Supporting Information for:

Decarbonylative Fluoroalkylation at Palladium(II): From Fundamental Organometallic Studies to Catalysis

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I. General information

All manipulations were performed inside an N₂-filled glovebox unless otherwise noted, and all glassware was ovendried for a minimum of 24 h in an oven at 150 °C before use. NMR spectra were obtained on a Varian VNMR 700 (699.76 MHz for 1 H; 175.95 MHz for ¹³C), Varian VNMR 500 (500.09 MHz for ¹H; 470.56 MHz for ¹⁹F; 125.75 MHz for ¹³C), or Varian VNMR 400 (401 MHz for ¹H; 376 MHz for ¹⁹F; 123 MHz for ¹³C) spectrometer. ¹H and ¹³C NMR chemical shifts are reported in parts per million (ppm) relative to TMS, with the residual solvent peak used as an internal reference. ¹⁹F NMR chemical shifts are reported in ppm and are referenced to 4-fluorotoluene (–119.85 ppm) or trifluoromethoxybenzene (–58.00 ppm). ¹³C NMR spectra are referenced to the residual CHCl₃ peak (77.16 ppm). Abbreviations used in the NMR data are as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad signal. Yields of reactions that generated fluorinated products were determined by ¹⁹F NMR spectroscopic analysis using a relaxation delay of 5 s with a 90° pulse angle. Mass spectral data were obtained on a Micromass Magnetic Sector Mass Spectrometer. Automated flash chromatography was performed using a Biotage Isolera One system with cartridges containing high performance silica gel.

Abbreviations: tetrahydrofuran (THF), dichloromethane (DCM), diethyl ether (Et₂O), difluoroacetyl fluoride (DFAF), difluoroacetic anhydride (DFAAn), trifluoroacetic anhydride (TFAAn), difluoroacetic acid (DFA), trifluoroacetic acid (TFA), pentafluoropropionic anhydride (PFPAn), pentafluoropropionic acid (PFPA), tetramethyl ammonium fluoride (TMAF), tetrabutylammonium fluoride (TBAF), room temperature (RT).

II. Materials and methods

All commercially available reagents were used as received unless otherwise stated. Pd[P(o-Tol)₃]₂ (Strem, Alfa Aesar), phosphine ligands (Strem, CombiBlocks, Sigma Aldrich, Oakwood Chemicals), cesium fluoride (Acros), tetramethylammonium fluoride (Manchester), and fluoroalkyl anhydrides (Oakwood Chemicals) were stored in a N₂atmosphere glovebox. Aryl boronic acid and aryl bromide precursors were purchased from commercial sources (Sigma Aldrich, Alfa Aesar, Matrix Scientific, Frontier Scientific, Synquest, TCI America, Oakwood Chemicals) and used as received, unless stated otherwise. Deuterated solvents were purchased from Cambridge Isotope Laboratories, Inc.

III. Compatibility experiments [Table 1 of manuscript]



A. Trifluoroacetic anhydride and diphenyl zinc. A THF solution (0.5 M) of trifluoroacetic anhydride was prepared, and 0.1 mL of this solution was added to a pre-weighed vial containing a stir bar and Ph₂Zn (11 mg, 0.05 mmol, 1.0 equiv). The solution was diluted to 0.4 mL with THF, sealed with a Teflon-lined cap, and stirred at room temperature for 1 h. 4-Fluorotoluene (0.025 mL, 2.0 M, 0.05 mmol, 1.0 equiv) was added as a ¹⁹F NMR standard, the solution was transferred to an NMR tube, and the reaction was analyzed via ¹⁹F NMR spectroscopy. 2,2,2-

trifluoromethyl acetophenone **A** was formed in 77% yield (signal at –72.4 ppm) with no remaining anhydride. A broad signal attributed to a zinc trifluoromethyl acetate was also observed at –76.2 ppm as shown below in Figure S1.



30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 f1 (ppm)

Figure S1. Incompatibility of Ph₂Zn with trifluoroacetic anhydride (TFAAn) in THF as shown by (top) ¹⁹F NMR spectrum of TFAAn with 4-fluorotoluene internal standard, and (bottom) ¹⁹F NMR spectrum of reaction of TFAAn with diphenyl zinc after 1 h in THF showing formation of ketone **A**.

- B. Trifluoroacetic anhydride and phenyl boronic acid. A CDCl₃ solution (1.0 M) of trifluoroacetic anhydride was prepared, and 0.3 mL was added to a pre-weighed vial containing a stir bar and PhB(OH)₂ (36 mg, 0.30 mmol, 1.0 equiv). The solution was diluted to 0.4 mL with CDCl₃, sealed with a Teflon-lined cap, and stirred at room temperature for 1 h. The reaction solution was transferred to an NMR tube and analyzed by ¹³C NMR spectroscopy. (¹³C rather than ¹⁹F NMR was used, because the former allows more clear differentiation of the CF₃-containing products, *vide infra*.) As shown in Figure S2, trifluoroacetic acid (B) is the major CF₃-containing product formed under these conditions. Conducting the experiment in THF or toluene with analysis by ¹⁹F NMR spectroscopy led to inconclusive results due to the poor resolution between TFAAn and TFA in the ¹⁹F NMR spectra.
- C. Trifluoroacetic anhydride with phenylboronic acid neopentylglycol ester. A CDCl₃ solution (1.0 M) of trifluoroacetic anhydride was prepared, and 0.3 mL was added to a pre-weighed vial containing a stir bar and phenylboronic acid neopentylglycol ester (54 mg, 0.30 mmol). The solution was diluted to 0.4 mL with CDCl₃, sealed with a Teflon-lined cap, and stirred at room temperature for 1 h. The solution was transferred to an NMR tube, and ¹³C NMR analysis was conducted. After 1 h or 3 h, ¹³C NMR spectroscopic analysis showed that neither A nor B was formed (Figure S2). Conducting the experiment in THF or toluene followed by analysis via ¹⁹F NMR

spectroscopy led to inconclusive results due to the poor resolution between TFAAn and TFA in the ¹⁹F NMR spectra.



Figure S2. Compatibility experiments of organoboron nucleophiles with trifluoroacetic anhydride after 1 h at room temperature in CDCl₃ as shown by ¹³C NMR spectroscopy.

D. Trifluoroacetic anhydride with phenylboronic acid neopentylglycol ester at 50 °C. A CDCl₃ solution (1.0 M) of trifluoroacetic anhydride was prepared, and 0.3 mL was added to a pre-weighed vial containing a stir bar and phenylboronic acid neopentylglycol ester (54 mg, 0.30 mmol). The solution was diluted to 0.4 mL with CDCl₃, sealed with a Teflon-lined cap, and stirred at 50 °C for 1 h. The reaction was cooled to room temperature. The solution was transferred to an NMR tube and ¹³C NMR analysis was conducted, showing a small amount of trifluoroacetic acid B (Figure S3).



Figure S3. ¹³C NMR spectrum showing minor amount of decomposition of trifluoroacetic anhydride to trifluoroacetic acid in the presence of phenylboronic acid neopentylglycol ester after 1 h at 50 °C in CDCl₃.

E. Trifluoroacetic anhydride with phenylboronic acid pinacol ester. A CDCl₃ solution (1.0 M) of trifluoroacetic anhydride was prepared, and 0.3 mL was added to a pre-weighed vial containing a stir bar and phenylboronic acid pinacol ester (61 mg, 0.30 mmol). The solution was diluted to 0.4 mL with CDCl₃, sealed with a Teflon-lined cap, and stirred at room temperature for 1 h. The mixture was transferred to an NMR tube and ¹³C NMR spectroscopic analysis was conducted. Neither A nor B was detected (Figure S4).



Figure S4. Compatibility of phenylboronic acid pinacol ester with trifluoroacetic anhydride after 1 h at room temperature in CDCl₃ as shown by ¹³C NMR spectroscopy.

- F. Difluoroacetic anhydride with phenyl boronic acid. A CDCl₃ solution (1.0 M) of difluoroacetic anhydride was prepared, and 0.15 mL was added to a pre-weighed vial containing a stir bar and PhB(OH)₂ (18 mg, 0.15 mmol, 1.0 equiv). The solution was diluted to 0.4 mL with CDCl₃, sealed with a Teflon-lined cap, and stirred at room temperature for 1 h at room temperature. The solution was transferred to an NMR tube and ¹³C NMR analysis was conducted, which showed complete hydrolysis of DFAAn to form difluoroacetic acid (DFA) (Figure S5).
- G. Difluoroacetic anhydride with phenylboronic acid neopentylglycol ester. A CDCl₃ solution (1.0 M) of difluoroacetic anhydride was prepared, and 0.15 mL was added to a pre-weighed vial containing a stir bar and phenyl boronate ester 1b (26 mg, 0.15 mmol). The solution was diluted to 0.4 mL with CDCl₃, sealed with a Teflon-lined cap, and stirred at room temperature for 1 h at room temperature. The solution was transferred to an NMR tube, and ¹³C NMR analysis was conducted, which showed no detectable DFA (Figure S5).



Figure S5. Compatibility experiments of organoboron nucleophiles with difluoroacetic anhydride after 1 h at room temperature in CDCl₃ as shown by ¹³C NMR spectroscopy.

- H. Difluoroacetyl fluoride with phenyl boronic acid. A solution of PhB(OH)₂ (12 mg, 0.10 mmol, 1.0 equiv) in 0.4 mL THF was prepared. To this was added a THF solution of DFAF (0.035 mL, 2.85 M, 0.10 mmol, 1.0 equiv), and the reaction was sealed with a Teflon-lined cap and stirred at room temperature for 1 h. 4-fluorotoluene (0.05 mL, 2.0 M, 0.10 mmol, 1.0 equiv) was added as an internal standard, the solution was transferred to an NMR tube, and ¹⁹F NMR analysis was conducted. No DFAF was detected in solution, as shown in Figure S6. Instead, a doublet at –128.45 ppm appeared, which corresponds to the hydrolysis product DFA.
- I. Difluoroacetyl fluoride with phenylboronic acid neopentylglycol ester. A solution of PhBneo (19 mg, 0.10 mmol, 1.0 equiv) in 0.4 mL THF was prepared. To this was added a THF solution of DFAF (0.035 mL, 2.85 M, 0.10 mmol, 1.0 equiv), and the reaction was sealed with a Teflon-lined cap and stirred at room temperature for 1 h. 4-fluorotoluene (0.05 mL, 2.0 M, 0.10 mmol, 1.0 equiv) was added as an internal standard, the solution was transferred to an NMR tube, and ¹⁹F NMR analysis was conducted. Less than 3% DFA was observed in solution, as shown in Figure S6.



Figure S6. Compatibility experiments of organoboron nucleophiles with DFAF after 1 h at room temperature in THF as shown by ¹⁹F NMR spectroscopy. NMR spectrum also contains trifluoromethoxybenzene and 4-fluorotoluene as internal standards. Referenced to 4-fluorotoluene as internal standard (-119.85 ppm).

IV. Procedure and spectral data for stoichiometric decarbonylation studies [Figures 1 and 2 in manuscript]

A. Stoichiometric reaction of trifluoroacetic anhydride with Pd/SPhos. A solution of trifluoroacetic anhydride (0.4 mL, 0.04 mmol, 0.1 M, 4 equiv) in anhydrous tetrahydrofuran was prepared. To this was added a THF-solution of 4-fluorotoluene (0.4 mL, 0.04 mmol, 0.1 M, 4.0 equiv). Pd[P(o-Tol)₃]₂ (21.3 mg, 0.03 mmol, 3 equiv) and SPhos (12.3 mg, 0.03 mmol, 3 equiv) were suspended in anhydrous THF (0.3 mL), and this mixture was stirred vigorously for 5 min. To the THF-suspension of Pd and SPhos was added 0.6 mL of the anhydride solution to generate a solution with 0.9 mL of total volume. The remaining 0.2 mL of anhydride solution was diluted and used as a ¹⁹F NMR reference (spectrum A in Figure S7). The reaction mixture was stirred briefly to ensure homogeneity and was then portioned evenly into three vials (*i, ii, iii*). A ¹⁹F NMR spectrum of sample *i* was acquired after 15 min and showed 98% yield of I-COCF₃ (Figure S7, B). A ¹⁹F NMR spectrum of sample *ii* was acquired after 4 h (Figure S7, C) and lacked the minor TFAAn peak still observed in the spectrum collected after 15 min, with no detectable II-CF₃. Sample *iii* was heated to 90 °C for 30 min and then analyzed by ¹⁹F NMR spectroscopy, which showed the formation of II-CF₃ in 90% yield (Figure S7, D). The associated

spectra are shown in Figure S7. The complexes were identified by ¹⁹F NMR spectroscopy based on the diagnostic locations of the observed peaks, which are in excellent agreement with our previous report^{1a} of an analogous transformation at (RuPhos)Pd(0).



