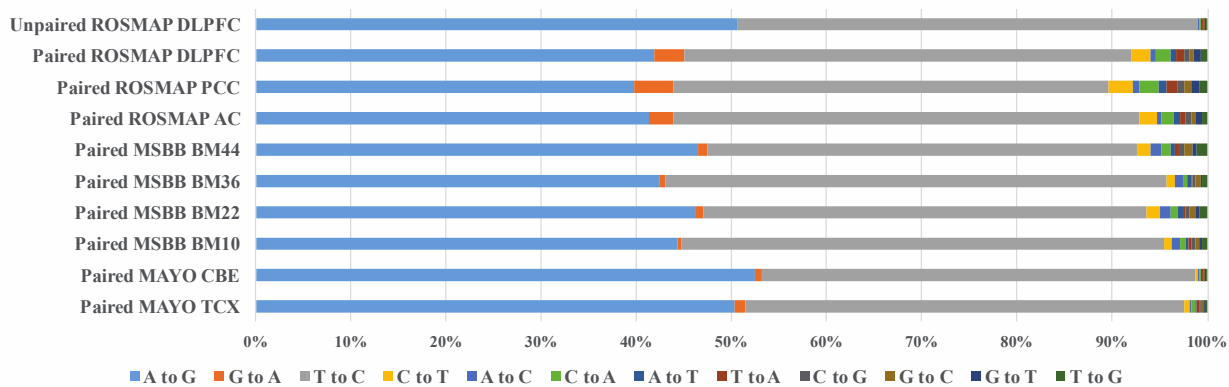


SUPPLEMENTARY MATERIALS

CONTENTS OF SUPPLEMENTARY MATERIAL:

- 1
- 2
- 3 Supplementary Data (excel sheet): Association results of those non-A-to-I editing events.
- 4 Fig. S1. Distributions of different types of RNA editing events across datasets.
- 5 Fig. S2. Tissue specificity of the known RNA editing events.
- 6 Fig. S3. Volcano plots of comparisons between different regions within each study.
- 7 Fig. S4. Differential expressions of *ADARs* across Alzheimer's disease (AD) status in ROSMAP
- 8 unpaired 635 DLPFC samples.
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- 10 Fig. S6. Distribution of frequent RNA recoding events across all datasets and their associations
- 11 with AD status compared to the non-coding events.
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- 13 events on the expressions of genes, isoforms and proteins.
- 14 Fig. S8. Mass-spectrometry figure of edited peptide (frequency $\geq 50\%$) hits in ROSMAP TMT
- 15 proteomic dataset.
- 16 Table S1. Characteristic of the cohorts and picard metrics of RNA-seq datasets.
- 17 Table S2. The recoding RNA editing events with proteomic peptide hits.
- 18 Table S3. Regional differences in gene expressions of *ADARs*.
- 19 Table S4. Top RNA editing events associated with AD status.
- 20 Table S5. Association of *ORAI2* editing event (chr7:102096952, hg19) on multiple traits in
- 21 ROSMAP unpaired DLPFC dataset (N=635).
- 22 Table S6. Comparisons between AD genes and non-AD genes.
- 23

24 Fig. S1. Distributions of different types of RNA editing events across datasets. Each dataset is
 25 presented as one stacked bar and different colors represent different types of editing events. On
 26 average, the majority of RNA editing events are the canonical A-to-I editing types, which are
 27 shown as the A-to-G and T-to-C editing types ($\geq 90\%$) and the C-to-T and G-to-A types (5%).

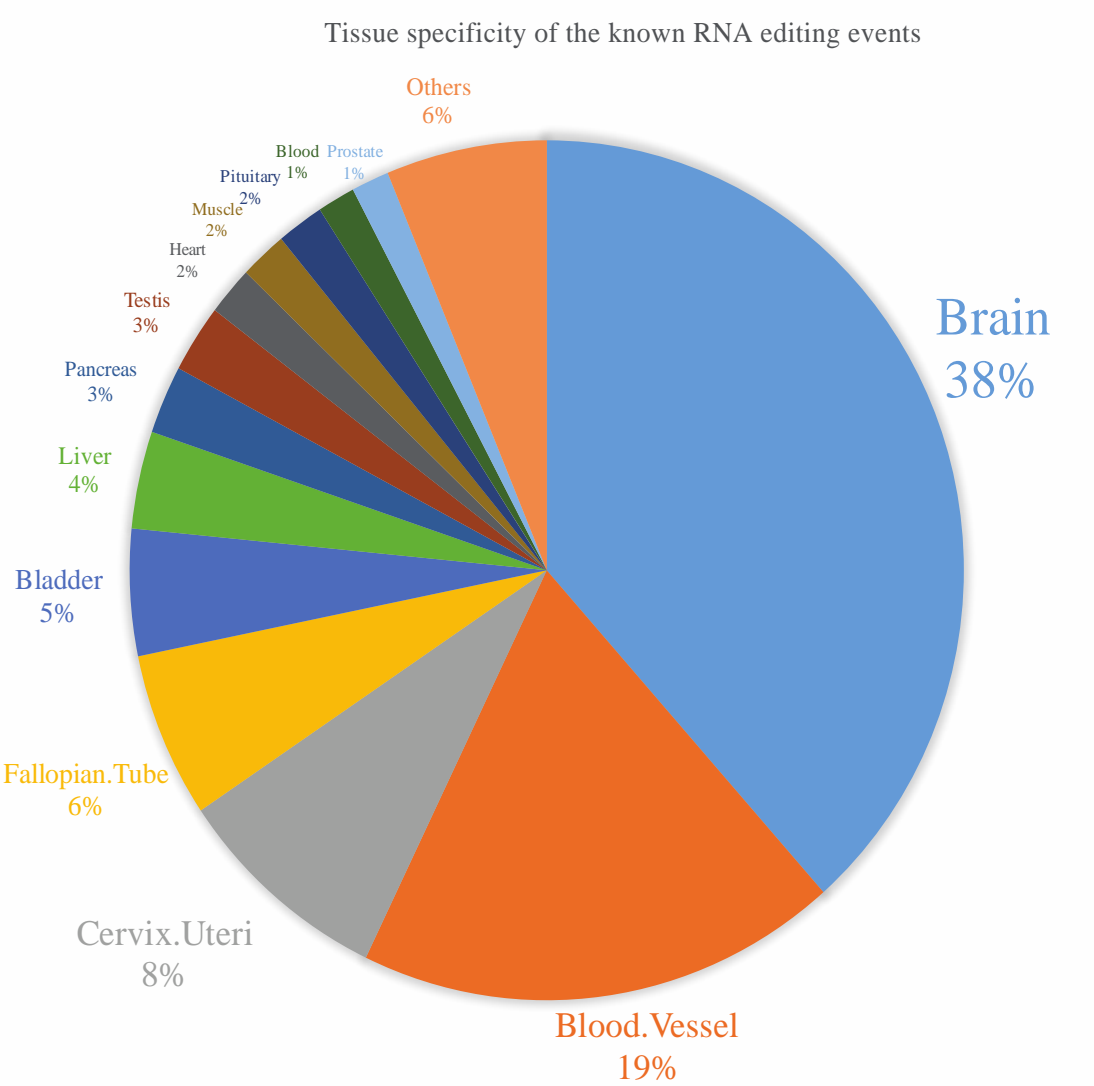


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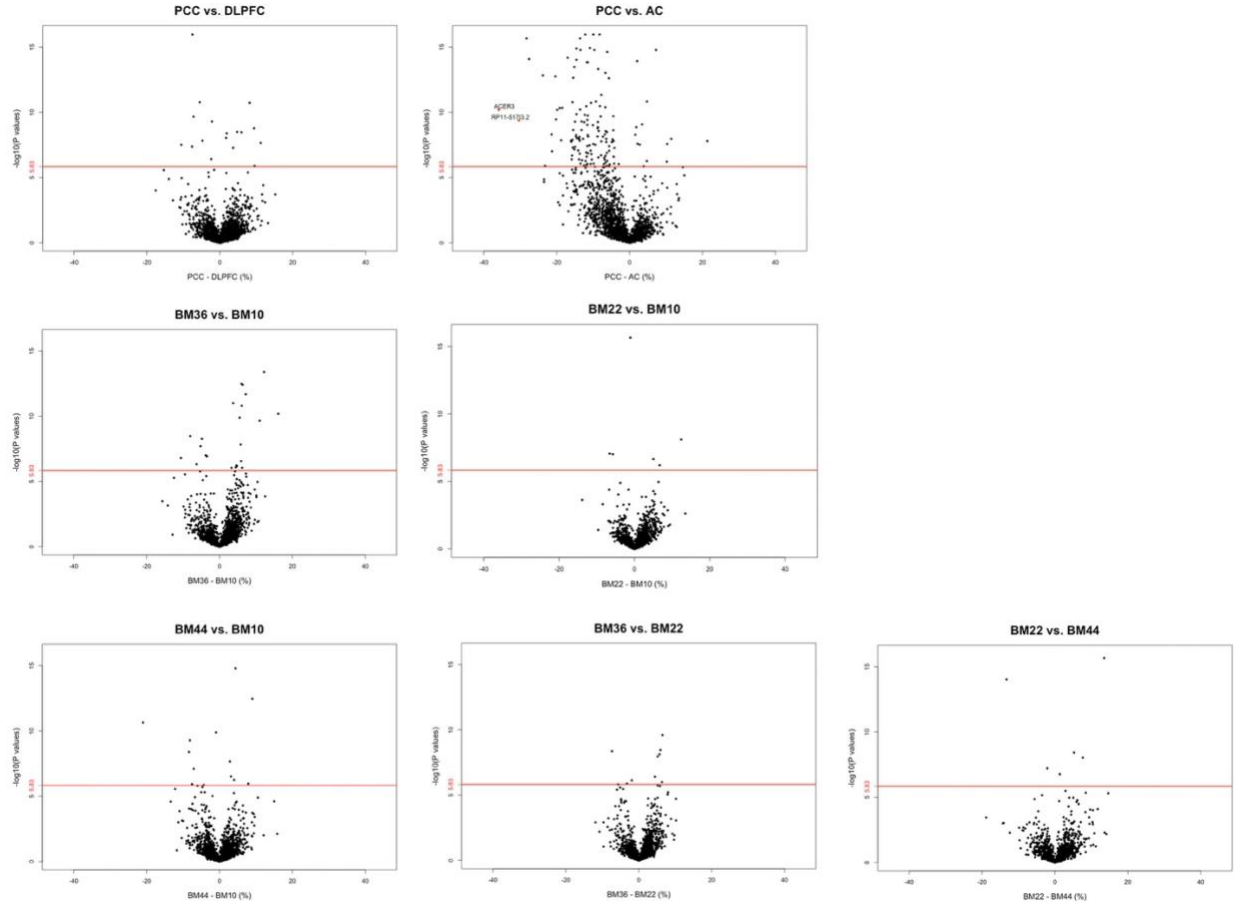
31 Fig. S2. Tissue specificity of the known RNA editing events. Pie chart showed the percentage of
32 the known RNA editing events across different tissue types.



