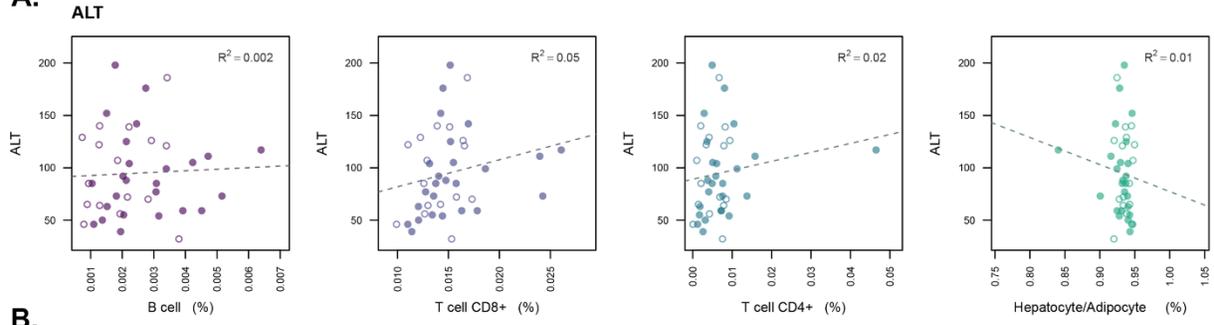
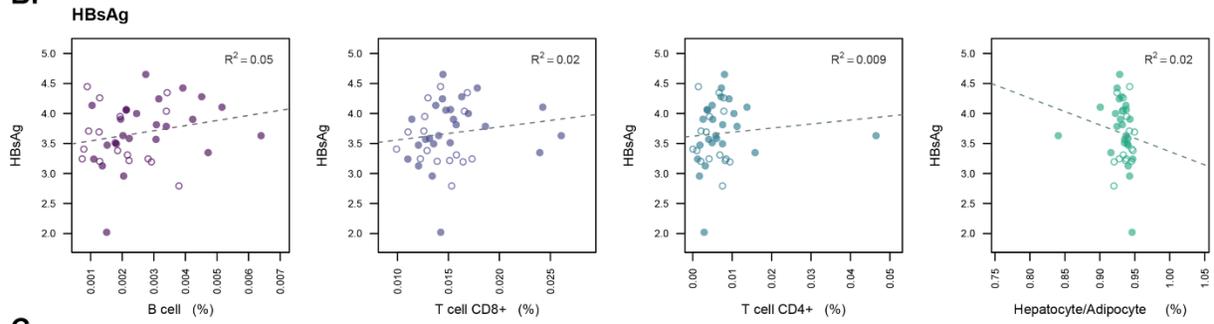
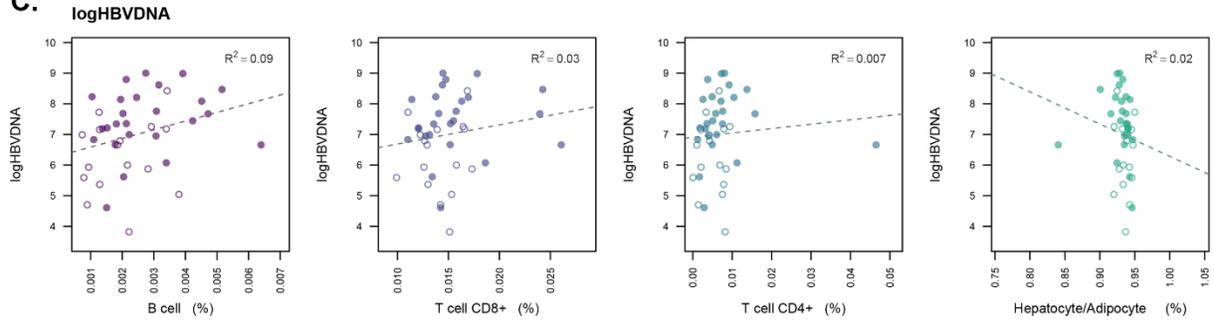
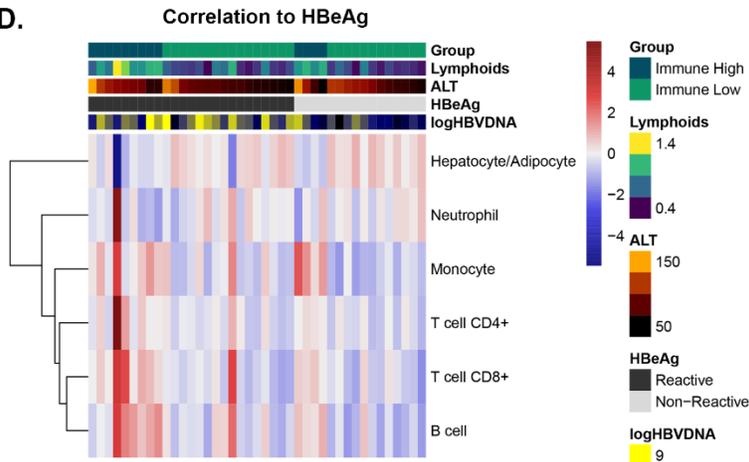


