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Supplemental information

Amino acid primed mTOR activity

is essential for heart regeneration

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Supplemental Figure 1: Presentation of MTZ to *vmhc*:mCherry-NTR fish results in cell death, related to Figure 1. A) Wild type AB zebrafish heart and *vmhc*:mCherry-NTR zebrafish heart shown in brightfield, merged, and mCherry fluorescence. The mCherry signal shows the tissue specific, ventricle, expression of the mCherry-NTR protein. The chambers of the heart are outlined and labeled: A – atrium, V- ventricle, and B – bulbus arteriosus. The scale bars are: 500 μ m. B) Confocal fluorescent microscopy image of a sectioned ventricle of the *vmhc*:mCherry-NTR line. mCherry staining is found throughout the ventricle. Red – mCherry-NTR and Blue – DAPI. Scale bar is 50 μ m. C) Confocal microscopy of vmhc:mCherry-NTR zebrafish heart after 48 hours of vehicle (DMSO, uninjured) or MTZ (injured). Injured fish show TUNEL positive cells, yellow arrow heads, while vehicle treated fish do not. During injury there is a loss of ventricular muscle. Red – mCherry-NTR, Green – TUNEL, and Blue – DAPI. Scale bars are 10 μ m.



Supplemental Figure 2: Wnt/ β -catenin signaling effects cardiomyocyte proliferation during the early stages of regeneration, related to Figure 1. A-C) Immunohistochemistry images of cardiomyocyte proliferation at 3 days post injury during Wnt/ β -catenin inhibition via hsDkk1b overexpression or no Wnt/ β -catenin inhibition via no heat shock or no hsDkk1bGFP construct present. D-E) Immunohistochemistry images of cardiomyocyte proliferation at 3 days post injury during Wnt/ β -catenin activation via hsWnt8a overexpression or no additional Wnt/ β -catenin activation via hsWnt8a overexpression or no additional Wnt/ β -catenin activation via hsWnt8a overexpression or no additional Wnt/ β -catenin activation via no heat shock and no hsWnt8aGFP construct present. F) Quantification of cardiomyocyte proliferation in the context of Wnt/ β -catenin activation via hsWnt8a. All animals were presented with MTZ. Scale bar: 25 µm. Blue-DAPI, Red-Pcna, and Green-Mef2c. Yellow arrow heads denote proliferating cardiomyocytes, Pcna and Mef2c positive.



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Supplemental Figure 3: Metabolism is dynamically regulated during the first week of adult zebrafish heart regeneration, related to Figure 2. A) RNA-sequencing data of cardiac transcription factors, structural, and metabolic gene expression during adult zebrafish heart regeneration and RT-qPCR validation of these genes. B) Principle component analysis of isolated young uninjured, 3-day old hearts, and adult zebrafish hearts as compared to Bednarek *et al.* adult zebrafish hearts. C) Principle component analysis of isolated young uninjured 3-day old hearts, uninjured adult hearts, and NTR ablated adult zebrafish hearts as compared to Bednarek *et al.* uninjured and cryo-injured adult zebrafish hearts. D) Venn diagrams for the genes that revert to a fetal like expression level during regeneration. E) Key cardiac pathways that are differentially expressed during adult zebrafish heart regeneration. Log2 fold change shown. F) Metabolic pathways enrichment for up- and down-regulated genes during the first week of adult zebrafish heart regeneration.

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Supplemental Figure 4: Proteomics analysis of adult zebrafish heart regeneration, related

to Figure 2. A) Schematic of adult zebrafish hearts used for proteomics analysis at UI adult, 3, or 7 dpi. B) Principle component analysis of full proteome analysis during the first week of heart regeneration. C) Volcano plots of protein expression comparing adult hearts at UI, 3, or 7 dpi. D) Gene ontology terms that were down-regulated at 3 dpi. E) Gene ontology terms that were up-regulated at 7 dpi. F) Protein validation of succinate dehydrogenase complex assembly factor 3 expression during the first week of adult zebrafish heart regeneration. G) RNA-sequencing expression of succinate dehydrogenase complex assembly factor 3. H) Venn diagrams showing the number of genes and proteins that were differentially regulated and found in both data sets during heart regeneration. Bar graphs error bars represent standard error of the mean.

