

Supplemental Information for

MEKK3-TGF β crosstalk regulates inward arterial remodeling

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Supplemental Figure Legends

Figures S1-10

Table S1

Supplemental Figure Legends

Figure S1. Loss of MEKK3 in ECs induces cardiac hypertrophy. (A) Q-PCR analysis of *Mekk3* expression in isolated lung ECs, aortic smooth muscle cells and cardiomyocytes from Ctrl and *Mekk3*^{iECKO} mice, n=3 per group. Data represent mean \pm SD. **** $P < 0.0001$, ns: not significant, calculated by unpaired *t*-test. (B) Representative higher magnification H&E images of RV and LV for Ctrl and *Mekk3*^{iECKO} mice at 4 weeks after tamoxifen injection. Scale bar: 500 μ m. (C) Representative right ventricle systolic pressure (RVSP) tracing for Ctrl and *Mekk3*^{iECKO} mice at 4 weeks after tamoxifen injection. (D) Representative left ventricle systolic pressure (LVSP) tracing for Ctrl and *Mekk3*^{iECKO} mice at 8 weeks after tamoxifen injection. (E) Representative systolic blood pressure (SBP) and diastolic blood pressure (DBP) tracing for Ctrl and *Mekk3*^{iECKO} mice at 8 weeks after tamoxifen injection.

Figure S2. Heart rate and ECG for Ctrl and *Mekk3*^{iECKO} mice. (A) Heart rate (BPM) for Ctrl and *Mekk3*^{iECKO} mice at 2, 4 and 8 weeks after tamoxifen injection. n=5 mice per time point and per group. Data represent mean \pm SEM. ns: not significant, calculated by two-way ANOVA with Tukey's multiple comparison tests. (B) Representative electrocardiogram (ECG) tracing for Ctrl and *Mekk3*^{iECKO} mice at 4 and 8 weeks after tamoxifen injection.

Figure S3. *Mekk2* knockout mice don't develop hypertension. (A) Western blot analysis of MEKK2 expression in Ctrl and *Mekk2*^{-/-} lungs. (B) RVSP of Ctrl and *Mekk2*^{-/-} mice at 6 months old. (C) LVSP of Ctrl and *Mekk2*^{-/-} mice at 6 months old. (B-C) n=4 male mice for each group. Data represent mean \pm SEM. ns: not significant, calculated by unpaired *t*-test.

Figure S4. MEKK3 deletion in ECs induces TGF β signaling. (A) Representative TGF β R2 staining of Ctrl and *Mekk3*^{iECKO} entire lungs at 4 weeks after tamoxifen

injection. Scale bar: 1mm. **(B-C)** Representative immunostaining and quantification of SM22 α **(B)**, and Collagen I **(C)** in lung sections from Ctrl and *Mekk3*^{ECKO} mice at 4 weeks after tamoxifen injection. Scale bar: 25 μ m. **(D)** Representative Smad2/3 immunostaining and nuclear translocation quantification in lung sections from Ctrl and *Mekk3*^{ECKO} mice at 4 weeks after tamoxifen injection. Scale bar: 25 μ m. n=4 mice per group. Data represent mean \pm SEM. **P* < 0.05, calculated by Mann-Whitney U-test.

Figure S5. Knockdown of MEKK3 in HPAECs induces EndMT. **(A)** Q-PCR analysis of EndMT markers expression in human pulmonary artery endothelial cells (HPAECs) treated with Ctrl or MEKK3 siRNA. n=3 independent experiments. Data represent mean \pm SD. ***P* < 0.01, ****P* < 0.001, calculated by unpaired *t*-test. **(B)** Western blot analysis of EndMT markers expression in HPAECs treated with Ctrl or MEKK3 siRNA. **(C)** Immunostaining of LIN28 in Ctrl and *Mekk3*^{ECKO} mice lungs. Scale bar: 50 μ m. Arrowheads point to endothelial cells expressing LIN28.