33

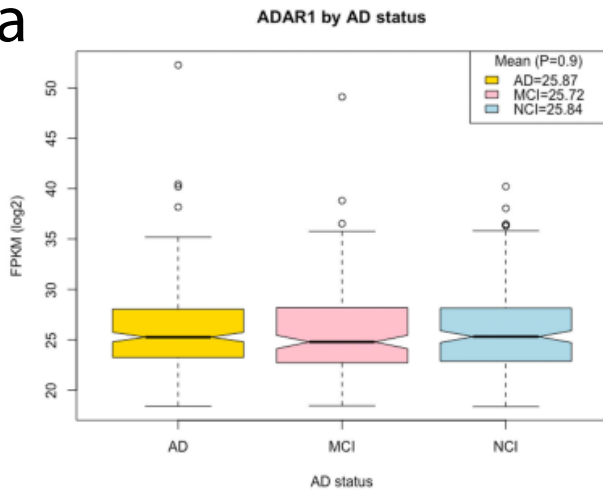
34

35 Fig. S3. Volcano plots of comparisons between different regions within each study. Comparisons
36 of the level (% alternative reads) of each RNA editing event between 2 brain regions within each
37 study were presented in a volcano plot. Each dot is one RNA editing event present in both brain
38 regions. X and Y axes display the regression coefficient and its corresponding $-\log_{10}$
39 transformed P value of the exposure variable of 2 brain regions in the general linear mixed
40 model which used subject as random effect, RNA editing level of each RNA editing event as
41 dependent variable and its covariates include age at death, sex, postmortem interval (PMI), and
42 RIN score using R lme package. The red horizontal line showed the Bonferroni-corrected P
43 value threshold of 1.49×10^{-6} ($0.05/33,641$). The red dots with annotated gene names are those
44 RNA editing event with $P \leq 1.49 \times 10^{-6}$ and regression coefficient of the differences between 2
45 brain regions $\geq 30\%$. X and Y axes display the regression coefficient and its corresponding -
46 \log_{10} transformed P value from the mixed linear regression model with the subject as random
47 effect and fixed covariates of age at death, sex, postmortem interval and RIN score. The
48 displayed P values were not adjusted by the multiple testings and were derived by two-sided
49 tests.

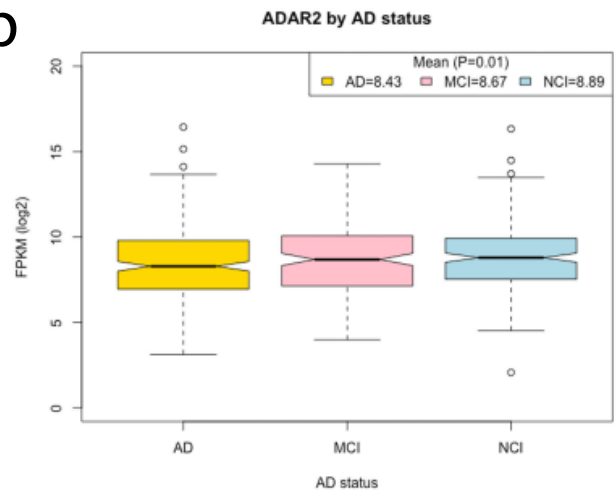


51 Fig. S4. Differential expressions of ADARs across Alzheimer's disease (AD) status in ROSMAP
52 unpaired 635 DLPFC samples. All the panels in this figure have the same sample size (N=635
53 subjects). We showed the level of RNA expressions of (a) *ADAR1*, (b) *ADAR2*, (c) *ADAR3* and
54 (d) the calculated value combining all three types of *ADARs* (*ADAR1+ADAR2-ADAR3*), based
55 on their previously reported potential functions on RNA editings (*ADAR1* and *ADAR2* increase
56 while *ADAR3* inhibits RNA editings), in subjects who cognitively non-impaired (NCI, blue),
57 mildly impaired (MCI, pink), or fulfill criteria for AD dementia (AD, yellow). The RNA
58 expression of *ADARs* (log₂ transformed FPKM) are displayed by the notched boxplot, in which
59 the lines from the bottom to the top represents the minimum value or the value <25% percentile,
60 25% percentile, median, 75% percentile, the maximum value or the value >75% percentile. The
61 "Notch" around the median represent the 95% confidence interval of the median, and the area
62 with color represent the interquartile with 50% of values. The open circle represents the potential
63 outliers. The mean *ADARs* expressions are displayed in the legend box on the right top corner,
64 in which the P value for the general linear regression model with *ADARs* expression as
65 dependent variable and 3 AD status as exposure variable. The displayed *P* values were not
66 adjusted by the multiple testings and were derived by two-sided tests.

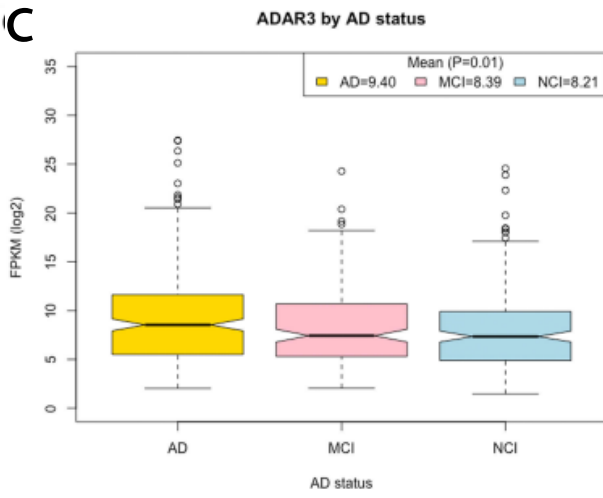
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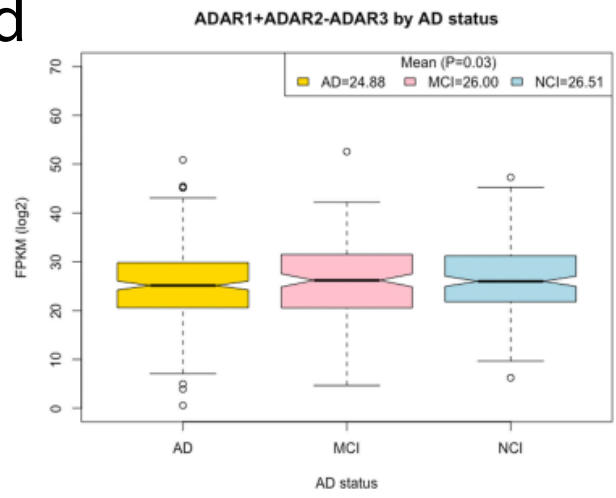
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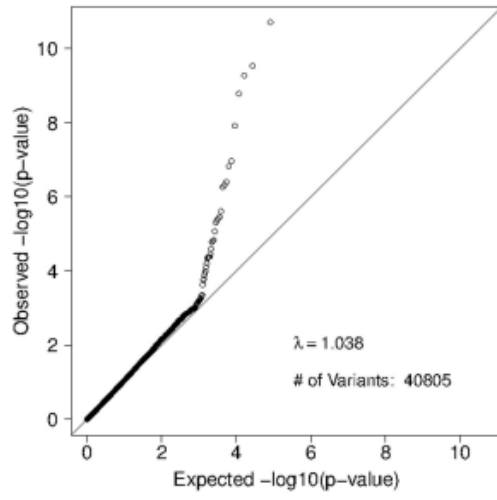


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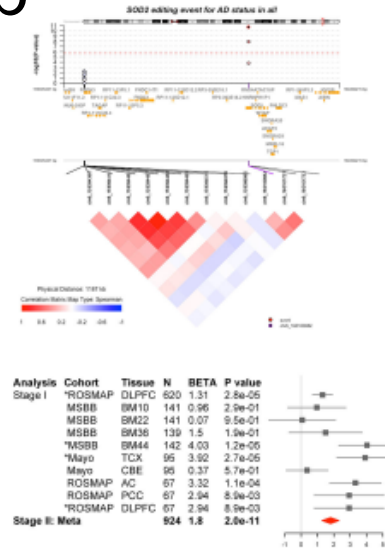
68

69 Fig. S5. QQ plot of stage II meta-analyzed results and regional and forest plots at top loci. (a)
70 QQ plot. (b-e) Regional (upper) and forest (lower) plots of top loci at *SOD2* (b), *MCURI* (c),
71 *HSDL2* (d) and *PFKP* (e). Regional plot shows the Stage I association results (upper). Each
72 circle represents a frequent RNA editing event with the top one in purple. The correlation (r)
73 between the top one and the others are shown with the color coding of: shallow to dark red for
74 $0 < r \leq 1$, white for $r=0$, and shallow to dark blue for $-1 \leq r < 0$. The correlation matrix between each
75 pair is shown in a triangle (lower) with the same color coding. Forest plot show results in each
76 dataset and Stage II meta-analyzed results. The estimated difference in the mean level of RNA
77 editing (% alternative reads) by clinical AD status (0 for normal controls, 1 for mild cognitive
78 impairment, and 2 for AD) and its 95% confidence interval were illustrated by the filled square
79 and horizontal line for each dataset or the filled red diamonds for the summaries. The displayed
80 P values were not adjusted by the multiple testings and were derived by two-sided tests.

