

S4 Table. Primers used for qPCR.

Locus	Primer Name	Primer Sequence (5' → 3')	Conc. each (μM)	Efficiency (%)	Amplicon Size (bp)		Comment	Reference
					cDNA	gDNA		
At1g02410	COX11 F	GATTGACATGCCGGTCTTCT	0.33	102.6	164	N/A	spans exon/exon	(Radin et al., 2015)
	COX11 R	TGGTTTCTTGAAGTGGAACAGA						
At1g13320	PP2A F	CCTGCGGTAATAACTGCATCT	0.33	95.5	142	356	spans 2 introns	(Czechowski et al., 2005)
	PP2A R	CTTCACTTAGCTCCACCAAGCA						
At3g18780	ACT2 F	GGTAACATTGTGCTCAGTGGTGG	0.33	100.5	108	108	spans no introns	(Czechowski et al., 2005)
	ACT2 R	AACGACCTTAATCTCATGCTGC						
At3g22370	AOX1a F	GGAGGGCTTCCTGCTGATGCGACA	0.16	102.7	134	214	spans 1 intron	(Radin et al., 2015)
	AOX1a R	AGCTGGAGCTTCCTTAGTTACCGACC						
At3g08950	HCC1 F	GGCCCCGATCTTACCGGGTTT	0.25	99.8	158	391	spans 1 intron	(Radin et al., 2015)
	HCC1 R	CAACGCCGTCTGTCAACGAG						
At4g23290	CRK21 F	TCGTCCAAGCATGTCCACAA	0.5	101.1	120	120	spans no introns	This work
	CRK21 R	CTCTCGGCTAATGGGTTCGG						
At3g15640	COX5b-1 F	GACAAGCGAATTGTGGCTG	0.25	104.3	112	238	spans 1 intron	(Radin et al., 2015)
	COX5b-1 R	AGTACTGAGTGCAAACCGGG						
At4g39740	HCC2 F	CGGATGTTGGACCTGAGCA	0.25	97.0	189	379	spans 2 introns	This work
	HCC2 R	TTGCACTTGCAGTCCCGGTTA						

The optimal final primer concentrations (conc.) and primer pair efficiencies were experimentally determined. The amplicon size of the cDNA and of the potential contamination genomic DNA (gDNA) are given. F = forward, R = reverse, N/A = not applicable.

Czechowski T, Stitt M, Altmann T, Udvardi MK, Scheible WRR. Genome-wide identification and testing of superior reference genes for transcript normalization in *Arabidopsis*. Plant Physiology. 2005;139: 5-17. doi: 10.1104/pp.105.063743

Radin I, Mansilla N, Rödel G, Steinebrunner I. The *Arabidopsis* COX11 Homolog is Essential for Cytochrome c Oxidase Activity. Frontiers in Plant Science. 2015;6:1091. doi: 10.3389/fpls.2015.01091