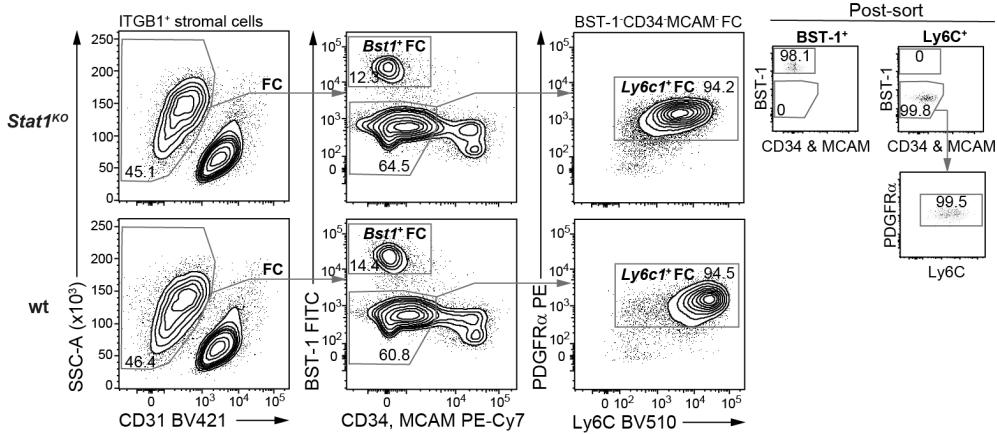
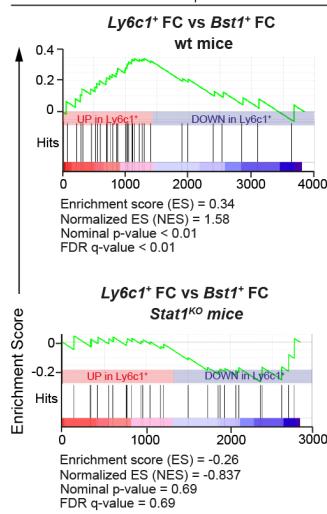
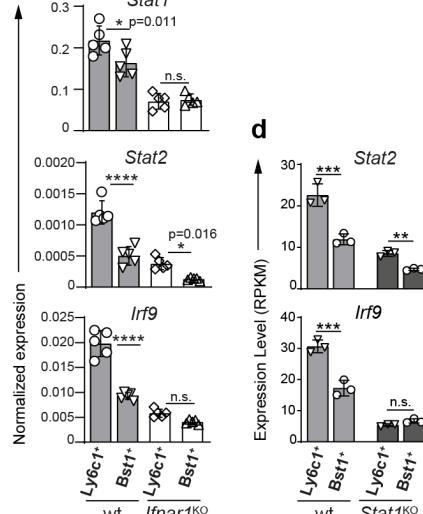
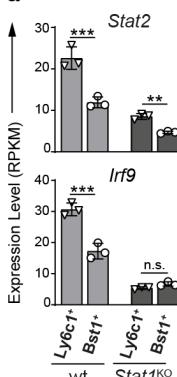


a**b**
ISG set collated for IFN- β -stimulated fibroblasts
**c****d**
Supplementary Figure 2. Related to Figures 5-6.

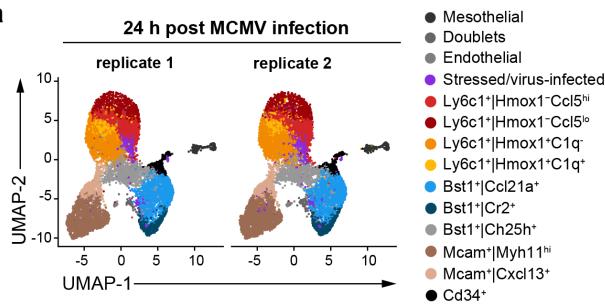
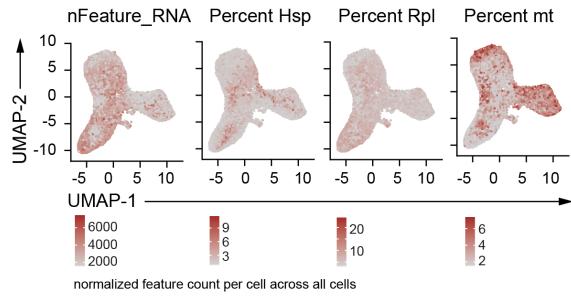
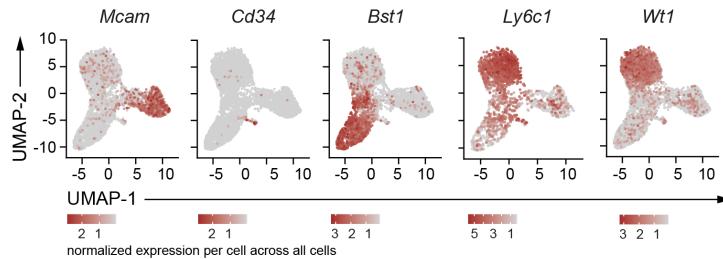
a FACS gating on *Ly6c1*⁺ FC and *Bst1*⁺ FC in *Stat1*^{KO} mice. Numbers are percentage of cells in the indicated gates.

