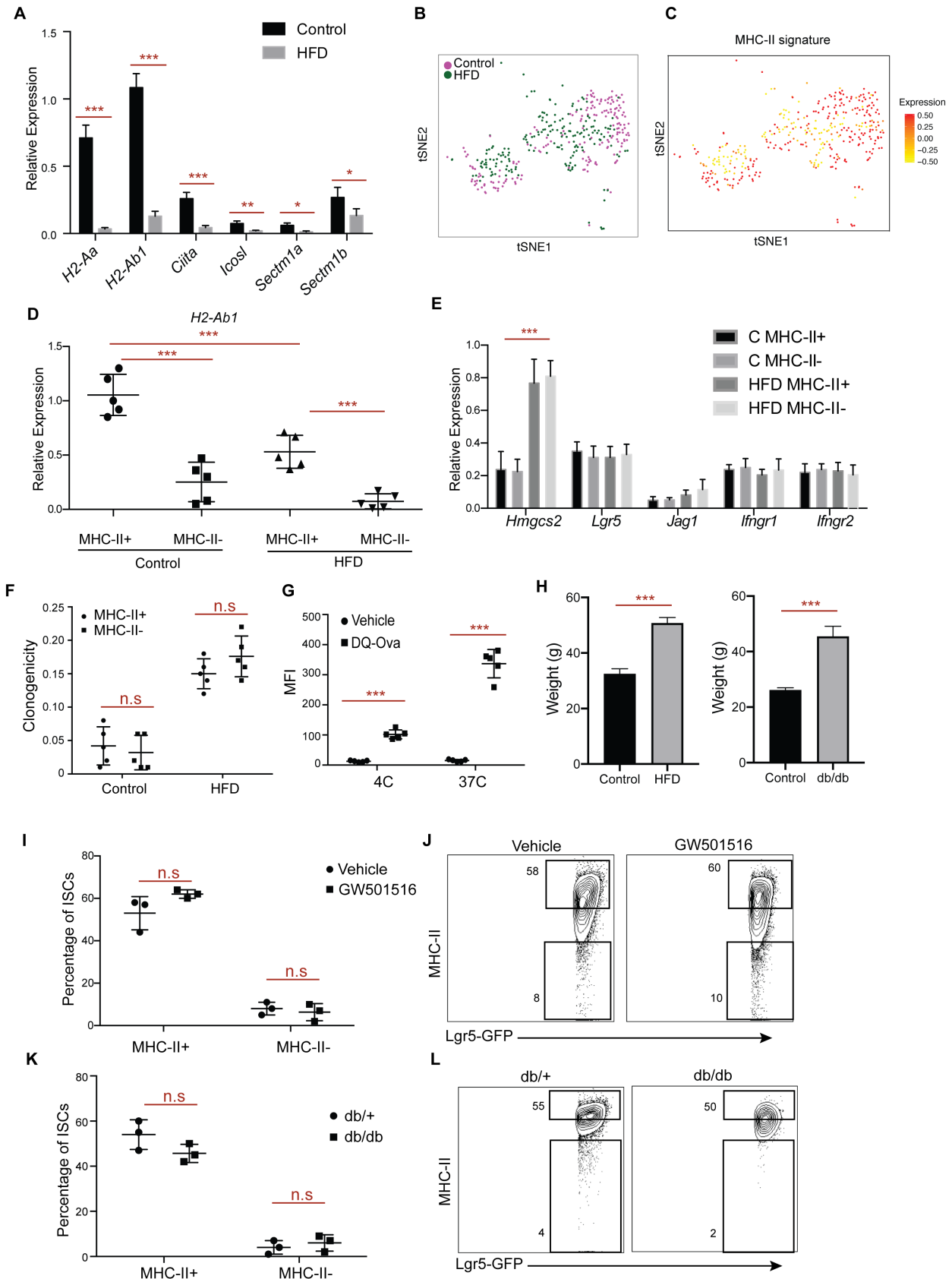


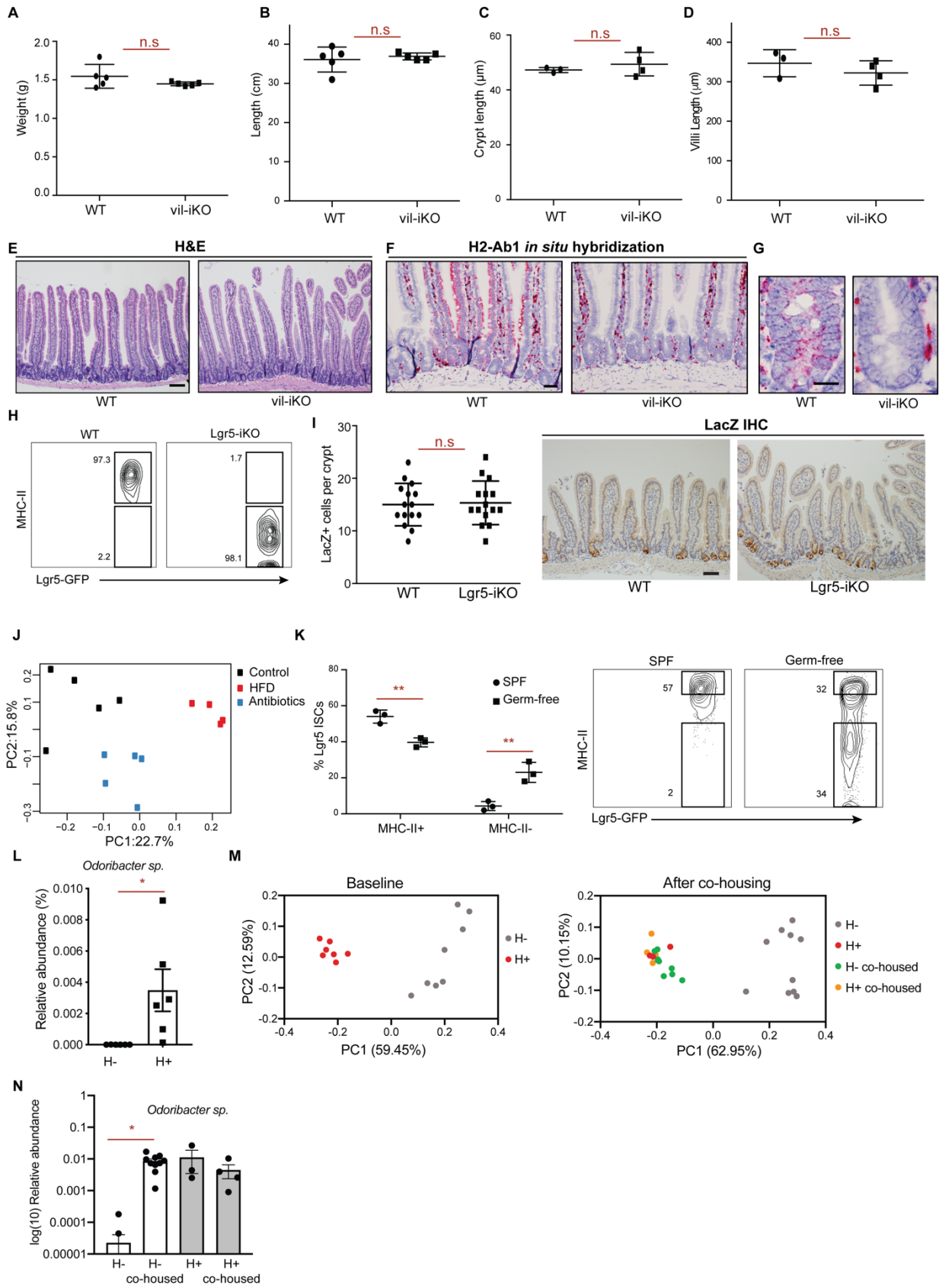
Figure S1



Supplementary Figure 1. Characterization of MHC-II expression in ISCs, related to Figure 1.

- A.** Relative expression of immunomodulatory genes in control and HFD Lgr5+ ISCs ($n=5$).
- B.** *t*-Distributed stochastic neighbour embedding (tSNE) analysis of single Lgr5+ ISCs isolated from control ($n=171$ cells, 2 independent experiments) or HFD mice ($n=144$ cells, 2 independent experiments).
- C.** tSNE analysis of single cells using MHC-II pathway signature genes (Supplementary Table 1)
- D.** Relative expression of MHC-II (*H2-Ab1*) in control and HFD MHC-II+ and MHC-II- Lgr5+ ISCs ($n=5$).
- E.** Relative expression of signature PPAR- δ target (*Hmgcs2*), stem cell marker (*Lgr5*), PPAR- δ - dependent B-catenin target (*Jag1*) and IFNGR genes in control and HFD MHC-II+ and MHC-II- Lgr5+ ISCs ($n=5$).
- F.** Organoid-initiating capacity of MHC-II+ and MHC-II- Lgr5+ ISCs from HFD mice ($n=5$).
- G.** Mean fluorescence intensity (MFI) of ISCs pulsed with either vehicle or DQ-Ovalbumin for 8 hours at 4C and 37C ($n=5$, 3 technical replicates per experiment).
- H.** Weight of mice used for diet-induced obesity (left: Control and HFD, $n=15$) and leptin receptor deficiency models of obesity (right, Control and db/db, $n=7$) in the study.
- I, J.** Frequency of MHC-II+ and MHC-II- Lgr5+ ISCs in vehicle- and PPAR- δ agonist GW501516-treated mice by flow cytometry (**I**, $n=3$). Representative flow cytometry plots of MHC-II in vehicle- and PPAR- δ agonist GW501516-treated Lgr5+ ISCs (**J**).
- K, L.** Frequency of MHC-II+ and MHC-II- Lgr5+ ISCs in lean db/+ and obese db/db mice (**K**, $n=3$). Representative flow cytometry plots of MHC-II+ and MHC-II- Lgr5+ ISCs (**L**, $n=3$).
- Unless otherwise indicated, data are mean \pm s.d. from n independent experiments; n.s.: not significant, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ (Student's *t*-tests).
- Related to Figure 1.

