KSHV transactivator-derived small peptide traps coactivators to attenuate MYC and inhibits leukemia and lymphoma cell growth

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S-Figure 1. Protein dissociated from RNAPII during reactivation. Average of peptides counts that are 1.5x less than non-reactivated samples were plotted with STRING.

Other herpesvirus & Bacteria	Alignment/protein seq.			
KSHV ORF50 (ADQ57929.1):	DDAL L S <mark>SILQ</mark> GLYQLDT			
LysR Bacteria TF (WP193632878.1):	KAQ-LPDILQGLYQLTQ			
Saimiriine GHV2 (CAC85015.1):	DDNI L AS ILQ D LY DLPA			
Macaca Nemerha Virus (YP010084594.1)	:DDDMLAAILQDLYGLQS			
Bovine GHV4 (NP076541.1):	EDAY L EL ILQGLY H L DE			
Retrofibro.GHV (YP010084411.1):	DDEL L ST ILQ GLYQLDE			
Colobine GHV1 (QDQ69256.1):	DEDL L SA ILQ GLYQLDE			
Rhesus radinovirus (AAF60029.1).	DDDM L AA ILO D LY G L OS			

b **BC3 cell** T->E T->S 100 S->T Viability (%) VGN50-Mut VGN50-d(1) VGN50-d(3) 50 0 20 0 40 60 80 μΜ

S-Figure 2. (a) Consensus sequences observed at ORF50 transactivation domain with other gamma-herpesviral homologous proteins and a bacterial transcriptional factor. Essential amino acid residue for MYC cell growth inhibition identified in this study is underlined. Conserved amino acid residues are also marked in bold case. (b) Exchangeable amino acid indicates conserved function with other gamma-herpesvirus family homologous protein. Amino acids residues that are exchangeable to corresponding amino acid in other herpesvirus without any loss of cell killing in MTT assays were indicated in red (T->E, T->S, S->T). Additionally changing two C-terminal amino acids to D-amino acid (d3) did not increase efficacies of peptide cell killing in vitro.







S-Figure 3. Flow cytometry analyses

a

(a) Apoptosis (%) was determined as AnnexinV+ 7AAD- population gated in red shown in representative FACS plots of BCBL-1 cells treated with PBS or VGN50 at 32 μ M. (b) Ki67+ (%) gating strategy. Representative FACS plots with ancestry gating strategy of Ki67+ population for BCBL-1 and BC-1 cells treated with PBS with an isotype control are shown. Mean +/-SD of Ki67% of total BCBL-1 and BC-1 cells analyzed are shown in Fig. 2e.



confirmed/excluded duplicated transcription ID manually and included for the analyses.

transcription ID had different ID name, and found

to be duplicated in the list. We



64 transcription regulators in VGN50
down-regulated gene promoters

S-Figure 4. List of putative target transcription regulators of VGN50. (a) Schematic analysis flow for identification of VGN50 targets. Transcription targets were determined by differential transcriptional output between VGN50Mut and VGN50Wt treated cells. After the transcription targets were filtered with indicated cut-off criteria, they were submitted to CSCAN to identify common regulators. The commonly targeted transcription regulators of 2 cell lines were shown as Venn diagram. **(b) Venn diagram and putative VGN50-target transcription regulators.** Number of protein identified is indicated in Venn diagram (left) and list of protein names that are commonly identified between two cell lines were depicted in table (right). Three protein that are known to play important role in PEL cell growth (c-Myc and IRF4) and a protein identified in the targeted coactivator complex (p300 in Fig. 1b) are indicated with bold face.