Figure S7. ¹⁹F NMR spectra associated with oxidative addition and carbonyl de-insertion of trifluoroacetic anhydride at Pd[P(*o*-Tol)₃]₂/SPhos. (A) TFAAn; (B) 0.25 h, RT; (C) 4 h, RT; and (D) 0.5 h, 90 °C. Referenced to 4-fluorotoluene as internal standard (-119.85 ppm).

In situ NMR characterization:

TFAAn: ¹⁹F NMR (376 MHz) δ -76.73 (s, 6F)

I-COCF₃: ¹⁹F NMR (376 MHz) δ -73.94 (s, 3F), -75.61 (s, 3F).

II-CF₃: ¹⁹**F NMR** (376 MHz) δ -11.60 (bs, 3F), -75.69 (s, 3F).

B. Stoichiometric decarbonylation of difluoroacetic anhydride with Pd/SPhos. A solution of difluoroacetic anhydride (0.4 mL, 0.04 mmol, 0.1 M, 4 equiv) in anhydrous tetrahydrofuran was prepared. To this was added a THF solution of 4-fluorotoluene (0.4 mL, 0.04 mmol, 0.1 M, 4.0 equiv). Pd[P(o-Tol)₃]₂ (21.3 mg, 0.03 mmol, 3 equiv) and SPhos (12.3 mg, 0.03 mmol, 3 equiv) were suspended in anhydrous THF (0.3 mL), and this mixture stirred vigorously for 5 min. To the THF suspension of Pd and SPhos was added 0.6 mL of the anhydride solution. The remaining 0.2 mL of anhydride solution was diluted and used as a ¹⁹F NMR reference (spectrum A in Figure S8). The Pd solution containing internal standard, ligand, and anhydride was stirred briefly to ensure homogeneity and then portioned evenly into three vials (*i*, *ii*, *iii*). A ¹⁹F NMR spectrum was acquired of sample *i* after 15 min at room temperature and showed 85% yield of I-COCHF₂ and 13% yield of II-CF₃ (Figure S8, B). A ¹⁹F NMR spectrum was acquired of sample *iii* after 10 h at room temperature and showed 91% yield of II-CHF₂ and 6% remaining I-COCHF₂ (Figure S8, C).



Figure S8. ¹⁹F NMR spectra associated with oxidative addition and decarbonylation of difluoroacetic anhydride at Pd[P(*o*-Tol)₃]₂/SPhos in THF. (A) DFAAn; (B) 0.25 h, RT; and (C) 10 h, RT. Black star represents 4-fluorotoluene internal standard (-119.85 ppm).

The spectra show a dramatically lower barrier for carbonyl de-insertion at **I-COCHF**₂ compared to **I-COCF**₃. After 10 h stirring at room temperature, **II-CHF**₂ is obtained in 91% yield with 6% acyl complex remaining based on the ¹⁹F NMR spectrum. This is in sharp contrast to the temperature requirement for carbonyl de-insertion of **I-COCF**₃. Notably,

reacting Pd[P(o-Tol)₃]₂/SPhos with difluoroacetic anhydride in an NMR tube without stirring led to significantly lower rates of carbonyl de-insertion.

C. Stoichiometric reaction of pentafluoropropionic anhydride (PFPAn) with Pd/SPhos. A THF-solution of pentafluoropropionic anhydride (0.4 mL, 0.05 mmol, 0.125 M, 5 equiv) was prepared. To this was added a THF-solution of 4-fluorotoluene (0.4 mL, 0.04 mmol, 0.1 M, 4 equiv). Pd[P(o-Tol)₃]₂ (21.3 mg, 0.03 mmol, 3 equiv) and SPhos (12.3 mg, 0.03 mmol, 3 equiv) were weighed in a separate 4-mL vial with a stirbar, 0.6 mL THF were added, and the mixture was stirred. To the stirred mixture of Pd and SPhos in THF was added 0.6 mL of the anhydride/4-fluorotoluene solution to generate a solution with a total volume of 1.2 mL. The remaining 0.2 mL of anhydride/4-fluorotoluene solution was diluted with THF and used as a ¹⁹F NMR reference (spectrum A in Figure S9). The reaction mixture was stirred briefly to ensure homogeneity and was then portioned evenly (0.4 mL) into three vials (*i*, *ii*, *iii*). A ¹⁹F NMR spectrum of sample *i* was acquired after 0.25 h and shows conversion of PFPAn to I-COCF2CF3 (Figure S9, B). Sample ii was also analyzed by ³¹P NMR spectroscopy (Figure S10, A). A ¹⁹F NMR spectrum of sample *ii* was acquired after 4 h and showed no significant change to the ¹⁹F NMR spectrum obtained from sample I (Figure S9, C). Sample iii was heated to 90 °C for 30 min and then analyzed by ¹⁹F NMR spectroscopy (Figure S9, D) and ³¹P spectroscopy (Figure S10, B), which support the quantitative conversion of I-COCF2CF3 to II-CF2CF3. The complexes were characterized in situ by ¹⁹F NMR and ³¹P NMR spectroscopy based on the diagnostic locations of the observed peaks^{1a}.



Figure S9. ¹⁹F NMR spectra associated with oxidative addition and decarbonylation of pentafluoropropionic anhydride at Pd[P(*o*-Tol)₃]₂/SPhos in THF. (A) PFPAn; (B) 15 min, RT; (C) 4 h, RT; and (D) 0.5 h, 90 °C. Black star represents 4-fluorotoluene internal standard (-119.85 ppm).



Figure S10. ³¹P NMR spectra associated with oxidative addition and decarbonylation of pentafluoropropionic anhydride at Pd[P(*o*-Tol)₃]₂/SPhos in THF. (A) 0.25 h, RT; (B) 0.5 h, 90 °C.

In situ NMR characterization:

PFPAn: ¹⁹F NMR (376 MHz) δ -84.07 (s, 6F), -123.33 (s, 4F).

I-COCF₂CF₃: ¹⁹F NMR (376 MHz) δ -80.96 (s, 3F), -83.28 (s, 3F), -112.00 (s, 2F), -119.25 (s, 2F). ³¹P NMR (162 MHz) δ 46.04 (s, 1P).

II-CF₂CF₃: ¹⁹F NMR (376 MHz) δ -76.49 (bs, 2F), -79.14 (s, 3F), -88.48 (s, 3F), -119.91* (s, 2F). ³¹P NMR (162 MHz) δ 52.98 (bs, 1P).

The data in Figures S9 and S10 show that the reactivity of PFPAn nearly identical to that observed for TFAAn (i.e. fast oxidative addition and slow carbonyl de-insertion at room temperature). Notably, both TFAAn and PFPAn lack the acidic hydrogen found in DFAAn and thus require high temperatures for carbonyl de-insertion.

V. Synthesis, isolation, and spectral data for complex II-CHF₂.



Synthesis of complex II-CHF2. A 20 mL vial equipped with a stir bar was charged with Pd[P(o-Tol)3]2 (340 mg, 0.48 mmol, 1.0 equiv), SPhos (195 mg, 0.48 mmol, 1.0 equiv), and THF (4 mL). To this stirring suspension was added difluoroacetic anhydride (87 mg, 0.5 mmol, 1.05 equiv). The reaction was stirred for 18 h at room temperature, then concentrated to ca. 1 mL in vacuo to yield a dark yellow oil. The oil was loaded onto a 2 cm tall celite plug in a disposable fritted funnel and eluted with THF (8-10 mL). The yellow filtrate was concentrated in vacuo, yielding a yellow, oily residue. After addition of pentanes, a pale-yellow precipitate formed. The yellow suspension was loaded onto a 2 cm tall celite plug in a disposable fritted funnel, and the solid was washed with diisopropyl ether (10 mL), then a minimal amount of cold anhydrous Et₂O. The pale solid was eluted with tetrahydrofuran, and the resultant yellow filtrate was concentrated in vacuo, yielding II-CHF₂ as an off-white solid (195 mg, 0.29 mmol, 61% yield) containing about 0.2 equiv of tetrahydrofuran, as determined by ¹H NMR spectroscopy. ¹⁹F NMR (470 MHz, Methylene Chloride-d₂) δ –75.29 (t, J = 46.1 Hz), -124.38 (d, J = 57.0 Hz). ³¹P NMR (202 MHz, Methylene Chloride-d₂) δ 46.05 (t, J = 37.6 Hz). ¹H NMR (700 MHz, Methylene Chloride-d₂) δ 7.73 (t, J = 7.1 Hz, 1H), 7.58-7.38 (multiple peaks, 3H), 6.78 (ddd, J = 7.6, 3.0, 1.4 Hz, 1H), 6.57 (d, J = 8.5 Hz, 2H), 6.26 (td, J = 53.0, 5.1 Hz, 1H), 5.60 (t, J = 55.7 Hz, 1H), 3.80 (s, 6H), 2.33-2.23 (m, 2H), 2.13 (d, J = 8.1 Hz, 2H), 1.95-1.76 (multiple peaks, 6H), 1.74-1.55 (multiple peaks, 4H), 1.41-1.16 (multiple peaks, 6H). ¹³C NMR (176 MHz, Methylene Chloride-d₂) δ 167.24 (t, J = 24.4 Hz), 161.94, 144.06 (d, J = 17.5 Hz), 137.15, 135.32 (d, J = 42.3 Hz), 132.58, 132.08, 132.01 (d, J = 3.0 Hz), 127.49 (d, J = 6.0 Hz), 119.50 (td, J = 314.1, 14.1 Hz), 110.20 (t, J = 249.6 Hz), 56.38, 35.85 (d, J = 26.9 Hz), 29.50, 29.16 (d, J = 1.8 Hz), 27.87 (dd, J = 29.1, 12.8 Hz), 26.62 (d, J = 1.7 Hz). HRMS (ESI+) calcd. for C₂₉H₃₇F₄O₄PPd [M+H] *m*/z 662.1400. Parent – OCOCHF₂ *m*/z 567.1456. Found 567.1470. X-ray quality crystals of II-CHF₂ were obtained by vapor diffusion of Et₂O /pentanes into a THF solution of II-CHF₂ at room temperature. An ORTEP diagram of II-CHF₂ is shown with select hydrogen atoms are omitted for clarity. Selected bond lengths (Å) and angle (deg): O3-Pd1 2.11, O4-Pd1 3.09, C29-Pd1 1.99, C1-Pd1 2.46; H29---O3 2.38; C29–Pd1–O3 81.7, C29–H29---O3 96.9. See section XVI for X-ray crystal structure data of II-CHF2.



Figure S11. ESI+ Scans for HRMS analysis of II-CHF₂–OCOCHF₂.

