

Table 1 Antibacterial efficacy of EOs nanoemulsion in foods

S. No.	EO/Component nanoemulsion	Effective constitutes	Biopolymer	Fabrication Method	Effective over/ Test organism	Key findings	References
<b>Gram (+)ve</b>							
1	<i>Piper nigrum</i>	$\beta$ - caryophyllene, limonene, $\delta$ -3-carene and pinene	Hydroxypropyl $\beta$ -cyclodextrin	Inclusion complex	<i>Staphylococcus aureus</i>	<ul style="list-style-type: none"> <li>• Minimum Inhibitory Concentration (MIC): MIC value of Nanoemulsion of, black pepper was improved 4 times against the <i>S. aureus</i> (from 1000 to 250 <math>\mu\text{g/mL}</math>).</li> <li>• Effective mechanism: Encapsulated oil primarily acted on cell membrane and inside the cytoplasm.</li> </ul>	Rakmai et al. (2017)

2	<i>Eugenia caryophyllata</i>	Eugenol and $\beta$ - caryophyllene	Chitosan	Ionic gelation		<ul style="list-style-type: none"> <li>• Minimum inhibitory volume (MIV) of oil loaded nanoparticle is 2 <math>\mu</math>L against <i>S. aureus</i> and <i>L. monocytogenes</i></li> <li>• Effective mechanism: Nano-sized Clove EO can penetrate bacterial cell wall and destroyed cell membrane.</li> </ul>	Hadidi et al. (2020)
3	<i>Origanum vulgare</i>  <i>Thymus capitatus</i>	Thymol, $\gamma$ - terpinene, and Cymene  Carvacrol, <i>p</i> -Cymene, $\beta$ - caryophyllene, and $\gamma$ - terpinene	Polymeric poly ( $\epsilon$ -caprolactone)	Nanoprecipitation	<i>S. aureus</i> and <i>L. monocytogenes</i>	<ul style="list-style-type: none"> <li>• MIC of <i>Thymus</i> nanoemulsion against <i>S. aureus</i> and <i>L. monocytogenes</i> was 0.5 and 0.25 mg/mL, respectively.</li> <li>• In case of <i>O. vulgare</i> nanoemulsion MIC was 0.5 and 1 mg/mL against <i>S. aureus</i> and <i>L. monocytogenes</i>, respectively.</li> </ul>	Granata et al. (2018)

						<ul style="list-style-type: none"> <li>• Effective mechanism: Effective constituents of EO encapsulated in polycaprolactone able to interact with outer cell membrane, increasing permeability and fluidity, which causes structural and functional damage to cell membrane.</li> </ul>	
4	<i>Moringa oleifera</i>	Palmitic acid and phytol	Chitosan	Ionic gelation		<ul style="list-style-type: none"> <li>• <i>M. oleifera</i> nanoemulsion acted as promising antibacterial agent against <i>S. aureus</i> and <i>L. monocytogenes</i> at 4 °C and 25 °C during 10 days storage of cheese without affecting</li> </ul>	Lin et al. (2019)

						<p>organoleptic properties.</p> <ul style="list-style-type: none"> <li>• Effective mechanism: After treatment bacteria with encapsulated oil disturbed the cell membrane resulting the leakage of cell contents.</li> </ul>	
5	d-Limonene	-	$\epsilon$ -polylysine ( $\epsilon$ -PL)	High pressure Homogenization	<i>S. aureus</i> <i>Bacillus subtilis</i> and <i>Saccharomyces cerevisiae</i>	<ul style="list-style-type: none"> <li>• d-limonene nanoemulsion encapsulated into <math>\epsilon</math>-PL resulted in synergistic antimicrobial activity against the wide range Gram +ve and Gram –ve bacteria.</li> </ul>	Zahi et al. (2017)
<b>Gram (-)ve</b>							
6	<i>Piper nigrum</i>	-	Hydroxypropyl $\beta$ -cyclodextrin	Inclusion complex		<ul style="list-style-type: none"> <li>• Minimum Inhibitory Concentration (MIC): Improvement in MIC</li> </ul>	Rakmai et al. (2017)

