

Additional file 1

A polygenic risk score predicts mosaic loss of chromosome Y in circulating blood cells

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Additional Methods

LOY estimation from SNP-array data

Whole blood DNA samples collected from 6,140 male ASPREE participants were genotyped using the Axiom 2.0 Precision Medicine Diversity Research Array (PMDA) following standard protocols (ThermoFisher). We followed best practice genotyping and quality control (QC) protocols from Thermo Fisher, starting from raw intensity CEL files, we used a command line custom script designed for the the Axiom PMDA array, mapped to human genome reference GRCh38, to produce variant call files.

We performed sample level QC using plink version 1.9, excluding samples for gender discordance (80 samples mismatched and excluded) using plink default F statistics threshold (≤ 0.2 female and ≥ 0.8 male), relatedness (124 individuals excluded) using default PI-HAT threshold >0.025 to exclude one sample from each related pair.

To estimate population structure in the ASPREE cohort we performed principal component analysis (PCA) using The 1000 Genomes Project as a reference population [1]. Directly genotyped data from ASPREE and The 1000 Genomes Project 1K phase 3 (liftover to GRCh38) were merged and LD pruned ($r^2 < 0.1$) using plink version 1.9 [2] followed by R package SNPrelate [3]. We calculated the Z score for first 2 principal component eigenvectors and excluded samples with $\pm 2SD$ (standard deviation) of Z score compared to their respective five reference superpopulation groups from the 1000 Genomes Project that included: Europeans, South Asians, East Asians, African American (African super population) and Hispanics (Ad Mixed American) (Figure S5). The final dataset of 12,815 samples (6,140 males) from Caucasians (Non-Finish Europeans) were selected from the ASPREE cohort for further analysis.

From the Axiom genotyping dataset we generated Log R Ratio (LRR) calls from the allele specific signal intensity data for each marker using a custom pipeline from the CEL files. Following this, the genomewide experimental quality for each sample was assessed and 340 samples were excluded based on published criteria for sample inclusion [4]. We also exclude 669 saliva samples from the data set. After the quality controls based on sex, relatedness, ancestry, blood/saliva sample and genotyping quality; a total of 5,131 male samples were retained for LOY analysis.

Following this, the level of LOY mosaicism in the 5,131 samples passing strict QC was estimated by calculation of the mLRRY, as described previously [5]. First, the mLRRY was calculated for each sample as the median of the Log R Ratio (LRR) values of the 488 Axiom probes located within the male specific part of chromosome Y (MSY, chrY:2.787.139-22.318.450; GRCh38/hg38). We also calculated an mLRRX value (median LRR of probes positioned in the non-PAR part of chromosome X) for each sample, and observed a technical covariation with mLRRY. To adjust for this potential bias for the LOY assessment, an adjustment of mLRRY values was performed by a new approach, based on the coefficients of a linear model estimated between mLRRY and mLRRX in the unadjusted dataset (Figure S6). Following this, the mLRRY values over the entire dataset was adjusted using a constant defined as the peak of the unadjusted mLRRY distribution. This operation shifts the peak of the mLRRY distribution so that samples without LOY align around zero, thus improving comparability between datasets. The constant applied in the present dataset (i.e. 0.0006827) was calculated as the peak of the local regression median of the unadjusted mLRRY distribution using kernel estimation in the *density* function in R and the smoothing bandwidth method “SJ”. We next defined a threshold in the adjusted mLRRY distribution that could be used for scoring samples with or without LOY. Samples with adjusted mLRRY values lower than the 99% confidence limit in a simulated distribution of technical mLRRY variation was

scored with LOY (Figure S7). As a final step to improve interpretability of our results, we used a modified version of a published formula [6] to translate individual mLRRY values into percentage of cells with LOY in every sample, as described in Figure S8. Finally, the concordance between LOY estimation from SNP-array and whole genome sequencing data was evaluated and shown in Figure S4. The distribution of mosaic LOY in male blood samples observed in ASPREE study in relation to age is shown in Figures S1 and S2.

Whole genome Sequencing and LOY estimation

We performed Whole Genome Sequencing (WGS) of 2,795 ASPREE participants [7, 8]. Samples were WGS on the Illumina HiSeq X system with an average of 30x sequencing coverage as described previously [8]. The sequence alignment and processing were performed using reference 1000 Genomes Phase 3 decoyed version of build 37 of the human genome through Genome analysis tool kit (GATK) pipeline [9]. The joint genotype calling was performed in a single batch using GTAK GenotypeGVCFs function. To estimate the WGS level of LOY, we used ASPREE WGS samples that passed MGRB quality control criteria that includes call rate > 98%, depth standard deviation < 10, VAF standard deviation at loci called heterozygous < 1, hetero:homo variant ratio of < 2, inbred coefficient from X chromosome values between < 0.2 or > 0.8 and Singleton rate < 0.001 [8]. Total 947 males from ASPREE WGS cohort were also genotyped on AXIOM SNP array chip and used to check the LOY concordance across two platforms.

To estimate LOY using WGS data, first, the program Control-FREEC (version 11.5) [10] was used to calculate a read depth ratio in 50,000 bp genomic windows on the UPPMAX Bianca cluster. The default parameters was used for other Control-FREEC settings. A mappability gem file for hg19 (read length 100 bp and up to 2 mismatches) was used in combination with the ASPREE-reference genome. To get a per chromosome read depth ratio, the median was calculated for all windows on each chromosome. Next, samples with deviating chromosomal-

ratios was removed, i.e. suspected female samples (median X-ratio > 0.7), samples with suspected XYY aberration (median Y-ratio > 0.75) as well as all samples showing different types of abnormal autosomal read depth ratios (read depth standard deviation 1.5 IQR outside the third quartile).