VI. Procedure and spectral data for stoichiometric transmetalation studies [Figures 6 and 7 in manuscript]

General procedure for the stoichiometric transmetalation of complex II-CHF² with various aryl boron-based *nucleophiles.* A 4 mL vial equipped with a stir bar was charged with **II-CHF**² (19.9 mg, 0.03 mmol, 3.0 equiv) and anhydrous tetrahydrofuran (0.6 mL). To the stirred solution, a THF solution of 4-fluorotoluene (0.3 mL, 0.1 M, 0.03 mmol, 3 equiv) was added. The solution was divided into three separate solutions (0.3 mL each), each in a 4 mL vial equipped with a stir bar. The first solution was transferred to a screw-cap NMR tube and used for ¹⁹F NMR analysis (t = 0). To each of the remaining solutions was added one of the following nucleophiles: 4-phenyl boronic acid **1a**, 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzonitrile, **1b** (2.2 mg, 0.01 mmol, 1.0 equiv) or 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzonitrile, **1b** (2.2 mg, 0.01 mmol, 1.0 equiv) or 4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzonitrile, **1b** (2.1 mg, 0.01 mmol, 1.0 equiv). Each solution was stirred for 0.25 h, transferred to a screw cap NMR tube, and analyzed via ¹⁹F NMR spectroscopy. The spectra are shown in Figure S12. After 0.25 h, 40% of product **1** was observed with the reaction with 45% of **II-CHF**² remaining. Trace product **1** was observed using neopentyl boronate ester **1b**, whereas no detectable product was observed in the reaction between **II-CHF**² and pinacol

boronate ester **1c.** Notably, the Pd–aryl intermediate is not observed, only the organic product **1** was observed at - 114.2 ppm (d, J = 56.4 Hz) by ¹⁹F NMR spectroscopy as result of transmetalation and reductive elimination at room temperature.



Figure S12. ¹⁹F NMR spectra associated with transmetalation of II-CHF₂ with (B) boronic acid 1a, (C) boronic ester 1b, and (D) boronic ester 1c.

General procedure for the stoichiometric transmetalation at II-CHF₂ with added NMe₄F. A 4 mL vial equipped with a stir bar was charged with **II-CHF**₂ (6.6 mg, 0.01 mmol, 0.5 equiv) and anhydrous tetrahydrofuran (0.2 mL). To this was added a THF solution of 4-fluorotoluene (0.1 mL, 0.1 M, 0.01 mmol, 1.0 equiv) then solid NMe₄F•^tAmyIOH (3.6 mg, 0.02 mmol, 2.0 equiv).^{1b} The suspension was stirred for 0.5 h, filtered through a syringe filter, then transferred to a screw cap NMR tube, sealed with a Teflon-lined cap, and analyzed by ¹⁹F NMR spectroscopy. The formation of a distinct Pd-F intermediate in 12% yield is observed, based on a diagnostic resonance at –349.5 ppm.² Moreover, several new peaks appear in the Pd-CHF₂ region (–86 to –93 ppm). Upon addition of substrate **1b** (2.2 mg, 0.01 mmol, 1.0 equiv) to this sample, the Pd–F intermediate disappears, and organic product **1** was formed in 27% yield (shown below in Figure S13).



Figure S13. ¹⁹F NMR spectra associated with fluoride-aided transmetalation of II-CHF₂ with substrate 1b.



Figure S14. ¹⁹F NMR spectra associated with fluoride-aided transmetalation of II-CHF₂ with substrates 1b, 19b, and 20b.

Transmetalation with 19b. The general procedure was followed resulting in 4% of Pd–F observed by ¹⁹F NMR with 4-fluorotoluene internal standard. After addition of substrate **19b** (2.2 mg, 0.01 mmol, 1.0 equiv) to this sample, the Pd–F signal was consumed and organic product **19** was formed in 26% yield by ¹⁹F NMR (Figure S14, D). Characterized product **19** in situ: ¹⁹F NMR (376 MHz) δ -108.96 (d, *J* = 56.7 Hz, 2F).

Transmetalation with 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane. The general procedure was followed resulting in 4% of Pd–F observed by ¹⁹F NMR with 4-fluorotoluene internal standard. After addition of 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane (2.0 mg, 0.01 mmol, 1.0 equiv) to this sample, the Pd–F signal was consumed and 1-(difluoromethyl)-4-fluorobenzene was formed in 30% yield by ¹⁹F NMR. The remaining mass balance is 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane (70%) observed at -111.47 ppm in the ¹⁹F NMR (Figure S14, E). Characterized 1-(difluoromethyl)-4-fluorobenzene in situ: ¹⁹F NMR (376 MHz) δ -110.47 (d, *J* = 56.4 Hz, 2F), -111.35 (m, 1F).

VII. Procedure for the synthesis of aryl neopentyl boronate esters



*General procedure for the synthesis of aryl neopentyl boronate esters from aryl boronic acids (Method A)*³*:* Under ambient conditions, the respective aryl boronic acid (2.0 mmol, 1.0 equiv), 2,2-dimethyl-1,3-propanediol (208 mg, 2.0 mmol, 1.0 equiv), and anhydrous magnesium sulfate (240 mg, 2.0 mmol, 1.0 equiv) were weighed into a 20 mL vial. The vial was equipped with a magnetic stirbar, and Et₂O (10 mL) was added. The vial was sealed with a Teflonlined screw cap, and the resulting suspension was stirred under ambient conditions for 18 h. The reaction mixture was then filtered through a 3 cm x 3 cm x 3 cm celite plug set with Et₂O. The celite plug was washed with Et₂O (100 mL). Solvent was removed *in vacuo* to afford the corresponding aryl neopentyl boronate ester as a white solid for use in catalysis without further purification, unless stated otherwise.³



*General procedure for synthesis of aryl neopentyl boronate esters from aryl bromides (Method B)*⁴*:*</sup> To a 20 mL vial equipped with a magnetic stirbar was added PdCl₂(DPPF) (58 mg, 0.08 mmol, 0.04 equiv), anhydrous potassium acetate (589 mg, 6.0 mmol, 3.0 equiv), bis(neopentyl glycolato)diboron (542 mg, 2.4 mmol, 1.2 equiv), and aryl bromide (2.0 mmol, 1.0 equiv). To these solids was added anhydrous dimethyl sulfoxide (14 mL), forming a red-orange

suspension. The reaction was sealed with a Telfon-lined cap, removed from the glovebox, and heated at 80 °C for 18 h. The reaction mixture was allowed to cool to room temperature, then poured into ice-cold deionized water (75 mL). The mixture was extracted with ethyl acetate (3 x 40 mL), and the combined organic extracts were washed with saturated solution of aqueous NaCl (50 mL), dried over anhydrous sodium sulfate, and concenrated *in vacuo*. The crude residue was dissolved in minimal dichloromethane and purified via flash chromatography on silica gel using ethyl acetate/hexanes gradient elution (5-50%). Removal of solvent afforded the aryl neopentyl boronate ester as a white solid to be used in catalysis without further purification, unless otherwise stated.⁴ The ¹³C NMR signal corresponding to the carbon of the C–B bond for all boronate esters below is not observed due to broadening.⁵



4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzonitrile (1b). Method A was followed and yielded a white solid (427 mg, 99% yield). ¹H NMR (700 MHz, CDCl₃) δ 7.87 (d, *J* = 7.7 Hz, 2H), 7.62 (d, *J* = 7.7 Hz, 2H), 3.78 (s, 4H), 1.03 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 134.36, 131.16, 119.26, 114.07, 72.57, 32.05, 21.98. **HRMS** (positive ion GC-APCI) calcd. for C₁₂H₁₄BNO₂ [M+H] *m*/z 216.1190. Found 216.1194.



4-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzonitrile (1c). Method A was followed, except 2,3-dimethylbutane-2,3-diol (1.0 equiv) was used instead of 2,2-dimethyl-1,3-propanediol. The aryl pinacol boronate ester was obtained as white solid (455 mg, 99% yield). ¹H NMR (700 MHz, CDCl₃) δ 7.88 (d, J = 7.9 Hz, 2H), 7.63 (d, J = 7.9 Hz, 2H), 1.35 (s, 12H). ¹³C NMR (176 MHz, CDCl₃) δ 135.22, 131.25, 118.99, 114.67, 84.62, 25.00. HRMS (positive ion GC-APCl) calcd. for C₁₃H₁₆BNO₂ [M+H] *m/z* 230.1347. Found 230.1354.



4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-2-methylbenzonitrile (2b). Method B was followed and yielded a white solid (302 mg, 66% yield). ¹H NMR (401 MHz, CDCl₃) δ 7.73 (s, 1H), 7.66 (d, *J* = 7.7 Hz, 1H), 7.56 (d, *J* = 7.7 Hz, 1H), 3.78 (s, 4H), 2.54 (s, 3H), 1.02 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 140.73, 135.52, 131.60, 131.44, 118.55, 114.49, 72.57, 32.05, 21.99, 20.49. HRMS (positive ion GC-APCI) calcd. for C₁₃H₁₆BNO₂ [M+H] *m/z* 230.1374. Found 230.1352.

(4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)phenyl)(phenyl)methanone (3b). Method A was followed and yielded a white solid (560 mg, 95% yield). ¹H NMR (401 MHz, CDCl₃) δ 7.91 (d, *J* = 8.2 Hz, 2H), 7.85-7.72 *(multiple peaks,* 4H), 7.62-7.55 (m, 1H), 7.48 (t, *J* = 7.6 Hz, 2H), 3.80 (s, 4H), 1.05 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 197.22, 139.42, 137.83, 133.80, 132.54, 130.25, 129.12, 128.39, 72.56, 32.07, 22.05. HRMS (ESI+) calcd. for C₁₈H₁₉BO₃ [M+H] *m/z* 295.1500. Found 295.1503.



1-(4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)phenyl)butan-1-one (4b). Method B was followed and yielded a white solid (422.6 mg, 81% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.13-7.76 (*multiple peaks*, 4H), 3.78 (s, 4H), 2.95 (t, *J* = 7.3 Hz, 2H), 1.81-1.72 (m, 2H), 1.08-0.98 (*multiple peaks*, 9H). ¹³C NMR (126 MHz, CDCl₃) δ 201.00, 138.77, 134.13, 127.09, 72.53, 40.84, 32.05, 22.03, 17.93, 14.04. HRMS (positive ion GC-APCI) calcd. for C₁₅H₂₁BO₃ [M+H] *m/z* 261.1657. Found 261.1661.



2-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)anthracene-9,10-dione (5b). Method B was followed and yielded a yellow solid (419 mg, 65% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.77-8.71 (m, 1H), 8.37-8.28 (*multiple peaks*, 2H), 8.27 (d, *J* = 7.6 Hz, 1H), 8.20 (dd, *J* = 7.7, 1.3 Hz, 1H), 7.83-7.75 (*multiple peaks*, 2H), 3.83 (s, 4H), 1.05 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 183.71, 183.51, 139.44, 134.91, 134.20, 134.07, 133.82, 133.78, 133.06, 132.54, 127.38, 127.29, 126.20, 72.62, 32.10, 22.04. HRMS (ESI+) calcd. for C₁₉H₁₇BO₄ [M+H] *m/z* 321.1293. Found 321.1306.



ethyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate (6b). Method A was followed and yielded a white solid (471 mg, 90% yield). ¹H NMR (401 MHz, CDCl₃) δ 8.01 (d, *J* = 8.2 Hz, 2H), 7.86 (d, *J* = 8.2 Hz, 2H), 4.38 (q, *J* = 7.1 Hz, 2H), 3.78 (s, 4H), 1.40 (t, *J* = 7.1 Hz, 3H), 1.03 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 166.99, 133.86, 132.32, 128.60, 72.53, 61.08, 32.05, 22.05, 14.49. HRMS (positive ion GC-APCI) calcd. for C₁₄H₁₉BO4 [M+] *m/z* 263.1449. Found 263.1454.



methyl 4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)benzoate (7b). Method A was followed and yielded a white solid (468 mg, 94% yield). ¹H NMR (401 MHz, CDCl₃) δ 8.01 (d, J = 8.2 Hz, 2H), 7.86 (d, J = 8.2 Hz, 2H), 3.91 (s, 3H), 3.78 (s, 4H), 1.03 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 167.46, 133.90, 131.95, 128.64, 72.53, 52.22, 32.04, 22.04. HRMS (positive ion GC-APCI) calcd. for C₁₃H₁₇BO₄ [M+H] *m/z* 249.1293. Found 249.1298.



diethyl 5-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)isophthalate (8b). Method A was followed and yielded a white solid (662 mg, 99% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.73 (s, 1H), 8.62 (s, 2H), 4.41 (q, *J* = 7.4 Hz, 4H), 3.80 (s, 4H), 1.41 (t, *J* = 7.5 Hz, 6H), 1.03 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 166.27, 139.11, 132.89, 130.33, 72.54, 61.31, 32.08, 22.01, 14.52. HRMS (positive ion GC-APCI) calcd. for C₁₇H₂₃BO₆ [M+H] *m/z* 335.166. Found 335.1164.