Figure S2



Supplementary Figure 2. Intestine-specific deletion of MHC-II does not alter intestinal physiology and microbiome regulates epithelial MHC-II expression, related to Figures 2, 3 and 4.

A-E, Intestinal weight (**A**, $n=5$), length (**B**, $n=5$), crypt (**C**, $n=5$), and villi length (**D**, $n=5$) of MHC-II wild type (WT) and MHC-II^{L/L}; Villin-CreERT2 (vil-iKO) mice one month after tamoxifen injection ($n=$ mice, mean \pm s.d.). Representative H&Es of WT and vil-iKO small intestine (**E**).

F, G, *In situ* hybridization for *H2-Ab1* in the intestine (**F**) and representative images of intestinal crypts (**G**) in WT and vil-iKO mice ($n=3$).

H, Frequencies of MHC-II⁺ and MHC-II- Lgr5⁺ ISCs in WT and MHC^{L/L}; Lgr5-CreERT2 (Lgr5-iKO) by flow cytometry ($n=5$ mice).

I, Lineage tracing of LacZ⁺ cells in the intestinal crypt after deletion of MHC-II in ISCs (left). Representative images of LacZ immunostain in WT and Lgr5-iKO small intestine (right) ($n=5$ mice).

J, Principal coordinate analysis (PCoA) of microbial composition in feces of mice fed control diet, HFD, and mice treated with antibiotics ($n=5$ mice).

K, Frequencies of MHC-II⁺ and MHC-II- Lgr5⁺ ISCs in specific-pathogen free (SPF) and germ-free mice by flow cytometry ($n=3$, mean \pm s.d.). Representative flow cytometry plots of MHC-II in SPF and germ-free ISCs.

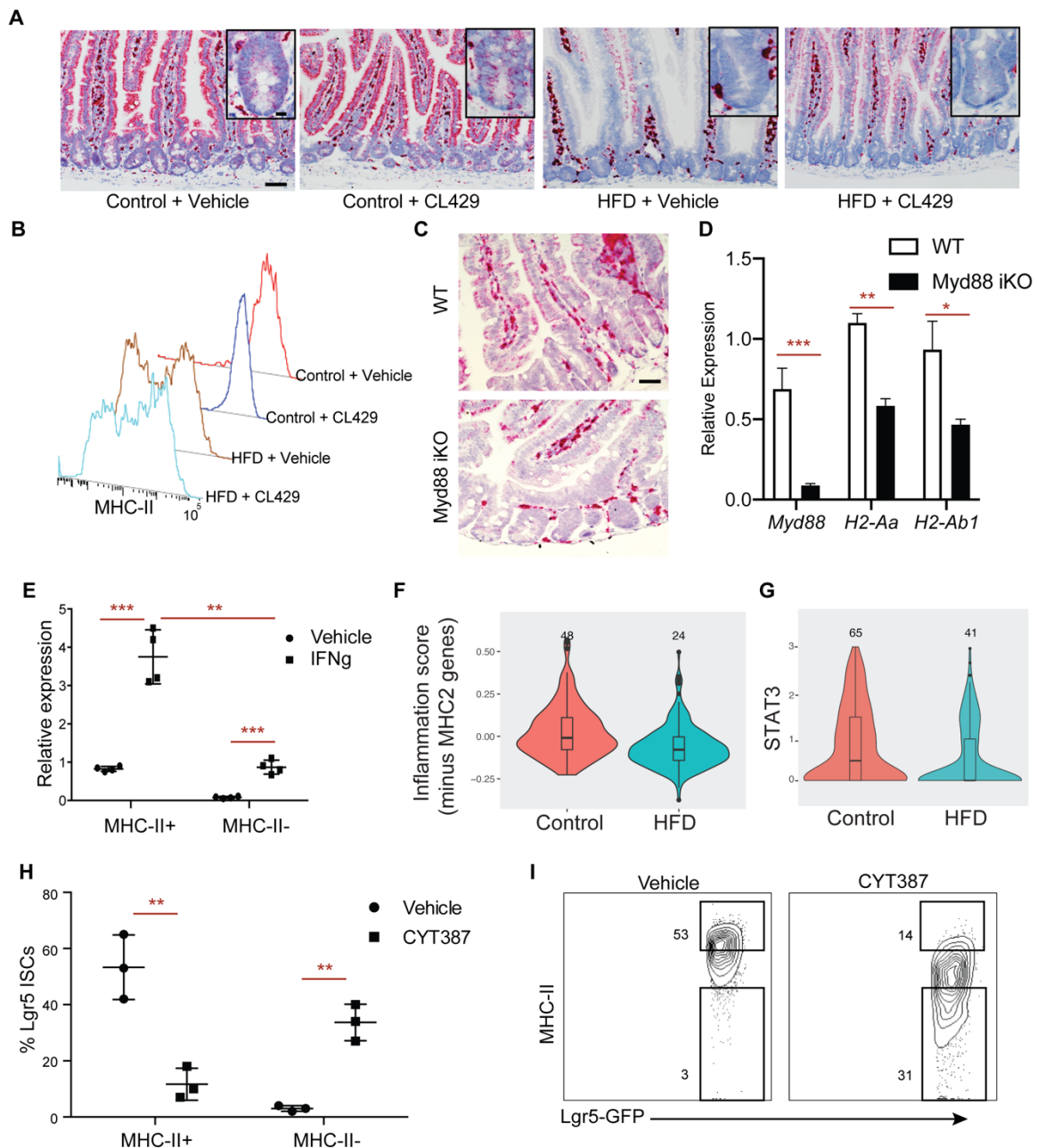
L, Relative abundance of *Odoribacter sp.* in mice housed in H- room or H+ room ($n=6$, mean \pm s.e.m).