					<i>Escherichia coli</i>	<p>value from against <i>E. coli</i> from 2000 to 500 µg/mL.</p> <ul style="list-style-type: none"> <li>• Effective mechanism- Encapsulated oil primarily acted on cell membrane and cytoplasm</li> </ul>	
7	<i>Thymus daenensis</i>	-	Tween 80 (2% w/w), and lecithin (0.001% w/w)	Sonication		<ul style="list-style-type: none"> <li>• Minimum Inhibitory Concentration (MIC): The nanoemulsion showed effective inhibitory action against <i>E. coli</i> with MIC 0.4 mg/mL.</li> <li>• Effective mechanism: The formulation inhibited the <i>E. coli</i> population by enhancing mechanism of action, bringing the EOs into</li> </ul>	Moghimi et al. (2016b)

						close proximity with the plasma membrane.	
8	<i>Thymus vulgaris</i>	Thymol, para-cymene, linalool, terpinene, $\alpha$ -pinene, and carvacrol	Casein and maltodextrin	Spray drying		<ul style="list-style-type: none"> <li>• Minimum Inhibitory Concentration (MIC): Encapsulated EO showed 0.1 mg/mL MIC value against <i>S. typhimurium</i>, <i>L. monocytogens</i>, <i>S. aureus</i> and <i>E. coli</i>.</li> <li>• Effective mechanism: Encapsulated EO showed higher antimicrobial activity in comparison to thyme EO through slow release of volatile components that protected by wall materials during <i>in situ</i> storage of hamburger like meat product.</li> </ul>	Radünz et al. (2020)

9	<i>Urtica dioica</i>	Carvacrol, carvone, naphthalene, and anethol	Chitosan	Ionic gelation		<ul style="list-style-type: none"> <li>The nanoencapsulated nettle EO boosted the antibacterial activity against the foodborne bacteria viz. <i>S. aureus</i>, <i>L. monocytogenes</i>, <i>S. typhi</i>, <i>E. coli</i>, and <i>B. cereus</i></li> </ul>	Bagheri et al. (2021)
10	<i>Melaleuca alternifolia</i>	Terpinene-4-ol, $\gamma$ -terpinene, and terpinene	$\beta$ -cyclodextrin	Inclusion complex		<ul style="list-style-type: none"> <li>TTO/ <math>\beta</math>-CD inclusion complex enhanced the shelf life of beef against <i>E. coli</i> and inhibited 99.99% food spoilage bacteria at 4 °C over 7 days storage.</li> </ul>	Cui et al. (2018)
11	<i>Cinnamomum zeylanicum</i>	Cinnamaldehyde	Medium-chain triglyceride	Emulsification		<ul style="list-style-type: none"> <li>MIC value of cinnamaldehyde nanoemulsion was 98 <math>\mu</math>g/mL against <i>E. coli</i>.</li> <li>Effective mechanism: Encapsulation of cinnamaldehyde</li> </ul>	Tian et al. (2016)

						showed strong inhibitory effect against <i>E. coli</i> within 9 h of treatment.	
12	Cinnamon oil	Eugenol and cinnamaldehyde	Coconut oil, and Tween 80	Microfluidization and Ultrasonication		<ul style="list-style-type: none"> <li>Nanoemulsion showed promising application in food sector against the <i>E.coli</i>.</li> </ul>	Yildirim et al. (2017)
13	Cinnamaldehyde	-	Chitosan	Ionic gelation	<i>Pseudomonas aeruginosa</i>	<ul style="list-style-type: none"> <li>MIC of cinnamaldehyde nanoemulsion against</li> </ul>	Subhaswaraj et al. (2018)



						<p><i>P. aeruginosa</i> was found to be 1000 µg/mL.</p> <ul style="list-style-type: none"> <li>• Cinnamaldehyde nanoemulsion inhibited quorum sensing activity and prevented food spoilage with controlled as well as slow release at specific sites.</li> </ul>	
14	Cinnamon oil	Cinnamaldehyde and α-pinene	Chitosan and whey protein	Nanoprecipitation	<i>E. coli</i> , <i>P. fragi</i> , and <i>Shewanella putrefaciens</i>	<ul style="list-style-type: none"> <li>• Antibacterial activity of cinnamon oil emulsion against <i>S. aureus</i> was observed at 0.041%.</li> <li>• Cinnamon oil emulsion acted as strong antimicrobial and long-lasting biological molecule</li> </ul>	Yang et al. (2021)

						against various food contaminating Gram +ve and Gram -ve bacteria indicating utilization as potential preservative in food industry.	
<b>15</b>	<i>Rosmarinus officinalis</i>	-	Chitosan (CS) and benzoic acid (BA)	Sonication	<i>Salmonella typhimurium</i>	<ul style="list-style-type: none"> <li>• Nanogel of <i>R. officinalis</i> showed the MIC value 5 µg/mL against the <i>S. typhimurium</i>.</li> <li>• Nanogel of <i>R. officinalis</i> applied on beef cutlet at the rate of 0.5 mg/g had positive effects on the color and shelf-life.</li> </ul>	Hadian et al. (2017)
<b>16</b>	<i>Cinnamodendron dinisii</i>	1,8-cineole	Chitosan and Zein	Nanoprecipitation	<i>S. typhimurium</i> and <i>Shigella flexneri</i>	<ul style="list-style-type: none"> <li>• Nanoemulsion of <i>C. dinisii</i> film showed greater antimicrobial activity against Gram</li> </ul>	Xavier et al. (2021)