2-(4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)phenyl)benzo[d]thiazole (9b). Method B was followed and yielded a white solid (595 mg, 91% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.08-7.90 (*multiple peaks*, 3H), 7.95-7.87 (*multiple peaks*, 3H), 7.49 (m, *J* = 7.7 Hz, 1H), 7.39 (t, *J* = 8.1 Hz, 1H), 3.80 (s, 4H), 1.05 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 168.42, 154.33, 135.48, 135.27, 134.62, 126.75, 126.45, 125.36, 123.43, 121.77, 72.55, 32.08, 22.08. HRMS (positive ion GC-APCI) calcd. for C₁₈H₁₈BNO₂S [M+H] *m/z* 324.1224. Found 324.1234.



3-(4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)phenyl)-5-methyl-1,2,4-oxadiazole (10b). Method B was followed and yielded a white solid (418 mg, 77% yield). ¹H NMR (401 MHz, CDCl₃) δ 8.04 (d, *J* = 8.1 Hz, 2H), 7.90 (d, *J* = 8.1 Hz, 2H), 3.79 (s, 4H), 2.65 (s, 3H), 1.04 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 176.63, 168.68, 134.42, 128.71, 126.51, 72.52, 32.06, 22.06, 12.56. HRMS (positive ion GC-APCI) calcd. for C₁₄H₁₇BN₂O₃ [M+H] *m/z* 273.1405. Found 273.1408.



2-(4-(isopropyIsulfonyI)phenyI)-5,5-dimethyI-1,3,2-dioxaborinane (11b). Method A was followed and yielded a white solid (569 mg, 96% yield). ¹H NMR (401 MHz, CDCl₃) δ 7.96 (d, J = 8.3 Hz, 2H), 7.82 (d, J = 8.3 Hz, 2H), 3.78 (s, 4H), 3.17 (hept, J = 6.9 Hz, 1H), 1.26 (d, J = 6.9 Hz, 6H), 1.02 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 138.59, 134.42, 127.97, 72.54, 55.61, 32.01, 21.95, 15.78. **HRMS** (positive ion GC-APCI) calcd. for C₁₄H₂₁BO₄S [M+NH₄] *m/z* 342.1592. Found 342.1909.



4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-N,N-dimethylbenzenesulfonamide (12b). Method A was followed and yielded a white solid (517 mg, 87% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.95 (d, J = 8.1 Hz, 2H), 7.81-7.69 (m, 2H), 3.79 (s, 4H), 2.68 (s, 6H), 1.03 (s, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 137.09, 134.46, 126.75, 72.57, 38.08, 32.05, 21.99. HRMS (positive ion GC-APCI) calcd. for C₁₃H₂₀BNO₄S [M+H] m/z 298.1279. Found 298.1292.



4-(5,5-dimethyl-1,3,2-dioxaborinan-2-yl)-N,N-dipropylbenzenesulfonamide (13b). Substrate was synthesized and isolated according to a previously reported procedure⁶ and yielded a white solid (80.5 mg, 76% yield). ¹H NMR (401 MHz, CDCl₃) δ 7.90 (d, J = 8.3 Hz, 2H), 7.76 (d, J = 8.3 Hz, 2H), 3.78 (s, 4H), 3.10-3.03 (m, 4H), 1.58-1.48 (m, 4H), 1.03 (s, 6H), 0.86 (t, J = 7.4 Hz, 6H). ¹³C NMR (176 MHz, CDCl₃) δ 141.92, 134.46, 126.08, 72.57, 50.12, 32.06, 22.11, 22.03, 11.34. HRMS (positive ion GC-APCI) calcd. for C₁₇H₂₈BNO₄S [M+H] *m/z* 354.1905. Found 354.1905.



5,5-dimethyl-2-(3-(methylsulfonyl)-5-(trifluoromethyl)phenyl)-1,3,2-dioxaborinane (14b). Method B was followed and yielded a white solid (544 mg, 81% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.54 (s, 1H), 8.31 (s, 1H), 8.25 (s, 1H), 3.81 (s, 4H), 3.09 (s, 3H), 1.04 (s, 6H). ¹⁹F NMR (377 MHz, CDCl₃) δ –62.90. ¹³C NMR (176 MHz, CDCl₃) δ 140.98, 135.97, 135.69 (q, *J* = 3.3 Hz), 131.40 (q, *J* = 33.3 Hz), 126.35 (q, *J* = 3.7 Hz), 123.50 (q, *J* = 273.0 Hz), 72.66, 44.55, 32.14, 21.95. HRMS (positive ion GC-APCI) calcd. for C₁₃H₁₆BF₃O₄S [M+H] *m/z* 337.0887. Found 337.0898.



2-(3,5-bis(trifluoromethyl)phenyl)-5,5-dimethyl-1,3,2-dioxaborinane (15b). Method A was followed and yielded a white solid (639 mg, 98% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.24 (s, 2H), 7.91 (s, 1H), 3.81 (s, 4H), 1.04 (s, 6H). ¹⁹F NMR (377 MHz, CDCl₃) δ –62.87. ¹³C NMR (176 MHz, CDCl₃) δ 133.99, 130.79 (q, *J* = 32.9 Hz), 124.30, 123.82 (q, *J* = 272.5 Hz), 72.64, 32.13, 21.96. HRMS (positive ion GC-APCI) calcd. for C₁₃H₁₃BF₆O₂ [M+H] *m/z* 327.0986. Found 327.0985.



5,5-dimethyl-2-(4-(trifluoromethyl)phenyl)-1,3,2-dioxaborinane (16b). Method A was followed and yielded a white solid (485 mg, 94% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.90 (d, *J* = 7.7 Hz, 2H), 7.60 (d, *J* = 7.6 Hz, 2H), 3.79 (s, 4H), 1.03 (s, 6H). ¹⁹F NMR (377 MHz, CDCl₃) δ –62.93. ¹³C NMR (176 MHz, CDCl₃) δ 134.24, 132.41 (q, *J* = 31.9 Hz), 124.44 (q, *J* = 272.2 Hz), 124.31 (q, *J* = 3.7 Hz), 72.54, 32.05, 22.00. **HRMS** (positive ion GC-APCI) calcd. for C₁₂H₁₄BF₃O₂ [M+H] *m/z* 259.1112. Found 259.1120.



2-(3,4-difluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane (17b). Method A was followed and yielded a yellow solid (408 mg, 90% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.62-7.49 (*multiple peaks*, 2H), 7.12 (dt, *J* = 10.5, 7.9 Hz, 1H), 3.76 (s, 4H), 1.02 (s, 6H). ¹⁹F NMR (471 MHz, CDCl₃) δ –135.53 (dtd, *J* = 20.0, 9.5, 4.8 Hz), –140.35 (ddd, *J* = 19.8, 11.1, 7.7 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 152.43 (dd, *J* = 251.1, 12.7 Hz), 150.31 (dd, *J* = 248.0, 11.9 Hz), 130.45 (dd, *J* = 6.6, 3.7 Hz), 122.54 (d, *J* = 15.2 Hz), 116.76 (d, *J* = 16.2 Hz), 72.50, 32.04, 22.00. HRMS (positive ion GC-APCI) calcd. for C₁₁H₁₃BF₂O₂ [M+H] *m/z* 227.1049. Found 227.1043.



5,5-dimethyl-2-(4-(trifluoromethoxy)phenyl)-1,3,2-dioxaborinane (18b). Method A was followed and yielded a white solid (521 mg, 95% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.83 (d, *J* = 8.5 Hz, 2H), 7.18 (d, *J* = 7.9 Hz, 2H), 3.77 (s, 4H), 1.02 (s, 6H). ¹⁹F NMR (471 MHz, CDCl₃) δ –57.84. ¹³C NMR (176 MHz, CDCl₃) δ 151.46, 135.71, 120.62 (q, *J* = 257.1 Hz), 119.86, 72.49, 32.03, 21.99. **HRMS** (positive ion GC-APCI) calcd. for C₁₂H₁₄BF₃O₃ [M+H] *m/z* 275.1061. Found 275.1066.



2-(4-methoxyphenyl)-5,5-dimethyl-1,3,2-dioxaborinane (19b). Method A was followed and yielded a white solid (406 mg, 93%). ¹H NMR (500 MHz, CDCl₃) δ 7.74 (d, *J* = 8.5 Hz, 2H), 6.89 (d, *J* = 8.6 Hz, 2H), 3.82 (s, 3H), 3.75 (s, 4H), 1.02 (s, 6H). ¹³C NMR (126 MHz, CDCl₃) δ 161.89, 135.65, 113.29, 72.41, 55.20, 32.05, 22.08. HRMS (positive ion GC-APCl) calcd. for C₁₂H₁₇BO₃ [M+] *m/z* 220.1271. Found 220.1277.

VIII. Procedure for catalytic reactions with fluoroalkyl anhydrides and fluoride salts (Figure 8 in manuscript)



General procedure for the catalytic reaction with DFAAn and fluoride salts: $Pd[P(o-Tol)_3]_2$ (4.1 mg, 0.005 mmol, 0.1 equiv, 3.6 mg) and SPhos (4.1 mg, 0.01 mmol, 0.2 equiv) were combined in THF (0.15 mL). The yellow suspension was stirred vigorously in a tall 10 mL vial for 15 min. To this solution was added a toluene-solution of DFAAn (0.2 mL, 0.75 M, 0.15 mmol, 3 equiv), then **1b** (10.8 mg, 0.05 mmol, 1 equiv), and finally CsF, NMe₄F, or NBu₄F (0.175 mmol, 3.5 equiv). The vial was sealed with a Teflon-lined screw cap with a septum, removed from the glovebox, and heated to the appropriate temperature for 3 h. After 3 h, the reaction mixture was allowed to cool to room temperature, and then 4-fluorotoluene (25 µL, 2.0 M in DCM, 1.0 equiv) was added as an internal standard, followed by dichloromethane (1.0 mL). The mixture was filtered through a plug of celite, and an aliquot of the solution was transferred to an NMR tube and analyzed by ¹⁹F NMR spectroscopy to determine the yield of **1** as well as the identity of major by-products.

Conducting the reaction at room temperature with 3 equiv of DFAAn and 3.6 equiv of CsF produced no observable quantity of **1**. However, in the crude ¹⁹F NMR spectra of these reactions, DFAF was observed as a major side product. In the reactions with NMe₄F or NBu₄F, DFAF was not observed, and DFA was a major side product.

Reactions conducted at 130 °C using 3 equiv of DFAAn and 3.6 equiv CsF produced modest but variable yields of 1 (17-31%). The range of 17-31% represents four reactions set up under identical conditions, with yields of 15%, 17%, 24%, and 31%, respectively, emphasizing the poor reproducibility of this transformation. Using equimolar amounts of CsF to DFAAn under otherwise analogous conditions resulted in significantly lower yields of 1 (4-9%). Reactions conducted using NMe₄F or NBu₄F produced no observable quantity of 1 or DFAF in these experiments, as determined by ¹⁹F NMR spectroscopy.



