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A framework for the investigation of rare genetic disorders in neuropsychiatry

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Supplementary Information for

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Materials and Methods

Estimating the effect size of copy number variants (CNVs)

Large samples of cases and controls provide an opportunity to estimate the effect size on specific neuropsychiatric symptoms of some of the more common rare genetic disorders. We extending the approach described previously¹, to estimate the frequency of these variants in defined cohorts systematically assessed for neurodevelopmental delay^{2,3}, autism spectrum disorder^{4,5}, and schizophrenia^{2,6,7} and in general population controls from UK Biobank⁸. These estimates of frequency in cases and controls were compared to estimate odds ratios for each neuropsychiatric diagnosis at each CNV locus. These numbers are shown in Table S2 and plotted as a polar plot in Fig. 2 of the main text. For highly penetrant CNVs we lack sufficient population controls to accurately define the odds ratio, therefore we limited the plot to a maximum odds ratio of ≥30.

Estimating the effect size of single gene rare genetic disorders (RGDs)

On average, the population frequency of a specific single gene RGD is lower than the population frequency of the most well-defined CNV loci, due to the higher mutation rate of non-allelic homologous recombination (NAHR). Accordingly, larger populations of control samples will be needed to accurately define the effect size by an odds ratio. Furthermore, the completeness of reporting for specific neuropsychiatric domains varies (Fig. 2). As an estimate of the relative impact of each single gene RGD on different neuropsychiatric symptoms, we used the data systematically collected from the literature in the Developmental Brain Disorder Genes Database hosted at Geisinger (https://dbd.geisingeradmi.org) to quantify the relative frequency of different symptoms. The raw data are shown in Table S4 and plotted as a polar plot in Extended Data Fig. 1.

Estimating the number of *de novo* **PTVs for genome-wide significance for a given population size.**

The rate at which germline *de novo* mutations arise in each gene is highly correlated with gene size (cDNA) and sequence context, especially GC content^{9,10}. We used previously published estimates of per gene mutation rates for protein-truncating variants (PTVs) in 18,224 protein-coding genes based on exome sequencing of over 60,000 general population samples in ExAC10–12. To estimate the expected number of *de novo* PTVs in controls, we assessed the total number of *de novo* PTVs identified in 1,807 unaffected siblings from the Simons Simplex Collection, identifying 187 such mutations (187 / 1807 = 0.103). The PTV mutations rates and estimate of *de novo* PTV rate in controls were combined to estimate the expected number of *de novo* PTVs per individual per gene. Multiplying these values by a given population size provided the expected number of *de novo* PTVs per gene in a cohort; this was used as the Lambda value (λ) for a Poisson distribution. The probability of observing a specific number of *de novo* PTVs in the population was assessed using the cumulative density function (CDF) for a Poisson distribution with the estimated λ value. To reach genome-wide significance, this probability had to be lower than 2.7x10⁻⁶ based on a Bonferroni correction for all genes assessed (p=0.05/18,224). We also considered searching only the 1,528 genes with high "probability loss-of-function intolerance" scores (pLI ≥ 0.995), giving a p-value threshold for high-pLI significance of 3.3x10⁻⁵ with Bonferroni correction (p=0.05/1,528), however we would advocate for using the genome-wide significant threshold or a more conservative threshold.

Using this approach, we searched across a range of observed *de novo* PTVs (3, 5, 10, 20) and eight p-value thresholds $(3.3x10^{-5}$ to $2.7x10^{-12})$ to find the cohort size at which such at observation would result in a probability lower than the p-value threshold. To limit the compute required, we down-sampled to one in 15 genes. The results are shown in Extended Data Fig. 2.

In the main text (Fig. 3), we show a simplified version of this approach using a single p-value threshold $(2.7x10^{-6})$ and three cohort size (50,000, 100,000, and 200,000) to represent the effective cohort size of existing clinical exome sequencing (i.e. the total number of cases who could be reported in the literature). We note that these are estimates of the true number of individuals sequencing for a neuropsychiatric phenotype, and that the exact number is unknown, however current research cohorts include over 27,000 ASD cases between the Autism

Sequencing Consortium¹² and SPARK (https://sparkforautism.org), over 14,000 cases of severe developmental delay13 (https://decipher.sanger.ac.uk/ddd), and over 5,000 cases of schizophrenia14. This sets a lower bar for the true number of cases sequenced at 46,000; adding smaller research cohorts and clinical exome sequencing leads to an estimate of 100,000 effective cases with neuropsychiatric symptoms and exome sequencing that could contribute to case reports. We also note that GeneDx describes an exome- and genome-sequencing database of over 155,000 samples (https://www.genedx.com), though many of these are likely to be controls or parents.

Table S3. Attendee list of the "Rare Genetic Disease Workshop: Window into Genomic Risk and Resilience of Mental Disorders" held at the NIMH Sept. 2017.

NIMH RARE GENETIC DISEASES WORKSHOP

Table S5. Stakeholders in rare genetic disorders.

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