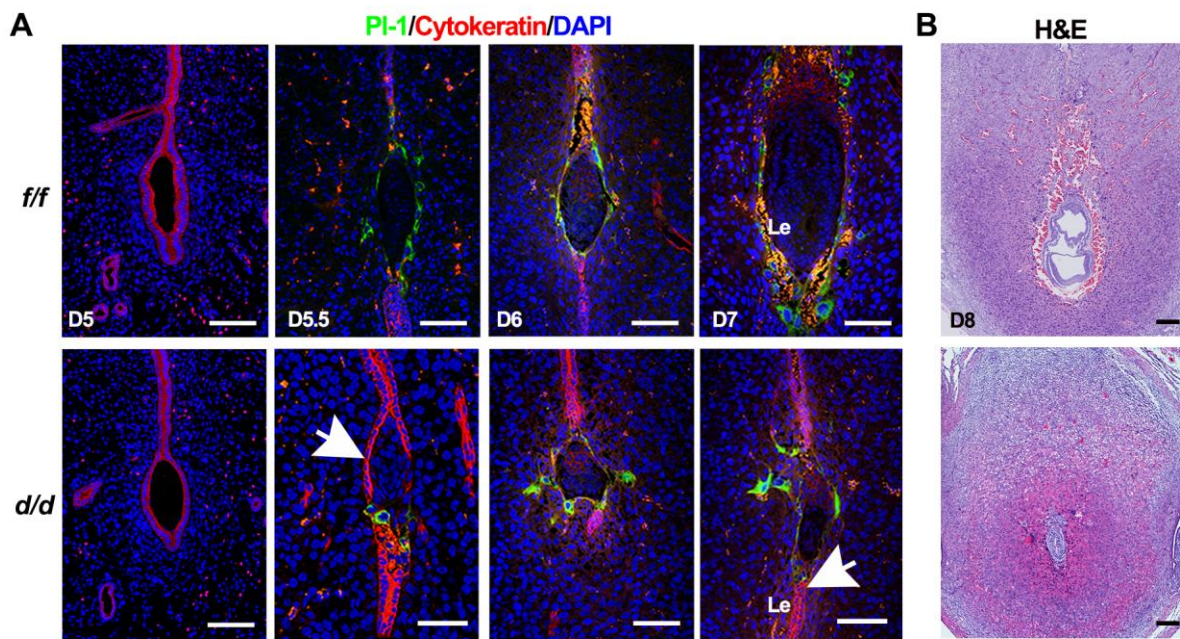


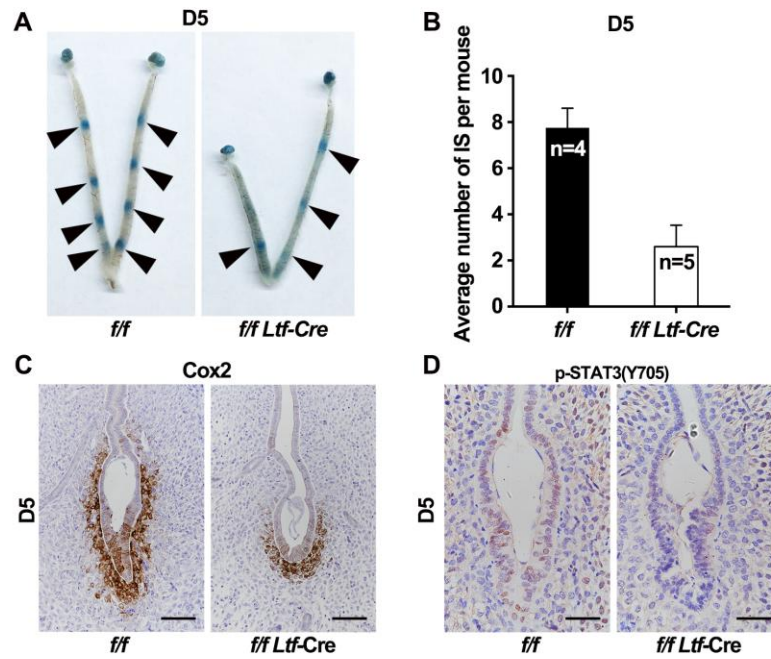
## Supplementary material

### Sequential activation of uterine epithelial IGF1R by stromal IGF1 and embryonic IGF2 directs normal uterine preparation for embryo implantation