Figure S6. Loss of MEKK3 in ECs impairs FGF2-ERK1/2-Let7 signaling pathway. **(A-B)** ERK1/2 activation upon **(A)** FGF2 (100ng/ml) and **(B)** VEGF165 (50ng/ml) treatment in HUVECs treated with Ctrl or MEKK3 siRNA. **(C-D)** Smad1/5/9 activation upon **(C)** BMP9 (10ng/ml) and **(D)** BMP6 (50ng/ml) treatment in HUVECs treated with Ctrl or MEKK3 siRNA. **(E)** Q-PCR analysis of FGFR1 (n=6) expression in HUVECs treated with Ctrl or MEKK3 siRNA. Data represent mean \pm SEM. **(F)** Western blot analysis of FGFR1 expression in HUVECs treated with Ctrl or MEKK3 siRNA. ns: not significant, calculated by unpaired *t*-test.

Figure S7. Additional images showing EndMT. **(A-C)** Representative GFP and SMA staining in lung **(A)**, kidney **(B)**, and liver **(C)** from mTmG Ctrl and mTmG *Mekk3*^{ECKO} mice at 4 weeks after tamoxifen injection. Scale bar: 50 μ m. Arrowheads point to endothelial cells expressing SMA.

Figure S8. F4/80 staining in atherosclerotic plaque.

(A) Representative F4/80 staining in brachiocephalic artery lesion from *Apoe*^{-/-} mice and *Apoe*^{-/-} *Mekk3*^{IECKO} mice. Scale bar: 100 μ m.

Figure S9. Suppression of TGF β R signaling rescues MEKK3-knockout-induced

EndMT. (A) Q-PCR analysis of MEKK3, SM22 α , fibronectin (FN) and N-Cadherin expression in HUVECs treated with Ctrl or MEKK3 siRNA in addition to TGF β R inhibitor. (B) Western Blot and (C) Q-PCR analysis of EndMT markers expression in HUVECs treated with Ctrl or MEKK3 siRNA in addition to TGF β R1/R2 siRNA. n=3 independent experiments. Data represent mean \pm SD. ***P* < 0.01, ****P* < 0.001, ns: not significant, calculated by one-way ANOVA with Tukey's multiple comparison tests.

Figure S10. Negative control for immunohistochemistry staining with non-

immune species-matched isotype IgG. Lung sections were blocked and incubated with indicated primary antibodies and its non-immune species-matched IgG at 4 $^{\circ}$ C overnight. Sections then were incubated with secondary antibodies at room temperature for 2h, finally mounted with DAPI. (A) Representative immunostaining of mouse isotype IgG, fibronectin and TGF β with CD31 and DAPI. (B) Representative immunostaining of rabbit isotype IgG, p-Smad2 Ser465/467 and p-Smad3 Ser423/425 with CD31 and DAPI. Scale bar: 25 μ m.