M, PCoA of microbial composition in feces at baseline level before co-housing experiment (left) and 10 days after co-housing H- mice with H+ mice in the H+ room.

N, Relative abundance of *Odoribacter sp.* in feces of mice housed either in H- room ($n=10$), H+ room ($n=3$), or after co-housing H- mice ($n=10$) with H+ mice ($n=4$) in H+ room (mean \pm s.e.m).

n.s.: not significant, * $P < 0.05$, ** $P < 0.01$ (Student's *t*-tests). Scale bars, 100 μ m (**E**), 50 μ m (**F, I**) and 20 μ m (**G**).

Figure S3



Supplementary Figure 3. Regulation of MHC-II expression in ISCs by PRR and IFN γ signaling, related to Figure 5.

A, B. *In situ* hybridization for H2-Ab1 in control and HFD mice treated with vehicle- and CL429-treated mice in proximal small intestine (**A**, $n=3$). Representative histogram plots of MHC-II in Lgr5⁺ ISCs in control and HFD mice treated with vehicle and CL429 (**B**, $n=3$).

C. *In situ* hybridization for H2-Ab1 in WT and Myd88 KO mice ($n=4$).

D. Relative expression of *Myd88* and MHC-II genes (*H2-Aa* and *H2-Ab1*) in the intestine from WT and Myd88 KO mice ($n=5$, mean \pm s.d.).

E. Relative expression of MHC-II (*H2-Ab1*) in MHC-II⁺ and MHC-II⁻ Lgr5⁺ ISCs isolated from HFD mice with or without IFN γ stimulation ($n=4$, mean \pm s.d.).