LOY calling in the remaining 1137 male samples was performed from the ControlFREEC output. First, the *density* function in R with bandwidth set to “SJ”, was used to find the highest density Y-ratio point (using all Y chromosome window ratios) and the Scales package was used to linearly rescale the Y-ratios based on the density value. This resulted in a first estimate of LOY percentage in each sample between 0 and 100% cells with Y loss aneuploidy. To further improve these estimates, we identified normal samples (without LOY) as those showing a level of LOY within 2 standard deviations in the distribution. In these non-LOY samples, we noticed some highly variable Y-regions that were considered less informative for LOY calling. Specifically, 50.000 bp windows on chromosome Y with a standard deviation larger than 1.5 IQR from third quartile was considered highly variable and excluded. After this, the median Y-ratio was re-calculated for all male samples using only the Y-regions windows that passed this additional QC step. Thus, a final estimate of LOY percentage was achieved using the *density* function in R (bandwidth=“SJ”) and rescaled as described above in the first step. Finally, a binary threshold used for LOY scoring of samples was calculated following the same principle as described for the SNP array data. Finally, the concordance between LOY estimation from whole genome sequencing and SNP-array data was evaluated and shown in Figure S4.

Figure S1.

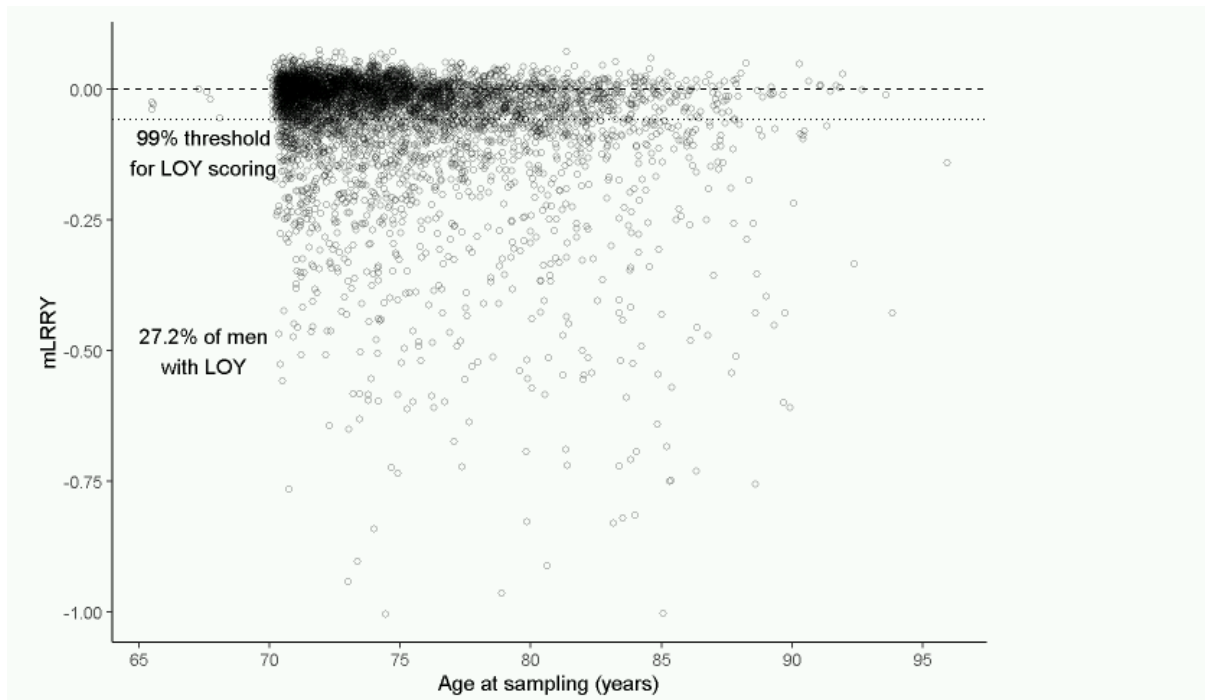


Figure S1. Observed level of LOY mosaicism in 5,131 ASPREE men, estimated from SNP array data and plotted in relation to age at blood sampling. The dotted line represents the threshold in the mLRRY distribution at the 99th percentile of experimental noise (mLRRY = -0.06) that was used for the scoring of individuals with or without LOY.

Figure S2.