						–ve bacteria and stabilized the deterioration reaction in ground beef leading to prolong shelf-life under refrigerated storage.	
17	Lime oil	-	Chitosan	Nanoprecipitation	<i>Shigella dysenteriae</i>	<ul style="list-style-type: none"> <li>Chitosan nanoparticles incorporating lime EO performed highest antibacterial activity against <i>S. dysenteriae</i> and inhibition halo (HI) value was 3.5 cm at 40 µL volume.</li> </ul>	Sotelo-Boyás et al. (2017)

Table 2. Antifungal efficacy of EOs nanoemulsion in food

S. No.	Effective over/ Test organism	EO/Component (nanoemulsion)	Effective constituents	Biopolymer	Method	Key findings	References
1	<i>A. flavus</i>	<i>Origanum majorana</i>	Terpinen-4-ol, $\alpha$ -terpineol, linalool, and caryophyllene	Chitosan	Ionic gelation	<ul style="list-style-type: none"> <li>MIC of <i>O. majorana</i> EO nanoemulsion was observed to be 1<math>\mu</math>L/mL against <i>A. flavus</i>.</li> <li>Effective mechanism: <i>O. majorana</i> EO nanoemulsion showed remarkable inhibitory efficacy against the <i>A. flavus</i> growth and AFB<sub>1</sub> production as compared to free <i>O. majorana</i> EO by enhancing reduction of ergosterol concentration.</li> </ul>	Chaudhari et al. (2020)
2		<i>Eugenia caryophyllata</i>	Eugenol and caryophyllene	Chitosan	Sonication	<ul style="list-style-type: none"> <li><i>E. caryophyllata</i> EO nanoemulsion showed remarkable antifungal (0.25 <math>\mu</math>L/mL) and aflatoxin B<sub>1</sub></li> </ul>	Kujur et al. (2021)

						<p>inhibitory (0.15 <math>\mu\text{L}/\text{mL}</math>) activity.</p> <ul style="list-style-type: none"> <li>• <i>E. caryophyllata</i> EO nanoemulsion exhibited promising fungitoxic, aflatoxin B<sub>1</sub> inhibitory activity and preserved the sensory properties of six month stored maize seeds.</li> </ul>	
3		<i>Cuminum cyminum</i>	-	Chitosan and Caffeic acid	Sonication	<ul style="list-style-type: none"> <li>• Encapsulated <i>C. cyminum</i> EO showed promising MIC (350 ppm) against <i>A. flavus</i> under sealed condition as compared to free oil (650 ppm).</li> </ul>	Zhaveh et al. (2015)
4		Orange oil	-	$\beta$ -cyclodextrin	Inclusion complex	<ul style="list-style-type: none"> <li>• Orange EO nanoemulsion showed complete inhibition at 900 mg/mL.</li> <li>• Nanoencapsulated EO improved the antimicrobial activity in contaminated cakes</li> </ul>	Kringel et al. (2021)

5		<i>Cananga odorata</i>	Linalool and benzyl acetate	Chitosan	Ionic gelation	<ul style="list-style-type: none"> <li>• <i>C. odorata</i> EO nanoemulsion reduced the <i>A. flavus</i> growth and aflatoxin B<sub>1</sub> secretion at 1.0 and 0.75 <math>\mu\text{L}/\text{mL}</math>, respectively.</li> </ul>	Upadhyay et al. (2021)
6		Eugenol	-	Chitosan	Ionic gelation	<ul style="list-style-type: none"> <li>• Eugenol nanoemulsion showed antifungal and aflatoxin B<sub>1</sub> inhibitory activity at 0.07 and 0.06 <math>\mu\text{L}/\text{mL}</math> concentration, respectively.</li> <li>• Eugenol-loaded chitosan nanoemulsion significantly protected the rice seeds from fungal, AFB<sub>1</sub> contamination, lipid peroxidation and maintained their mineral contents and macronutrients</li> </ul>	Das et al. (2021a)
7	<i>A. niger</i>	<i>Eugenia caryophyllata</i>	Eugenol	Chitosan	Ionic gelation	<ul style="list-style-type: none"> <li>• Inhibition of <i>A. niger</i> mycelial growth was 1.5 mg/mL.</li> </ul>	Hasheminejad et al. (2019)