b Gene set enrichment analysis for IFN- β -stimulated gene set collated for primary fibroblasts (extracted from the Interferome database v2.0 using the following search criteria: fold change > 2.5; p-val < 0.05; max. 6 hours post stimulation (<http://www.interferome.org>)) performed on DEGs ($\log_2(\text{fold change}) > 0.8$, adj. p-val < 0.05) from the comparison between *Ly6c1*⁺ FC versus *Bst1*⁺ FC in wt or *Stat1*^{KO} mice.

c Expression of indicated IFNAR signalling mediators, based on RT-qPCR analysis of *Ly6c1*⁺ FC and *Bst1*⁺ FC isolated from the spleens of wt or *Ifnar1*^{KO} mice. Bars denote mean \pm SD of n = 5 mice (depicted as symbols). Statistical significance was calculated using one-way ANOVA with Sidak's multiple comparison test.

d Expression of indicated IFNAR signalling mediators, based on RNA-seq analysis of *Ly6c1*⁺ FC and *Bst1*⁺ FC isolated from the spleens of wt mice or *Stat1*^{KO} mice. Bars denote mean RPKM \pm SD of n = 3 biological replicates (depicted as symbols) using cells sorted from 2-4 mice per replicate. Shown are adjusted p-values from the comparison between *Ly6c1*⁺ FC versus *Bst1*⁺ FC transcriptomes in the wt or the *Stat1*^{KO} condition.

c, d * p<0.05; ** p<0.01; *** p<0.001; **** p<0.0001; n.s. denotes p>0.05

a**b****c**

Supplementary Figure 3. Related to Figure 7.

a-c scRNA-seq analysis of splenic FC 24 hours post infection with 10^6 PFU of MCMV *i.p.* **a** UMAP embedding before removal of doublets, endothelial-, mesothelial- and virus-infected cells. **b** Number of detected genes (nFeature), percentage of reads for heat-shock proteins (Hsp), percentage of reads mapping to large ribosomal subunit proteins (Rpl) or percentage of mitochondrial reads (mt) per cell superimposed on UMAP embedding. **c** Gene expression on UMAP embedding in the infected condition.