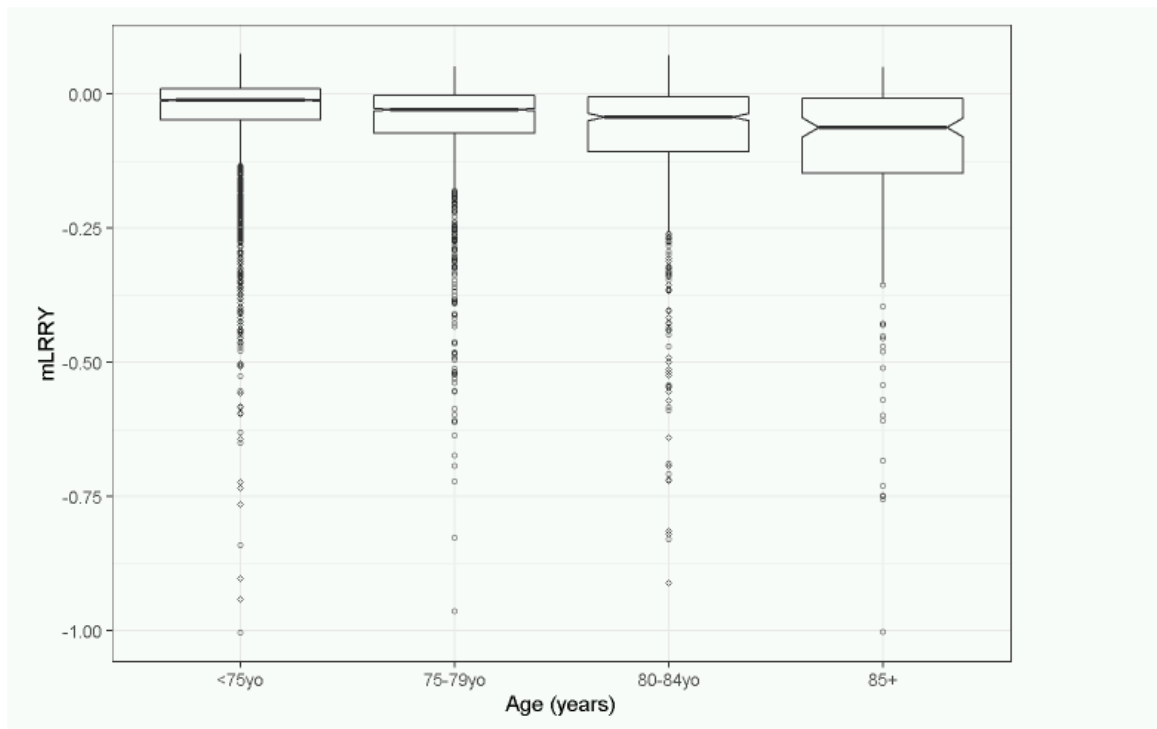
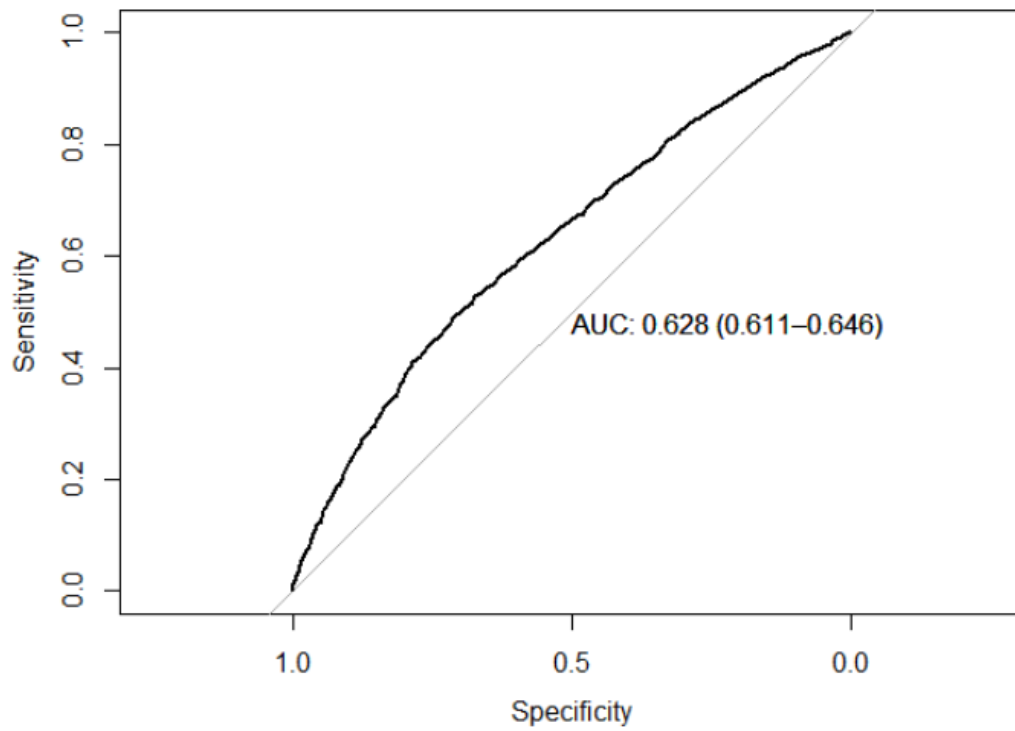


Figure S2. The frequency of mosaic LOY in blood samples of 5.131 ASPREE male participants in relation to age. The box and Whisker plot was generated in four age groups, the middle line in the box shows the median around the interquartile ranges and top and bottom vertical lines are 95% confidence intervals.

Figure S3.

a)



b)

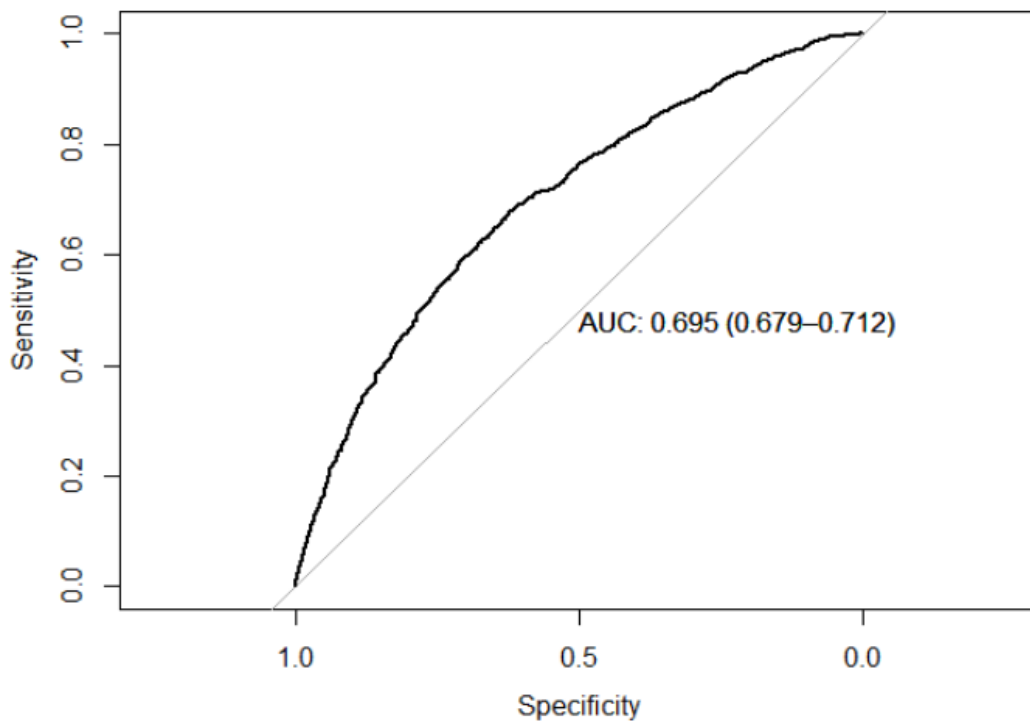


Figure S3. A) ROC curve for LOY for age, smoking and alcohol use. B) ROC curve for LOY for age, smoking alcohol use and PRS.