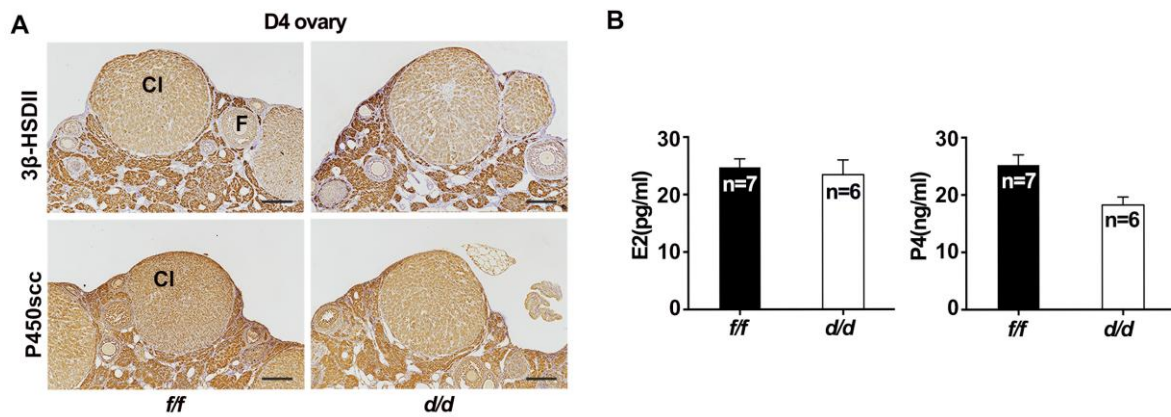


#### Supplementary Figure S1. Post-implantation embryo development in control and knockout mice.

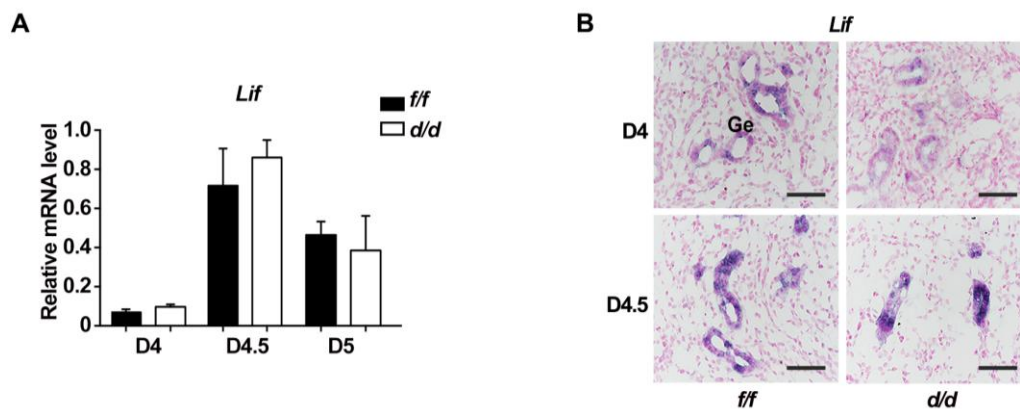
(A) Cytokeratin and PI-1 were used to mark uterine epithelium and the outermost layer trophoblast of fetus separately from D5-D7 of pregnancy, and the fluorescent results indicated developmental retardation of a few embryos in KO uterus. (B) HE staining of D8 implantation sites of uteri, and the embryos had stopped growth and been absorbed. Scale bars, 50  $\mu$ m. Le, luminal epithelium; the arrow indicted the persistent epithelium in the implantation chamber surrounding the embryos.



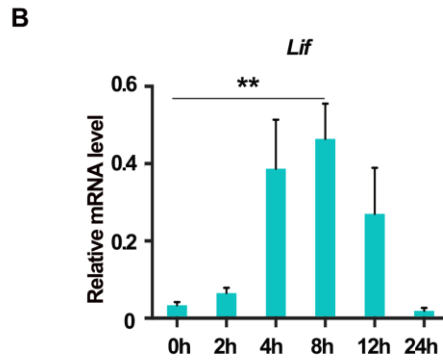
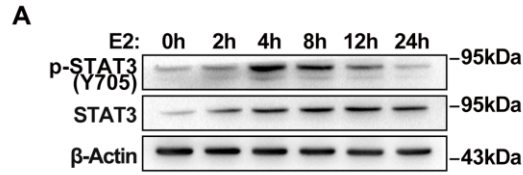
**Supplementary Figure S2. Uterine epithelial-specific deletion of IGF1R impaired embryo implantation.** (A and B) The average number of implantation sites (IS) showed an apparent decrease in *Igf1r<sup>f/f</sup>/Ltf-cre* mice on D5. (C) Immunohistochemical analysis of Cox2 protein in *Igf1r<sup>f/f</sup>* and *Igf1r<sup>f/f</sup>/Ltf-cre* uteri with a blastocyst on D5 morning. scale bars, 100  $\mu$ m. (D) Immunohistochemical analysis of pStat3 (Y705) in uteri with a blastocyst on D5 morning. Scale bars, 50  $\mu$ m.