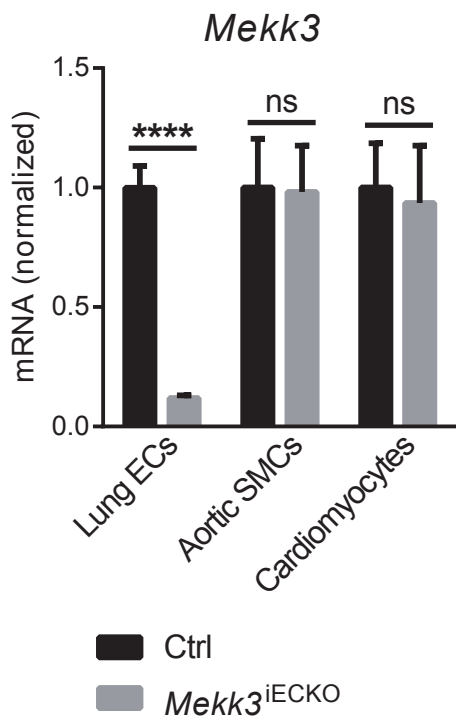
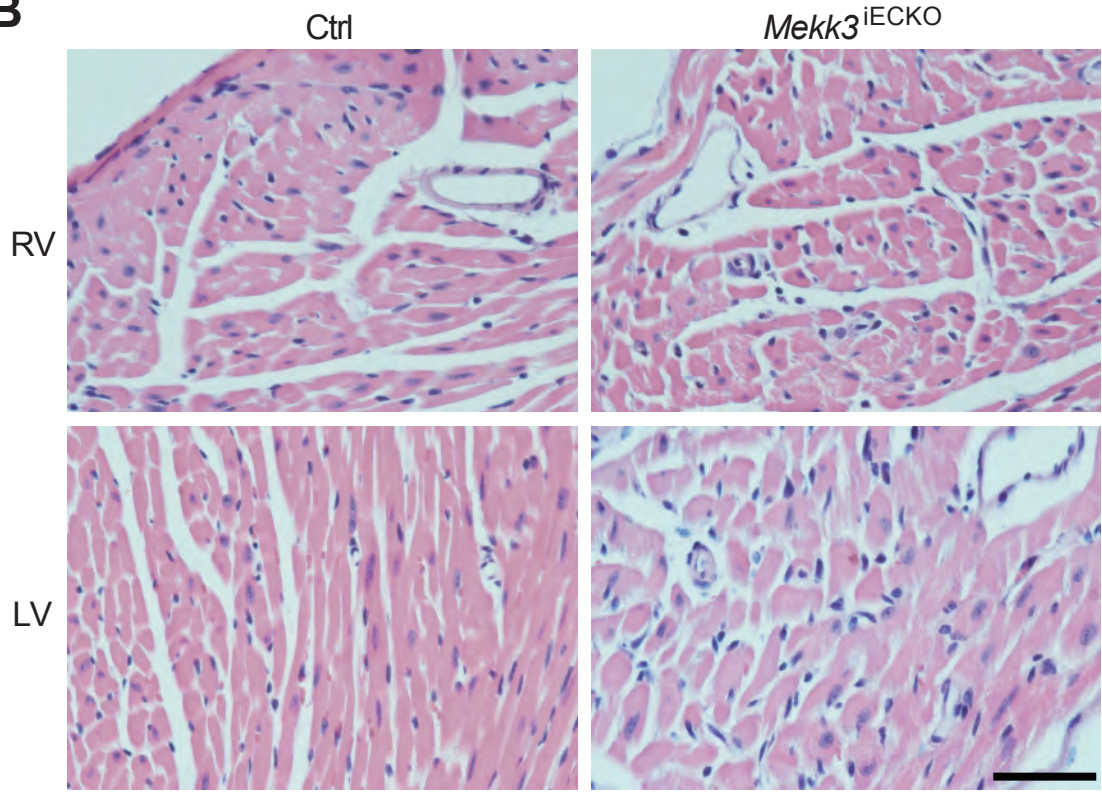
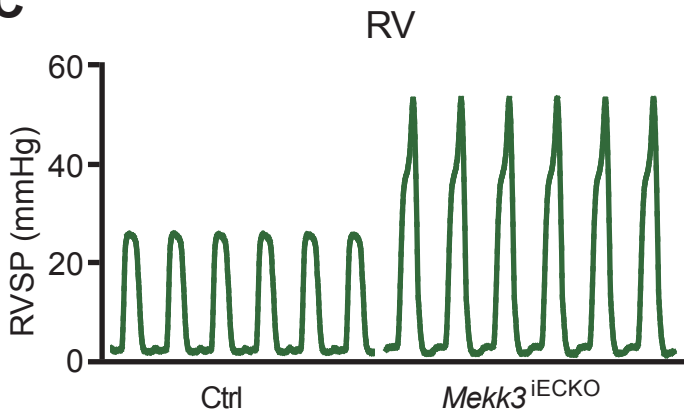