8		Cinnamon leaf oil	Eugenol, benzyl benzoate, caryophyllene, and aceteeugenol	Whey protein	High pressure homogenization	<ul style="list-style-type: none"> <li>• 0.25 µg/mL concentration of CEO inhibited 75% fungal population, whereas, the same concentration in nanoemulsion form was noted to completely inhibit <i>A. niger</i> population.</li> </ul>	Ribes et al. (2017)
9	<i>A. parasiticus</i>	<i>Schinus molle</i>	D-limonene, o-cimene, β-phellandrene, δ-cadinene and caryophyllene	Chitosan	Ionotropic gelation	<ul style="list-style-type: none"> <li>• Nanoencapsulated <i>S. molle</i> EO at 500 µg/mL concentration showed strong antifungal and antiaflatoxigenic activity.</li> </ul>	López-Meneses et al. (2018)
10		<i>Schinus molle</i>	D-limonene, β-phellandrene, and o-cimene	Chitosan	Nanoprecipitation	<ul style="list-style-type: none"> <li>• <i>S. molle</i> EO nanoemulsion effectively reduced (40-50%) viable spores of <i>A. parasiticus</i>.</li> <li>• Antifungal effect of emulsion due to cytoplasmic coagulation, protoplast leakage and loss of conidiation.</li> </ul>	Luque-Alcaraz et al. (2016)

11	<i>Alternaria alternata</i>	<i>Origanum vulgare</i>	$\alpha$ -pinene, $\beta$ -myrcene, carvacrol, borneol, caryophyllene, $\gamma$ -terpinene, $\rho$ -cymene and terpinolene	Chitosan	Electrospraying deposition	<ul style="list-style-type: none"> <li>• Oil loaded chitosan emulsion showed antifungal activity associated with destruction of cell membrane permeability as well as remarkable depolarization of membrane.</li> <li>• Oregano EO loaded nanoemulsion can be used in agriculture and food industry as a promising fungicidal agent against <i>A. alternata</i>.</li> </ul>	Yilmaz et al. (2019)
12		$\alpha$ -pinene	-	Chitosan	Nanoprecipitation	<ul style="list-style-type: none"> <li>• Nanoemulsion inhibited <i>A. alternata</i> growth during the cold storage of bell peppers and maintained the physiochemical property</li> </ul>	Hernández-López et al. (2020)
13	<i>Penicillium italicum</i> <i>P. digitatum</i> and	<i>Litsea cubeba</i>	Neral and limonene	$\beta$ -cyclodextrin	Inclusion complex	<ul style="list-style-type: none"> <li>• MIC values of <i>L. cubeba</i> EO nanoemulsion was found to be 3.13, 1.56, and 0.78 mg/mL against <i>P. italicum</i>, <i>P.</i></li> </ul>	Wang et al. (2020)



	<i>Geotrichum citri-aurantii</i>					<p><i>digitatum</i> and <i>G. candidum</i>, respectively.</p> <ul style="list-style-type: none"> <li>• <i>L. cubeba</i> EO nanoemulsion exhibited antifungal activity and preserved citrus fruits from postharvest decay and disease.</li> </ul>	
<b>14</b>	<i>P. chrysogenum</i> and <i>A. flavus</i>	<i>Ocimum basilicum</i>	Methyl chavicol and linalool	Saponin	Ultrasonication	<ul style="list-style-type: none"> <li>• The nanoemulsion of basil oil had higher antifungal activity against food spoilage fungi. Nanoemulsion at 1000 ppm concentration displayed 64-67% inhibitory activity over <i>P. chrysogenum</i> and <i>A. flavus</i>.</li> </ul>	Gundewadi et al. (2018)
<b>15</b>	<i>Fusarium graminearum</i>	<i>Cymbopogon martini</i>	Geraniol, geranial, geranyl propionate and geranyl acetone	Chitosan	Ionic gelation	<ul style="list-style-type: none"> <li>• <i>C. martini</i> EO nanoemulsion completely inhibited the growth of <i>F. graminearum</i> and mycotoxins at 700 and 900 ppm.</li> <li>• <i>C. martini</i> EO nanoemulsion showed remarkable</li> </ul>	Kalagatur et al. (2018)

						antifungal and antimycotoxigenic activity against <i>F. graminearum</i> during 28 days storage of maize grain.	
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Table 3. Application of nanoencapsulated EO in active packaging of food products