Figure S4.

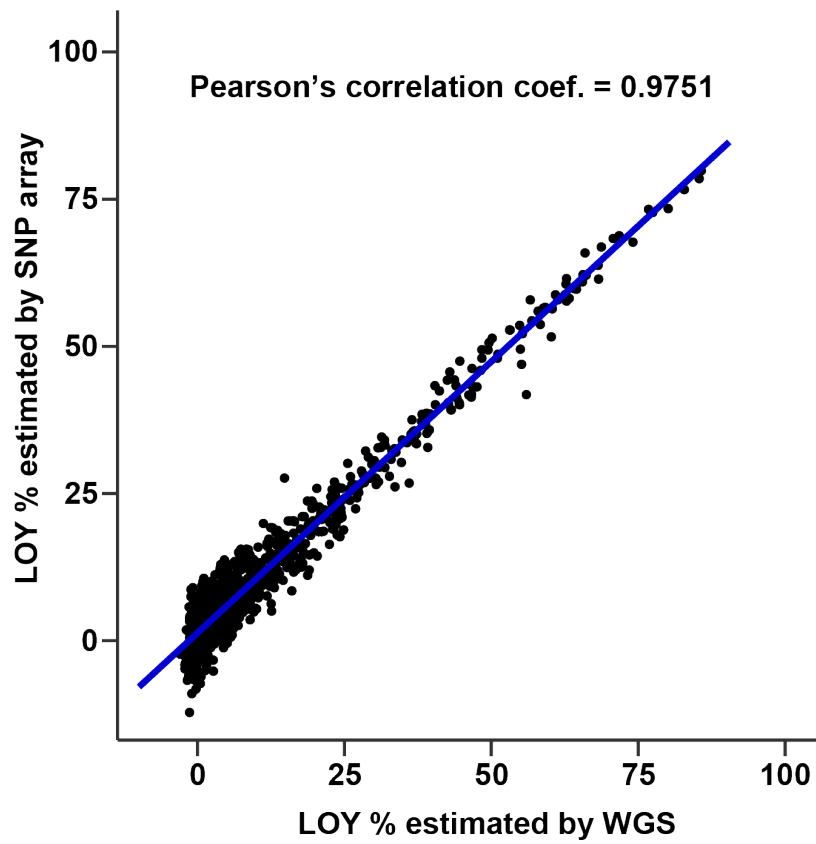


Figure S4. Correlation plot comparing microarray-derived LOY calls (Y axis) versus WGS-derived LOY calls (X-axis).

Figure S5.

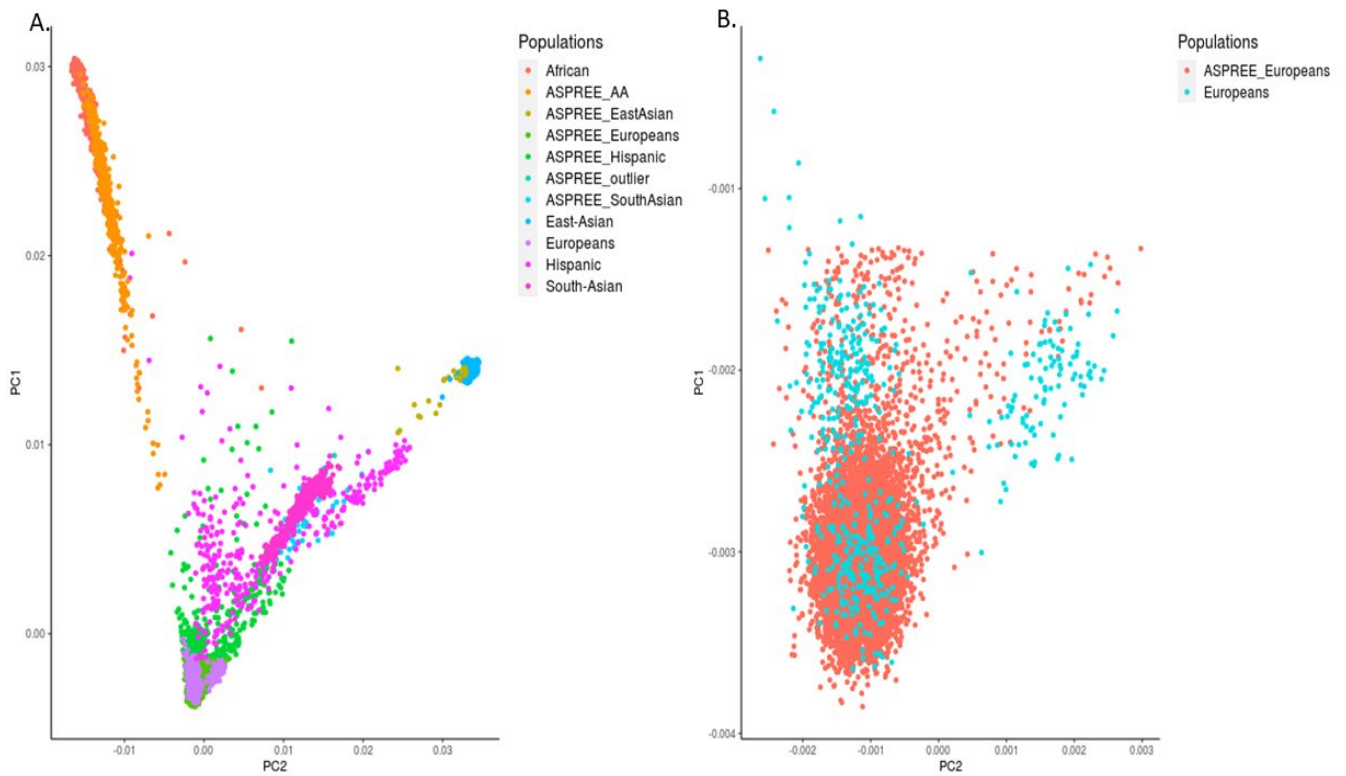


Figure S5. Principal component analysis (PCA) of the ASPREE cohort compared with the 1000 Genomes Project. A) PCA plot of all ASPREE participants projected onto 1000 Genome populations. B). PCA plot of ASPREE Europeans samples projected onto 1000 Genome Europeans samples that were included in this study. ASPREE_AA represents African American samples.