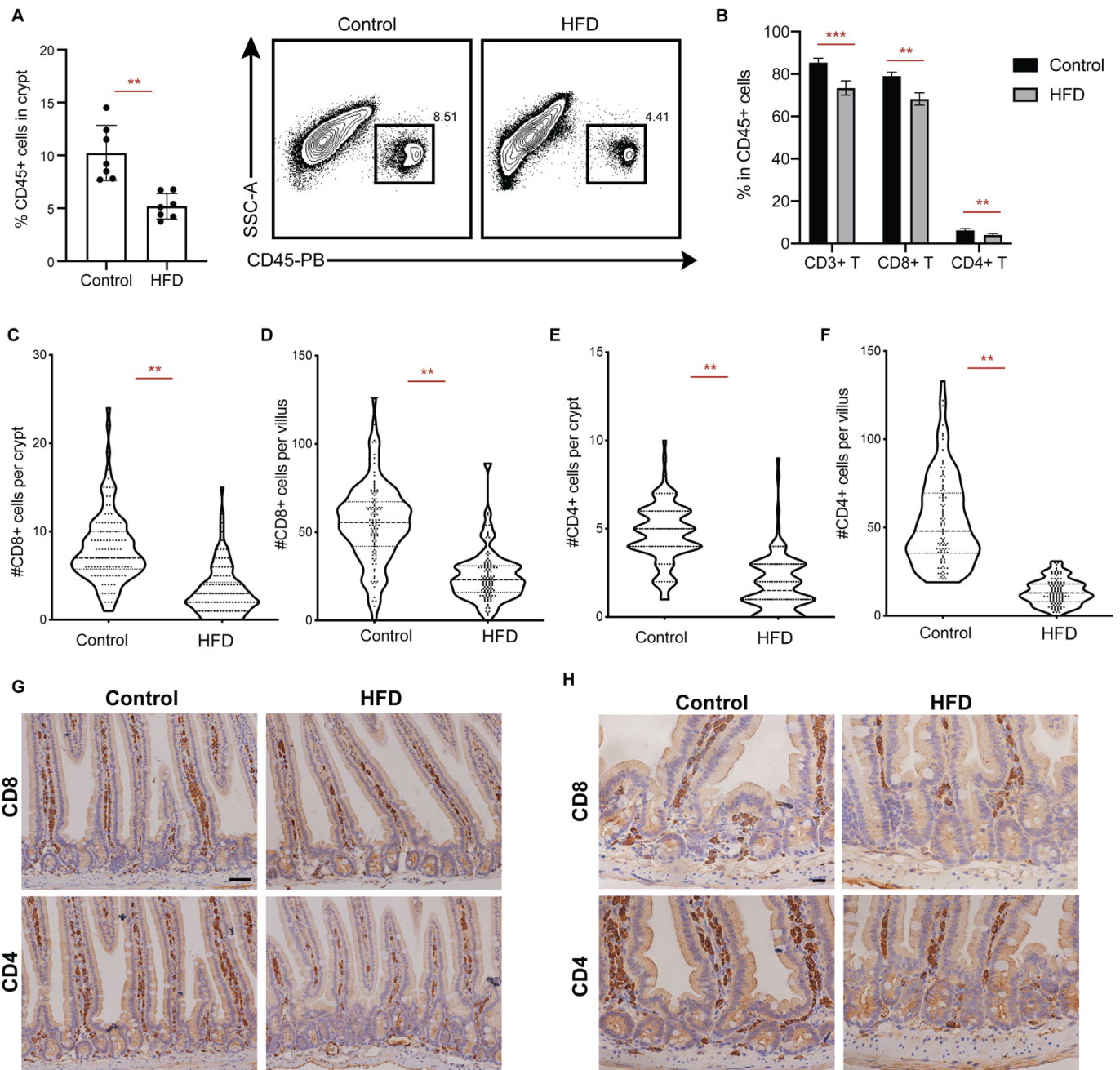
F. Violin plots demonstrating the levels of IFN γ -induced genes excluding MHC-II pathway genes in control and HFD Lgr5⁺ ISCs by scRNA-seq.

G. Violin plots demonstrating the levels of STAT3 in control and HFD Lgr5+ ISCs by scRNA-seq.

H, I. Frequencies of MHC-II+ and MHC-II- Lgr5+ ISCs in vehicle- and Jak/Stat inhibitor CYT387-treated mice ($n=3$). Representative flow cytometry plots of MHC-II in vehicle- and CYT387-treated Lgr5+ ISCs (**I**).

Unless otherwise indicated, data are from n independent experiments; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ (Student's t -tests). Scale bars, 50 μm (**A, C**) and 20 μm (insets, **A**).