S. No.	Essential oil or their active ingredients	Packaging types/ Emulsifier/Carrier	Food products	Type of microorganism	Biological activity	Reference
1	Citral	Tween 80/Sodium alginate	Pineapple fruit	<i>Salmonella enterica</i> S. <i>typhimurium</i> and <i>L. monocytogenes</i>	<ul style="list-style-type: none"> <li>• Sodium alginate based edible coating of different concentration of citral nanoemulsion i.e. 0.1% (NE1), 0.5% (NE2), and 1% (NE3) significantly reduced the microbial load of fresh cut pineapples during storage for 12 days.</li> <li>• High concentration of citral in NE3 led to improvement in the sensory quality and texture of pineapple, thus the NE2 could be explored as for commercial application.</li> </ul>	Prakash et al. (2020)
2	Carvacrol	Tween 80/ Starch Polymer/Montmorillonite	Vegetables and fruits	<i>E. coli</i>	<ul style="list-style-type: none"> <li>• Carvacrol nanoemulsion showed strong antimicrobial efficacy in food packaging</li> </ul>	de Souza et al. (2020)

					against <i>E. coli</i> due to destabilization and partial destruction of bacterial cell membrane, electron transport, respiration and permeability.	
3	Linalool	Tween 80	Pineapple fruit	<i>S. typhimurium</i>	<ul style="list-style-type: none"> <li>• Nanoemulsion biofilm on the surface of cut pineapple effectively reduced the <i>S. typhimurium</i> and increased the shelf life of cut pineapple.</li> <li>• Nanoemulsion enhanced twofold antibacterial activity against <i>S. typhimurium</i> by increasing ability to cell membrane integrity.</li> </ul>	Prakash et al. (2019)
4	<i>Origanum virens</i>	Whey protien	Sausages	<i>L. monocytogenes</i>	<ul style="list-style-type: none"> <li>• Coating of Portuguese sausage delayed the lipid peroxidation and inhibited the growth of <i>L. monocytogenes</i> during storage condition.</li> <li>• EO coating enhanced the antibacterial activity of oil via</li> </ul>	Catarino et al. (2017)

					sustained release as well as increased the solubility of oil.	
5	<i>Rosmarinus officinalis</i>	Chitosan/ $\epsilon$ -Poly-L-lysine	Carbonado chicken	All types of microorganism viz. Coliform, molds and Yeast	<ul style="list-style-type: none"> <li>• Total bacterial count reduced effectively by nanoemulsion coating and enhances the shelf life of carbonado chicken meat during 6 days storage.</li> <li>• Similar results found in case of Yeast and molds.</li> </ul>	Huang et al. (2020)
6	<i>Zataria multiflora</i> and <i>Bunium persicum</i>	Tween 80/ Chitosan	Turkey fillet	<i>S. enteritidis</i> and <i>L. monocytogens</i>	<ul style="list-style-type: none"> <li>• Shelf life of turkey fillet could be extended to 9 days by using nanoemulsion containing oil during 20 days storage at refrigeration condition.</li> <li>• Turkey meat coated with nanoemulsion of <i>Zataria multiflora</i> EO (1%) showed best antibacterial activity.</li> </ul>	Keykhosravy et al. (2020)
7	<i>Thymus vulgaris</i>	Tween 80/ Calcium alginates	Ground beef	<i>Staphylococcus aureus</i> , <i>E. coli</i> , Yeast and Molds	<ul style="list-style-type: none"> <li>• Active biopolymer films revealed high antimicrobial activity in agar test agar</li> </ul>	Almasi et al. (2021)

					diffusion tests, when apply to package ground beef.	
8	Lemmon grass oil	Tween 80/ Chitosan	Grape berries	<i>S. typhimurium</i>	<ul style="list-style-type: none"> <li>The nanoemulsion coating showed greater efficiency in microbial safety against <i>Salmonella</i> and preserving ready-to-eat grape berries.</li> </ul>	Oh et al. (2017)
9	<i>Ferulago angulate</i>	Gelatin-Chitosan	Turkey Meat	<i>S. aureus</i> and Coliform	<ul style="list-style-type: none"> <li>Turkey meat packed with gelatin-chitosan film containing 0.5% <i>Ferulago angulate</i> EO reduced the microbial growth and enhances the shelf life of preserved the turkey meat.</li> </ul>	Naseri et al. (2020)
10	<i>Cuminum cyminum</i> and <i>Piper nigrum</i>	Tween 80	Fruits and vegetables	<i>E. coli</i> , and <i>S. enterica</i>	<ul style="list-style-type: none"> <li>Cumin and pepper oil nanoemulsion inhibited the bacterial population on fruits and vegetable by forming biofilm.</li> <li>MIC of Cumin oil nanoemulsion against <i>E. coli</i>,</li> </ul>	Amrutha et al. (2017)