General procedure for the catalytic reaction with TFAAn and CsF: $Pd[P(o-Tol)_3]_2$ (4.1 mg, 0.005 mmol, 0.1 equiv, 3.6 mg) and SPhos (4.1 mg, 0.01 mmol, 0.2 equiv) were combined in THF (0.15 mL). The yellow suspension was stirred vigorously in a tall 10 mL vial for 15 min. To this solution was added a toluene-solution of TFAAn (0.2 mL, 0.75 M, 0.15 mmol, 3 equiv), then **1b** (10.8 mg, 0.05 mmol, 1 equiv), and finally CsF, (26.6 mg, 0.175 mmol, 3.5 equiv). The vial was sealed with a Teflon-lined screw cap with a septum, removed from the glovebox, and heated to the appropriate temperature for 3 h. After 3 h, the reaction mixture was allowed to cool to room temperature, and then 4-fluorotoluene (25 µL, 2.0 M in DCM, 1.0 equiv) was added as an internal standard, followed by dichloromethane (1.0 mL). The mixture was filtered through a plug of celite, and an aliquot of the solution was transferred to an NMR tube and analyzed by ¹⁹F NMR spectroscopy to determine the yield of 4-trifluoromethylbenzonitrile, **1-CF**₃. Spiking in an authentic sample of **1-CF**₃ to the crude reaction mixture confirmed its presence in the crude reaction mixture. The associated ¹⁹F NMR spectra are shown below in Figure S15.



Figure S15. (Top) ¹⁹F NMR of crude reaction with 4-fluorotoluene internal standard showing 4trifluoromethylbenzonitrile, **1-CF**₃ and (bottom) ¹⁹F NMR of crude reaction after spiking in an authentic sample of **1-CF**₃.



General procedure for the catalytic reaction with PFPAn and CsF: $Pd[P(o-Tol)_3]_2$ (4.1 mg, 0.005 mmol, 0.1 equiv, 3.6 mg) and SPhos (4.1 mg, 0.01 mmol, 0.2 equiv) were combined in THF (0.15 mL) and toluene (0.2 mL). The yellow suspension was stirred vigorously in a tall 10 mL vial for 15 min. To this solution was added a PFPAn (46.5 mg, 0.15 mmol, 3 equiv), then **1b** (10.8 mg, 0.05 mmol, 1 equiv), and finally CsF, (26.6 mg, 0.175 mmol, 3.5 equiv). The vial was sealed with a Teflon-lined screw cap with a septum, removed from the glovebox, and heated to the appropriate temperature for 3 h. After 3 h, the reaction mixture was allowed to cool to room temperature, and then 4-fluorotoluene (25 μ L, 2.0 M in DCM, 1.0 equiv) was added as an internal standard, followed by dichloromethane (1.0 mL). The mixture was filtered through a plug of celite, and an aliquot of the solution was transferred to an NMR tube and analyzed by ¹⁹F NMR spectroscopy (Figure S16) to determine the yield of **1-CF₂CF₃**. Spectral data for this product in this reaction (¹⁹F)

NMR: -85.51 ppm (s, 3F), -116.55 ppm (s, 2F)) are consistent with our previous report⁷ and other reports⁸ in the literature. Pentafluoropropionic acid (PFPA) was observed (55%) in the crude reaction.



Figure S16. ¹⁹F NMR of crude reaction with 4-fluorotoluene internal standard indicating formation of 1-CF₂CF₃.

IX. Synthesis of DFAF solution in THF



General procedure for the distillation of difluoroacetyl fluoride. A 20 mL vial containing THF (7 mL) was placed in the freezer at –36 °C and cooled for 15 min. In a 4 mL vial, a solution of difluoroacetic anhydride (4.35 g, 25 mmol, 1.0 equiv) in tetrahydrofuran (1 mL) was prepared and cooled to –36 °C in the glovebox freezer for 15 min. While cooling, solid cesium fluoride (5.32 g, 35 mmol, 1.4 equiv) was added to a 25 mL round bottom flask equipped with a medium-sized stirbar. Cooled THF (5 mL) was added to the reaction flask. The cooled solution of anhydride in THF was then added to the flask containing CsF. The flask was quickly sealed with a rubber septum, removed from the glovebox, and allowed to stir at room temperature for 15 min. A short-path distillation apparatus equipped with nitrogen flow, water circulation, and a thermoprobe was fitted to a 25 mL round bottom collection flask cooled to 0 °C in an ice water bath.

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The reaction flask was quickly connected to the distillation apparatus and heated from 25 °C to 80 °C in a water bath over a period of 30-45 min. The distillation was determined complete when minimal solvent remained in the reaction flask, the temperature of the gas in the apparatus as determined by the thermoprobe dropped below 45 °C, and condensation into the collection flask slowed dramatically. Once complete, the collection flask containing the product in tetrahydrofuran was quickly sealed with a rubber septum, wrapped securely with black electrical tape, and shortcycled into the glovebox. In a 20 mL vial, trifluoromethoxybenzene (54 mg, 0.33 mmol, 1 equiv in F) was weighed as an internal standard, and a portion of the DFAF solution was added via syringe to the vial with total volume recorded. The trifluoromethoxybenzene was treated as approximately 0.05 mL in volume. After determining the total volume to be 5.4 mL and ensuring homogeneity, a sample of this solution was transferred to a screw cap NMR tube, diluted with 0.5 mL tetrahydrofuran, and analyzed via ¹⁹F NMR spectroscopy. The bulk solution was sealed with a Teflon-lined screw cap and stored in the glovebox freezer. The spectrum is shown in Figure S19 with DFAF characterized in situ. Trace difluoroacetic acid is observed in all cases. The concentration of DFAF was calculated as mmol DFAF divided by the volume (mL) of solution as determined by ¹⁹F NMR spectroscopy (Figure S17). A sample calculcation is shown under Figure S12. In some cases, 4-fluorotoluene (110 mg, 1 mmol, treated as 0.11 mL in volume) or fluorobenzene (96 mg, 1 mmol, treated at 0.095 mL in volume) was used as the internal standard to assess the yield from DFAF synthesis.



Figure S17. Representative ¹⁹F NMR spectrum obtained after synthesis and distillation of DFAF containing 0.33 mmol of trifluoromethoxybenzene internal standard. ¹⁹F NMR (377 MHz, THF) δ 18.82 (m), –128.89 (dd, *J* = 51.9, 8.9 Hz).

Sample calculation of DFAF concentration: [16.00 + (31.96/2)]/2 = 15.99 mmol DFAF 15.99 mmol / 5.4 mL = 2.96 M 15.99 mmol / 25 mmol = 64% yield

THF was identified to be the most suitable reaction solvent and distillation solvent due to the relatively low boiling point of THF and the high solubility of CsF in THF. While a higher boiling solvent, such as mesitylene or xylenes, could in principle enable us to raise the temperature of the catalytic reaction, the poor solubility of CsF in these solvents limited the acid fluoride generation step in thees media. Further, use of aromatic sovlents, such as toluene, introduces additional challenges in the co-distillation of low boiling DFAF in these high boiling solvents.

X. Optimization for catalytic aryl difluoromethylation with DFAF



General procedure for the optimization of catalytic decarbonylative aryl difluoromethylation: A Pd⁰ source (0.005 mmol, 0.1 equiv) was dissolved with a phosphine ligand (0.01 mmol, 0.2 equiv) in 0.2 mL of anhydrous co-solvent and 0.05 mL THF. The yellow suspension was stirred in a tall 10 mL vial until homogenous, then **1b** (10.8 mg, 0.05 mmol, 1 equiv) was added. To the resulting mixture was added a cold (–36 °C) solution of DFAF (0.085 mL, 0.25 mmol, 2.96 M) in anhydrous THF. The reaction mixture was then diluted with THF to a total volume of 0.35 mL. The vial was sealed with a Teflon-lined screw cap with a septum (Figure S18), removed from the glovebox, and heated to a given temperature for 3 h. After 3 h, the reaction mixture was allowed to cool to room temperature. To it was added 4-fluorotoluene (25 μ L, 2.0 M in DCM, 1.0 equiv) as an internal standard, followed by dichloromethane (1.0 mL). An aliquot of the solution was transferred to an NMR tube and analyzed by ¹⁹F NMR spectroscopy. Yields reported are an average of three runs.



Figure S18. Reaction solution in tall 10 mL vial before heating in THF/toluene.

NC 1b (0	.05 mmol	$F_{2}HC = F \frac{10 \text{ mo}}{20}$	I % Pd[P(o-Tol) ₃] ₂ mol % SPhos nditions, 3 h	CHF ₃
Entry	х	temperature (°C)	solvent(s)	Yield (%) ^a
1	1	25	THF	<1
2	1	100	THF	29
3	1	130	THF:toluene (1:1)	51
4	3	130	THF:toluene (1:1)	58
5	5	130	THF:toluene (1:1)	67
6	5	150	THF:toluene (1:1)	86
7	5	150	THF:dioxane (1:1)	72
8 ^b	5	150	THF:toluene (1:1)	93
9 ^b	5	150	THF:toluene (1:2)	92

^aYields determined by ¹⁹F NMR with internal standard

^bReaction was run for 5 h

Table S1. Optimization of reaction temperature, equivalents, and solvent, and time for catalytic decarbonylative difluoromethylation of neopentyl boronate ester **1b**.

NC 1b (0.05 mmol)	$F_{2}HC F = 5 equiv$ $10 \text{ mol\% Pd[P(o-Tol)_{3}]_{2}}{10 \text{ mol\% SPhos}} NC$	CHF ₂
Entry	Change of reaction conditions	Yield (%) ^a
1	none	86
2	RuPhos instead of SPhos	76
3	DavePhos instead of SPhos	78
4	XantPhos instead of SPhos	<1
5	BrettPhos instead of SPhos	36
6	$Pd(dba)_2$ instead of $Pd[P(oTol)_3]_2$	68
7	4-CNPhB(OH) ₂ instead of 1a	<1
8	4-CNPhBpin instead of 1a	29
9	10 mol% Pd[P(oTol) ₃] ₂ /10 mol% SPhos	12
10	5 mol% Pd[P(oTol) ₃] ₂ /10 mol% SPhos	38
12	no SPhos	<1
13	no Pd[P(o-Tol) ₃] ₂	<1

^aYields determined by ¹⁹F NMR with internal standard

 Table S2. Impact of catalyst, nucleophile, and ligand loading on reaction yields.



Procedure for determining mass balance of catalytic reaction using 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2dioxaborinane. In a 4-mL vial with stirbar, Pd[P(o-Tol)₃]₂ (21.5 mg, 0.03 mmol, 0.6 equiv) and SPhos (24.6 mg, 0.06 mmol, 1.2 equiv) were combined with 1.2 mL toluene and 0.3 mL THF. The mixture was stirred for fifteen minutes. In each of five tall 10 mL vials equipped with a stir bar, 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane (10 mg, 0.05 mmol, 1.0 equiv) was added. To each vial containing substrate 20b was added an aliquot (0.25 mL) of the Pd/SPhos mixture. A THF solution of difluoroacetyl fluoride (0.095 mL, 0.25 mmol, 5 equiv) was added via syringe to each vial, and the vials were sealed with Teflon-lined screw caps. The vials were removed from the glovebox and heated at 150 °C for 3 h. The reaction mixtures were allowed to cool to room temperature and then were combined. To the combined reaction mixture was added 4-fluorotoluene (125 µL, 2.0 M in DCM, 5.0 equiv) as an internal standard. An aliquot (0.4 mL) of the solution was transferred to an NMR tube and analyzed via ¹⁹F NMR spectroscopy (Figure S19). The crude NMR yield of 1-(difluoromethyl)-4-fluorobenzene after 3 h was 17% as determined by ¹⁹F NMR. After initial analysis, five additional NMR samples were prepared (0.4 mL each), and the following authentic standards were spiked in: 4fluorophenyl 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane, boronic acid, 4-fluorobenzene and 4,4'difluorobiphenyl. Then, ¹⁹F NMR analysis was conducted and the associated spectra are shown below in Figure S20.



Figure S19. ¹⁹F NMR spectrum obtained from crude reaction of 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane under optimized conditions. Reaction was ran for 3 h.

Figure S20. ¹⁹F NMR spectra obtained from spiking authentic samples into the crude reaction of 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane.

While the mass balance of the reaction cannot be totally accounted for, spiking experiments revealed the presence of 14% of 4-fluorobenzene and 43% of 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane remaining. Similar to the stoichiometric transmetalation experiment using 2-(4-fluorophenyl)-5,5-dimethyl-1,3,2-dioxaborinane, 4-fluorophenyl boronic acid and 4,4'difluorobiphenyl were not observed.