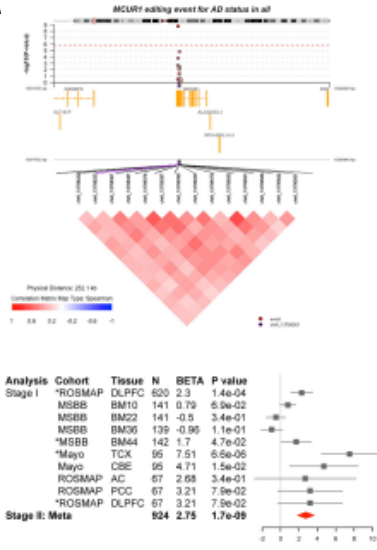
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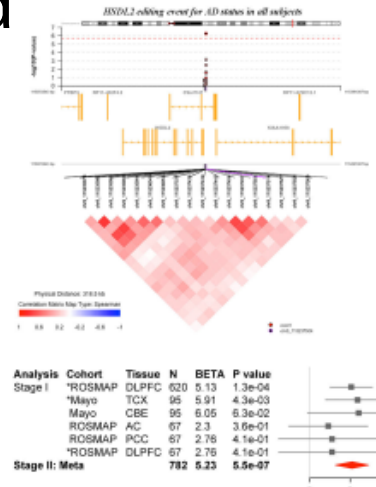
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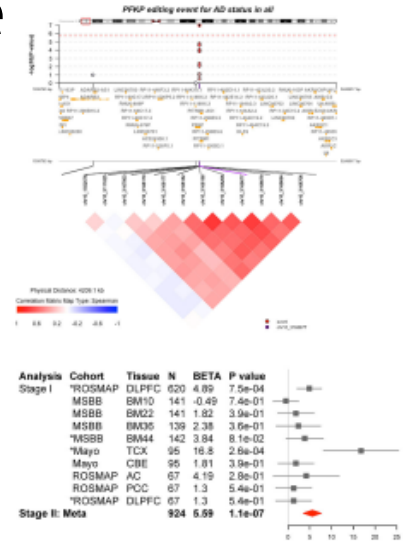
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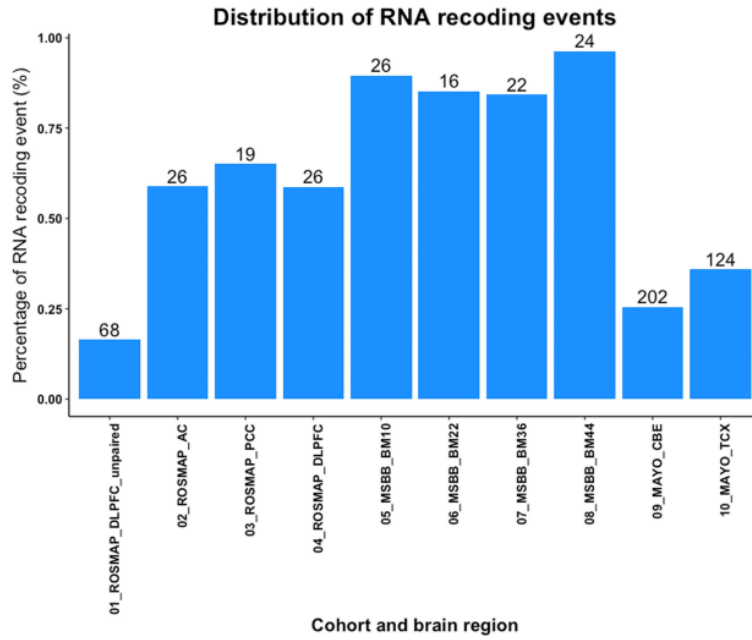


e

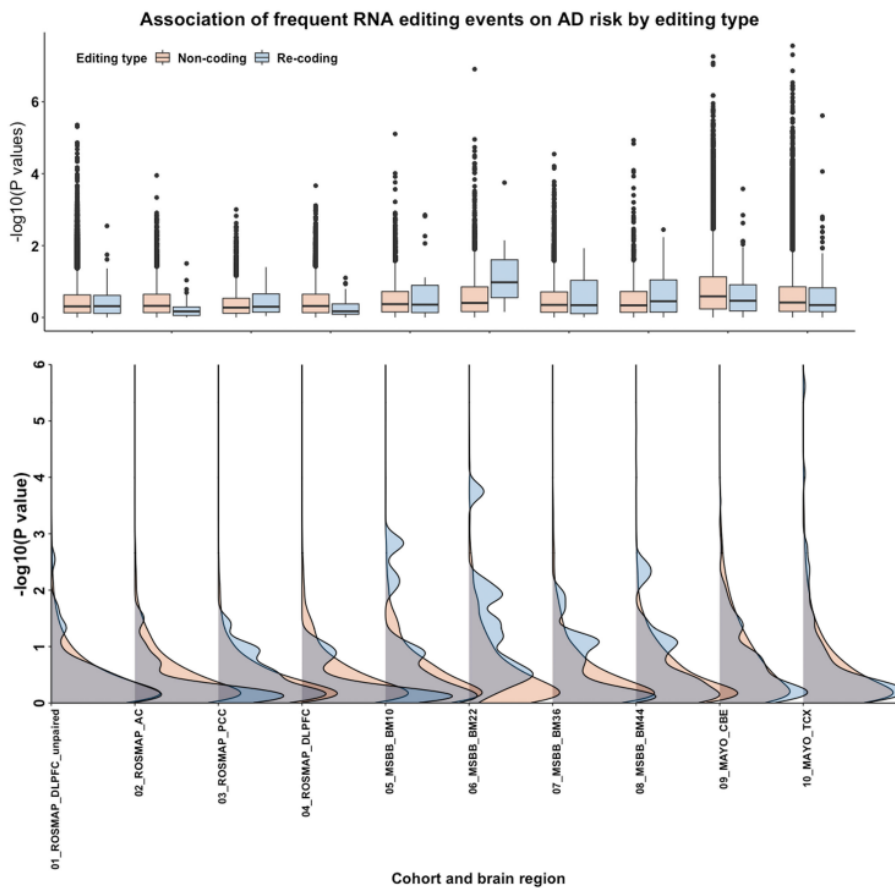


82 Fig. S6. Distribution of frequent RNA recoding events across all datasets and their associations
83 with AD status compared to the non-coding events. (a), Percentage and number of RNA recoding
84 events called in different RNA-seq datasets. The percentages are shown in blue bars and the
85 number on each bar represent the counts of RNA recoding event. (b), Box plots (upper panel)
86 and density plots (lower panel) showed the comparisons of the AD association P values ($-\log_{10}$)
87 of the non-coding (gold) and re-coding events (blue) called across all different studies with
88 RNA-seq datasets. The samples size (N) for each study are 635 subjects in the unpaired
89 ROSMAP DLPFC dataset, 68 subjects in the paired ROSMAP datasets, 142 subjects in the
90 paired MSBB datasets, and 229 subjects in the paired MAYO datasets. Y axis showed their
91 meta-analyzed P values for the general linear model (glm) with RNA editing level (% alternative
92 reads) as dependent variable, clinical AD status (0, 1, and 2 represent normal controls, mild
93 cognitive impairment (MCI), and AD patients) as exposure variable, and covariates include sex,
94 age at death, PMI, RIN, experimental batch, study (in 2 of the ROSMAP DLPFC datasets), race
95 (in MSBB BM44 dataset), and tissue source (in MAYO temporal cortex dataset). The
96 distributions of the P values by different types of RNA editing events across all datasets are
97 represented by boxplots where the minimum and the maximum values are represented by the
98 lowest and highest end of the vertical line passing the center of the box, and the first quartile
99 (25%), the median (50%), the third quartile (75%) of the values are represented as the lower,
100 center, and the higher horizontal lines of the box.