					<p>and <i>S. enterica</i> was 100 and 80 <math>\mu\text{L}/\text{mL}</math> receptively.</p> <ul style="list-style-type: none"> <li>• MIC of Pepper oil against <i>E. coli</i>, and <i>S. enterica</i> was 120 and 140 <math>\mu\text{L}/\text{mL}</math>, receptively.</li> </ul>	
11	<i>Eryngium campestre</i>	Tween 80/Chitosan	Cherry fruit	Mesophilic aerobic bacteria	<ul style="list-style-type: none"> <li>• Coating on cherries decreased the microbial population and weight loss compared to control during storage.</li> <li>• Coating enhanced the firmness, pH, antioxidant activity and total phenolic content.</li> </ul>	Arabpoor et al. (2021)
12	Thymol	Poly-(lactic acid/poly (butylene-succinateco-adipate) (PLA/PBSA) blend film	Bread	<i>Aspergillus</i> spp. and <i>Penicillium</i> spp.	<ul style="list-style-type: none"> <li>• The antifungal activity of thymol coated film was very efficacious against the <i>Aspergillus</i> and <i>Penicillium</i> spp. in both packed bread and <i>in vitro</i> condition.</li> </ul>	Suwanamornlert et al. (2020)
13	Cinnamon Oil	Tween 80	Bread	<i>P. digitatum</i>	<ul style="list-style-type: none"> <li>• The film having 1.5 and 3% Cinnamon oil showed no</li> </ul>	Fasihi et al. (2019)

		/Carboxymethyl cellulose/polyvinyl alcohol			<p>mycelial growth of <i>P. digitatum</i> over 60 days incubation of bread.</p> <ul style="list-style-type: none"> <li>• Active coating at 0.5, 1.5 and 3% exhibited complete fungal growth inhibition on bread slices.</li> <li>• Thus, coating showed the higher effectivity in comparison to films during inhibition of fungal growth in bread slices.</li> </ul>	
14	<i>Origanum vulgare</i>	Tween 20/ Basil Seed Gum (BSG)	Apricot	Yeast and Molds	<ul style="list-style-type: none"> <li>• Coating significantly improved the EO dispersibility.</li> <li>• Coatings of BSG + 6% <i>Origanum</i> oil was showed the most effective in reducing yeast and molds population, extending the shelf life of apricot cuts.</li> </ul>	Hashemi et al. (2017)



15	<i>Mentha piperata</i>	Chitosan	Grape fruits	<i>A. niger</i> , <i>Botrytis cinerea</i> , <i>P. expansum</i> and <i>R. stolonifer</i>	<ul style="list-style-type: none"> <li>• The combined effect of chitosan and <i>Mentha</i> oil effectively delayed the rate of grape decay during storage at room and low temperature.</li> <li>• Coatings of grape barriers strongly reduced the mold infections by increasing antibacterial activity of coatings.</li> </ul>	Guerra et al. (2016)
16	Cinnamon Oil	Chitosan	Cherry Fruits	<i>P. citrinum</i> and <i>A. flavus</i>	<ul style="list-style-type: none"> <li>• Cinnamon oil improved the growth inhibition of <i>P. citrinum</i> from 33.17 mm to 36.23 mm with increasing concentration from 1% to 4% respectively.</li> <li>• Similar result obtained in case of <i>A. flavus</i>. Growth inhibition from 31.12 mm to 35.28 mm with increasing concentration from 1% to 4% respectively.</li> </ul>	Xing et al. (2016)

17	Lemmon grass oil	Flaxseed (FSG)	Gum	Pomegranate Arils	Yeast and Molds	<ul style="list-style-type: none"> <li>• The coating enhanced the potential of oil to enter the cell and improves the interaction between cell and oil.</li> <li>• The coatings significantly reduce the number of microbes and maintain the intrinsic colour of pomegranate arils.</li> </ul>	Yousuf, and Srivastava (2017)
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