Procedure generating CsOCOCHF² in the optimized reaction. In a tall 10 mL vial equipped with a stir bar, Pd[P(o-Tol)₃]² (3.6 mg, 0.005 mmol, 0.1 equiv) and SPhos (4.2 mg, 0.01 mmol, 0.2 equiv) were combined in toluene (~0.2 mL) and THF (0.05 mL), and the resulting mixture was stirred vigorously for fifteen minutes at room temperature. A THF solution of difluoroacetyl fluoride (0.25 mmol, 5 equiv) was added via syringe. To this solution was added aryl neopentyl

boronate ester **1b-19b** (0.05 mmol, 1.0 equiv), then difluoroacetic anhydride (1.7 mg, 0.01 mmol, 0.2 equiv), and CsF (1.5 mg, 0.01 mmol, 0.2 equiv). The vial was sealed with a Teflon-lined screw cap, removed from the glovebox, and heated at 150 °C for 3 h. The reaction mixture was allowed to cool to room temperature. To it was added 4-fluorotoluene (25 µL, 2.0 M in DCM, 1.0 equiv) as an internal standard, followed by dichloromethane (1.0 mL). An aliquot of the solution was transferred to an NMR tube and analyzed via ¹⁹F NMR spectroscopy. The yield was determined to be 11% after 3 h compared to 86% under standard conditions at 3 h (see Tables S1 and S2).

XI. Catalysis procedure

General procedure for the Pd-catalyzed decarbonylative difluoromethylation of aryl neopentyl boronate esters with difluoroacetyl fluoride. In a tall 10 mL vial equipped with a stir bar, Pd[P(o-Tol)₃]₂ (3.6 mg, 0.005 mmol, 0.1 equiv) and SPhos (4.2 mg, 0.01 mmol, 0.2 equiv) were combined in toluene (~0.2 mL) and THF (0.05 mL), and the resulting mixture was stirred vigorously for fifteen minutes at room temperature. A THF solution of difluoroacetyl fluoride (0.25 mmol, 5 equiv) was added via syringe, and the vial was sealed with a Teflon-lined screw cap and shaken gently to ensure homogeneity (total 0.35 mL volume). To this solution was added aryl neopentyl boronate ester **1b-19b** (0.05 mmol, 1.0 equiv). The vial was sealed with a Teflon-lined screw cap, removed from the glovebox, and heated at 150 °C for 4-10 h. The reaction mixture was allowed to cool to room temperature. To it was added 4-fluorotoluene (25 μL, 2.0 M in DCM, 1.0 equiv) as an internal standard, followed by dichloromethane (1.0 mL). An aliquot of the solution was transferred to an NMR tube and analyzed via ¹⁹F NMR spectroscopy.

These reactions were observed to be highly sensitive to the purity of all components as well as the reaction vessel used. Efforts to scale the reaction using reaction vials, Schlenk glassware, pressure tubes, and microwave tubes of various volumes resulted in significantly diminished yields. For isolation of products **1-13**, six reactions were conducted in parallel to total 0.3 mmol scale. After the reaction time, the reaction mixtures were allowed to cool to room temperature and diluted with DCM and Et₂O. The mixture was filtered through a pad of silica and concentrated *in vacuo*. The crude material was then purified via silica gel chromatography using a gradient elution with solvent mixture A/B (%); A = 15% chloroform in diethyl ether and B = hexanes, unless otherwise stated. Fractions containing product were collected and carefully concentrated *in vacuo* to yield pure difluoromethyl arene product.

NC CHF2

4-(difluoromethyl)benzonitrile (1). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **1b**. Reactions were run for 5 h. Reactions were carefully concentrated to due to the volatity of the product. Purification by flash chromatography on silica gel (0-10% A/B; B = pentanes) afforded the product as a low-melting white solid (35 mg, 77% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.77 (d, *J* = 7.9 Hz, 2H), 7.64 (d, *J* = 7.9 Hz, 2H), 6.69 (t, *J* = 55.8 Hz, 1H). ¹⁹F NMR (471 MHz, CDCl₃) δ –113.26 (d, *J* = 55.8 Hz). ¹³C NMR (176 MHz, CDCl₃) δ

138.69 (t, J = 22.9 Hz), 132.73, 126.54 (t, J = 6.1 Hz), 118.02, 114.93 (t, J = 2.0 Hz), 113.45 (t, J = 240.7 Hz). HRMS (positive ion GC-APCI) calcd. for C₈H₅F₂N [M+H] m/z 154.0463. Found 154.0457.

4-(difluoromethyl)-2-methylbenzonitrile (2). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **2b**. Reactions were run for 5 h. Purification by flash chromatography on silica gel (0-10% A/B; B = pentanes) afforded the purified product as a colorless oil (34 mg, 68% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.69 (d, *J* = 8.0 Hz, 1H), 7.47 (s, 1H), 7.42 (d, *J* = 8.0 Hz, 1H), 6.64 (t, *J* = 55.9 Hz, 1H), 2.60 (s, 3H). ¹⁹F NMR (377 MHz, CDCl₃) δ –113.12 (d, *J* = 55.9 Hz). ¹³C NMR (126 MHz, CDCl₃) δ 142.90, 138.48 (t, *J* = 22.6 Hz), 133.09, 127.46 (t, *J* = 6.1 Hz), 123.59 (t, *J* = 6.1 Hz), 117.36, 115.26 (t, *J* = 2.1 Hz), 113.58 (t, *J* = 240.3 Hz), 20.65. HRMS (positive ion GC-APCI) calcd. for C₉H₇F₂N [M+H] *m*/z 168.0619. Found 168.0623.

(4-(difluoromethyl)phenyl)(phenyl)methanone (3). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate 3b. Reactions were run for 8 h. Purification by flash chromatography on silica gel (0-10% A/B) afforded the purified product as a white solid (36 mg, 51% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.88 (d, *J* = 7.0 Hz, 2H), 7.81 (d, *J* = 7.7 Hz, 2H), 7.71-7.59 (*multiple peaks*, 3H), 7.51 (t, *J* = 6.9 Hz, 2H), 6.73 (t, *J* = 56.1 Hz, 1H). ¹⁹F NMR (471 MHz, CDCl₃) δ –112.12 (d, *J* = 56.1 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 196.04, 139.88, 137.95 (t, *J* = 22.5 Hz), 137.17, 133.02, 130.37, 130.23, 114.18 (t, *J* = 239.7 Hz). HRMS (positive ion GC-APCI) calcd. for C₁₄H₁₀F₂O [M+H] *m/z* 233.0772. Found 233.0777.

1-(4-(difluoromethyl)phenyl)butan-1-one (4). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **4b** and 15 mol % Pd[P(*o*-Tol)₃]₂ and 30 mol % SPhos. Reactions were run for 10 h. Purification by flash chromatography on silica gel (0-10% A/B) afforded the purified product as a white solid (30 mg, 50% yield). Small amounts of grease were present in the purified product but could not be successfully removed via washing with HPLC-grade hexanes. Under the optimized conditions (Entry 1, Table S2), the ¹⁹F NMR yield of product **4** as determined with 4-fluorotoluene internal standard is 37% after 3 h. ¹H NMR (500 MHz, CDCl₃) δ 8.03 (d, *J* = 8.2 Hz, 2H), 7.60 (d, *J* = 8.1 Hz, 2H), 6.69 (t, *J* = 56.1 Hz, 1H), 2.96 (t, *J* = 7.3 Hz, 2H), 1.78 (m, 2H), 1.01 (t, *J* = 7.4 Hz, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ –112.52 (d, *J* = 56.1 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 199.79, 139.05, 138.41 (t, *J* = 22.4 Hz), 128.50, 126.00 (t, *J* = 6.0 Hz), 114.14 (t, *J* = 239.7 Hz), 40.88, 17.77, 13.97. HRMS (positive ion GC-APCl) calcd. for C₁₁H₁₂F₂O [M+H] *m/z* 199.0929. Found 199.0931.

2-(difluoromethyl)anthracene-9,10-dione (5). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **5b**. Reactions were run for 6 h. Purification by flash chromatography on silica gel (0-10% A/B) afforded the purified product as a pale yellow solid (55 mg, 72% yield). Small amounts of grease were present in the purified product but could not be successfully removed via washing with HPLC-grade hexanes. ¹H NMR (500 MHz, CDCl₃) δ 8.46-8.37 (*multiple peaks*, 2H), 8.32 (dt, *J* = 6.0, 3.1 Hz, 2H), 7.94 (d, *J* = 8.0 Hz, 1H), 7.83 (dt, *J* = 5.7, 2.4 Hz, 2H), 6.79 (t, *J* = 55.8 Hz, 1H). ¹⁹F NMR (377 MHz, CDCl₃) δ -112.97 (d, *J* = 55.8 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 182.51, 182.38, 139.87 (t, *J* = 23.0 Hz), 135.04 (t, *J* = 1.7 Hz), 134.60, 133.94, 133.45, 133.44, 130.90 (t, *J* = 5.6 Hz), 128.12, 127.57, 127.55, 124.97 (t, *J* = 6.4 Hz), 113.60 (t, *J* = 240.8 Hz). HRMS (positive ion GC-APCI) calcd. for C₁₅H₈F₂O₂ [M+H] *m/z* 259.0565. Found 259.0571.

ethyl 4-(difluoromethyl)benzoate (6). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate 6b. Reactions were run for 6 h. Reaction was carefully concentrated to due to volatity of product. Purification by flash chromatography on silica gel (0-20% A/B; B = pentanes) afforded the purified product as a white solid (35 mg, 58% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.13 (d, *J* = 7.9 Hz, 2H), 7.59 (d, *J* = 7.9 Hz, 2H), 6.69 (t, *J* = 55.8 Hz, 1H), 4.41 (q, *J* = 7.1, 2H), 1.41 (t *J* = 7.1, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -112.27 (d, *J* = 55.8 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 165.88, 138.47 (t, *J* = 22.4 Hz), 132.81 (t, *J* = 2.0 Hz), 130.03, 125.71 (t, *J* = 6.0 Hz), 114.16 (t, *J* = 239.6 Hz), 61.47, 14.39. HRMS (positive ion GC-APCI) calcd. for C₁₀H₁₀F₂O₂ [M+H] *m/z* 201.0722. Found 201.0727.

methyl 4-(difluoromethyl)benzoate (7). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate 7b. Reactions were run for 6 h. Reactions were carefully concentrated to due to the volatity of the product. Purification by flash chromatography on silica gel (0-20% A/B; B = pentanes) afforded the purified product as a white solid (27 mg, 49% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.13 (d, *J* = 7.9 Hz, 2H), 7.59 (d, *J* = 7.9 Hz, 2H), 6.69 (t, *J* = 56.1 Hz, 1H), 3.95 (s, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -112.36 (d, *J* = 56.1 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 166.37, 138.59 (t, *J* = 22.4 Hz), 132.46, 130.09, 125.77 (t, *J* = 6.0 Hz), 114.14 (t, *J* = 239.8 Hz), 52.52. HRMS (positive ion GC-APCI) calcd. for C₉H₈F₂O₂ [M+H] *m/z* 187.0565. Found 187.0569.


diethyl 5-(difluoromethyl)isophthalate (8). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate 8b. Reactions were run for 6 h. Purification by flash chromatography on silica gel (0-10% A/B) afforded the purified product as a white solid (41 mg, 51% yield). ¹H NMR (401 MHz, CDCl₃) δ 8.78 (s, 1H), 8.48-8.07 (*multiple peaks*, 2H), 6.73 (t, *J* = 55.4 Hz, 1H), 4.43 (q, *J* = 7.1 Hz, 4H), 1.42 (t, *J* = 7.1 Hz, 6H). ¹⁹F NMR (377 MHz, CDCl₃) δ –111.73 (d, *J* = 55.4 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 165.05, 135.34 (t, *J* = 23.3 Hz), 132.77 (t, *J* = 1.7 Hz), 131.89, 130.86 (t, *J* = 6.0 Hz), 113.69 (t, *J* = 240.3 Hz), 61.89, 14.43. HRMS (positive ion GC-APCI) calcd. for C₁₃H₁₄F₂O₄ [M+H] *m/z* 273.0933. Found 273.0935.