	Wt mice		Stat1 ^{KO} mice	
ISGs	log ₂ (fold change) Ly6c1 ⁺ FC vs Bst1 ⁺ FC	adj. p-val	log ₂ (fold change) Ly6c1 ⁺ FC vs Bst1 ⁺ FC	adj. p-val
<i>Rtp4</i>	3.776	6.93E-43	6.285	2.17E-27
<i>Ifi13</i>	3.235	4.69E-28	2.129	4.30E-49
<i>Ifi11</i>	3.119	4.18E-32	1.686	0.001317706
<i>Plac8</i>	2.731	3.25E-22	1.817	4.24E-23
<i>Cmpk2</i>	2.420	8.27E-74	2.381	1.97E-66
<i>Oas1a</i>	2.239	3.14E-22	0.000	1
<i>Isg15</i>	2.165	9.57E-19	-0.329	0.286874755
<i>Gbp3</i>	2.109	2.41E-30	1.724	3.73E-16
<i>Xaf1</i>	2.073	1.96E-20	0.962	1.02E-10
<i>Irif7</i>	1.930	4.69E-16	1.698	1.72E-29
<i>Spats2l</i>	1.892	1.08E-29	1.932	4.79E-11
<i>Rsd2</i>	1.798	1.39E-12	3.469	7.3905E-05
<i>Lgals9</i>	1.790	3.29E-24	0.737	8.28E-18
<i>Tmem140</i>	1.759	5.77E-12	1.542	0.000102696
<i>Zbp1</i>	1.755	4.42E-12	-0.750	0.008939974
<i>Ifitm3</i>	1.577	2.56E-09	0.847	7.93E-21
<i>Gbp4</i>	1.550	7.92E-24	0.396	0.581391389
<i>Rnf213</i>	1.435	1.20E-11	0.456	0.152926136
<i>Ifi44</i>	1.421	4.35E-11	-5.897	0.005285034
<i>Ly6e</i>	1.416	8.48E-14	0.432	4.55343E-11
<i>Ddx60</i>	1.404	1.93E-12	0.356	0.35772351
<i>Ifitm2</i>	1.392	5.10E-08	0.682	5.61E-12
<i>Agrn</i>	1.350	0.003024882	0.806	0.002022399
<i>Parp14</i>	1.325	6.00E-08	1.268	4.76171E-05
<i>Zcchc24</i>	1.295	3.39E-06	1.397	2.60E-38
<i>Mx2</i>	1.293	0.000751404	1.719	0.008415425
<i>Ifit2</i>	1.263	2.20E-08	0.625	0.0581921
<i>Phf11b</i>	1.243	0.000476852	0.669	0.249007739
<i>Trim25</i>	1.213	1.85E-06	0.929	0.001642548
<i>Ifitm1</i>	1.195	0.000276969	0.037	0.821690635
<i>Mikl</i>	1.173	1.03E-11	0.674	0.051807094
<i>Parp12</i>	1.169	1.93E-28	0.254	0.699968757
<i>Rnf114</i>	1.146	2.32E-18	0.504	3.58415E-05
<i>Usp18</i>	1.137	0.00015923	-0.765	0.025493647
<i>Sifn5</i>	1.077	5.70E-05	1.322	2.35E-14
<i>Phf11d</i>	1.075	1.84E-09	-0.278	0.198582662
<i>Ifi35</i>	1.025	2.25E-05	0.279	0.075168912
<i>Tnfsf10</i>	0.960	0.04825153	0.398	0.679097299
<i>Stat2</i>	0.923	2.60E-09	0.861	0.001569826
<i>Trim21</i>	0.899	2.13E-07	2.980	0.005067653
<i>Ifi1h1</i>	0.869	1.20E-11	0.222	0.322400559
<i>Eif2ak2</i>	0.844	0.000660729	0.107	0.819661336
<i>Samd9l</i>	0.841	8.66E-10	-0.166	0.601906348
<i>Dhx58</i>	0.832	0.000126611	0.035	0.972571033
<i>Irif9</i>	0.829	2.10E-06	-0.312	0.212880211
<i>Satt1</i>	0.822	1.16E-11	1.558	8.33E-31
<i>Parp10</i>	0.753	5.15E-06	0.930	0.025892372
<i>Cnp</i>	0.703	6.38E-06	0.944	1.31E-08
<i>Parp9</i>	0.683	1.17E-07	-0.107	0.793685179
<i>Atp10a</i>	0.676	0.245699711	0.964	0.564600977