Figure S6.

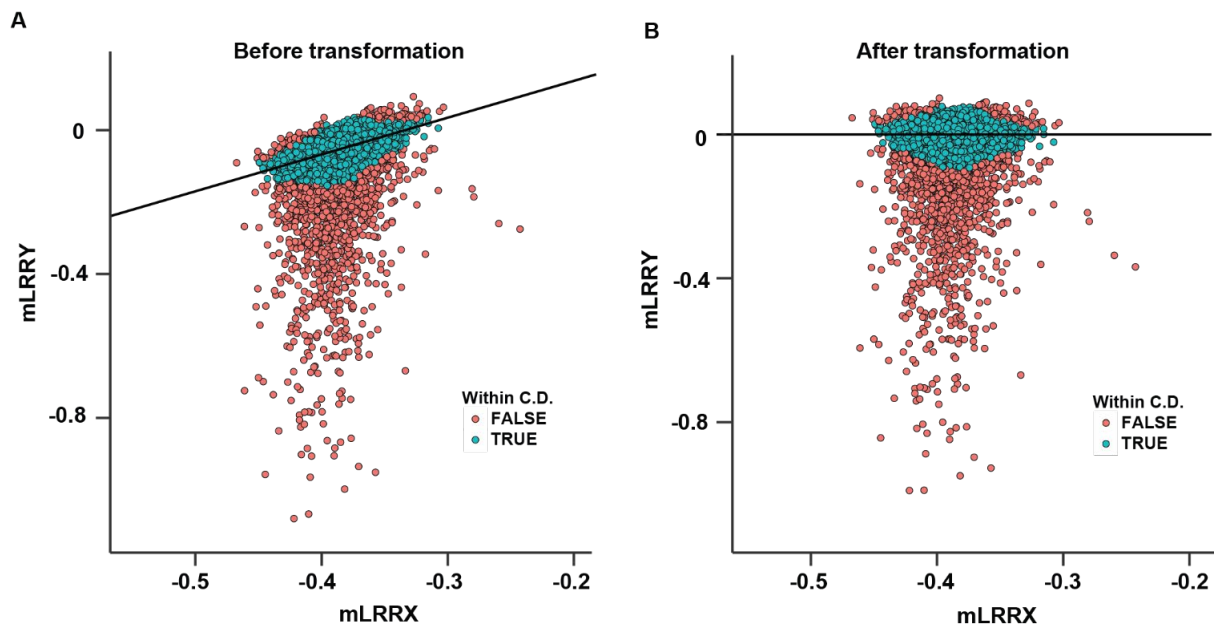


Figure S6. Transformation of mLRRY values by adjusting for technical variation as estimated by mLRRX, panels A and B show the unadjusted adjusted datasets, respectively. The correction was based on the coefficients of a linear model estimated between mLRRY and mLRRX in the unadjusted dataset (panel A). To optimize the model only samples showing a value of mLRRY > -0.2 in the unadjusted dataset and samples within a Cook's distance (i.e. similarity of mLRRY and mLRRX estimates between samples, abbreviated C.D.) less than 1.5 IQR of the third quartile, were included in the model. The coefficients of the produced linear regression model could thereafter be used to adjust the mLRRY values [mLRRY_adj = (mLRRY_unadj - 1.023611 · mLRRX) - 0.341685].

Figure S7.