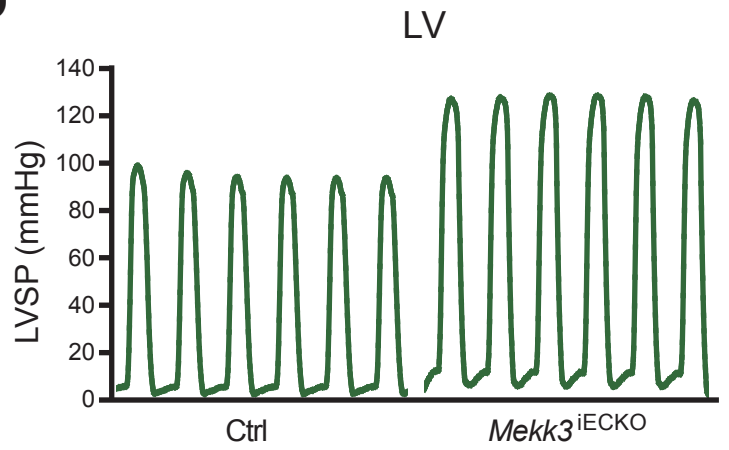
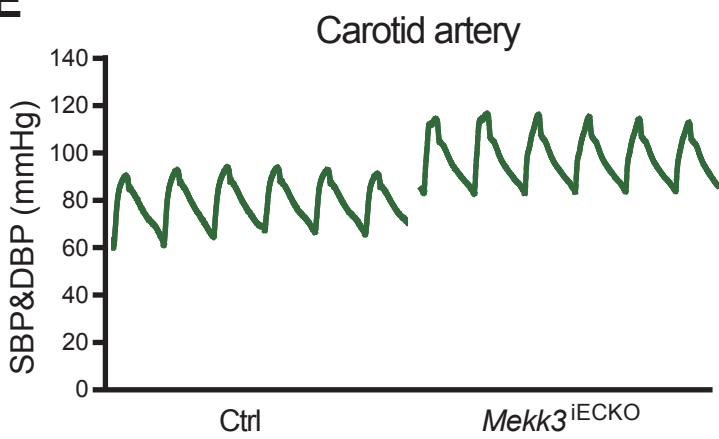
Figure S1**A****B****C****D****E**