a



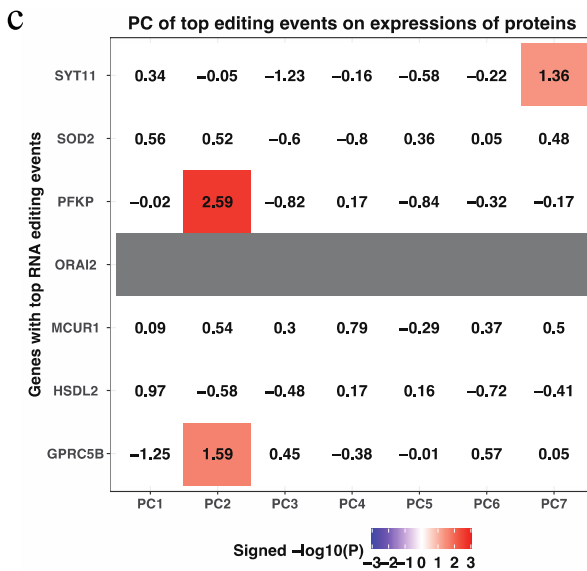
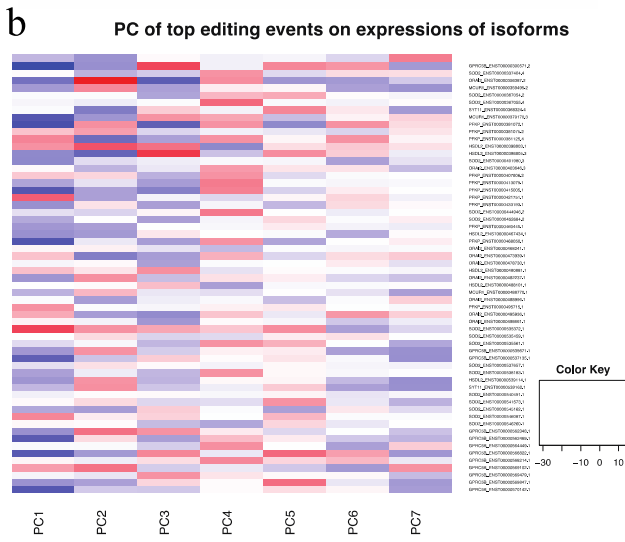
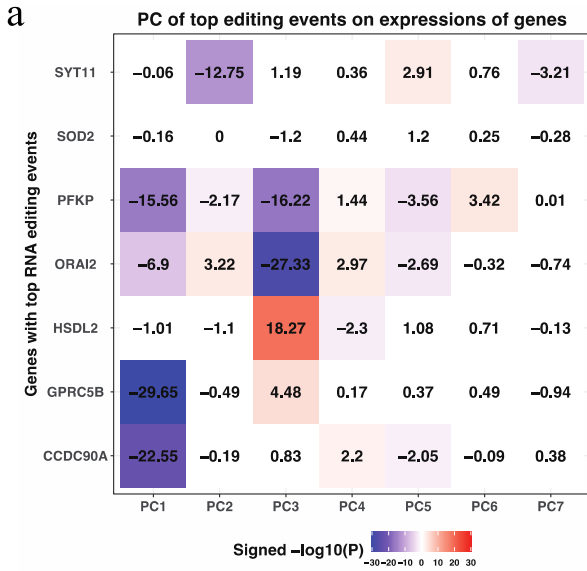
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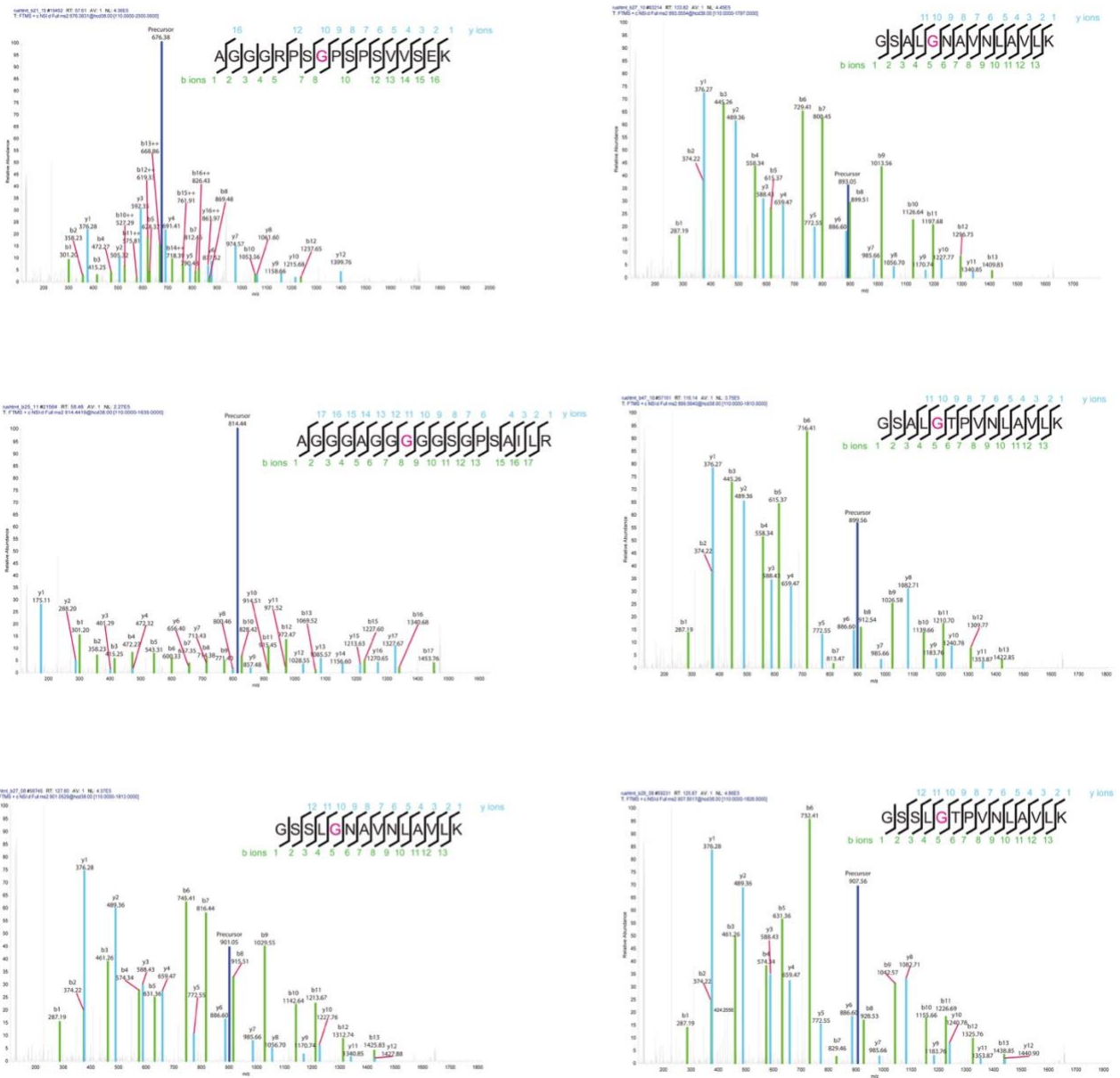
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102

103 Fig. S7. Functions of the principal components derived from the top AD associated editing
104 events on the expressions of genes, isoforms and proteins. The matrix plot shows the associations
105 of the 7 principal components (PCs) derived from the 7 top RNA editing events with expressions
106 of the mRNA and protein expressions of the genes (a,c) and isoforms (b) harboring the editing
107 event. The signed $-\log_{10}(P)$ values were presented and coded for different colors where white
108 was for values between -1.3 to 1.3 ($P=0.05$), darkening blue was for negative values from 0, and
109 darkening red was for positive values from 0. The displayed P values were not adjusted by the
110 multiple testings and were derived by two-sided tests.



112 Fig. S8. Mass-spectrometry figure of edited peptide (frequency $\geq 50\%$) hits in ROSMAP TMT
 113 proteomic dataset. The edited peptide hits are mapped to RNA editing events at *CADPS*
 114 (chr3:62860434, upper left), *GRIA3* (chrX:122598962, upper right), *CACNG8* (chr19:54485579,
 115 middle left), *GRIA3* (chrX:122598962, middle right), *GRIA2* (chr4:158281294, bottom left and
 116 right). Within each mass-spectrometry plot, X axis represents the ratio of mass to charge number
 117 of y and b ions, and the Y axis represent the relative abundance of each peak.



118

119 Table S1. Characteristic of the cohorts and picard metrics of RNA-seq datasets.

Characteristics of cohorts						
Data type	Study	Brain region	AD status	N	Age at death (years)*	Female (N, %)*
Paired	MAYO	CBE	All	229	79.62 (8.23)	112, 48.91
		TCX	Normal controls	62	82.55 (8.30)	29, 46.77
			MCI	0	NA	NA
		AD cases		65	82.29 (7.16)	38, 58.46
			PSP		102	76.14 (7.52)
	MSBB	BM10	All	142	83.05 (7.85)	93, 65.49
		BM22	Normal controls	20	82.45 (8.15)	12, 60
		BM36	MCI	24	79.29 (10.53)	11, 45.83
		BM44	AD cases	98	84.1 (6.76)	70, 71.43
	ROSMAP	AC	All	68	91.35 (6.78)	42, 61.76
		DLPFC	Normal controls	23	90.67 (6.55)	11, 47.83
			MCI	15	95.05 (4.63)	11, 73.33
		AD cases		29	89.81 (7.39)	19, 65.52
	Unpaired	ROSMAP	DLPFC	All	635	88.71 (6.65)
Normal controls				200	86.11 (6.75)	122, 61
MCI				167	88.84 (6.04)	105, 62.87
AD cases				255	90.73 (6.26)	172, 67.45
Picard metrics of RNA-seq datasets						
Picard metrics / Data type	Study	Brain region	Mean (SD)#	Comparison to MAYO CBE		
				BETA#	SE#	p#
Total reads (N)						
Paired	MAYO	CBE	45762142(8917944)	reference	reference	reference
		TCX	45002272(8941394)	-759869.8	732325	3.00E-01
	MSBB	BM10	17530795(5048660)	-28231347.4	837011.9	6.72E-195
		BM22	16583964(4761792)	-29178177.8	837011.9	3.23E-205
		BM36	15810183(4292940)	-29951959.3	837011.9	1.06E-213
		BM44	21690336(7302082)	-24071806.6	837011.9	1.01E-150
	ROSMAP	AC	36312847(5129869)	-9449295.4	1082212.7	5.50E-18
		DLPFC	39105695(10822802)	-6656447.6	1082212.7	9.42E-10
			PCC	37943854(5685988)	-7818288.2	1082212.7
		Unpaired	ROSMAP	DLPFC	28278275(8778985)	-17483867.5
Aligned reads (N)						
Paired	MAYO	CBE	45185927(8770988)	reference	reference	reference