<i>Dtx3l</i>	0.675	0.049529815	0.069	0.936476322
<i>Scarb2</i>	0.674	4.35E-06	0.574	1.99E-14
<i>Fbxo6</i>	0.655	0.005837012	0.433	0.120868253
<i>Gbp2</i>	0.566	7.18E-06	0.500	0.004266245
<i>Zc3HAV1</i>	0.527	3.96E-08	0.153	0.147412801
<i>Psme2</i>	0.514	0.05725131	-0.056	0.866395447
<i>Chmp5</i>	0.504	0.000257715	0.269	0.103556815
<i>Adar</i>	0.464	0.244573091	0.036	0.981364291
<i>Acot9</i>	0.458	0.015137002	0.167	0.630351861
<i>Tmem106a</i>	0.446	0.042776103	0.196	0.207427517
<i>Psme1</i>	0.396	0.009505137	-0.027	0.920311469
<i>Gbp6</i>	0.391	0.048587847	-0.691	0.088445103
<i>Gbp5</i>	0.336	0.230637709	0.387	0.378482139
<i>Uncb301</i>	0.333	0.006382631	-0.196	0.65845475
<i>Myd88</i>	0.321	0.062127335	0.220	0.345129693
<i>Ube2l6</i>	0.270	0.304295277	-0.442	0.000950503
<i>Gstdmd</i>	0.258	0.06256598	0.311	0.158642212
<i>Rab24</i>	0.203	0.467691547	-1.139	0.022104957
<i>Ankfy1</i>	0.182	0.78942167	0.113	0.785947397
<i>Sp140</i>	0.167	0.680294345	0.023	0.971973367
<i>Samhd1</i>	0.156	0.312458174	-0.111	0.536120397
<i>Bst2</i>	0.132	0.577588507	-1.517	3.81E-26
<i>B2m</i>	0.118	0.541853297	-0.975	3.90E-16
<i>Irif1</i>	0.118	0.579921157	1.399	2.70E-19
<i>Lap3</i>	0.079	0.69339683	0.054	0.877140009
<i>Tdrd7</i>	0.077	0.775181231	-0.187	0.726626325
<i>Sp110</i>	-0.024	0.932055876	-0.538	0.066958384
<i>Bag1</i>	-0.026	0.9167317	-0.674	0.135394211
<i>Ddx58</i>	-0.054	0.798735084	-0.590	2.79188E-05
<i>Znfx1</i>	-0.077	0.913316284	-0.222	0.720211365
<i>Lgals3bp</i>	-0.080	0.644637116	-1.911	4.27E-85
<i>Nod1</i>	-0.212	0.368998841	-0.159	0.676973253
<i>Psmb8</i>	-0.219	0.496817363	-0.995	2.00E-31
<i>N4bp1</i>	-0.288	0.245197809	-0.080	0.930491534
<i>Psmb9</i>	-0.387	0.10403244	-0.816	4.10E-10
<i>Nub1</i>	-0.463	2.54E-07	-0.491	0.082969968
<i>Traf1</i>	-0.729	3.18E-08	-1.200	5.08E-08
<i>Tap1</i>	-0.762	8.75E-10	-1.476	3.75E-28
<i>Ly96</i>	-0.924	3.05E-08	-1.578	2.29E-14
<i>Casp1</i>	-0.978	1.08E-08	-1.011	3.26617E-05
<i>Serpinb9</i>	-1.175	7.71E-24	-1.776	9.1971E-125
<i>Epst1</i>	-1.191	0.052868151	-0.602	0.719587324
<i>Casp4</i>	-1.266	1.82E-10	-1.676	1.27E-13
<i>Isg20</i>	-1.288	1.43E-06	-2.206	2.51E-31
<i>Stampb1</i>	-1.551	1.87E-20	-1.243	0.00599388
<i>Vrk2</i>	-1.625	6.18E-26	-1.417	3.44E-06
<i>Lysmd2</i>	-1.836	6.17E-15	-1.635	5.23E-40
<i>Map3k8</i>	-2.089	1.19E-32	-0.859	0.000234561
<i>Abtb2</i>	-2.249	2.74E-13	-1.497	0.26401582
<i>Lcp2</i>	-3.622	3.32E-36	-2.891	1.26925E-09

