaceae</i>         |
| <b>WP_062456638.1</b> | <i>Vibrio shilonii</i>                    | Gammaproteobacteria   | Vibrionales           | <i>Vibrionaceae</i>               |
| <b>WP_086940253.1</b> | <i>Thaumasiovibrio occultus</i>           | Gammaproteobacteria   | Vibrionales           | <i>Vibrionaceae</i>               |
| <b>WP_008485098.1</b> | <i>Gallaecimonas xiamenensis</i>          | Gammaproteobacteria   | <i>incertae sedis</i> | -                                 |
| <b>WP_087036954.1</b> | <i>Oceanisphaera profunda</i>             | Gammaproteobacteria   | Aeromonadales         | <i>Aeromonadaceae</i>             |
| <b>TXH95157.1</b>     | <i>Pseudorhodobacter sp.</i>              | Alphaproteobacteria   | Rhodobacterales       | <i>Rhodobacteraceae</i>           |
| <b>WP_028109905.1</b> | <i>Ferrimonas futtsuensis</i>             | Gammaproteobacteria   | Alteromonadales       | <i>Ferrimonadaceae</i>            |
| <b>WP_031567826.1</b> | <i>Pararheinheimera texensis</i>          | Gammaproteobacteria   | Chromatiales          | <i>Chromatiaceae</i>              |
| <b>WP_011073304.1</b> | <i>Shewanella oneidensis</i>              | Gammaproteobacteria   | Alteromonadales       | <i>Shewanellaceae</i>             |
| <b>WP_011495637.1</b> | <i>Shewanella denitrificans</i>           | Gammaproteobacteria   | Alteromonadales       | <i>Shewanellaceae</i>             |
| <b>WP_089349171.1</b> | <i>Pseudoalteromonas espejiana</i>        | Gammaproteobacteria   | Alteromonadales       | <i>Pseudoalteromonadaceae</i>     |
| <b>WP_018692378.1</b> | <i>Algicola sagamiensis</i>               | Gammaproteobacteria   | Alteromonadales       | <i>Pseudoalteromonadaceae</i>     |
| <b>WP_054581066.1</b> | <i>Acinetobacter equi</i>                 | Gammaproteobacteria   | Pseudomonadales       | <i>Moraxellaceae</i>              |
| <b>WP_016657436.1</b> | <i>Acinetobacter rufus</i>                | Gammaproteobacteria   | Pseudomonadales       | <i>Moraxellaceae</i>              |
| <b>WP_004924459.1</b> | <i>Acinetobacter baylyi</i>               | Gammaproteobacteria   | Pseudomonadales       | <i>Moraxellaceae</i>              |
| <b>WP_004854861.1</b> | <i>Acinetobacter gernerri</i>             | Gammaproteobacteria   | Pseudomonadales       | <i>Moraxellaceae</i>              |

|                       |                                      |                     |                 |                      |
|-----------------------|--------------------------------------|---------------------|-----------------|----------------------|
| <b>WP_134244962.1</b> | <i>Alkanindiges illinoiensis</i>     | Gammaproteobacteria | Pseudomonadales | <i>Moraxellaceae</i> |
| <b>WP_066804633.1</b> | <i>Moraxella oblonga</i>             | Gammaproteobacteria | Pseudomonadales | <i>Moraxellaceae</i> |
| <b>STY87967.1</b>     | <i>Moraxella ovis</i>                | Gammaproteobacteria | Pseudomonadales | <i>Moraxellaceae</i> |
| <b>WP_078307184.1</b> | <i>Moraxella lincolnii</i>           | Gammaproteobacteria | Pseudomonadales | <i>Moraxellaceae</i> |
| <b>WP_028858284.1</b> | <i>Psychrobacter phenylpyruvicus</i> | Gammaproteobacteria | Pseudomonadales | <i>Moraxellaceae</i> |
| <b>WP_011279654.1</b> | <i>Psychrobacter arcticus</i>        | Gammaproteobacteria | Pseudomonadales | <i>Moraxellaceae</i> |
| <b>WP_093067772.1</b> | <i>Psychrobacter pacificensis</i>    | Gammaproteobacteria | Pseudomonadales | <i>Moraxellaceae</i> |