Figure S2

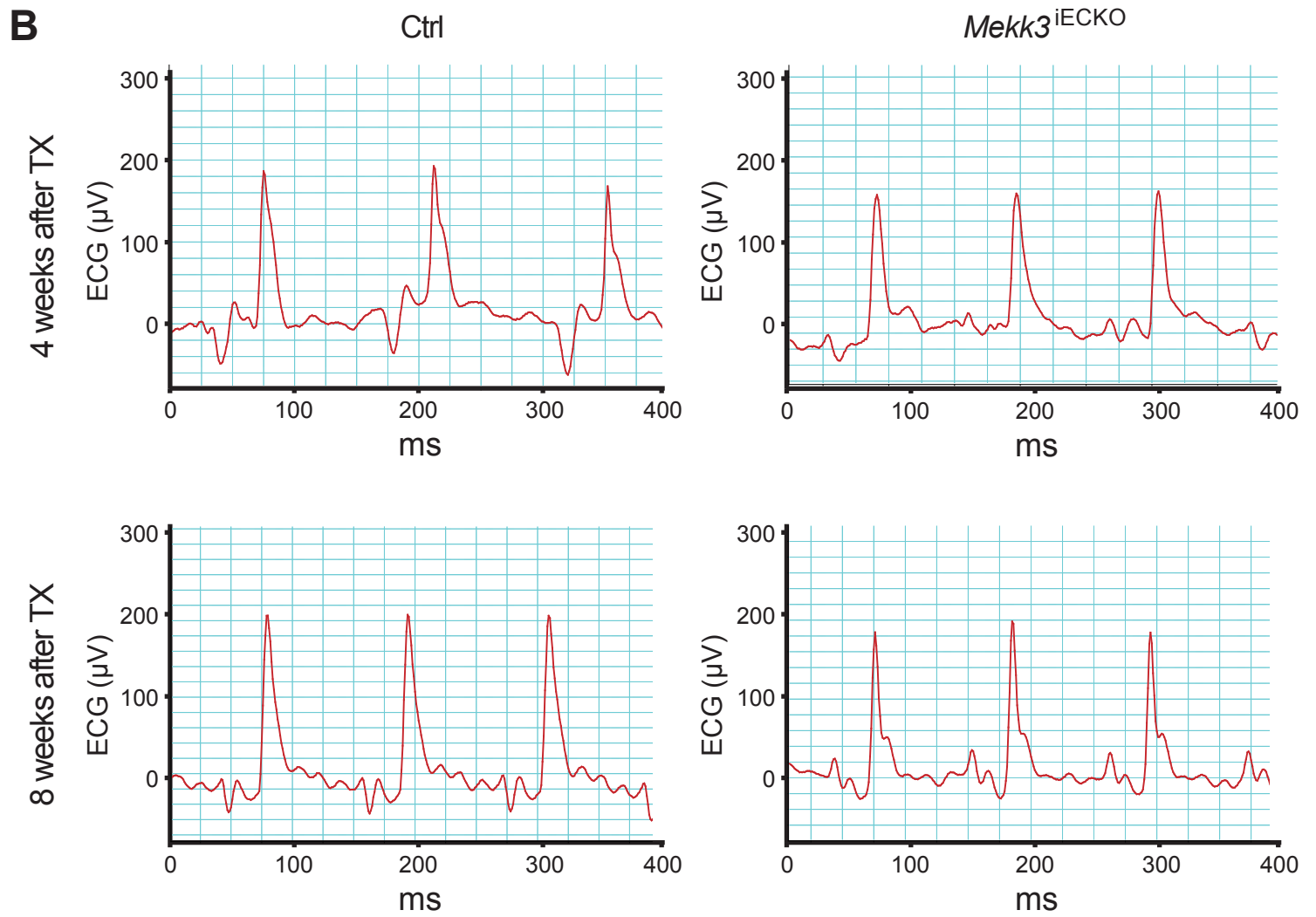
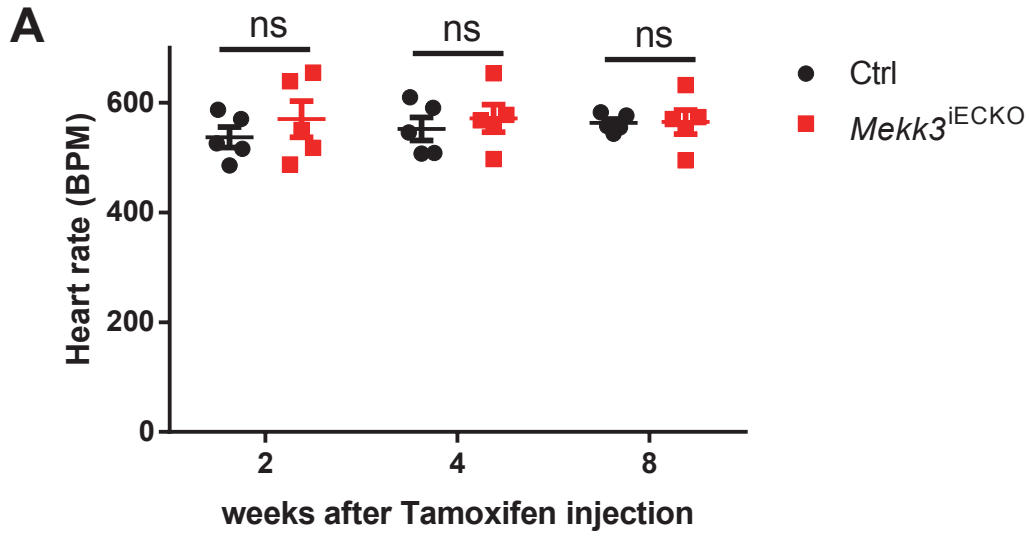
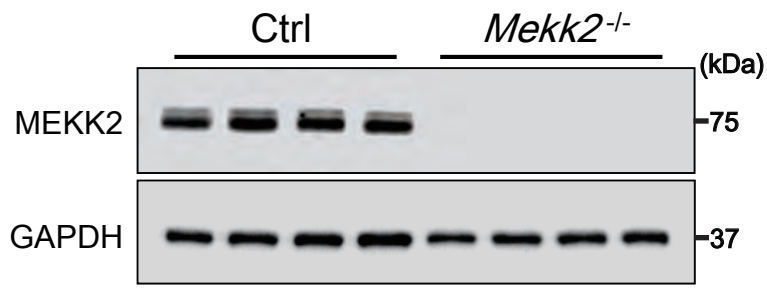
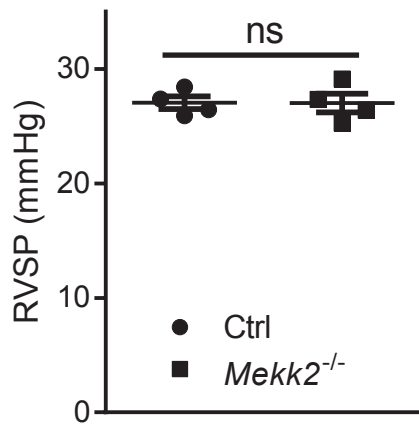