		TCX	44501837(8813148)	-684090.2	717854.6	3.41E-01
		BM10	17314608(5038827)	-27871318.7	820473	4.31E-197
	MSBB	BM22	16412370(4752375)	-28773557.2	820473	3.88E-207
		BM36	15656050(4260491)	-29529876.6	820473	1.31E-215
		BM44	21518386(7297171)	-23667541.2	820473	1.78E-151
		AC	34444757(5010850)	-10741170	1060828.7	1.72E-23
	ROSMAP	DLPFC	37204812(10833602)	-7981115.3	1060828.7	8.27E-14
		PCC	35986334(5548391)	-9199592.6	1060828.7	9.09E-18
Unpaired	ROSMAP	DLPFC	27160757(8495879)	-18025170	592095.1	1.93E-165

Uniquely aligned reads (N)

		CBE	41769995(8130814)	reference	reference	reference
	MAYO	TCX	40915115(8147291)	-854880.4	661521.3	1.96E-01
		BM10	14178561(4104213)	-27591434.6	756086.7	3.14E-220
	MSBB	BM22	13279733(4045273)	-28490262.6	756086.7	3.13E-231
Paired		BM36	12834868(3397540)	-28935126.9	756086.7	1.08E-236
		BM44	16667204(5682457)	-25102791.3	756086.7	4.12E-190
		AC	32777825(4724429)	-8992169.8	977580.6	9.48E-20
	ROSMAP	DLPFC	35330631(10327461)	-6439364.4	977580.6	5.83E-11
		PCC	34154475(5230901)	-7615519.8	977580.6	1.11E-14
Unpaired	ROSMAP	DLPFC	25841144(8081045)	-15928851	545630.7	1.72E-154

Ribosomal bases (%)

		CBE	1.79(0.64)	reference	reference	reference
	MAYO	TCX	2.65(0.97)	0.8643061	0.1707043	4.53E-07
		BM10	0.74(3.44)	-1.0519114	0.1951067	7.88E-08
	MSBB	BM22	0.83(1.54)	-0.9601917	0.1951067	9.35E-07
Paired		BM36	0.37(0.46)	-1.4234333	0.1951067	4.38E-13
		BM44	1.27(4.76)	-0.5242833	0.1951067	7.27E-03
		AC	0.6(0.18)	-1.1944694	0.2522628	2.36E-06
	ROSMAP	DLPFC	0.7(0.21)	-1.0902018	0.2522628	1.63E-05
		PCC	0.58(0.24)	-1.2066312	0.2522628	1.86E-06
Unpaired	ROSMAP	DLPFC	2.89(1)	1.1036106	0.140799	7.64E-15

Median 3' bias

		CBE	0.68(0.07)	reference	reference	reference
	MAYO	TCX	0.7(0.07)	0.01959141	0.01411924	1.65E-01
		BM10	0.96(0.07)	0.27558379	0.01613761	7.66E-61
Paired	MSBB	BM22	0.94(0.07)	0.25440768	0.01613761	1.21E-52
		BM36	0.93(0.05)	0.24507238	0.01613761	3.48E-49
		BM44	0.86(0.07)	0.18117453	0.01613761	2.41E-28
	ROSMAP	AC	0.56(0.04)	-0.12711676	0.02086508	1.35E-09

		DLPFC	0.63(0.05)	-0.05676807	0.02086508	6.57E-03
		PCC	0.61(0.05)	-0.07113098	0.02086508	6.66E-04
Unpaired	ROSMAP	DLPFC	1.3(0.24)	0.61886072	0.01164572	0.00E+00

Correlation between number of editing events and total reads

Data type	Study	Brain region	N of subjects	N of editing events (mean,SD)	R	P
			229	27060.24, 10538.6	0.5	7.57E-16
	MAYO	CBE				
		TCX	229	12490.45, 3330.1	0.69	< 2.2E-16
		BM10	142	857.95, 397.52	0.78	< 2.2E-16
Paired	MSBB	BM22	142	602.19, 198.07	0.78	< 2.2E-16
		BM36	142	771.55, 302.95	0.78	< 2.2E-16
		BM44	142	872.48, 344.30	0.86	< 2.2E-16
		AC	68	1544.57, 613.03	0.16	0.19
	ROSMAP	DLPFC	68	1474.37, 675.52	0.29	0.01
		PCC	68	881.37, 446.84	0.02	0.86
Unpaired	ROSMAP	DLPFC	635	12257.07, 7006.66	0.31	3.00E-15

120 *Age at death is represented by mean (SD) and female is represented by number (N) and percentage (%)
121 #BETA represent the difference in each picard metric between MAYO CBE dataset compared to all the other 9
122 datasets, and the SE and P represent the corresponding standard error and P value. The displayed P values were not
123 adjusted by the multiple testings and were derived by two-sided tests.
124 Abbreviation: SD, standard deviation; MAYO, Mayo RNAseq study; CBE, cerebellum; TCX, temporal cortex;
125 MSBB, Mount Sinai Brain Bank; AC, anterior cingulate cortex; DLPFC, dorsolateral prefrontal cortex; PCC,
126 posterior cingulate cortex; PSP, progressive supranuclear palsy.
127

128

129 Table S2. The recoding RNA editing events with proteomic peptide hits.

Annotation^					
Unique identifier	NM	CDS	Exon number	DNA substitute	Amino acid substitute
chr1_16133978_UQCRHL	NM_001089591.1	51 to 326	1	A65G	E22G
chr1_110256304_GSTM5_peptide1	NM_000851.3	72 to 728	5	A281G	K94R
chr1_110256304_GSTM5_peptide2					
chr1_160302244_COPA	NM_001098398.1	395 to 4096	6	A490G	I164V
chr3_62423807_CADPS	NM_183393.2	361 to 4185	25	A3512G	E1171G
chr3_62860434_CADPS	NM_183393.2	361 to 4185	1	A271G	S91G
chr4_158281294_GRIA2_peptide1	NM_001083619.1	460 to 3111	13	A2290G	R764G
chr4_158281294_GRIA2_peptide2	NM_000826.3 / NM_001083620.1	460 to 3111 / 112 to 2622	13	A2290G / A2149G	R764G / R717G
chr6_26104432_HIST1H4C	NM_003542.3	1 to 312	1	A257G	D86G
chr19_47152737_DACT3	NM_001301046.1	442 to 1656	4	A217G	S73G
chr19_54485579_CACNG8	NM_031895.5	108 to 1385	4	A754G	S252G
chrX_122598962_GRIA3_peptide1	NM_000828.4	294 to 2978	13	A2323G	R775G

chrX_122598962_GRIA3_peptide2

NM_007325.4

294 to 2978

13

A2323G

R775G

Annotation[^]

Unique identifier	RNA sequence ^{&}	Edited peptide sequence	Non-edited peptide sequence
chr1_16133978_UQCRHL	75atgctaccgaatccggagatcctgagggaggaagaggA[G]agaggaggaattagtgatccccta acaacagtgagag253	9MLTESGDPEEEEEEGEELVDPLTTVR24	9MLTESGDPEEEEEEEELVDPLTTVR24
chr1_110256304_GSTM5_peptide1	316caagcacaacctgtgtggggagacagaagaggagaA[G]ga354	83KHNLCGETEEEEK94	83KHNLCGETEEEEK94
chr1_110256304_GSTM5_peptide2		84HNLCEEEEEK94	84HNLCEEEEEK94
chr1_160302244_COPA	875gtttggatA[G]tttctgtctgagg898	161VWDVVSGLR168	161VWDISGLR168
chr3_62423807_CADPS	3863aggtcaatgA[G]ggagatgtacatagaaggt3892	1169VNGEMYIER1177	1169VNEEMYIER1177
chr3_62860434_CADPS	610gctggcggcggccgcccctcA[G]gccccagcccgtcgggtgtgagcagagaag660	84AGGGRPSGPSPSVSEK100	84AGGGRPSGPSPSVSEK100
chr4_158281294_GRIA2_peptide1	2737ggatcctcattaaA[G]aatgctgttaacctgcagactactaaaa2778	760GSSLGNVAVNLAVLK773	765NAVNLAVLK773
chr4_158281294_GRIA2_peptide2	2737/2248ggatcctcattaaA[G]aaccccagtaaatcttgcagattgaaa2778/2289	760/713GSSLGTPVNLAVLK773/726	765/718NAVNLAVLK773/726
chr6_26104432_HIST1H4C	240aaactgtcacagccatggA[G]ttagtatatgccttaaac277	81TVTAMGVVYALK92	81TVTAMDVVYALK92
chr19_47152737_DACT3	625cagcggcggccggcggcgtctccgccccggcA[G]gcgcgcgaccggcggg675	62QRPPDASPSPGGARPAR78	62QRPPDASPSPGARSARPAR78
chr19_54485579_CACNG8	840gccggcggggcgccggcgccA[G]gtggcggggcggccccctggccatcctcgt893	245AGGGAGGGGGSGPSAILR262	245AGGGAGGGGGSGPSAILR262
chrX_122598962_GRIA3_peptide1	2604ggetcagcattagA[G]aatgctgttaacctgcagattataaa2645	771GSALGNVAVNLAVLK784	776NAVNLAVLK784
chrX_122598962_GRIA3_peptide2	2604ggetcagcattagA[G]aacgcctgtaacctgcagattgaaa2645		776TPVNLAVLK784