Figure S4



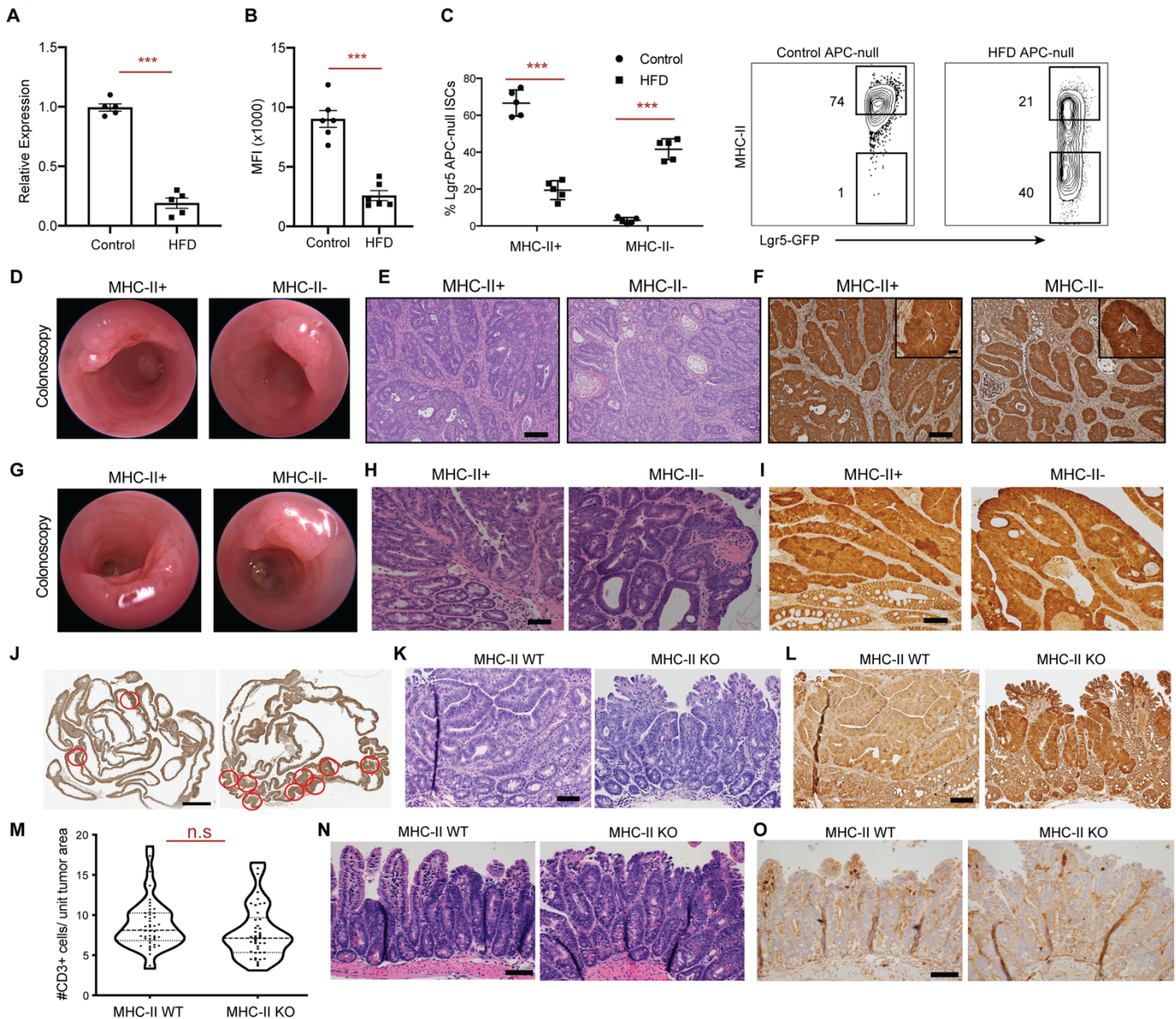
Supplementary Figure 4. The effect of HFD on intestinal immune cells, related to Figure 5.

A. Frequency of CD45+ cells in intestinal crypts isolated from control or HFD mice ($n=7$).

B. Frequency of CD3+ T cells, CD8+ T cells and CD4+ T cells among CD45+ cells in intestinal crypts isolated from control or HFD mice ($n=5$).

C-H. Numbers of CD8+ cells (**C**, crypt; **D**, villus) and CD4+ cells (**E**, crypt; **F**, villus) in the intestines of control or HFD mice. Representative images of CD4 and CD8 immunostaining in control or HFD intestines (**G**, **H**) ($n=5$). $**P < 0.01$, $***P < 0.001$ (Student's t -tests). Scale bars, 50 μm (**G**) and 20 μm (**H**).

Figure S5



Supplementary Figure 5. The effect of Lgr5+ ISC MHC-II expression on tumor formation, related to Figure 6.

A. Relative expression of MHC-II (*H2-Ab1*) in Lgr5+ *Apc*-null pre-malignant ISCs isolated from control or HFD mice 3 days post tamoxifen injection ($n=5$).

B. Mean fluorescence intensity (MFI) of MHC-II in Lgr5+ *Apc*-null pre-malignant ISCs isolated from control or HFD mice 3 days post tamoxifen injection ($n=6$).

C. Frequency of MHC-II+ and MHC-II- Lgr5+ *Apc*-null ISCs in control and HFD mice by flow cytometry ($n=5$). Representative flow cytometry plots of MHC-II in HFD *Apc*-null ISCs.