b



	PBS		VGN50			Mutant	
			10	0 MG/KG	1	0 MG/KG	
	AVERAGE	STDEV	AVERAGE	STDEV	AVERAGE	STDEV	
Alanine Transaminase U/L	118.43	101.94	66.45	5.39	82.85	38.16	0-403
Aspartate Transaminase U/L	554.43	183.82	500.33	54.75	651.13	336.08	0-552
Albumin g/dL	3.89	0.11	3.74	0.07	3.67	0.20	2.9-4.0
Alkaline PhosphataseU/L	96.53	7.12	111.00	18.89	129.20	29.09	49-172
Amylase U/L	1789.88	217.38	1669.23	179.42	1624.05	13.13	2463-6660
Blood Urea Nitrogen mg/dL	19.78	1.38	16.43	1.25	20.03	2.53	15.2-34.7
Calcium mg/dL	10.24	0.20	9.00	1.63	9.39	0.60	9.6-11-5
Chloride mmol/L	112.23	1.27	112.88	0.65	112.13	2.05	105-118
Creatinine mg/dL	0.20	0.04	0.25	0.02	0.20	0.06	0.0-0.3
Glucose mg/dL	176.45	6.58	183.45	33.13	194.73	35.16	130-254
Potassium mmol/L	6.59	1.22	5.84	0.75	6.90	0.68	6.9-10.0
Sodium mmol/L	147.50	1.29	147.75	2.50	145.25	1.26	150-160
Phosphorus mg/dL	7.22	1.09	5.83	1.10	7.46	0.95	7.5-10.7
Total Bilirubin mg/dL	0.05	0.02	0.09	0.01	0.05	0.01	0.0-0.2
Total Protein g/dL	5.03	0.21	4.74	0.09	4.81	0.18	4.7-6.1
	AVERAGE	31DEV 2.10	AVERAGE	31DEV	AVERAGE	31DEV	5 1 1/ 7
Absolute Neutrophilcells (K/ul)	1.53	2.10	1.81	2.85	4.22	2.04	J.1-14.7 1 0_3 7
Absolute Lymphocyte cells (K/ul)	3 72	1 47	3 35	1 64	2 54	1 16	4 3-10 6
Absolute Monocyte cells (K/ul)	0.56	0.37	0.49	0.29	0.43	0.12	0.2-1.0
Absolute Eosinophil cells (K/ul)	0.19	0.05	0.16	0.16	0.14	0.06	0.0-0.1
Absolute Basophil cells (K/ul)	0.08	0.05	0.05	0.04	0.06	0.02	0.0-0.1
Neutrophil %	26.04	4.53	32.01	4.24	16.70	14.43	12.5-31.2
Lymphocyte %	60.34	3.27	56.72	4.52	31.21	31.58	62.9-82.7
Monocyte %	8.95	3.20	8.34	1.60	5.52	3.67	2.5-7.5
Eosinophil %	3.36	1.39	2.23	1.57	2.14	0.89	0.0-0.8
Basophil %	1.31	0.34	0.71	0.43	0.70	0.44	0.0-0.5
RBC (M/ul)	8.48	1.41	8.77	0.74	4.85	4.37	8.12-10.55
Hemoglobin (g/dL)	11.85	2.19	13.03	1.23	7.07	6.22	11.7-16.2
Hematocrit %	48.60	8.70	49.38	4.19	27.72	24.63	38.3-54.0
MCV (fL)	57.25	2.04	56.33	0.53	29.04	32.05	45.3-53.6
MCH (pg)	13.98	1.39	14.88	0.25	7.62	7.88	13.3-16.7
MCHC (g/dL)	24.40	1.64	26.38	0.29	13.18	14.14	27.6-32.9
RDW %	19.85	0.66	19.15	0.68	10.08	10.88	12.4-27.0
Platelets (K/uL)	353.75	155.55	524.00	299.84	333.29	152.20	574-1079
MPV (fL)	5.93	0.25	5.43	0.62	3.05	3.04	5.0-20.0

Extended Data Figure 5. BALB/c mice (8 weeks old, female, n=5/group) were injected with 10mg/kg of VGN50 or mutant peptide in 200 uL i.p. with a 5 day-on 2 day off schedule for two weeks. Body weight changes were measured daily (a). At the end of the experiment, fresh blood and serum collected were subjected for complete blood counts and serum biochemical properties, respectively (b).



S-Figure 6. Cell atrophy by VGN50. A PEL gate was set based on FSC-A and SSC-A and applied for each ascites sample with a different treatment. A representative overlay FACS plot and histograms for FSC-A and SSC-A are shown. Mean geometric fluorescent intensity (gMFI) +/- SD for FSC-A and SSC-A are shown in Fig. 6C.



S-Figure 7. Ascites cytokines were measured by Olink assay using inflammatory cytokine panel as shown in STable. Among 92 cytokines, 15 cytokines significantly changed between the groups are shown. * <0.05, ** <0.01, *** <0.001.

	GS	NES	NOM p-val	FDR q-val
1	HALLMARK_TNFA_SIGNALING_VIA_NFKB	1.74	0.001	0.034
2	HALLMARK_ESTROGEN_RESPONSE_EARLY	1.69	0.001	0.036
3	HALLMARK_G2M_CHECKPOINT	1.58	0.000	0.068
4	HALLMARK_MITOTIC_SPINDLE	1.57	0.002	0.060
5	HALLMARK_ALLOGRAFT_REJECTION	1.55	0.016	0.061
6	HALLMARK_APOPTOSIS	1.54	0.015	0.055
7	HALLMARK_E2F_TARGETS	1.51	0.001	0.066
8	HALLMARK_INTERFERON_GAMMA_RESPONSE	1.48	0.021	0.071
9	HALLMARK_ESTROGEN_RESPONSE_LATE	1.47	0.020	0.068
10	HALLMARK_IL2_STAT5_SIGNALING	1.39	0.065	0.128
11	HALLMARK_MTORC1_SIGNALING	1.37	0.048	0.137
12	HALLMARK_MYC_TARGETS_V1	1.36	0.053	0.138

S-Figure 8. Total RNA-seq of long-term treated xenograft BCBL-1 cells. Xenograft BCBL-1 cells. Xenograft BCBL-1 cells were isolated from ascites fluid and total RNA was extracted and sequenced. Results of GSEA over mutant peptide treated samples is presented in Table (N=3/group).

Fig. 1c

Fig. 1g