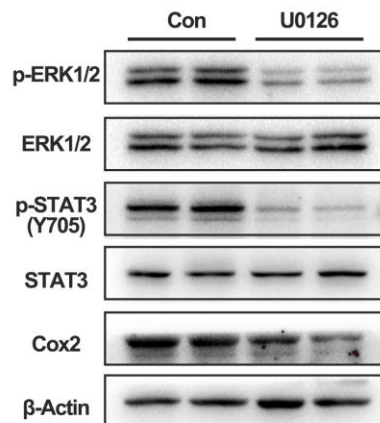
**Supplementary Figure S3. Hormone levels are comparable between *Igf1r<sup>f/f</sup>* and *Igf1r<sup>d/d</sup>* mice on D4. (A)** Immunohistochemical staining of 3 $\beta$ -HSDII and P450scc in D4 *Igf1r<sup>f/f</sup>* and *Igf1r<sup>d/d</sup>* ovaries. CL, corpus luteum; F, follicle. Scale bars, 200  $\mu$ m. **(B)** Serum levels of E<sub>2</sub> and P<sub>4</sub> in *Igf1r<sup>f/f</sup>* and *Igf1r<sup>d/d</sup>* mice on D4 of pregnancy.



**Supplementary Figure S4. The expression of LIF was comparable between *Igf1r<sup>f/f</sup>* and *Igf1r<sup>d/d</sup>* uteri. (A)** The *Lif* mRNA level in *Igf1r<sup>f/f</sup>* and *Igf1r<sup>d/d</sup>* uteri from D4 to D5 as detected by quantitative real-time PCR. The values are normalized to the *Gapdh* expression level and indicated as the mean  $\pm$  SEM, n=3. \*,  $P < 0.05$ . **(B)** *In situ* hybridization of *Lif* in *Igf1r<sup>f/f</sup>* and *Igf1r<sup>d/d</sup>* uteri on D4 and D4.5. Ge, glandular epithelium; Bl, blastocyst; S, stroma. Scale bars, 50  $\mu$ m.



**Supplementary Figure S5. E<sub>2</sub> increased the levels of STAT3 activation and *Lif* expression.** (A) The expression level of p-STAT3 (Y705) and STAT3 proteins in D4 WT mouse uteri in response to E<sub>2</sub> treatment. Actin served as the loading control. (B) Quantitative real-time PCR analysis of *Lif* mRNA expression in WT mouse uteri in response to E<sub>2</sub> treatment. The values are normalized to the *gapdh* expression level and indicated as the mean ± SEM. n=3.



**Supplementary Figure S6. U0126 treatment reduced ERK and STAT3 activation.** Western blot analysis for the indicated protein in D4 uteri. The uteri were treated with IGF2 recombinase protein, combined with the control or U0126 injection for 6 hours. Actin served as the loading control.

**Supplementary Table S1. Quantitative real-time PCR primers.**

<b>Primer name</b>	<b>Sequence</b>
Msx1 F	CTTCCTCCTGGTTGTCGCT
Msx1 R	CTCTTGGCCTCTGCACCCTTAGTTT
Lif F	GAGAGATTTCTGTCTCACTC
Lif R	CTTCCAAAGTTTCTCTGAGA
CLDN1 F	CTGGTAGAGGTAATGTGAGT
CLDN1 R	GAGATACAGAACAGTTGAAGG
CLDN3 F	CCAGGAGAGGAGCCGTTAAG
CLDN3 R	GCTGGACCTGGGAATCAACT
CLDN4 F	CCACTCTGTCCACATTGCCT
CLDN4 R	CTTTGCACAGTCCGGGTTTG
CLDN7 F	GCTATGACTGGAGGCATT
CLDN7 R	GTGACAATCTGATGACCAATC
CLDN8 F	GCCCTCTACATAGGCTGGAC
CLDN8 R	GAAACTCCGTTGAGTGGTGC
CLDN15 F	CGTGGGCAACATGGATCTCT
CLDN15 R	CCACGAGATAGCCACCATCC
Occludin F	TGCTGCTGATGAATATAATAGAC
Occludin R	ATCCTCTTGATGTGCGATAA
Igf1r F	GCAGACACTACTACTACAAAG
Igf1r R	ACTCATCGTCGTGGATAA
Igf1 F	CGCTCTGCTTGCTCACCTTAC
Igf1 R	AATGCTGGAGCCATAGCCTGTG
Cdh1 F	GAGACCAGTTTCCTCGTCCG
Cdh1 R	AGCAGCTCTGGGTTGGATTC
Ctnna1 F	ACGGTCTGGAGAAGGAAGGT
Ctnna1 R	ACACGAGCCCGAATAAAGCA
Ctnnb1 F	AGGATGATCCCAGCTACCGT
Ctnnb1 R	AGATCAGGCAGCCCATCAAC
Ptgs2 F	GTCTGGTGCCTGGTCTGATGAT
Ptgs2 R	GTGGTAACCGCTCAGGTGTTG
Bmp2 F	GATCTGTACCGCAGGCACTCA
Bmp2 R	AGTTCCTCCACGGCTTCTTCG