Concordance, correlation, and expression levels

Unique identifier	Concordance [§]				Correlation [*]		Expression level [#]		
	N1, %	N2, %	N3, %	N4, %	r	P	Edited RNA reads (%)	Edited peptide quantity (abundance)	Non_edited peptide quantity (abundance)
chr1_16133978_UQCRHL	13, 7.6 %	84, 49.12 %	60, 35.09 %	14, 8.19 %	0.07	0.39	33.67±40.05	13.5±2.73	7.7±1.87
chr1_110256304_GSTM5_peptide1	0, 0 %	164, 95.91 %	5, 2.92 %	2, 1.17 %	0.02	0.83	0.46±2.34	56.48±37.39	95.97±75.13
chr1_110256304_GSTM5_peptide2	2, 1.17 %	89, 52.05 %	3, 1.75 %	77, 45.03 %	0.03	0.70		44.98±14.25	585.09±446.40

chr1_160302244_COPA	23, 13.45 %	54, 31.58 %	74, 43.27 %	20, 11.7 %	0.08	0.30	13.27±13.80	127.3±8.2	239.06±20.12
chr3_62423807_CADPS	47, 27.49 %	30, 17.54 %	82, 47.95 %	12, 7.02 %	-	0.05	14.67±9.95	177.36±17.56	507.38±45.40
chr3_62860434_CADPS	0, 0 %	19, 11.11 %	0, 0 %	152, 88.89 %	NA	NA	0.03±0.73	229.97±23.26	929.77±49.22
chr4_158281294_GRIA2_peptide1	114, 66.67 %	2, 1.17 %	15, 8.77 %	40, 23.39 %	-	0.09	0.30±0.20	738.08±62.41	581.38±48.07
chr4_158281294_GRIA2_peptide2	114, 66.67 %	2, 1.17 %	15, 8.77 %	40, 23.39 %	-	0.08	500.35±69.07	197.64±20.95	
chr6_26104432_HIST1H4C	1, 0.58 %	167, 97.66 %	0, 0 %	3, 1.75 %	-	0.29	0.07±1.83	54.57±52.73	2003.41±916.83
chr19_47152737_DACT3	2, 1.17 %	157, 91.81 %	5, 2.92 %	7, 4.09 %	0.24	1.40E-03	3.82±15.58	18.04±4.62	NA
chr19_54485579_CACNG8	18, 10.53 %	15, 8.77 %	2, 1.17 %	136, 79.53 %	0.02	0.78	2.15±6.33	85.89±11.66	501.95±48.83
chrX_122598962_GRIA3_peptide1	83, 48.54 %	6, 3.51 %	11, 6.43 %	71, 41.52 %	0.21	6.74E-03	49.78±45.77	577.77±44.71	581.38±48.07
chrX_122598962_GRIA3_peptide2	83, 48.54 %	6, 3.51 %	11, 6.43 %	71, 41.52 %	-	0.07	545.44±64.34	197.64±20.95	

AD association@

Unique identifier	Reported event	Reported AD association	Peptide			RNA			
			BETA	STDERR	P	N	BETA	STDERR	P
chr1_16133978_UQCRHL	No	No	-0.007	0.03	0.84	269	-2.22	1.71	0.20
chr1_110256304_GSTM5_peptide1	Yes	No	0.004	0.01	0.66	25	0.32	0.12	6.40E-03
chr1_110256304_GSTM5_peptide2			0.492	0.20	0.01				
chr1_160302244_COPA	Yes	Yes	-0.047	0.04	0.22	346	1.31	0.65	0.04
chr3_62423807_CADPS	Yes	No	0.000	0.07	0.995	469	-0.85	0.43	0.05
chr3_62860434_CADPS	Yes	No	0.024	0.07	0.72	1	-0.05	0.04	0.22
chr4_158281294_GRIA2_peptide1	Yes	Yes	-0.014	0.02	0.46	458	-1.15	0.85	0.18
chr4_158281294_GRIA2_peptide2			-0.022	0.05	0.67				
chr6_26104432_HIST1H4C	No	No	-0.093	0.10	0.33	1	-0.07	0.09	0.47
chr19_47152737_DACT3	Yes	No	-0.156	0.09	0.09	38	-0.18	0.75	0.81
chr19_54485579_CACNG8	Yes	No	-0.010	0.08	0.91	72	-0.04	0.30	0.89
chrX_122598962_GRIA3_peptide1	Yes	Yes	-0.022	0.01	0.04	346	-5.40	2.24	0.02

130 ^On both the RNA and peptide level, the nucleotide and amino acid at the editing site were represented by the two red characters
131 separated by slash. The left character is the non-edited one while the right character in the bracket is the edited one.
132 \$N1, %: Number and percentage of subjects with detection of editings on both RNA and peptide level; N2, %: Number and
133 percentage of subjects without detection of editings on either RNA or peptide level; N3, %: Number and percentage of subjects with
134 detection of editing on RNA but not peptide level; N4, %: Number and percentage of subjects with detection of editing on peptide but
135 not RNA level. Chi-square and corresponding P values for the contingency table of the four numbers are displayed.
136 *r and P represent the correlation coefficient and corresponding P value between fraction of edited RNA read and abundance.
137 #Values are represented by mean \pm SD. Edited RNA reads were represented by the percentage of alternative read over the total
138 amount of reads at that locus. On the peptide level, unit are shown in both the normalized abundance.
139 @N represent the number of subjects (out of total 635) carrying the editing event. BETA, STDERR, and P represent the regression
140 coefficient, standard error and corresponding P values of the generalized linear model of RNA editing level (log2 of the ratio of the
141 normalized abundance of the edited peptide over the nonedited peptide and % alternative allele on RNA level) as the dependent
142 variable and clinical AD status (0, 1, and 2 represent normal controls, mild cognitive impairment (MCI), and AD patients) as exposure
143 variable, and covariates include sex, age at death, postmortem interval (PMI), study (ROS or MAP), RIN and experimental batch (for
144 RNA dataset). Peptide analysis is within 166 subjects with available data of all the variables included in the analysis and also those
145 subjects with RNA-seq datasets. RNA-seq analysis include 620 subjects with RNA-seq datasets and all the included variables. All the
146 statistical tests are two-sided.
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Table S3. Regional differences in gene expressions of *ADARs*.

Study	Comparison	ADAR genes	N	BETA ^{&}	STDERR ^{&}	p ^{&}
ROSMAP multi-region# (N=68)	AC - DLPFC	<i>ADAR1</i>	68	-0.64	1.21	0.6
		<i>ADAR2</i>	68	-2.16	0.32	1.20E-11
		<i>ADAR3</i>	68	0.25	0.36	0.49
		<i>ADARs</i> [§]	68	-3.01	1.44	0.04
	PCC - DLPFC	<i>ADAR1</i>	68	-8.14	1.06	1.80E-14
		<i>ADAR2</i>	68	-2.24	0.3	6.20E-14
		<i>ADAR3</i>	68	-2.25	0.32	1.00E-12
		<i>ADARs</i> [§]	68	-8.11	1.34	1.40E-09
	PCC - AC	<i>ADAR1</i>	68	-7.07	1.18	1.90E-09
		<i>ADAR2</i>	68	-0.07	0.28	0.79
		<i>ADAR3</i>	68	-2.45	0.36	1.30E-11
		<i>ADARs</i> [§]	68	-4.68	1.41	9.30E-04
MSBB* (N=142)	BM36 - BM44	<i>ADAR1</i>	142	-966	252	1.20E-04
		<i>ADAR2</i>	142	-160	49.7	1.30E-03
		<i>ADAR3</i>	142	-341	57.7	3.60E-09
		<i>ADARs</i> [§]	142	-778	264	3.30E-03
	BM36 - BM10	<i>ADAR1</i>	142	-146	193	0.45
		<i>ADAR2</i>	142	-148	35.8	3.90E-05
		<i>ADAR3</i>	142	104	37.1	4.90E-03
		<i>ADARs</i> [§]	142	-396	198	0.05
	BM36 - BM22	<i>ADAR1</i>	142	121	142	0.39
		<i>ADAR2</i>	142	-132	31.6	3.00E-05
		<i>ADAR3</i>	142	26.3	30.8	0.39
		<i>ADARs</i> [§]	142	-41.5	150	0.78
BM22 - BM10	<i>ADAR1</i>	142	-238	183	0.19	
	<i>ADAR2</i>	142	-17.8	37.9	0.64	
	<i>ADAR3</i>	142	93.7	32.1	3.50E-03	
	<i>ADARs</i> [§]	142	-350	195	0.07	
BM44 - BM10	<i>ADAR1</i>	142	937	254	2.20E-04	
	<i>ADAR2</i>	142	17.9	49.3	0.72	
	<i>ADAR3</i>	142	457	54.9	1.10E-16	
	<i>ADARs</i> [§]	142	491	260	0.06	
BM22 - BM44	<i>ADAR1</i>	142	-1030	231	9.10E-06	
	<i>ADAR2</i>	142	-32.1	51.2	0.53	
	<i>ADAR3</i>	142	-318	50.2	2.40E-10	

		<i>ADARs</i> [§]	142	-767	244	1.70E-03
	TCX - CBE	<i>ADAR1</i>	229	-2250	325	3.90E-12
MAYO*		<i>ADAR2</i>	229	-7500	263	<2.2E-16
(N=229)		<i>ADAR3</i>	229	1460	145	<2.2E-16
		<i>ADARs</i> [§]	229	-11200	578	<2.2E-16

155 #Gene expression units for ROSMAP multi-region study are represented by transcripts per
156 million (TPM) values obtained through RSEM tool with the reference transcriptome of
157 GENCODE v24.