# **Characterization of the liver immune microenvironment in liver biopsies from patients with chronic HBV infection**

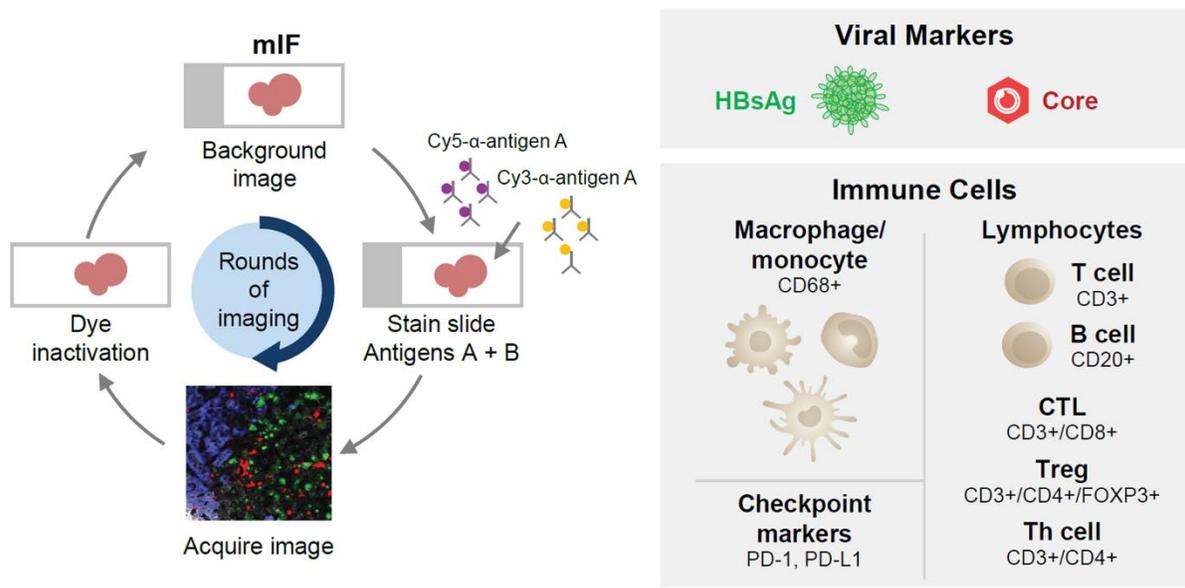
Nicholas van Buuren, Ricardo Ramirez, Scott Turner, Diana Chen, Vithika Suri, Abhishek Aggarwal, Christina Moon, Sam Kim, Dmytro Korniyev, Nam Bui, Neeru Bhardwaj, Henry LY Chan, Patrick Marcellin, Maria Buti, Jeffrey Wallin, Anuj Gaggar, Simon P. Fletcher, Lauri Diehl, Li Li, Hongmei Mo, Becket Feierbach