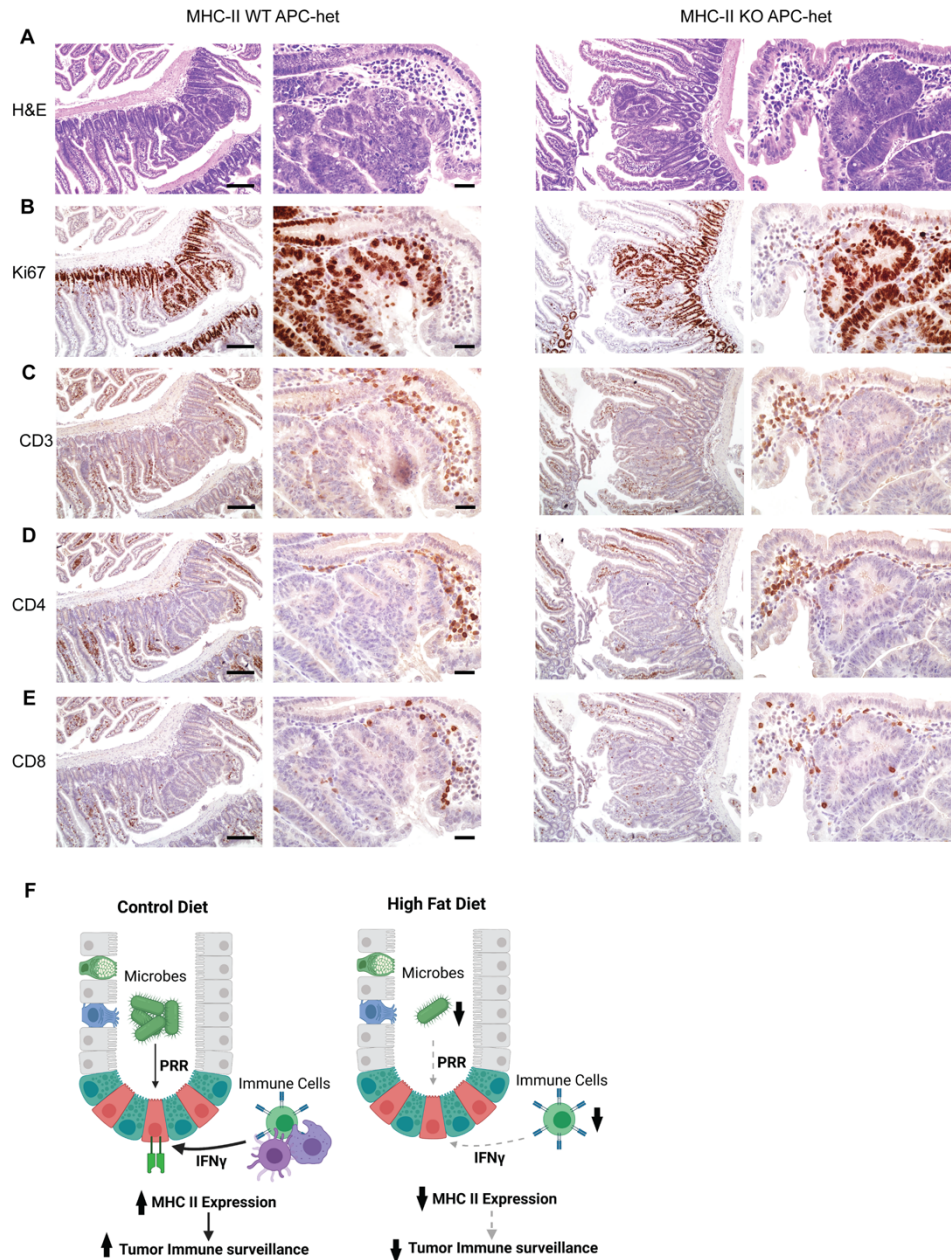
D-F. Characterization of orthotopically transplanted MHC-II+ and MHC-II- *Apc*-null Lgr5+ ISC-derived tumors three months after transplantation into immunocompetent syngeneic hosts. Optical colonoscopy images (**D**), Hematoxylin and eosin (**E**) and beta-catenin immunostain (**F**) of tumors.

G-I. Characterization of orthotopically transplanted MHC-II+ and MHC-II- *Apc*-null Lgr5+ ISC-derived tumors three months after transplantation into immunodeficient Rag2-KO hosts. Optical colonoscopy images (**G**), H&E (**H**) and beta-catenin immunostain (**I**) of tumors.

J-L. Tumors in Lgr5-CreERT2 MHC-II^{L/+}, APC^{L/+} ($n=9$, MHC-II WT APC-het) and Lgr5-CreERT2 MHC-II^{L/L}, APC^{L/+} ($n=9$, MHC-II KO APC-het). Representative images of tumors that arise in MHC-II WT (**J**, left) and MHC-II KO (**J**, right) mice. Representative H&E (**K**) and beta-catenin immunostain (**L**) images from tumors in MHC-II WT and MHC-II KO mice.

M-O. Numbers of CD3+ T cells per tumor area (**M**). Representative H&E (**N**) and CD3 immunostain (**O**) images from tumors ($n=5$). n.s.: not significant, $***P < 0.001$ (Student's *t*-tests). Scale bars, 100 μm (**E**, **F**, **H**, **I**, **K**, **L**, **N**, **O**) and 20 μm (insets, **F**).

Figure S6



Supplementary Figure 6. Immune cell profile of tumors that arise upon intestine specific deletion of MHC-II and *Apc*, related to Figure 6.