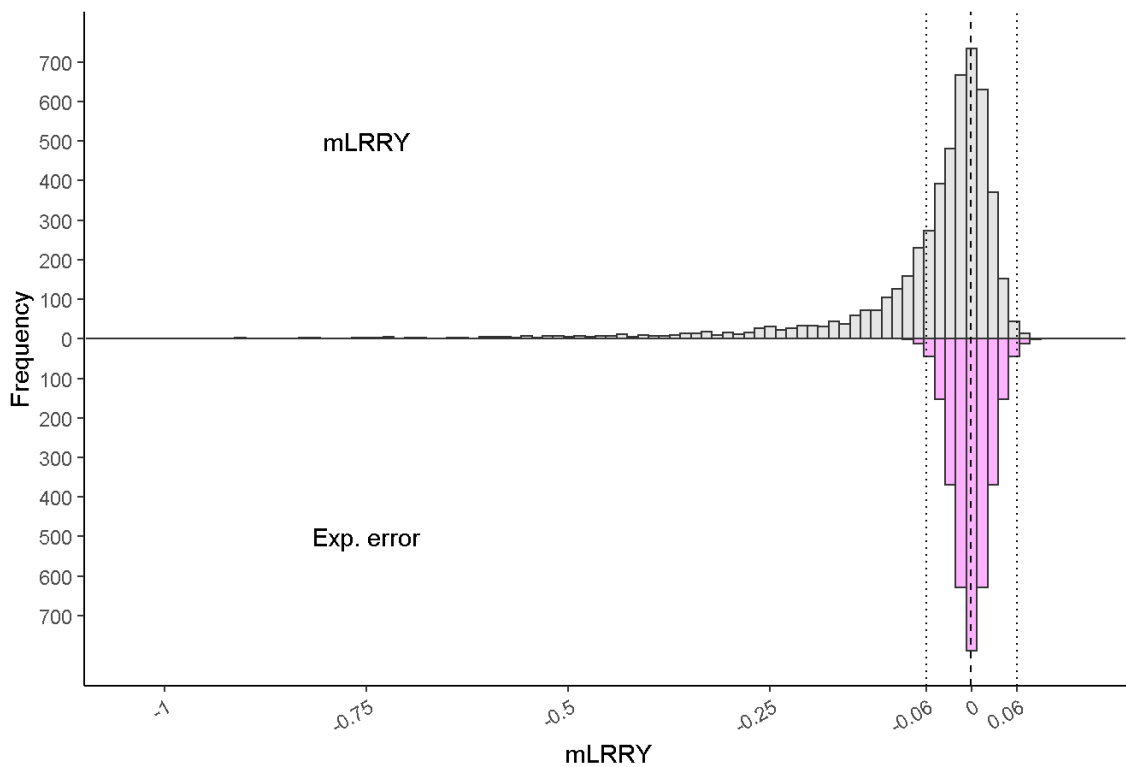


Figure S7. Distribution and estimation of mosaic loss of chromosome in ASPREE study from SNP array data. The grey bars show the distribution of mLRRY values observed in the 5.131 investigated samples. The pink bars represents the part of the total variation originating from technical factors, generated by imposing the observed variation in the positive part of the grey tail into a reflected negative tail, while assuming that this variation was distributed symmetrically. The dotted lines marks the 99% confidence limits of the latter distribution and samples were defined as having LOY when the mLRRY value was below the lower 99% confidence limit (-0.06) of this error distribution.

Figure S8.

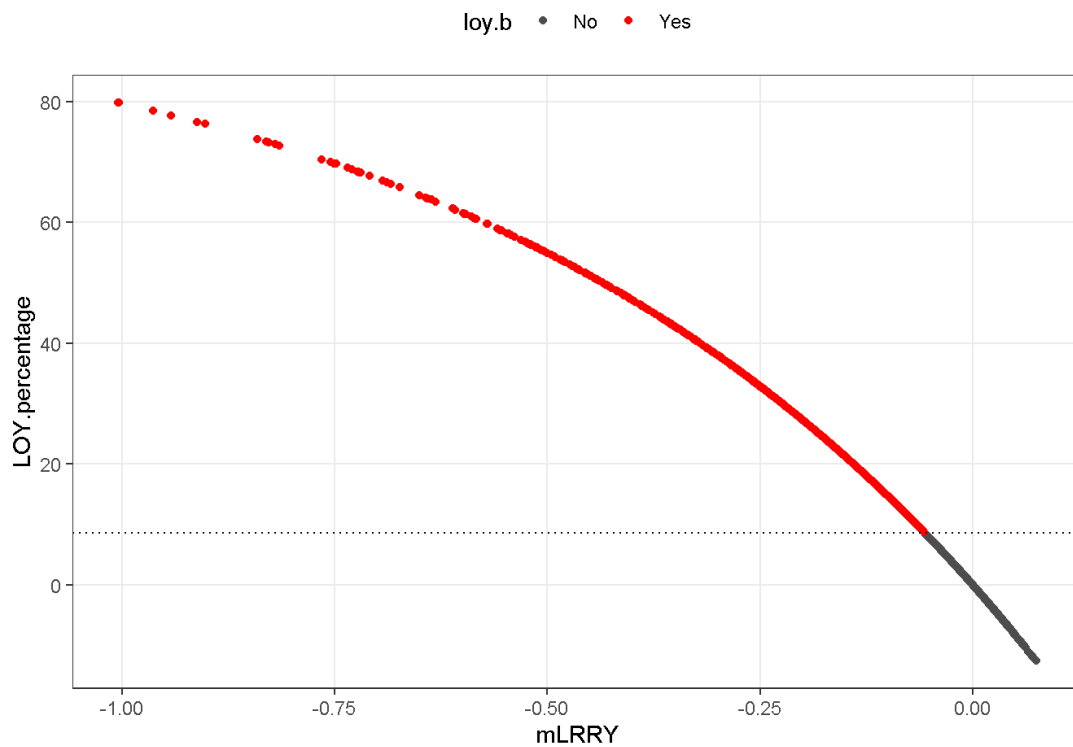


Figure S8. Transformation of mLRRY to percentage of cells with LOY. The mLRRY values were converted to LOY percentage using the formula $[\text{LOY}(\%) = 100 \cdot (1 - 2^{2.3 \cdot \text{mLRRY}})]$. This is a modified version of the published formula $[\text{LOY}(\%) = 100 \cdot (1 - 2^{\text{mLRRY}})]$ that was optimized from data generated by Illumina arrays. The mLRRY to LOY percentage transformation was derived empirically by optimizing alpha and beta in $[(\text{LOY percentage from WGS}) = 100 \cdot (1 - \alpha^{\text{beta} \cdot \text{mLRRY}})]$ using the data in the present analysis. In this model, alpha and beta was originally estimated as 1.006543 and 2.325276, respectively. To avoid overfitting, both values were rounded to produce the modified formula used here. Samples scored with LOY in at least 8.57% blood cells without chromosome Y (i.e. $\text{mLRRY} > -0.06$) are plotted with red dots.

Table S1. Genome wide significant SNPs used to generate the LOY-PRS in ASPREE.