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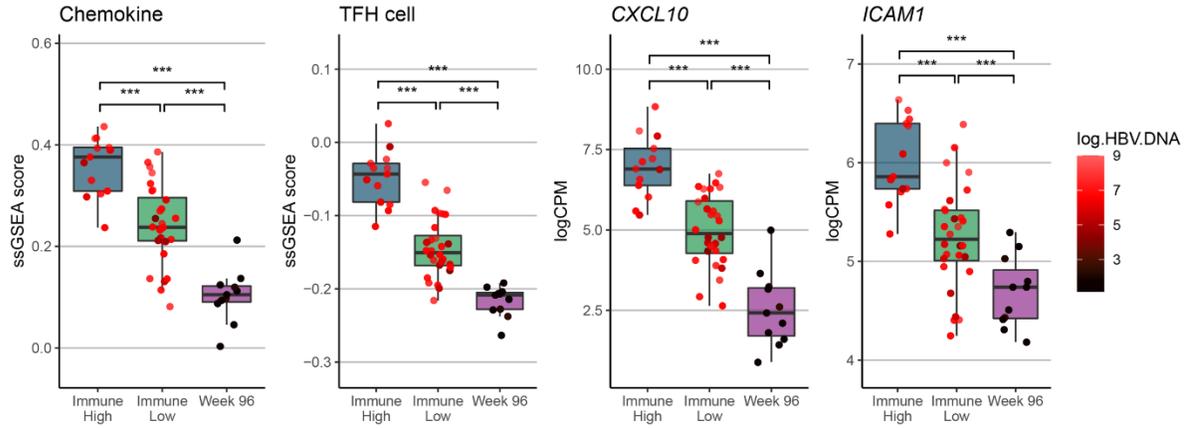
**A.****B.****C.****D.**

**Fig. S1: Liver immune signatures independent of serum HBV DNA, ALT or HBeAg status.** Individual cell type scores from EPIC cell deconvolution were correlated to peripheral biomarkers **(A)** ALT, **(B)** HBsAg, or **(C)** HBV DNA. **(D)** HBeAg status was used to cluster EPIC cell scores in a heatmap. No significant correlations were observed between immune cell signatures and these peripheral biomarkers.

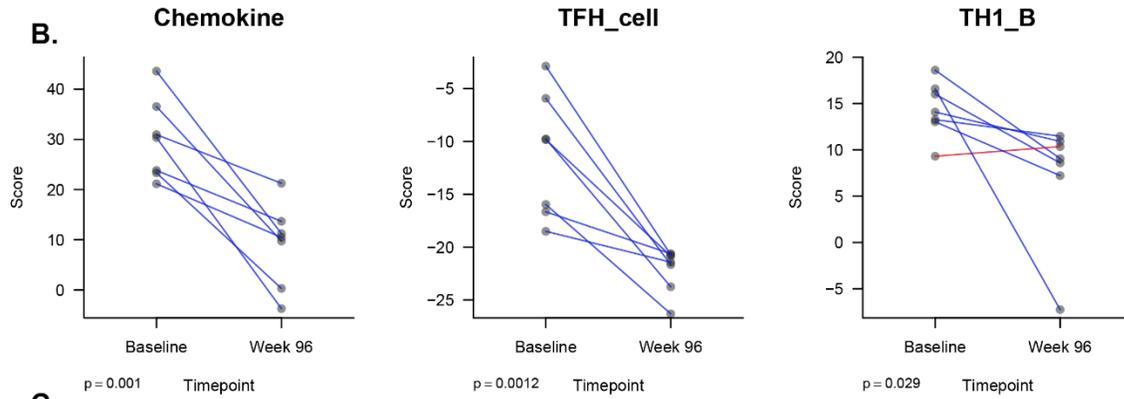


**Fig. S2: Neogenomics mIF platform.** All antibodies were conjugated to Cy5 and Cy3 dyes in pairs and liver biopsies were subjected to 7 rounds of staining, imaging and dye-inactivation. DAPI was used to align subsequent rounds of staining. The markers used for this analysis include HBcAg, HBsAg, CD3, CD4, CD8, CD68, FoxP3, Ki67, PD-1, PD-L1, CD299, and NaKATPase.

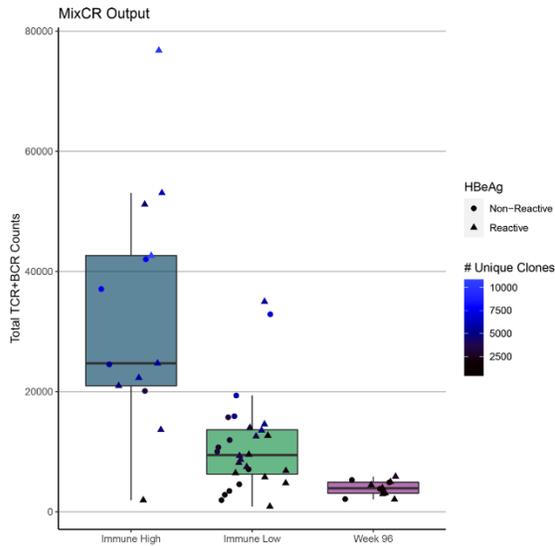
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**B.**