A-E. Representative H&E (A), Ki67 (B), CD3 (C), CD4 (D) and CD8 (E) immunostain images of tumors in *Lgr5-CreERT2 MHC-II^{L/+}, APC^{L/+}* (MHC-II WT APC-het) and *Lgr5-CreERT2 MHC-II^{L/L}, APC^{L/+}* (MHC-II KO APC-het) mice. Scale bars, 500 μ m (left, A – E), 100 μ m (right, A – E).

F. Model of dietary regulation of intestinal tumor initiation through perturbing microbe – stem cell – immune cell crosstalk. Intestinal microbiome, Pattern Recognition Receptor (PRR) and IFN γ signaling regulate epithelial MHC-II expression and tumor immune surveillance in control diet fed mice (left). A HFD leads to microbial dysbiosis and dampened MHC-II expression that results in impaired tumor immune surveillance and increased tumor initiation.

Supplementary Table S1: The list of genes defining the stem cell MHC-II signature. Related to Figure 1.

Stem Cell MHC-II Signature
<i>H2-Ab1</i>
<i>H2-DMb1</i>
<i>H2-DMA</i>
<i>H2-Aa</i>
<i>CD74</i>
<i>Ciita</i>
<i>H2-Eb1</i>
<i>Ceacam10</i>
<i>Cd177</i>
<i>Cd320</i>

Supplementary Table S2: The list of microbes that colonize mice in H+ room. Related to Figure 4.

	H- Room	H+ Room
<i>Helicobacter spp.</i>	-	+
<i>Helicobacter mastomyrinus</i>	-	+
<i>Helicobacter typhlonius</i>	-	+

Supplementary Table S3: The list of qRT-PCR primers. Related to the STAR Methods.

TLR1(F): TGAGGGTCCTGATAATGTCCTAC
TLR1(R): AGAGGTCCAAATGCTTGAGGC
TLR2(F): GCAAACGCTGTTCTGCTCAG
TLR2(R): AGGCGTCTCCCTCTATTGTATT
TLR3(F): GTGAGATACAACGTAGCTGACTG
TLR3(R): TCCTGCATCCAAGATAGCAAGT
TLR4(F): ATGGCATGGCTTACACCACC
TLR4(R): GAGGCCAATTTTGTCTCCACA
TLR5(F): GCAGGATCATGGCATGTCAAC
TLR5(R): ATCTGGGTGAGGTTACAGCCT
TLR6(F): TGAGCCAAGACAGAAAACCCA
TLR6(R): GGGACATGAGTAAGGTTCTGTT
TLR7(F): ATGTGGACACGGAAGAGACAA
TLR7(R): GGTAAGGGTAAGATTGGTGGTG
TLR8(F): GAAAACATGCCCCCTCAGTCA
TLR8(R): CGTCACAAGGATAGCTTCTGGAA
TLR9(F): ATGGTTCTCCGTCGAAGGACT
TLR9(R): GAGGCTTCAGCTCACAGGG
Nod1(F): GAAGGCACCCCATTGGGTT
Nod1(R): AATCTCTGCATCTTCGGCTGA
Nod2(F): CAGGTCTCCGAGAGGGTACTG
Nod2(R): GCTACGGATGAGCCAAATGAAG
Sectm1a (F): TCAGTGCCTGCTATCCCTACC
Sectm1a (R): GGGGCTTTTTATCGAAGATGGT
Sectm1b (F): AGCCTCCCTGAATGCCTATAA
Sectm1b (R): ACGTCTCTGAGATTGTTGGAGAT
Icosl (F): TAAAGTGTCCCTGTTTTGTGTCC
Icosl (R): ATTGCACCGACTTCAGTCTCT
Ifngr1 (F): CTGGCAGGATGATTCTGCTGG

Ifngr1 (R): GCATACGACAGGGTTCAAGTTAT
Ifngr2 (F): TCCTCGCCAGACTCGTTTTTC
Ifngr2 (R): GTCTTGGGTCATTGCTGGAAG
Hmgcs2 (F): ATACCACCAACGCCTGTTATGG
Hmgcs2 (R): CAATGTCACCACAGACCACCAG
H2-Ab1 (F): AGCCCCATCACTGTGGAGT
H2-Ab1 (R): GATGCCGCTCAACATCTTGC
H2-Aa (F): TCAGTCGCAGACGGTGTTTAT
H2-Aa (R): GGGGGCTGGAATCTCAGGT
Jag1 (F): CCTCGGGTCAGTTTGAGCTG
Jag1 (R): CCTTGAGGCACACTTTGAAGTA
Lgr5 (F): CCTACTCGAAGACTTACCCAGT
Lgr5 (R): GCATTGGGGTGAATGATAGCA