158 *Gene expression units for MSSB and MAYO RNA-seq study are represented by counts of the
159 reads of the gene using STAR tool by setting quantMode as GeneCounts according to the
160 reference transcriptome of GENCODE v24.

161 § ADARs = ADAR1+ADAR2-ADAR3.

162 & The BETA, STDERR, and *P* represent the regression coefficient, standard error, and the
163 corresponding P values for the exposure variable of region (coded as 1 for one brain region and 0
164 for the other brain region) in the general linear mixed model using R lme package with
165 dependent variable of gene expression levels of ADARs, random effect of subject ID, and fixed
166 effects of the covariates including age at death, sex, postmortem interval (PMI), and RIN score.
167 The displayed *P* values were not adjusted by the multiple testings and were derived by two-sided
168 tests.

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171 Table S4. Top RNA editing events associated with AD status.

Associations with AD status in Stage I and II [#]								
Gene: top RNA editing	Outcome	Stage I			Stage II			
		N	BETA (STDERR)	P	N	BETA (STDERR)	Direction	P
<i>SYT11:chr1_155851645</i>	AD status in all	620	1.36 (0.32)	2.09E-05	924	1.66 (0.26)	"++++"	2.94E-10
<i>MCUR1:chr6_13788361</i>	AD status in all	620	2.30 (0.60)	1.36E-04	924	2.75 (0.46)	"++++"	1.66E-09
<i>SOD2:chr6_13788361</i>	AD status in all	620	1.31 (0.31)	2.75E-05	924	1.80 (0.27)	"++++"	1.96E-11
<i>ORAI2:chr7_102096952</i>	AD status in all	620	2.94 (0.64)	4.64E-06	782	2.74 (0.55)	"+x++"	4.80E-07
<i>HSDL2:chr9_115237504</i>	AD status in all	620	5.13 (1.33)	1.25E-04	782	5.23 (1.04)	"+x++"	5.49E-07
<i>PFKP:chr10_3168677</i>	AD status in all	620	4.89 (1.44)	7.54E-04	924	5.59 (1.05)	"++++"	1.08E-07
<i>SYT11:chr1_155851645</i>	AD status in females	398	1.98 (0.41)	1.65E-06	581	2.23 (0.34)	"++++"	3.60E-11
<i>MCUR1:chr6_13788361</i>	AD status in females	398	2.74 (0.78)	5.22E-04	581	2.98 (0.57)	"++++"	2.10E-07
<i>SOD2:chr6_13788361</i>	AD status in females	398	1.81 (0.39)	4.33E-06	581	2.13 (0.34)	"++++"	3.08E-10
<i>GPRC5B:chr16_19876365</i>	AD status in females	398	5.59 (1.17)	2.68E-06	448	5.87 (1.14)	"+x+x"	2.79E-07
Functional associations in STAGE I unpaired ROSMAP DLPFC samples*								
Gene: top RNA editing	Outcome Category	Outcome	N	BETA	STDERR	P		
<i>SYT11:chr1_155851645</i>	ADARs gene expression	<i>ADAR1</i>	505	0.0427	0.0164	0.00938		
		<i>ADAR2</i>	505	-0.169	0.0197	1.14E-16		
		<i>ADAR3</i>	505	0.308	0.0538	1.75E-08		
	Target gene expression	<i>SYT11</i>	505	0.166	0.0334	9.60E-07		
	Target isoform expression	<i>ENST00000368324.4</i>	526	4.37	1.46	0.00284		
	Target protein expression	<i>SYT11_Q9BT88</i>	78	-0.06	0.02	0.01		
<i>SOD2:chr6_13788361</i>	ADARs gene expression	<i>ADAR1</i>	505	0.00502	0.0175	0.774		
		<i>ADAR2</i>	505	-0.0824	0.0221	0.00021		
		<i>ADAR3</i>	505	0.19	0.0582	0.00119		
	Target gene expression	<i>SOD2</i>	505	0.0517	0.0493	0.295		
	Target isoform expression	<i>ENST00000546260.1</i>	526	-0.0528	0.0252	0.0369		
	Target protein expression	<i>SOD2_P04179</i>	78	-0.02	0.03	0.54		
<i>PFKP:chr10_3168677</i>	ADARs gene expression	<i>ADAR1</i>	505	0.0481	0.0164	0.00346		
		<i>ADAR2</i>	505	0.0158	0.0211	0.455		

		<i>ADAR3</i>	505	0.0959	0.0554	0.0841
	Target gene expression	<i>PFKP</i>	505	0.388	0.0376	9.17E-23
	Target isoform expression	<i>ENST00000381072.1</i>	526	22.3	1.97	1.00E-26
	Target protein expression	<i>PFKP_Q01813</i>	78	-0.01	0.02	0.61
<i>ORAI2:chr7_102096952</i>		<i>ADAR1</i>	505	-0.0412	0.0175	0.0188
	ADARs gene expression	<i>ADAR2</i>	505	0.0101	0.0225	0.654
		<i>ADAR3</i>	505	0.146	0.0588	0.0132
	Target gene expression	<i>ORAI2</i>	505	0.254	0.0334	1.43E-13
	Target isoform expression	<i>ENST00000356387.2</i>	526	2.43	0.221	2.04E-25
	Target protein expression	Protein	78	NA	NA	NA
<i>MCUR1:chr6_13788361</i>		<i>ADAR1</i>	505	0.0568	0.0163	0.000555
	ADARs gene expression	<i>ADAR2</i>	505	-0.0426	0.0211	0.0438
		<i>ADAR3</i>	505	0.11	0.0554	0.0468
	Target gene expression	<i>MCUR1</i>	505	0.0663	0.0197	0.000823
	Target isoform expression	<i>ENST00000379170.3</i>	526	1.22	0.216	2.40E-08
	Target protein expression	<i>MCUR1_Q96AQ8</i>	78	-0.08	0.04	0.05
<i>HSDL2:chr9_115237504</i>		<i>ADAR1</i>	505	-0.043	0.0189	0.0233
	ADARs gene expression	<i>ADAR2</i>	505	-0.126	0.0237	1.67E-07
		<i>ADAR3</i>	505	0.175	0.0635	0.00594
	Target gene expression	<i>HSDL2</i>	505	0.201	0.0367	6.81E-08
	Target isoform expression	<i>ENST00000398803.1</i>	526	1.25	0.305	4.93E-05
	Target protein expression	<i>HSDL2_Q6YN16</i>	78	-0.04	0.03	0.13
<i>GPRC5B:chr16_19876365</i>		<i>ADAR1</i>	505	-0.0318	0.0167	0.0573
	ADARs gene expression	<i>ADAR2</i>	505	-0.0568	0.0213	0.00792
		<i>ADAR3</i>	505	0.233	0.0555	3.26E-05
	Target gene expression	<i>GPRC5B</i>	505	0.155	0.0438	0.000422
	Target isoform expression	<i>ENST00000300571.2</i>	526	17.8	3.62	1.16E-06
	Target protein expression	<i>GPRC5B.Q9NZH0.2</i>	78	0.16	0.08	0.05

Associations in STAGE I unpaired ROSMAP DLPFC samples before and after adjustment of the neuron proportion or total reads

Gene: top RNA editing	Outcome	Unadjusted <i>P</i>	Adjusted neuron transcription module			Adjusted total reads		
			N	BETA (STDERR)	<i>P</i>	N	BETA (STDERR)	<i>P</i>

<i>SYT11:chr1_155851645</i>	AD status in all [#]	2.09E-05	493	1.21 (0.36)	7.05E-04	614	1.34 (0.32)	3.27E-05
<i>MCUR1:chr6_13788361</i>		1.36E-04	493	2.29 (0.69)	9.99E-04	614	2.21 (0.60)	2.32E-04
<i>SOD2:chr6_13788361</i>		2.75E-05	493	1.12 (0.35)	1.24E-03	614	1.31 (0.31)	3.30E-05
<i>ORAI2:chr7_102096952</i>		4.64E-06	493	2.49 (0.71)	4.90E-04	614	3.02 (0.63)	2.07E-06
<i>HSDL2:chr9_115237504</i>		1.25E-04	493	4.81 (1.49)	1.38E-03	614	0.52 (0.01)	1.01E-04
<i>PFKP:chr10_3168677</i>		7.54E-04	493	4.39 (1.65)	8.04E-03	614	0.48 (0.01)	9.38E-04
<i>GPRC5B:chr16_19876365</i>	AD status in females [#]	2.68E-06	306	5.45 (1.42)	1.48E-04	392	0.57 (0.01)	2.40E-06
<i>ORAI2:chr7_102096952</i>	PHFtau in all*	4.72E-08	500	0.11 (0.03)	3.66E-05	622	0.13 (0.02)	8.07E-08
<i>KCNIP2:chr10_103596067</i>		6.72E-07	500	0.06 (0.01)	1.10E-05	622	0.07 (0.01)	1.08E-06
<i>GPRC5B:chr16_19874115</i>		5.54E-07	500	0.12 (0.03)	1.14E-04	622	0.15 (0.03)	6.59E-07
<i>YPEL1:chr22_22078228</i>		1.77E-07	500	0.17 (0.03)	1.03E-06	622	0.18 (0.03)	9.39E-08
<i>AC174470.1:chr17_79780692</i>	β -amyloid in all*	9.38E-07	500	-0.08 (0.02)	2.23E-04	622	-0.09 (0.02)	8.51E-07
<i>ORAI2:chr7_102096952</i>	Neuritic plaque burden in all*	8.47E-07	505	0.008 (0.002)	1.59E-05	628	8.37E-03 (1.61E-04)	2.97E-07
<i>CABP1:chr12_121078907</i>		2.74E-07	505	-0.009 (0.002)	2.08E-05	628	-9.34E-03 (1.84E-04)	5.41E-07
<i>AC174470.1:chr17_79780692</i>	Cognition decline*	5.76E-07	481	0.002 (6E-04)	1.82E-03	593	2.35E-03 (4.7E-04)	7.44E-07
<i>MUM1:chr19_1371887</i>		8.50E-07	481	-7E-04 (1E-04)	9.95E-06	593	-6.88E-04 (1.37E-04)	6.65E-07