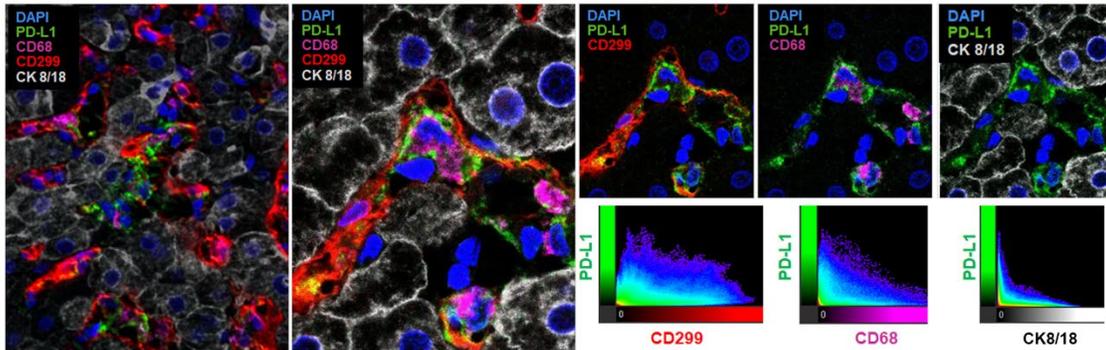


**C.**

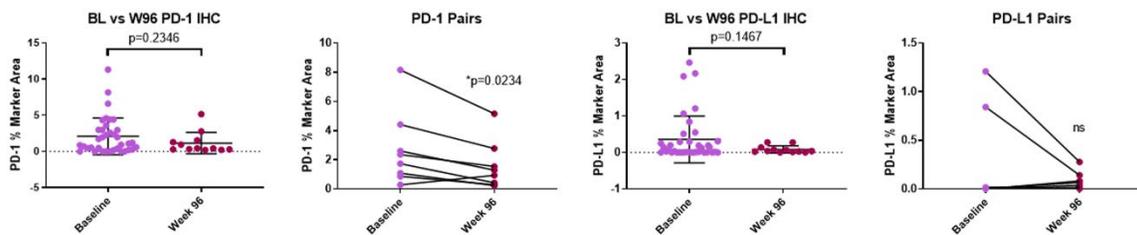


**Fig. S3: TLS signatures reduced by treatment.** **A.** Previously published TLS gene signatures were assessed in all liver biopsies [1-4]. **B.** TLS signatures were also quantified in seven longitudinal pairs. **C.** BCR and TCR clonality was quantified from all RNA-Seq data.