Igf2 F	TGCTGCATCGCTGCTTACGG
Igf2 R	GACGGTTGGCACGGCTTGAA
Esr1 F	TGCCTCTGGCTACCATTAT
Esr1 R	TGCCCACTTCGTAACACTT
Pgr F	ACCTGATCTAATCCTAAATGA
Pgr R	ATTGTGTTAAGAAGTAGTAAGAC
Ihh F	CATCTTCAAGGACGAGGAGAACA
Ihh R	CATGACAGAGATGGCCAGTGA
Nr2f2 F	CTTTGGAAGAGTACGTTAGG
Nr2f2 R	AACAATTGCTCTATGACTGA
Hand2 F	TCGGTTATCTAGTGCTGTC
Hand2 R	ATACTTACAATGTTTACACCTTCA
Hoxa10 F	GGCAGTTCCAAAGGCGAAAA
Hoxa10 R	CAAAAAAAGCCAGAACAAC
Wnt7b F	TGAGGCGGGCAGAAAGG
Wnt7b R	CCTGACACACCGTGACACTTACA
Muc1 F	ATTGTGTTAAGAAGTAGTAAGAC
Muc1 R	AAGTGGTCACCACAGCTGGG
Coch F	GTGCAGCAAAACCTGCTACAA
Coch R	AGCTAGGACGTTCTCTTTGGT
GAPDH F	ATGGTGAAGGTCGGTGTGA
GAPDH R	TGAGTGGAGTCATACTGGAACAT

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**Supplementary Table S2. Primers for vector construct.**

<b>Primer name</b>	<b>Sequence</b>
STAT3-pBiFC-VC155-F	GCCGAATTCATGGCTCAGTGGAAACCAGCTG
STAT3-pBiFC-VC155-R	GAAGGTACCTCACATGGGGGAGGTAGCAC
ERK1- pBiFC-VN173-F	GAAGTCGACATGGCGGCGGCGGCGG
ERK1- pBiFC-VN173-R	GCCGGTACCTTAAGATCTGTATCCTGGCTGGAATCTAG
STAT3-Zsgreen-F	GCCGAATTCATGGCTCAGTGGAAACCAGCTG
STAT3-Zsgreen-R	GAAGGTACCTCACATGGGGGAGGTAGCAC
Cox2-P1-Luciferase-F	GGGGGTACCTTCGCTACTCCATCCTCACACC
Cox2-P1-Luciferase-R	GGGAAGCTTGCTCCACTTCATCGGAATGCTA
Cox2-P2-Luciferase-F	GGGGGTACCCGCAGACTCAGCGAAC
Cox2-P2-Luciferase-R	GGGAAGCTTTTTCCGCTTAGGCTTTCC
Cox2-P3-Luciferase-F	GGGGGTACCCACCAGTACAGATGTGGACCCT
Cox2-P3-Luciferase-R	GGGAAGCTTGCTCAAGAGTGTCACAGCTTCC

**Supplementary Table S3. Primers for *in situ* DIG-labelled probes.**

<b>Primer name</b>	<b>Sequence</b>
Igf1r-DIG-F	GATTGATCCTAATGTATG
Igf1r-DIG-R	TTATTTCTCTTTCTATGG
Msx1-DIG-F	CCTGACTTAGGTGGGTCCAG
Msx1-DIG-R	GTCCTTTTGGCCTCTGGTCT
Igf1-DIG-F	ACAATAATAAGTCCAATAACAT
Igf1-DIG-R	GAAGAGGTGAAGATAAGG
Ptgs2 (Cox2)-DIG-F	ACTCTGCTCCGAAGAATCTC
Ptgs2 (Cox2)-DIG-R	ACATCCCTGAGAACCTGCAG
Igf2-DIG-F	ATGGGGATCCCAGTGGGGAA
Igf2-DIG-R	GTCCAGCAACCATCAGGAAT

**Supplementary Table S4. ChIP-PCR primers.**

<b>Primer name</b>	<b>Sequence</b>
STAT3-ChIP-P1-F	TTCGCTACTCCATCCTCACACC
STAT3-ChIP-P1-R	GCTCCACTTCATCGGAATGCTA
STAT3-ChIP-P2-F	CGCAGACTCAGCGAACCA
STAT3-ChIP-P2-R	TTTCCGCTTAGGCTTTCC
STAT3-ChIP-P3-F	CACCAGTACAGATGTGGACCCT
STAT3-ChIP-P3-R	GCTCAAGAGTGTCACAGCTTCC