Supplementary Table 1. Related to Fig. 6

Shown are fold changes in the expression of individual ISGs between *Ly6c1⁺* FC versus *Bst1⁺* FC in wt or *Stat1*^{KO} mice. ISGs that are no longer overexpressed in *Ly6c1⁺* FC upon loss of *Stat1* are highlighted in grey.

Antibody	Source	Cat. No.
APC/Cy7 anti-mouse ITGB1 (HMβ1-1)	BioLegend	Cat# 102226
FITC anti-mouse ITGB1 (HMβ1-1)	BioLegend	Cat# 102205
Biotin anti-mouse ITGB1 (HMβ1-1)	BioLegend	Cat# 102203
Alexa Fluor 700 anti-mouse CD45.2 (104)	BioLegend	Cat# 109822
Brilliant Violet 510 anti-mouse CD45.2 (104)	BioLegend	Cat# 109838
Biotin anti-mouse CD34 (RAM34)	Thermo Fisher	Cat# 13-0341-82
eFluor 660 anti-mouse CD34 (RAM34)	Thermo Fisher	Cat# 50-0341-82
PE/Cy7 anti-mouse MCAM (ME-9F1)	BioLegend	Cat# 134714
Alexa Fluor 647 anti-mouse MCAM (ME-9F1)	BioLegend	Cat# 134718
Purified anti-mouse BST-1 (BP-3)	BioLegend	Cat# 140202
PE anti-mouse BST-1 (BP-3)	BioLegend	Cat# 140204
APC anti-mouse BST-1 (BP-3)	BioLegend	Cat# 140207
FITC anti-mouse BST-1 (KT157)	Thermo Fisher	Cat# MA5-17948
PE anti-mouse PDGFR α (APA5)	BioLegend	Cat# 135906
PE/Cy7 anti-mouse CR2/CR1 (7E9)	BioLegend	Cat# 123419
PE/Cy7 anti-mouse Ly6C (HK1.4)	BioLegend	Cat# 128017
Brilliant Violet 510 anti-mouse Ly6C (HK1.4)	BioLegend	Cat# 128033
Brilliant Violet 785 anti-mouse Ly6C (HK1.4)	BioLegend	Cat# 128041
Biotin anti-mouse Ly-6C (HK1.4)	BioLegend	Cat# 128003
BV421 anti-mouse/human CD45R/B220 (RA3-6B2)	BioLegend	Cat# 103251
APC anti-mouse CD71 (RI7217)	BioLegend	Cat# 113819
BV421 anti-mouse CD31 (MEC13.3)	BD Horizon	Cat# 562939
Biotin anti-mouse Pan-endothelial Cell Antigen (MECA-32)	BioLegend	Cat# 120504
Alexa Fluor 488 anti-mouse Pan-endothelial Cell Antigen Antibody (MECA-32)	BioLegend	Cat# 120506
PE Rat IgG2a Isotype Ctrl (RTK2758)	BioLegend	Cat# 400507
PE/Cy7 Rat IgG2a, κ Isotype Ctrl (RTK2758)	BioLegend	Cat# 400521
APC Rat IgG2a, κ Isotype Ctrl (RTK2758)	BioLegend	Cat# 400511
PE/Cy7 Rat IgG2c, κ Isotype Ctrl (RTK4174)	BioLegend	Cat# 400721
Purified Mouse IgG1, κ Isotype Ctrl Antibody (MOPC-21)	BioLegend	Cat# 400101
Polyclonal anti-mouse CCL21	LifeSpan BioSciences	Cat# LS-C104634-50
FITC anti-alpha smooth muscle actin (1A4)	Abcam	Cat# ab8211
Purified HMOX1 Monoclonal Antibody (HO-1-1)	ThermoFisher	Cat# MA1-112
APC CXCL13 Monoclonal Antibody (DS8CX13)	ThermoFisher	Cat# 17-7981-82
Desmin Polyclonal Antibody	ThermoFisher	Cat# PA5-16705
anti-mouse WT1 (SC06-41)	LIFE Technologies	Cat# MA532215
Rabbit IgG Isotype Control	Thermo Fisher	Cat# 10500C
Goat anti-Rabbit IgG (H+L) Secondary, Alexa Fluor 647	Thermo Fisher	Cat# A-21245
Brilliant Violet 785 Streptavidin	BioLegend	Cat# 405249
PE/Cyanine7 Streptavidin	BioLegend	Cat# 405206

Supplementary Table 2. Related to Methods

Antibodies used in the study.

	Forward (5' - 3')	Reverse (5' - 3')
<i>Gapdh</i>	TGTGTCCGTCGTGGATCTGA	CCTGCTTCACCACCTTCTTGAT
<i>Oas1a</i>	GTCCTGGCTCATGTTAATACTCCA	GCTCCGTGAAGCAGGTAGAG
<i>Ifit1</i>	GAGCAGAGAGTCAGGCAGG	CACCATCAGCATTCTCTCCCCA
<i>Ifi44</i>	TGGCATTCTGCATTGGCTT	TCCAGCTTGGACTTCACAGG
<i>Ifih1</i>	TGCTTATCGCTACGACGGTG	TTCATCAGCTGGCTCGAC
<i>Eif2ak2</i>	TCGTGACCGGAGTGGAGTAT	GTTGCAAGGCCAAAGTCTCC
<i>Irf7</i>	GGGACCTCTGCTTCAGGTT	AGGGTTCCCTCGTAAACACGG
<i>Usp18</i>	CGGACAGACGTGTTGCCCTA	TCCGAGGCACTGTTATCCTCT
<i>Stat1</i>	AGAGGTGTTGAGTTGGCAGT	ACCATCAGGCCAGCATTAG
<i>Stat2</i>	CTATTGCTGCCCAAGCTGGA	AACTTGCTCCAGCCGTCAA
<i>Irf9</i>	AGCAACTGCAACTCTGAGCTA	GTGAGCAGCAGCGAGTAGT
<i>Wt1</i>	CATCCGCAACCAAGGATACAG	TGAAGGAATGGTTGGGAAC

Supplementary Table 3. Related to Methods

Primer sequences.