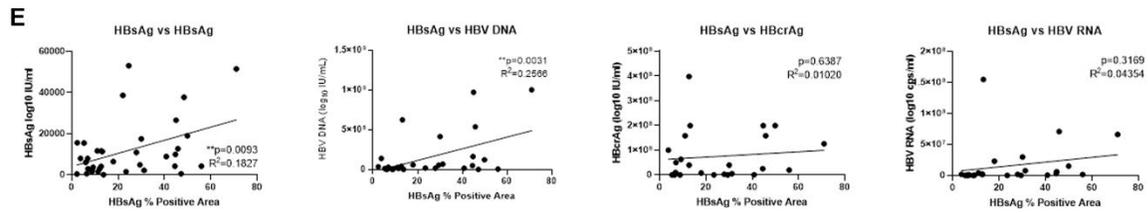
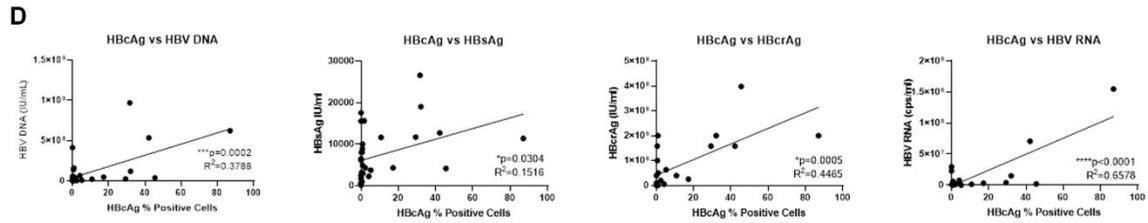
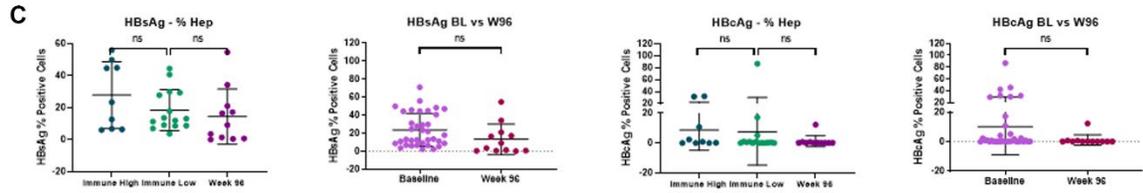
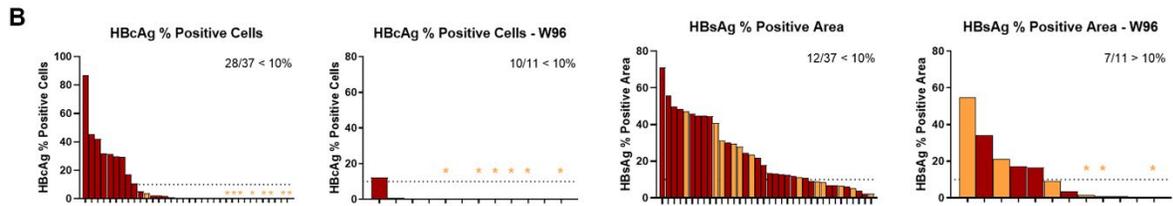
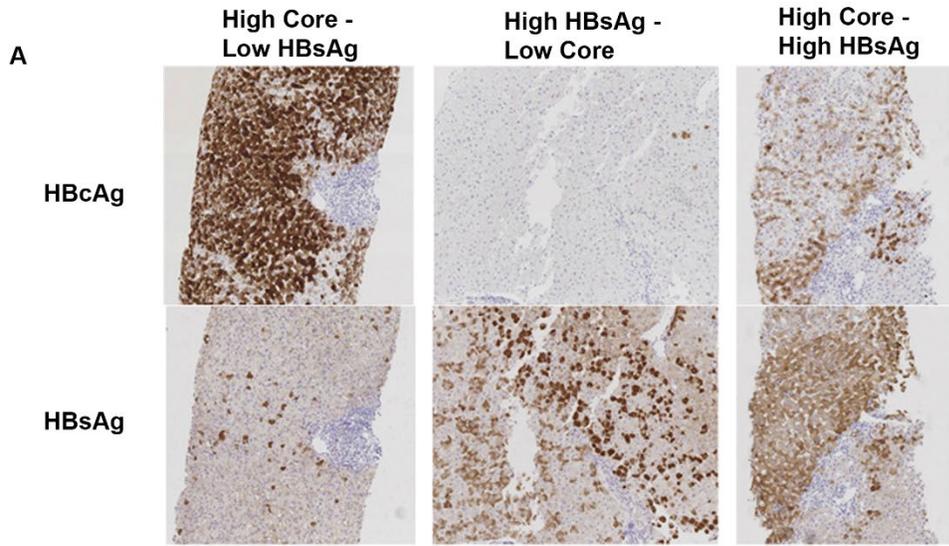
A.