TCA cycle RNA-seq



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Supplemental Figure 5: Dynamic transcriptional remodeling of metabolic enzymes during the first week of heart regeneration, related to Figure 2. A) Heat map of TCA cycle enzyme gene expression from RNA-sequencing data.



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Supplemental Figure 6: Cardiomyocytes and epicardial cells have higher TOR activity during zebrafish heart regeneration related to Figure 3. A) Western blot of TOR activity target p-Ulk1 and loading control protein alpha-tubulin. B) Quantification of protein abundance in A shows increased Tor activity at 3 dpi. C-E) Whole heart immunohistochemistry images assessing cardiomyocyte proliferation at 3 dpi in the uninjured, injured, and injured with rapamycin treated conditions. Scale bars: 100 µm. F-H) Immunohistochemistry assessing TOR activity at 3 dpi of heart regeneration. Positive TOR cardiomyocytes are double positive for pS6 and NTR-mCherry (yellow arrows). Positive TOR epicardial cells are positive for pS6 and were determined as the cells along the border of the heart. DAPI - blue, phosphorylation of S6 -Green, and NTR-mCherry – Red. Full heart image scale bars are 100 µm, magnified inset image scale bars are 20 µm. I) Quantification of the percentage of pS6 cardiomyocytes, cardiomyocytes determined by vmhc::mCherry-NTR. One-way ANOVA performed. p=0.023 for uninjured (-Rapa -MTZ) vs injured (-Rapa +MTZ) hearts. p=0.024 for injured (-Rapa +MTZ) vs rapamycin treated injured hearts (+Rapa +MTZ). N=2-3 biological replicates. J) Quantification of the percentage of pS6 epicardial cells. p=0.029 for uninjured (-Rapa -MTZ) vs injured (-Rapa +MTZ) hearts. p=0.050 for injured (-Rapa +MTZ) vs rapamycin treated injured hearts (+Rapa +MTZ). N=4 biological replicates. K-M) Whole heart immunohistochemistry images assessing cardiomyocyte TOR activation at 3 dpi in the uninjured, injured, and injured with rapamycin treated conditions. Scale bars: 100 µm. DAPI – blue, phosphorylation of S6 – Green, and MF20 – Red. N) Quantification of the percentage of pS6 epicardial cells. p < 0.001for uninjured (-Rapa -MTZ) vs injured (-Rapa +MTZ) hearts. p<0.001 for injured (-Rapa +MTZ) vs rapamycin treated injured hearts (+Rapa +MTZ). One-Way ANOVA performed. N=3-4 biological replicates. Bar graphs show individual data points with error bars representing standard error.



Supplemental Figure 7: Single cell RNA-sequencing of the adult regenerating zebrafish heart identifies transient cell populations, related to Figure 4. A) Principle component analysis separated by day. B) Plot showing ventricular cardiomyocyte markers. C) Gene expression of *lamtor5* in cluster 1 cells at different time points. D) Plot showing bulbus arteriosus markers. E) Gene expression of *tgfb3* in each of the four clusters. F) Plot showing epicardial markers. G) Plot showing activated epicardial markers. H) Plot showing fibroblast markers. I) Plot showing endocardial markers. J) Plot showing atrial cardiomyocyte markers. K) Plot showing cells which are expressing the gene for the Wnt ligand *wnt11r*.

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Mouse Explanted CM

2 condition 0hr 1 24hr 48hr 0 72hr

-1

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Supplemental Figure 8: Adult mouse cardiomyocyte de-differentiation resembles a regenerative transcript profile, related to Figure 5. A) Schematic of adult mouse cardiomyocyte isolation and *in vitro* culture. B) Heat map of cell proliferation, Wnt ligands and Wnt/ β -catenin targets during cardiomyocyte de-differentiation. Heatmap shows Log₂ fold change. C) Heat map of the transcriptional change in metabolic pathways during cardiomyocyte de-differentiation. Heatmap shows Log₂ fold change. D) Schematic of glutamine transport and mTORC1 activation in a cell.