2-(4-(difluoromethyl)phenyl)benzo[d]thiazole (9). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **9b**. Reactions were run for 8 h. Purification by flash chromatography on silica gel (0-10% A/B) afforded the purified product as a white solid (36 mg, 46% yield). Small amounts of grease were present in the purified product but could not be successfully removed via washing with HPLC-grade hexanes. ¹H NMR (401 MHz, CDCl₃) δ 8.19 (d, *J* = 8.2 Hz, 2H), 8.10 (d, *J* = 8.2 Hz, 1H), 7.93 (d, *J* = 7.9 Hz, 1H), 7.65 (d, *J* = 8.2 Hz, 2H), 7.52 (ddd, *J* = 8.4, 7.2, 1.3 Hz, 1H), 7.42 (ddd, *J* = 8.3, 7.2, 1.2 Hz, 1H), 6.72 (t, *J* = 56.2 Hz, 1H). ¹⁹F NMR (377 MHz, CDCl₃) δ -111.85 (d, *J* = 56.2 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 166.80, 154.22, 136.68 (t, *J* = 22.5 Hz), 135.94, 135.31, 127.97, 126.71, 126.45 (t, *J* = 6.1 Hz), 125.76, 123.66, 121.86, 114.32 (t, *J* = 239.4 Hz). HRMS (positive ion GC-APCI) calcd. for C₁₄H₉F₂NS [M+H] *m/z* 262.0497. Found 262.0503.



3-(4-(difluoromethyl)phenyl)-5-methyl-1,2,4-oxadiazole (10). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **10b**. Reactions were run for 10 h. Purification by flash chromatography on silica gel (0-10% A/B) afforded the purified product as a white solid (10 mg, 15% yield). ¹H NMR (500 MHz, CDCl₃) δ 8.16 (d, *J* = 8.0 Hz, 2H), 7.63 (d, *J* = 8.1 Hz, 2H), 6.70 (t, *J* = 56.2 Hz, 1H), 2.67 (s, 3H). ¹⁹F NMR (471 MHz, CDCl₃) δ -111.78 (d, *J* = 56.2 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 177.02, 167.85, 136.96 (t, *J* = 22.5 Hz), 129.30 (t, *J* = 1.8 Hz), 127.83, 126.27 (t, *J* = 6.1 Hz), 114.30 (t, *J* = 239.5 Hz), 12.56. HRMS (positive ion GC-APCI) calcd. for C₁₀H₈F₂N₂O [M+H] *m/z* 211.0677. Found 211.0680.



1-(difluoromethyl)-4-(isopropylsulfonyl)benzene (11). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **11b**. Reactions were run for 6 h. Purification by flash chromatography on silica gel (0-20% A/B) afforded the purified product as a white solid (41 mg, 58% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.99 (d, J = 8.1, 2H), 7.72 (d, J = 8.1 Hz, 2H), 6.73 (t, J = 55.9 Hz, 1H), 3.21 (hept, J = 6.8 Hz, 1H), 1.31 (d, J = 6.8 Hz, 6H). ¹⁹F NMR (471 MHz, CDCl₃) δ –113.19 (d, J = 55.9 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 139.57 (t, J = 1.7 Hz), 139.53 (t, J = 22.7 Hz), 129.75, 126.55 (t, J = 6.0 Hz), 113.56 (t, J = 240.6 Hz), 55.81, 15.80. HRMS (positive ion GC-APCI) calcd. for C₁₀H₁₂F₂O₂S [M+H] *m/z* 235.0599. Found 235.0607.



4-(difluoromethyl)-N,N-dimethylbenzenesulfonamide (12). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **12b**. Reactions were run for 6 h. Purification by flash chromatography on silica gel (0-50% A/B) afforded the purified product as a colorless oil (36 mg, 51% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.88 (d, *J* = 8.0 Hz, 2H), 7.70 (d, *J* = 8.0 Hz, 2H), 6.72 (t, *J* = 55.9 Hz, 1H), 2.74 (s, 6H). ¹⁹F NMR (471 MHz, CDCl₃) δ -112.68 (d, *J* = 55.9 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 138.54 (t, *J* = 22.8 Hz), 138.21 (t, *J* = 2.0 Hz), 128.24, 126.53 (t, *J* = 6.0 Hz), 113.67 (t, *J* = 240.3 Hz), 37.97. HRMS (positive ion GC-APCI) calcd. for C₉H₁₁F₂NO₂S [M+H] *m/z* 236.0551. Found 236.0556.



4-(difluoromethyl)-N,N-dipropylbenzenesulfonamide (13). Reaction was conducted for isolation scale (6 reactions in parallel, 0.05 mmol scale each) using substrate **13b**. Reactions were run for 6 h. Purification by flash chromatography on silica gel (0-20% A/B) afforded the purified product as a white solid (45 mg, 52% yield). ¹H NMR (500 MHz, CDCl₃) δ 7.90 (d, *J* = 7.9 Hz, 2H), 7.65 (d, *J* = 7.9 Hz, 2H), 6.70 (t, *J* = 56.0 Hz, 1H), 3.10 (t *J* = 7.6, 4H), 1.61-1.51 (m, 4H), 0.88 (t, *J* = 7.4, 6H).¹⁹F NMR (471 MHz, CDCl₃) δ –112.51 (d, *J* = 56.0 Hz). ¹³C NMR (176 MHz, CDCl₃) δ 142.60, 137.90 (t, *J* = 22.7 Hz), 127.40, 126.29 (t, *J* = 6.0 Hz), 113.57 (t, *J* = 240.2 Hz), 49.98, 21.97, 11.12. HRMS (positive ion GC-APCl) calcd. for C₁₃H₁₉F₂NO₂S [M+H] *m/z* 292.1177. Found 292.1182.

XII. Other substrates explored

Fluorine- and fluoroalky-substituted ArBneo substrates **14b-18b** were also explored. ¹⁹F NMR spectroscopic analysis of the crude reaction mixtures implicates the formation of the ArCHF₂ products **14-18**. The ¹⁹F NMR chemical shifts that are assigned as -**CHF**₂ resonance for each product are listed below, along with a ¹⁹F NMR yields (versus 4-fluorotoluene as an internal standard). None of these products were isolated due to their volatility.



1-(difluoromethyl)-3-(methylsulfonyl)-5-(trifluoromethyl)benzene (14). The reaction was conducted as a single run using **14b**, and the yield of **14** was determined by ¹⁹F NMR spectroscopy with 4-fluorotoluene as an internal standard. Reaction time: 4 h; yield: 65%. ¹⁹F NMR (377 MHz) δ –113.77 (d, *J* = 56.4 Hz).



1-(difluoromethyl)-3,5-bis(trifluoromethyl)benzene (15). The reaction was conducted as a single run using **15b**, and the yield of **15** was determined by ¹⁹F NMR spectroscopy with 4-fluorotoluene as an internal standard. Reaction time: 4 h; yield: 94%. A second trial using substrate **15b** yielded 86% of **15**.¹⁹F NMR (471 MHz) δ –113.64 (d, *J* = 55.5 Hz).



1-(difluoromethyl)-4-(trifluoromethyl)benzene (16). The reaction was conducted as a single run using **16b**, and the yield of **16** was determined by ¹⁹F NMR spectroscopy with trifluoromethoxybenzene as an internal standard. Reaction time: 4 h; yield: 58%. A second trial using substrate **16b** yielded 50% of **16**. ¹⁹F NMR (471 MHz) δ –113.13 (d, *J* = 56.1 Hz).

F CHF₂

1-(difluoromethyl)-4-(trifluoromethyl)benzene (17). The reaction was conducted as a single run using **17b**, and the yield of **17** was determined by ¹⁹F NMR trifluoromethoxybenzene as an internal standard. Reaction time: 4 h; yield: 60%. ¹⁹F NMR (377 MHz) δ –110.39 (d, *J* = 55.9 Hz).



1-(difluoromethyl)-4-(trifluoromethyl)benzene (18). The reaction was conducted as a single run using **18b**, and the yield of **18** was determined ¹⁹F NMR spectroscopy with 4-fluorotoluene as an internal standard. Reaction time: 4 h; yield: 69%. ¹⁹F NMR (471 MHz) δ –117.08 (d, *J* = 55.6 Hz).



Table S3. Other substrates explored for palladium-catalyzed difluoromethylation using difluoroacetyl fluoride.

XIII. Computational Methods

A parallel molecular mechanics (MM) and the advanced quantum mechanical (QM) calculations of complexes, intermediates, transition states and products were carried out using Jaguar programs, as implemented in Schrödinger Material Suite 2019-19 as well as Gaussian software.¹⁰ The MM based conformational search was carried out on the geometry of the pre-complexes and final products to obtain a preliminary insight into the different possible conformers. Top unique conformers were selected and were further optimized applying density functional theory (DFT) calculations. The geometry optimizations of all the structures were performed using the M06 functional with the combined LANL2DZ basis set for Pd, and P atoms, and 6-311G** is selected for C, H, O and F atoms. The polarization functions of Pd((t) = 1.472,14, and $P(\zeta d)$ = 0.340 were also included.¹¹ The optimized geometries were further confirmed by frequency calculations at the same level of theory as minima (zero imaginary frequencies) or transition states (one imaginary frequency). The thermochemical quantities were analyzed at both 298 K and 363 K (the experimental temperatures), and the intrinsic reaction coordinate¹² (IRC) was applied to obtain the minima of products and the starting materials on either side of each optimized transition state in gas phase. The energies were further corrected with BPV86 functional using larger basis sets and different solvation models using the self-consistent reaction field (SCRF) method with the conductor-like polarizable (CPCM) model and UAHF radii¹³ applying the default solvent parameters for THF (the applied experimental solvent) using Gaussian 09 package on Compute Canada systems and Great Lakes implemented at University of Michigan. The applied methods were chosen based on balancing computational time and accuracy to properly treat the key dispersion interactions. The structural visualizations were carried out using GaussView v5.0.8.4. To analyze in detail the electronic and structural perspectives, natural bond orbital analysis (NBO) using the same level of theory as solvation models as well as non-covalent interaction (NCI) analysis was performed.¹⁴ We limited calculations to the C-C bond cleavage/carbonyl reorientation process, which is a key step in the potential energy surface for the carbonyl de-insertion reaction. Optimal structures of intermediates in the decarbonylation process were found via conformational sampling of difluoroacetic anhydride and trifluoroacetic anhydride complexes. The most promising conformers were further optimized by the M06 functional with the combined LANL2DZ basis set for Pd, and P atoms, and 6-311G(d,p) for C, H, O and F atoms for further structural and electronic analysis (distortion interactions, non-covalent interactions, electrostatic potential surfaces). To account for the entropic contributions in the activation free energy barrier, we also computationally analyzed the temperature-dependent activation barrier of decarbonylation process at a high experimental temperature (363 K). The same trends are observed at this higher temperature and qualitatively reflected the experimental findings.

XIV. Computational Data and Discussion

A. Molecular Mechanics (MM)-based conformational analysis



Figure S21. The superimposed conformers of **I-COCHF**₂ and **I-COCF**₃ and transition state derived complexes, **TS1-CHF**₂ and **TS1-CF**₃, determined to be within 10 kcal/mol of the global minimum. The lowest energy conformer in each case is shown in green.

MM-based conformational analyses suggest that **I-COCF**₃ (Figure S21, C) exists within a narrower conformational range than **I-COCHF**₂. Few optimized conformers of **I-COCF**₃ were found 2-10 kcal/mol above the global minimum with low conformational diversity (as indicated by the green conformer of **I-COCHF**₂ (Fig. S21, C), that is nearly conformationally indistinct and overlapped with a second conformer in gray. For **I-COCHF**₂, a large mixture of thermally accessible conformational families is present and could be reactive under the applied conditions (Fig. S21, A). We propose that this larger conformational diversity near the energetic minimum relates to the presence of an acidic hydrogen atom that stabilizes various conformations of **I-COCHF**₂. A similar observation can be made for the transition states **TS1-R**_F (Figure S21, B and D), in which a greater number of low energy conformations are present for **TS1-CHF**₂ compared to **TS1-CF**₃. From the DFT-based optimized conformers, the two lowest energy conformers (**I-COR**_F **A** and **B** and their associated transition states) were selected for further computational analysis.