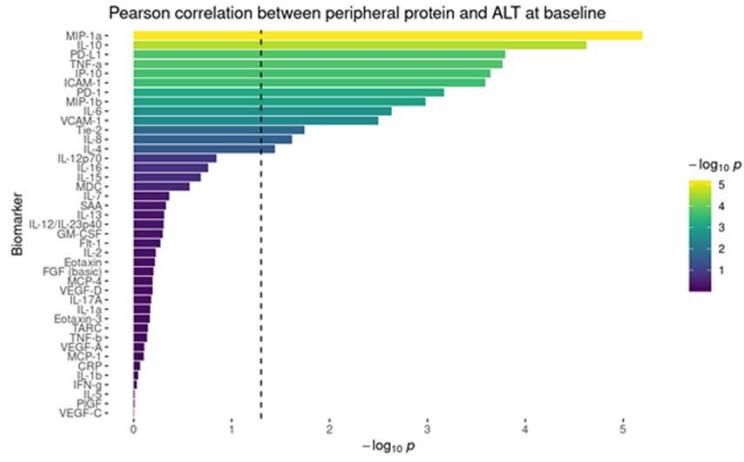
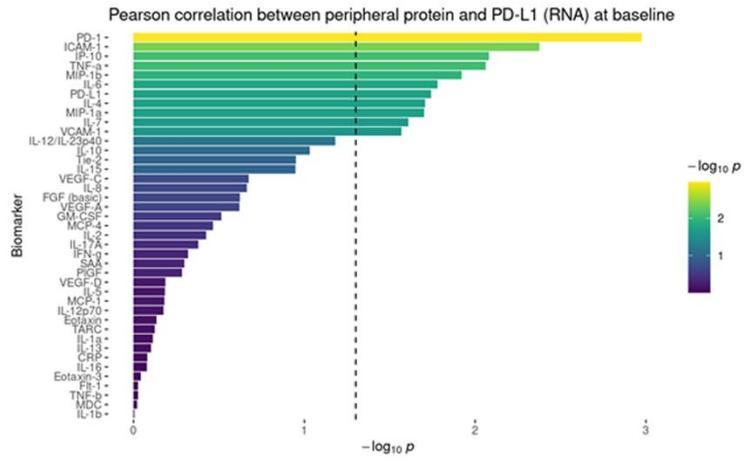
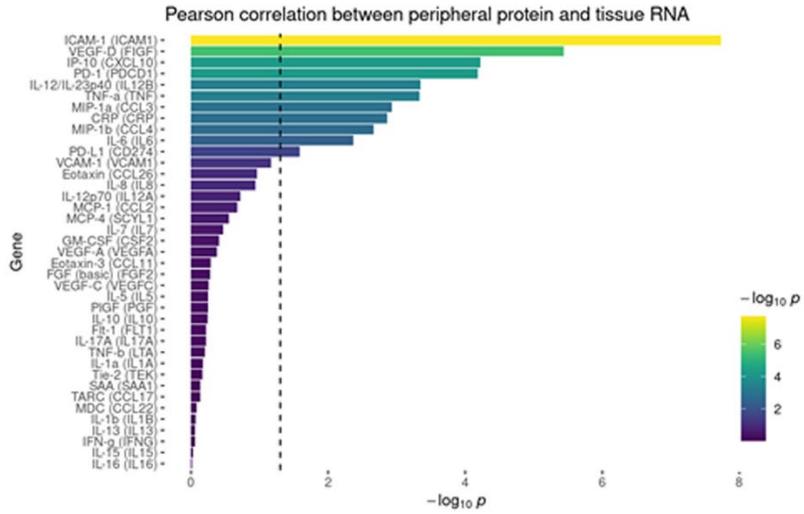
B.



**Fig. S4: PD-L1 is localized to liver sinusoidal endothelial cells and Kupffer cells and not hepatocytes.** A. 4-plex mIF was developed to determine which liver cells express PD-L1. Biopsies with elevated PD-L1 by IHC were subjected to staining for CD68, CD299, PanCK, PD-L1 and DAPI. Image overlays represent a combination of all markers or individual PD-L1 overlays with CD68, CD299 and PanCK. Pearson’s correlation analysis was performed to demonstrated colocalization with PD-L1. B. PD-L1 and PD-1 quantification by single-plex chromogenic staining. Only those with corresponding liver transcriptomes were presented in Figure 4B; this figure represents the comprehensive data set.



**Fig. S5: Analysis of HBcAg and HBsAg burden in liver biopsies using IHC.** **A.** Heterogeneity was observed for HBcAg and HBsAg burden. Antigen burden was scored as % hepatocytes positive for HBcAg or HBsAg and representative images are shown. Each HBcAg image is a matched serial section to the HBsAg image shown below. **B.** HBcAg and HBsAg were quantified for each patient sample imaged by single-plex IHC. Each bar represents an individual patient sample. HBeAg-positive samples are red and HBeAg-negative samples are orange. **C.** All data was plotted based on sampling time point. A subset of the entire collection had complementary intrahepatic transcriptome analyses that were used to classify samples by immune group. Statistical analysis was performed using unpaired t-tests. IHC scores for HBcAg (**D**) and HBsAg (**E**) were correlated to peripheral viral antigen levels including HBV DNA, HBV RNA, HBsAg and HBcrAg. Linear regression was used to quantify the correlation between liver biopsy staining scores and peripheral viral biomarkers.

**A****B****C**

**Fig. S6: Correlation between peripheral cytokine and chemokine biomarkers with serum ALT and liver PD-L1 expression.** 41 analytes quantified by MSD were compared to **(A)** ALT, **(B)** intrahepatic PD-L1 gene expression and **(C)** their own intrahepatic gene expression using Pearson's correlation and ranked by p value.