172 #Top RNA editing events associated with AD risk in Stage I and II genome-wide analysis in all subjects and female only (Stage II meta-P $\leq 1.2 \times 10^{-6}$). Stage I
173 dataset include ROSMAP unpaired samples of dorsolateral prefrontal cortex (DLPFC), and the Stage II datasets include samples from Stage I and BM44 of
174 MSBB study (MSBB_BM44), temporal cortex (TCX) of MAYO study (MAYO_TCX), and DLPFC samples of independent set of subjects participating in the
175 ROSMAP multi-region study (ROSMAP multi-region DLPFC). BETA, STDERR, and P represent the regression coefficient, standard error and corresponding P
176 values of the generalized linear model of RNA editing level (% alternative read) as the dependent variable and clinical AD status (0, 1, and 2 represent normal
177 controls, mild cognitive impairment (MCI), and AD patients) as exposure variable, and covariates include sex, age at death, postmortem interval (PMI), RIN,
178 experimental batch, study (in 2 of the ROSMAP DLPFC datasets), race (in MSBB BM44 dataset), and tissue source (in MAYO temporal cortex dataset). The
179 direction column in Stage II showed the sign of the BETA from each included dataset in the order of ROSMAP unpaired DLPFC, MSBB_BM44, MAYO_TCX,
180 and ROSMAP multi-region DLPFC. The displayed P values were not adjusted by the multiple testings and were derived by two-sided tests.

181 * BETA, STDERR, and P represent the regression coefficient, standard error and corresponding P values of the generalized linear model of RNA editing level
182 (% alternative allele for analysis of expressions of gene and isoforms and continuous outcome of AD neuropathological traits) or RNA editing status (alternative
183 allele is present or not for protein analysis) as the exposure and each of the trait (expression levels of genes, isoforms and proteins) as the outcomes with the
184 adjustment of covariates of age at death, sex, RIN, postmortem interval, study (ROS or MAP) and experimental batches. The expression of genes were
185 represented by log₂ of FPKM, the expression of isoforms were represented by FPKM, and the protein expressions were represented by the log₂-transformed ratio
186 (over the pooled standard following normalization). The displayed P values were not adjusted by the multiple testings and were derived by two-sided tests.
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190 Table S5. Association of *ORAI2* editing event (chr7:102096952, hg19) on multiple traits in ROSMAP unpaired DLPFC dataset
 191 (N=635).

Associations with multiple AD-related traits				
Traits	N	BETA	STDERR	<i>P</i>
PHFtau*	500	12.90	2.33	4.72E-08
β-amyloid*	500	4.16	1.22	7.31E-04
Neuritic plaque burden*	505	0.79	0.16	8.47E-07
Cognition decline*	481	-0.14	0.03	3.07E-06
Clinical AD status [#]	620	0.03	0.01	4.64E-06

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 193 *The BETA, STDERR, and *P* represent the regression coefficient and its corresponding standard error and *P* values for the exposure
 194 variable of *ORAI2* editing level (% alternative reads) in the generalized linear model with dependent variable of each trait and
 195 covariates of age at death, sex, RIN score, postmortem interval, study (ROS or MAP) and experimental batches. The displayed *P*
 196 values were not adjusted by the multiple testings and were derived by two-sided tests.

197 [#]The BETA, STDERR, and *P* represent the regression coefficient and its corresponding standard error and *P* values the general linear
 198 model of RNA editing level (% alternative read) as the dependent variable and clinical AD status (0, 1, and 2 represent normal
 199 controls, mild cognitive impairment (MCI), and AD patients) as exposure variable, and covariates include sex, age at death,
 200 postmortem interval, RIN, experimental batch, and study (ROS or MAP). The displayed *P* values were not adjusted by the multiple
 201 testings and were derived by two-sided tests.

202 Abbreviation: PHFtau, abnormally phosphorylated Tau protein, AT8.; Cog. dec., global cognitive decline (longitudinal).

203

204 Table S6. Comparisons between AD genes and non-AD genes.

Types	Genes	N of RNA editing events	BETA (minimum to maximum)	P values (minimum to maximum)
Non-AD genes	*	40,692	-5.250E-04 to 5.587E-04	1.9638E-11 to 1
AD genes	CACNA2D3, CELF1, CLU, MEF2C, PICALM, PILRA	113	-0.00016 to 3.65E-04	0.01 to 0.99
	APOE,CR1,BIN1,INPP5D,FBXL7,CD2AP,HLA-DRB5, HLA-DRB1,EPHA1,NME8,ZCWPW1,NYAP1,PTK2B,	0		
	ECHDC3,SPI1,MS4A2,MS4A6A,SORL1,SLC24A4,RIN3,	0		
	FERMT2,ACE,DSG2,ABCA7,CD33,HMHA1,GRIN3B,CASS4	0		

205 The primary analysis was conducted within each of the 10 dataset, and a general linear model (glm) was utilized to analyze the
 206 associations between RNA editing levels (% alternative reads) and AD status (0, 1, and 2 represent normal controls, mild cognitive
 207 impairment (MCI), and AD patients). we conducted the meta-analysis on the largest cohort including samples from each study of:
 208 ROSMAP unpaired DLPFC, the DLPFC dataset of the paired ROSMAP multi-region RNA-seq project, BM44 dataset of the paired
 209 MSBB RNA-seq project, and the temporal cortex dataset of the paired MAYO RNA-seq project. The beta estimates and standard
 210 errors from each dataset were meta-analyzed using an inverse variance-weighted, fixed-effects approach implemented in METAL.
 211 The displayed P values were not adjusted by the multiple testings and were derived by two-sided tests.

212 REFERENCES

- 213 1. Allen M, *et al.* Human whole genome genotype and transcriptome data for Alzheimer's and
214 other neurodegenerative diseases. *Sci Data* **3**, 160089 (2016).
- 215 2. De Jager PL, *et al.* A multi-omic atlas of the human frontal cortex for aging and Alzheimer's
216 disease research. *Sci Data* **5**, 180142 (2018).
- 217 3. Wang M, *et al.* The Mount Sinai cohort of large-scale genomic, transcriptomic and proteomic
218 data in Alzheimer's disease. *Sci Data* **5**, 180185 (2018).
- 219 4. Allen M, *et al.* Conserved brain myelination networks are altered in Alzheimer's and other
220 neurodegenerative diseases. *Alzheimers Dement* **14**, 352-366 (2018).
- 221 5. Allen M, *et al.* Divergent brain gene expression patterns associate with distinct cell-specific tau
222 neuropathology traits in progressive supranuclear palsy. *Acta Neuropathol* **136**, 709-727 (2018).
- 223 6. Li B, Dewey CN. RSEM: accurate transcript quantification from RNA-Seq data with or without a
224 reference genome. *BMC Bioinformatics* **12**, 323 (2011).
- 225 7. Dobin A, *et al.* STAR: ultrafast universal RNA-seq aligner. *Bioinformatics* **29**, 15-21 (2013).
- 226 8. Tan MH, *et al.* Dynamic landscape and regulation of RNA editing in mammals. *Nature* **550**, 249-
227 254 (2017).
- 228 9. Mertins P, *et al.* Reproducible workflow for multiplexed deep-scale proteome and
229 phosphoproteome analysis of tumor tissues by liquid chromatography-mass spectrometry. *Nat*
230 *Protoc* **13**, 1632-1661 (2018).
- 231 10. Mertins P, *et al.* Reproducible workflow for multiplexed deep-scale proteome and
232 phosphoproteome analysis of tumor tissues by liquid chromatography-mass spectrometry.
233 *Nat Protoc* **13**, 1632-1661 (2018).

- 234 11. Wingo TS, *et al.* Integrating Next-Generation Genomic Sequencing and Mass Spectrometry To
235 Estimate Allele-Specific Protein Abundance in Human Brain. *J Proteome Res* **16**, 3336-3347
236 (2017).
- 237 12. Tukey JW. *Exploratory Data Analysis*. Addison-Wesley (1977).
- 238 13. McKenzie AT, *et al.* Multiscale network modeling of oligodendrocytes reveals molecular
239 components of myelin dysregulation in Alzheimer's disease. *Molecular Neurodegeneration* **12**,
240 82 (2017).
- 241 14. Callister SJ, *et al.* Normalization approaches for removing systematic biases associated with
242 mass spectrometry and label-free proteomics. *J Proteome Res* **5**, 277-286 (2006).
- 243
- 244