Energetics were calculated only for the 1,1-de-insertion reaction of I-COR_F to form (CO)Pd–R_F proceeding through TS1-R_F as this is proposed to be the more demanding step for the decarbonylation process (rather than the release of CO). This in agreement with our stoichiometric results, in which we do not observe intermediate (CO)Pd–R_F for either -CF₃ or -CHF₂.



Figure S22. The DFT-based optimized geometries of the optimized CF_3 -containing reactant complexes (I-COCF₃ A and B), transition states (TS1-CF₃ A and B), and intermediates (CO)Pd–CF₃ A and B are shown above with energy values. The activation free energies for the C_(CO)–CF₃ bond cleavage, and subsequent CO-reorientation process are presented. The evaluation of the effect of functional, basis set and solvation models on the energetic trends at both 298 K and 363 K are shown. Enthalpic contributions to the free energy values shown in parentheses.

The energetics shown above in Figure S22 are reflective of the experimental findings, in which elevated temperatures are required for the decarbonylation of I-COCF₃ to form II-CF₃. Going forward, the energetic values corresponding to I-COCF₃ A, TS1-CF₃ A, and (CO)Pd–CF₃ were selected for further discussion.



Figure S23. The DFT-based optimized geometries of the CHF₂-containing reactant complexes (**I-COCHF₂ A** and **B**), transition states (**TS1-CHF₂ A** and **B**), and intermediates (**CO)Pd–CHF₂ A** and **B** are shown above with energy values. The activation free energies for the $C_{(CO)}$ –CF₃ bond cleavage, and subsequent CO-reorientation process are presented. The evaluation of the effect of functional, basis set and solvation models on the energetic trends at both 298 K and 363 K are shown. Enthalpic contributions to the free energy values shown are in parentheses.

The energetics shown above in Figure S23 are reflective of the experimental findings, in which the decarbonylation of **I-COCHF**₂ to form **II-CHF**₂ proceeds at room temperature. Going forward, the energetic values corresponding to **I-COCHF**₂ **A**, **TS1-CHF**₂ **A**, and **(CO)Pd–CHF**₂ were selected for further discussion.



Α

В

-(COCF₃A	TS1-CF₃A	$\Delta (TS1-CF_3A-I-COCF_3)$
$\begin{array}{l} Pd_{\dots}C_{Ar} = \\ Pd_{\dots}C_{OCH3}(\textit{back}) = \\ Pd_{\dots}C_{OCH3}(\textit{front}) = \\ Pd_{\dots}OCH_3(\textit{back}) = \\ Pd_{\dots}OCH_3(\textit{front}) = \end{array}$	3.16 Å	3.31 Å	+0.15 Å
	3.40 Å	3.73 Å	+0.33 Å
	3.26 Å	3.18 Å	-0.08 Å
	4.05 Å	4.47 Å	+0.42 Å
	3.82 Å	3.57 Å	-0.25 Å
CF ₃ Pd =	2.94 Å	2.50 Å	-0.44 Å
CF ₃ C=O =	1.56 Å	2.03 Å	+0.47 Å
PdC=O =	1.94 Å	1.85 Å	-0.09 Å
α =	70.6°	67.3°	-3.3°
β =	97.5°	123.6°	+26.1°
γ =	91.4°	104.0°	+12.6°
δ =	114.2°	95.3°	-18.9°



PdOCH ₃ (<i>front</i>)=	3.39 Å	3.45 Å	+0.06 Å
CHF ₂ Pd =	2.72 Å	2.40 Å	-0.32 Å
CHF ₂ C=O =	1.57 Å	1.91 Å	+0.34 Å
PdC=O =	1.95 Å	1.86 Å	-0.09 Å
α =	69.3°	66.3°	$\alpha = -3.0^{\circ}$
β =	154.9°	148.8°	$\beta = -6.1^{\circ}$
γ =	96.3°	105.5°	$\gamma = +9.2^{\circ}$
δ =	103.3°	101.1°	$\overline{o} = -2.2^{\circ}$

Figure S24. The calculated bond lengths, distances, and angles for intermediates (A) I-COCF₃ and transition state TS1-CF₃ and (B) I-COCHF₂ and TS-CHF₂. Angles α , β , γ , and δ are defined by the three atoms involved and are conserved in both structures (e.g., ß is defined as the angle between the two fluoroalkyl ligands with Pd as the vertex in both structures).

The data above highlight specific bond lengths, distances, and angles which show that the optimized conformer of **I**-**COCHF**₂ is greatly distorted away from the expected square planar geometry. Comparatively, the ground state geometry of **I**-**COCF**₃ shows angles much closer to a typical Pd^{II} square planar complex. Given that carbonyl deinsertion at Pd^{II} is proposed to occur via three-coordinate Pd complexes¹⁵, we propose that the distortion of the coordination geometry at **I**-**COCHF**₂ contributes to a barrier-lowering effect to proceed through **TS1-CHF**₂.

B. Analysis of dispersive repulsive electrostatic interactions

Differential electrostatic potential maps visualized electrostatic charge distribution differences between these complexes. For CF₃-derived skeletons, the plotted electrostatic potential surfaces of the complexes show greater and closer proximity of the strongly negatively polarized groups in the CF₃-substituted complexes in comparison to the analogous -CHF₂ complexes. Specifically, these dispersive, repulsive electrostatic interactions are more pronounced for the transition states than for the reactants in CF₃-derived complexes resulting in a higher activation barrier for 1,1-de-insertion via **TS1-CF₃**.





Our calculations show the existence of several stabilizing non-covalent interactions and weakly attractive bonding interactions in CHF₂-derived complexes, such as London dispersion forces and weak attractive van der Waals interactions that stabilize **TS1-CHF**₂ compared to **TS1-CF**₃. Accordingly, second sphere components in these complexes, such as the cyclohexyl rings and methoxy groups of SPhos, synergistically benefit from their mutual interactions with -CHF₂. These non-covalent and donor-acceptor stabilizing interactions contribute to the distinct conformation of CHF₂ derived complexes, producing a barrier-lowering effect for carbonyl de-insertion. In contrast, **TS1-CF**₃ lacks the stabilizing electrostatic interaction with the same degree and features fewer attractive interactions.



Figure S26. (A) The DFT-based optimized **II-CHF**₂ and **II-CF**₃ conformers with change in free energies shown relative to **I-COCHF**₂ and **I-COCF**₃, respectively. (B) the calculated NCI gradient iso-surfaces with s = 0.3 au representing the attractive non-covalent interactions, colored according to the values of sign(λ^2) ρ between -0.5 to 0.5 au. In this coloring system, green and yellow iso-surfaces are related to weakly attractive and weakly repulsive interactions respectively; blue boundaries show attractive interactions; and red boundaries show repulsive interactions. Highly favorable interactions are shown as disks of purple color.



Figure S27. The sum of predicted NBO derived donor-acceptor orbital interactions for (A) **II-CHF**₂ and (B) **II-CF**₃. The sum of NBO interaction energies is greater for **II-CHF**₂ versus **II-CF**₃ indicating a greater stabilization of **II-CHF**₂.

The combined computational data above reveal that a series attractive interactions (orbital interaction, attractive electrostatic interactions, and non-covalent interactions) were highly influential to the observed decarbonylative reactivity as shown by our combined application of quantum mechanical techniques such as natural bond orbital (NBO) analysis, non-covalent interactions (NCI) index analysis, and the molecular electrostatic potential (MEP) surface maps.

According to our study, the structural, energetic, and electronic parameters within the optimized geometries of CF₃and CHF₂-bearing complexes highlighted the presence of several attractive interactions comprised of 1) the orbital interaction that accounts for charge transfer (*i.e.*, donor–acceptor interactions); 2) a series of weakly attractive noncovalent bonding interactions such as induced dipole-induced dipole (London dispersion forces) interactions; and 3) attractive electrostatic interaction that accounts for the electrostatic charge distribution to recognize the reactive sites with positive and negative electron density.

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XVI. X-Ray Crystallography Data for II-CHF₂. Structure determination of II-CHF₂



Yellow blocks of **II-CHF**² were grown via vapor diffusion of pentane/diethyl ether solution into tetrahydrofuran solution of the compound at 22 °C. A crystal of dimensions 0.22 x 0.20 x 0.16 mm was mounted on a Rigaku AFC10K Saturn 944+ CCD-based X-ray diffractometer equipped with a low temperature device and Micromax-007HF Cu-target micro-focus rotating anode (I = 1.54187 A) operated at 1.2 kW power (40 kV, 30 mA). The X-ray intensities were measured at 85(1) K with the detector placed at a distance 42.00 mm from the crystal. A total of 2028 images were collected with an oscillation width of 1.0° in w. The exposure times were 1 sec. for the low angle images, 3 sec. for high angle. Rigaku d*trek images were exported to CrysAlisPro for processing and corrected for absorption. The integration of the data yielded a total of 42225 reflections to a maximum 2q value of 138.80° of which 5201 were independent and 5166 were greater than 2s(I). The final cell constants (Table S4) were based on the xyz centroids of 29274 reflections above 10s(I). Analysis of the data showed negligible decay during data collection. The structure was solved and refined with

the Bruker SHELXTL (version 2018/3) software package, using the space group P2(1)/c with Z = 4 for the formula C29H37O4F4PPd. All non-hydrogen atoms were refined anisotropically with the hydrogen atoms placed in idealized positions. The difluoroacetato group is rotationally disordered. Full matrix least-squares refinement based on F2 converged at R1 = 0.0317 and wR2 = 0.0851 [based on I > 2sigma(I)], R1 = 0.0319 and wR2 = 0.0853 for all data. Additional details are presented in Table 4 and are given as Supporting Information in a CIF file. Acknowledgement is made for funding from NSF grant CHE-0840456 for X-ray instrumentation.

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Empirical formula	C ₂₉ H ₃₇ F ₄ O ₄ PPd
Formula weight	662.95
Temperature	85(2) K
Wavelength	1.54184 A
Crystal system	Monoclinic
space group	P2(1)/c
Unit cell dimensions	a = 9.68570(10) A α = 90 °
	b = 18.93400(10) A ß = 91.4640(10)°
	c = 15.37910(10) A γ = 90 °
Volume	2819.44(4) A^3
Z	4
Calculated density	1.562 Mg/m^3
Absorption coefficient	6.373 mm^-1
F(000)	1360
Crystal size	0.220 x 0.200 x 0.160 mm
Theta range for data collection	3.703 to 69.400 deg.
Limiting indices	-11<=h<=11, -22<=k<=22, -17<=l<=18
Reflections collected / unique	42225 / 5201 [R(int) = 0.0734]
Completeness to theta	67.684 (99.0 %)
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.00000 and 0.64949
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	5201 / 6 / 366
Goodness-of-fit on F^2	1.054
Final R indices [I>2σ(I)]	R1 = 0.0317, wR2 = 0.0851
R indices (all data)	R1 = 0.0319, wR2 = 0.0853
Largest diff. peak and hole	0.799 and -0.744 e.A ⁻³

Table S4. Addition details regarding crystals of II-CHF2.

XVII. ¹H, ¹³C, ¹⁹F and ³¹P NMR spectral data





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230	220	210	200	190	180	170	160	150	140	130	120	110	100	90	80	70	60	50	40	30	20	10	0	-10
f1 (ppm)																								

































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											f	1 (ppm)											









S71




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230	220	210	200	190	180	170	160	150	140	130	120	110 f1 (ppm	100 1)	90	80	70	60	50	40	30	20	10	0	-10

























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30	20	10	0	-10	-20	-30	-40	-50	-60	-70	-80 f1 (p	-90 opm)	-100	-110	-120	-130	-140	-150	-160	-170	-180	-190	-2















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f1 (ppm)