	Immune High (N=14)	Immune Low (N=27)	Total (N=41)	p value
<b>Age</b>				
N	14	27	41	0.4655
Mean (SD)	36 (6.1)	34 (11.2)	35 (9.7)	
Min, Max	25, 49	19, 61	19, 61	
<b>Sex</b>				
F	5 (35.7%)	10 (37.0%)	15 (36.6%)	0.9344
M	9 (64.3%)	17 (63.0%)	26 (63.4%)	
<b>BMI</b>				
N	14	27	41	0.8798
Mean (SD)	24.8 (5.53)	24.1 (4.61)	24.3 (4.88)	
Min, Max	17.9, 35.7	15.7, 34.1	15.7, 35.7	
<b>Race</b>				
Asian	12 (85.7%)	19 (70.4%)	31 (75.6%)	0.2839
White	2 (14.3%)	8 (29.6%)	10 (24.4%)	
<b>Genotype</b>				
A	0	2 (7.4%)	2 (4.9%)	0.2520
B	3 (21.4%)	8 (29.6%)	11 (26.8%)	
C	10 (71.4%)	11 (40.7%)	21 (51.2%)	
D	1 (7.1%)	6 (22.2%)	7 (17.1%)	
<b>Baseline HBV DNA (log<sub>10</sub> IU/ml)</b>				
N	14	27	41	0.1413
Mean (SD)	7.1 (1.26)	6.6 (1.49)	6.8 (1.43)	
Min, Max	4.4, 9.0	3.8, 8.9	3.8, 9.0	
<b>Baseline HBsAg (log<sub>10</sub> IU/ml)</b>				
N	14	27	41	0.7519
Mean (SD)	3.7 (0.59)	3.6 (0.60)	3.6 (0.50)	
Min, Max	2.1, 4.4	2.0, 4.8	2.0, 4.8	
<b>Baseline ALT (IU/ml)</b>				
N	14	27	41	0.1270
Mean (SD)	132 (78.1)	101 (74.4)	112 (76.2)	
Min, Max	49, 347	6, 341	6, 347	
<b>Baseline HBeAg Status</b>				
Non-Reactive	4 (28.6%)	12 (44.4%)	16 (39.0%)	0.3291
Reactive	10 (71.4%)	15 (55.6%)	25 (61.0%)	
<b>Treatment Arm</b>				
Screen Failure	3 (21.4%)	4 (14.8%)	7 (17.1%)	0.3589
PEG	3 (21.4%)	8 (29.6%)	11 (26.8%)	
TDF	4 (28.6%)	3 (11.1%)	7 (17.1%)	
TDF + PEG	3 (21.4%)	4 (14.8%)	7 (17.1%)	
[TDF + PEG] - TDF	1 (7.1%)	8 (29.6%)	9 (22.0%)	
<b>Baseline HBV RNA (log<sub>10</sub> copies/ml)</b>				
N	11	21	32	0.8118
Mean (SD)	5.3 (1.52)	5.4 (1.62)	5.4 (1.56)	
Min, Max	2.6, 7.2	2.5, 8.2	2.5, 8.2	
<b>Baseline HBcrAg (log<sub>10</sub> IU/ml)</b>				
N	10	21	31	0.9158
Mean (SD)	6.7 (1.36)	6.5 (1.74)	6.5 (1.61)	
Min, Max	4.2, 8.3	3.1, 8.3	3.1, 8.3	

**Table S1: Patient Characteristics from the 174-0149 clinical trial liver biopsy sub-study.** Baseline clinical parameters associated with immune high and immune low samples are shown. No clinical parameters or viral biomarkers differentiated immune high versus immune low subsets. P values were calculated using Cochran-Mantel-Haenszel test and Wilcoxon test for categorical data and continuous data, respectively.

### Supplementary references

- [1] Hennequin A, Derangere V, Boidot R, Apetoh L, Vincent J, Orry D, et al. Tumor infiltration by Tbet+ effector T cells and CD20+ B cells is associated with survival in gastric cancer patients. *Oncoimmunology* 2016;5:e1054598.
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- [3] Messina JL, Fenstermacher DA, Eschrich S, Qu X, Berglund AE, Lloyd MC, et al. 12-Chemokine gene signature identifies lymph node-like structures in melanoma: potential for patient selection for immunotherapy? *Sci Rep* 2012;2:765.
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