Figure S3

A



B



C

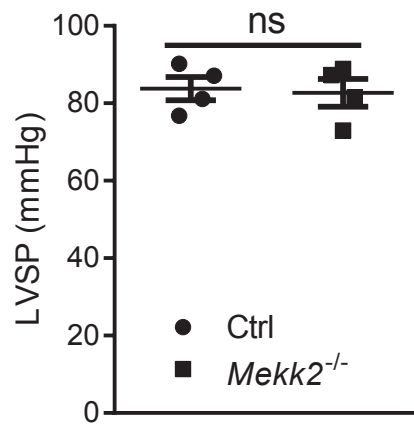


Figure S4

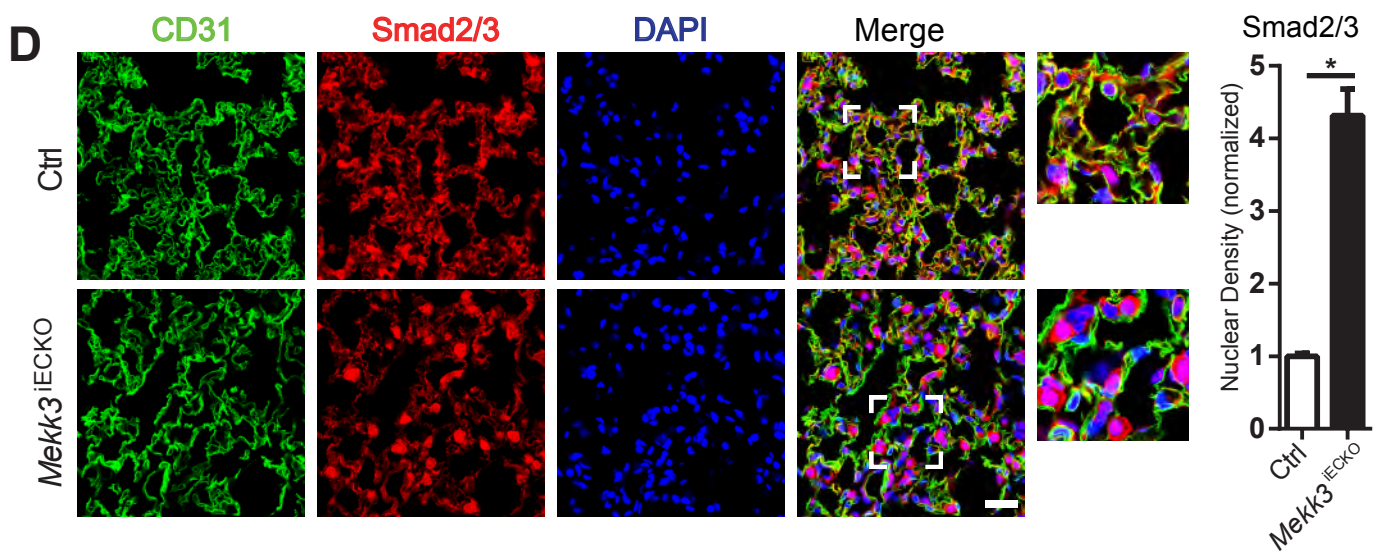