Consensus Gene	SNP	CHR	BP hg38	Risk Allele	OR [Confidence Intervals]
ARHGAP25	rs10048745	2	68735005	A	1.1 [1.08-1.12]
CTSK	rs10305667	1	150859632	C	1.05 [1.04-1.07]
CCND3 / GUCA1B	rs10456506	6	42048508	T	1.14 [1.12-1.17]
CCND2	rs1049612	12	4303596	A	1.05 [1.04-1.07]
PMF1	rs1052053	1	156232382	G	1.16 [1.14-1.18]
SESN3	rs10831321	11	95231179	G	1.05 [1.03-1.06]
LTBR	rs10849448	12	6384185	G	1.09 [1.07-1.1]
PARP11	rs11062924	12	3933197	T	1.08 [1.06-1.1]
SETBP1	rs11082396	18	44500755	C	1.23 [1.2-1.25]
KRBA1	rs111725880	7	149706135	C	1.15 [1.12-1.17]
MDM2 / NUP107	rs11177383	12	68815509	C	1.05 [1.04-1.07]
BEND7	rs11258419	10	13494086	G	1.04 [1.03-1.06]
HDAC7 / VDR	rs113736796	12	47819937	G	1.12 [1.08-1.16]
HEATR3	rs11642909	16	50022787	A	1.05 [1.03-1.07]
SPDL1	rs116483731	5	169588475	G	1.42 [1.31-1.53]
CXCR4	rs11679328	2	136126891	T	1.15 [1.11-1.18]
FLT3	rs117145034	13	28100660	A	1.25 [1.18-1.33]
IKZF1	rs11769630	7	50218107	A	1.14 [1.11-1.17]
FAM117A / SPOP	rs118035610	17	49726945	C	1.26 [1.22-1.3]
MRPS18A / VEGFA / MAD2L1BP	rs11965885	6	43725357	G	1.05 [1.03-1.06]
SLC25A37	rs12549737	8	23536948	T	1.07 [1.05-1.09]
ZBTB20	rs12695310	3	114855180	G	1.07 [1.06-1.09]
SENP7 / PCNP	rs13062095	3	101548541	C	1.12 [1.1-1.13]
SETD2 / NBEAL2	rs13063578	3	47046347	A	1.06 [1.05-1.08]
ANAPC5	rs13141	12	121318281	G	1.25 [1.17-1.34]
HABP4	rs13286011	9	96492112	C	1.13 [1.08-1.17]
SETBP1	rs141777833	18	44295910	C	1.39 [1.28-1.51]
TSPAN9	rs147764594	12	3125211	AT	1.12 [1.09-1.14]
ITPR2	rs149752564	12	26435050	A	1.12 [1.09-1.15]
DAP	rs1531842	5	10675654	T	1.05 [1.03-1.07]
PRDM16	rs1569419	1	3080038	T	1.06 [1.05-1.08]
FAM49A	rs16982394	2	16443481	A	1.11 [1.1-1.13]
AHI1	rs17064495	6	135419391	C	1.15 [1.1-1.2]
RPN1 / GATA2	rs17255991	3	128637371	C	1.12 [1.09-1.15]
BCL2	rs17758695	18	63253621	C	1.75 [1.67-1.82]
BAX	rs1805419	19	48955847	G	1.07 [1.05-1.09]
SETBP1	rs1849209	18	44581678	T	1.19 [1.17-1.21]
CHEK2	rs186430430	22	28707610	C	2.02 [1.72-2.38]
CTBP2	rs1926785	10	125161466	A	1.07 [1.05-1.09]
TET2	rs199741557	4	104943372	ACT	1.13 [1.09-1.18]
KIT	rs218264	4	54542708	A	1.06 [1.04-1.07]
MAD1L1	rs2280548	7	1937031	T	1.13 [1.11-1.14]

MADD / DDB2	rs2291119	11	47276650	C	1.05 [1.03-1.07]
MECOM	rs2293661	3	169120620	G	1.07 [1.06-1.09]
NFKB1	rs230533	4	102528926	G	1.05 [1.04-1.07]
MBD5	rs2382230	2	148545695	T	1.05 [1.03-1.06]
CCDC26	rs2395902	8	129599841	G	1.06 [1.04-1.07]
PHF11 / RCBTB1	rs2407797	13	49684885	A	1.06 [1.05-1.08]
AZGP1	rs2527884	7	99965219	G	1.05 [1.03-1.06]
PRDM16	rs2651932	1	3182097	G	1.08 [1.05-1.11]
SGMS1	rs2688887	10	50302463	T	1.05 [1.03-1.06]
TRPT1	rs2701539	11	64195906	G	1.09 [1.06-1.13]
SETBP1	rs2852780	18	44697023	A	1.08 [1.06-1.1]
TCL1A	rs2887399	14	95714358	G	1.25 [1.22-1.27]
MAD2L1	rs2908986	4	120063721	A	1.05 [1.03-1.06]
CDK5RAP1	rs291699	20	33394393	T	1.12 [1.09-1.14]
RBPM5	rs2979469	8	30427575	C	1.09 [1.08-1.11]
TAL1	rs34087210	1	47224170	G	1.05 [1.03-1.06]
TESC	rs747692613 merged into rs34344826	12	117040962	TTCC	1.05 [1.03-1.07]
TP53INP1	rs34624977	8	94954676	CT	1.06 [1.05-1.08]
RPN1 / GATA2	rs34890930	3	128610380	T	1.06 [1.04-1.07]
NFE2 / HNRNPA1	rs35979828	12	54292096	T	1.14 [1.11-1.17]
HMHA1	rs36084354	19	1079960	A	1.11 [1.08-1.14]
CENPN	rs3743503	16	81022836	G	1.14 [1.11-1.16]
H2AFY	rs3756364	5	135386511	G	1.08 [1.06-1.11]
BCL2L1	rs3761704	2	111146121	G	1.07 [1.05-1.09]
TRIM58	rs3811444	1	247876149	C	1.06 [1.04-1.08]
QKI	rs381500	6	164057356	C	1.16 [1.14-1.18]
MYB	rs397687328	6	135201210	AT	1.06 [1.05-1.08]
TSC22D2	rs59633341 merged into rs397689554	3	150301093	A	1.24 [1.22-1.27]
CEBPA	rs79966604 merged into rs398101182	19	33261752	G	1.07 [1.05-1.09]
USP28 / HTR3A	rs4288784	11	113874886	G	1.09 [1.06-1.12]
CDKN1C	rs458069	11	2837570	G	1.05 [1.03-1.06]
GACAT3	rs4669037	2	16088807	T	1.06 [1.05-1.08]
SPRED2	rs4671127	2	65335836	C	1.05 [1.03-1.06]
CCND3 / GUCA1B	rs4714550	6	42018535	C	1.1 [1.08-1.12]
PHGDH / NOTCH2	rs478093	1	119712503	G	1.05 [1.03-1.06]
PSMA6 / NFKBIA	rs4981287	14	35358992	A	1.08 [1.06-1.1]
USP35 / GAB2	rs760622583 merged into rs55765495	11	78213610	T	1.07 [1.05-1.09]
NREP	rs56116444	5	111726150	G	1.18 [1.15-1.21]
TERT	rs56345976	5	1276758	G	1.04 [1.03-1.06]
KCNA3 / RP11-284N8.3	rs56795609	1	110666096	A	1.06 [1.04-1.08]
HMGA1	rs57026767	6	34251921	C	1.06 [1.04-1.08]
SNTA1	rs575653337	20	33419196	T	1.24 [1.17-1.31]
SYNGR1	rs5757613	22	39319257	A	1.05 [1.03-1.07]
CEBPB	rs6020413	20	50279908	T	1.05 [1.03-1.07]

PMAIP1	rs60781426	18	59893578	A	1.07 [1.04-1.09]
CDKN1C	rs60808706	11	2836003	G	1.11 [1.07-1.15]
HM13 / BCL2L1	rs6089050	20	31725142	C	1.17 [1.15-1.2]
PARP11	rs609018	12	3962735	G	1.09 [1.08-1.11]
TSC1	rs621940	9	132994743	C	1.09 [1.06-1.11]
UQCRC1	rs62618742	3	48601368	C	1.28 [1.22-1.33]
SMC2	rs6479226	9	104210591	T	1.05 [1.03-1.06]
KLHL1	rs670180	13	70662479	A	1.05 [1.04-1.07]
FANCL	rs6731121	2	58764733	G	1.05 [1.04-1.07]
JARID2	rs6924733	6	15385050	G	1.05 [1.03-1.06]
ARID1B	rs6939093	6	156655411	A	1.12 [1.08-1.16]
KANK1	rs7035770	9	668887	A	1.04 [1.03-1.06]
ESYT2 / NCAPG2	rs710422	7	158753852	G	1.07 [1.05-1.09]
NPAT / ATM	rs7129527	11	108174268	G	1.1 [1.08-1.11]
ATP8B4	rs7172615	15	50065546	G	1.07 [1.05-1.08]
DLK1 / MEG3	rs72698720	14	100712378	C	1.18 [1.15-1.2]
HM13 / BCL2L1	rs7271671	20	31577985	C	1.22 [1.18-1.26]
SETBP1	rs72899729	18	44461166	G	1.45 [1.36-1.55]
LY75	rs72955755	2	159861821	G	1.06 [1.04-1.07]
ACVR1B	rs73111522	12	51910937	A	1.15 [1.11-1.19]
ELF1 / RGS17P1	rs73176930	13	41005673	A	1.1 [1.08-1.12]
STMP1	rs73721669	7	135662359	C	1.16 [1.14-1.19]
TM6SF2	rs739846	19	19308262	A	1.09 [1.06-1.12]
ZBED4	rs7410534	22	49896277	T	1.09 [1.06-1.12]
UGCG	rs74845559	9	111896656	C	1.09 [1.06-1.11]
KDELC2 / ATM	rs74911261	11	108486410	A	1.2 [1.15-1.26]
FLT3	rs76428106	13	28029870	C	1.28 [1.2-1.37]
ZWILCH / MAP2K1	rs76428668	15	66546944	G	1.06 [1.04-1.07]
IKZF1	rs7781977	7	50306538	C	1.07 [1.05-1.08]
TP53	rs78378222	17	7668434	G	1.77 [1.65-1.88]
SYT4	rs80036149	18	43486613	A	1.07 [1.04-1.09]
MUC22	rs9262570	6	31044708	T	1.18 [1.13-1.24]
CD164 / CCDC162P	rs9400267	6	109278961	T	1.13 [1.12-1.15]
KITLG	rs980381	12	88360793	T	1.06 [1.04-1.08]
RPN1 / GATA2	rs9844706	3	128661938	G	1.11 [1.09-1.14]
CCDC102A	rs9939347	16	57536792	G	1.06 [1.05-1.08]
PARP1	rs9943081	1	226350353	C	1.06 [1.04-1.08]

Table S2. Frequency of mosaic LOY in blood leukocytes among 5.131 ASPREE male participants, stratified by age groups.

	Age Categories			
	70-74 years (N = 3144)	75-79 years (N = 1280)	80-84 years (N = 551)	>=85 years (N = 156)
LOY Yes	668 (21.0%)	403 (31.5%)	214 (43.7%)	80 (51.3%)
LOY No	2476 (79.0%)	877 (68.5%)	310 (56.3%)	76 (48.7%)

Table S3. Area under the curve for each variable for mLRRY derived LOY risk prediction.

Variable	*AUC (95% confidence interval)
Age	0.620 (0.60; 0.63)
Smoking	0.526 (0.51; 0.54)
Alcohol use	0.510 (0.50; 0.52)
LOY-PRS	0.642 (0.62; 0.65)

*Area under the curve using ROC package in R.

Table S4. Association of a polygenic risk score for LOY predisposition (LOY-PRS) as a categorical variable (low, middle, high) with mosaic LOY modelled as a continuous variable.

	*OR (95% CI)	P value
LOY low risk PRS	Reference	
LOY middle risk PRS	1.45 (1.27; 1.66)	<0.0001
LOY high risk PRS	2.90 (2.46; 3.41)	<0.0001
Age, (years)	1.10 (1.08; 1.11)	<0.0001
Smoking		
Never/Former	Reference	
Current	2.19 (1.65; 2.90)	<0.0001
Alcohol		
Never/Former	Reference	
Current	1.22 (1.05; 1.41)	0.006

*Estimated change in the mean of LOY using linear regression. The PRS risk group define as: low risk PRS <20%, Middle risk 30-60% and high risk is >80% PRS distribution.

SUPPLEMENTARY REFERENCES

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