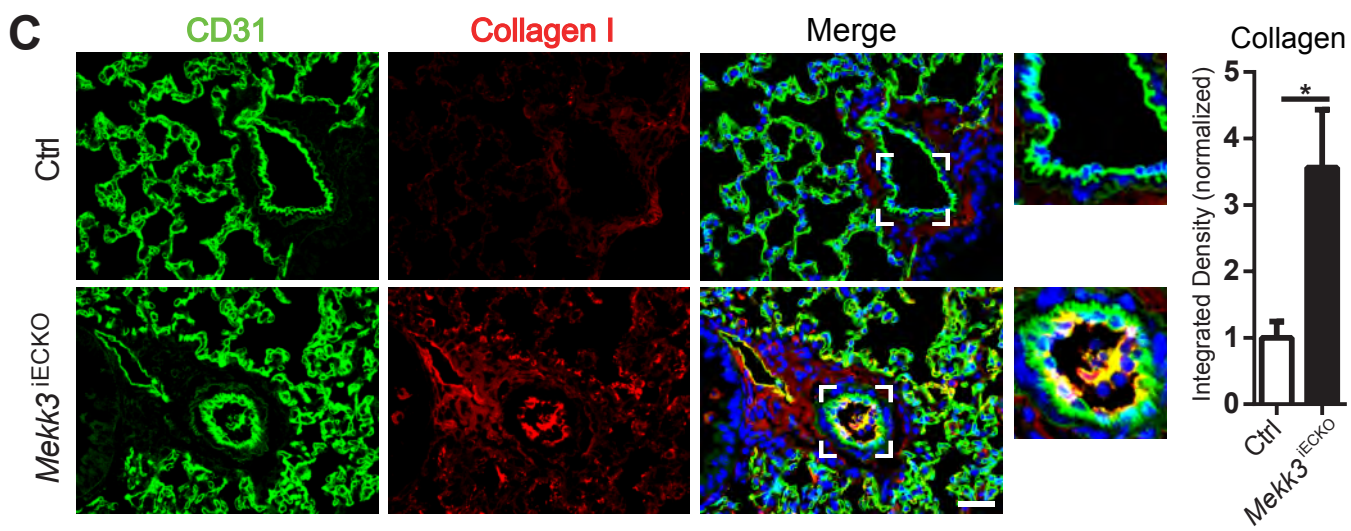
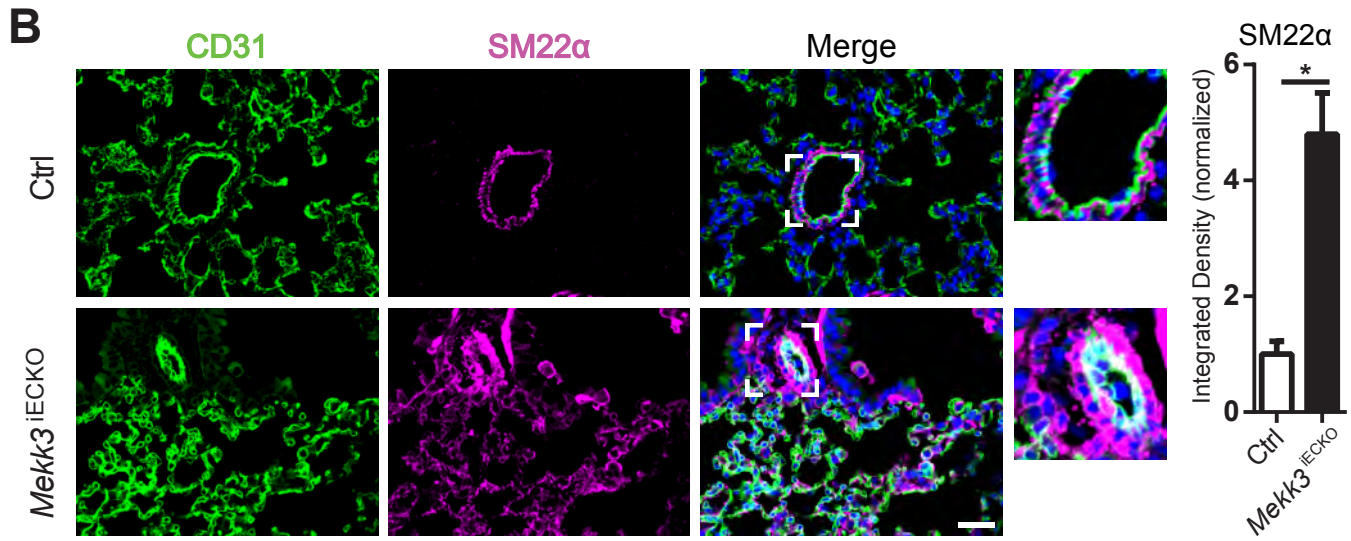
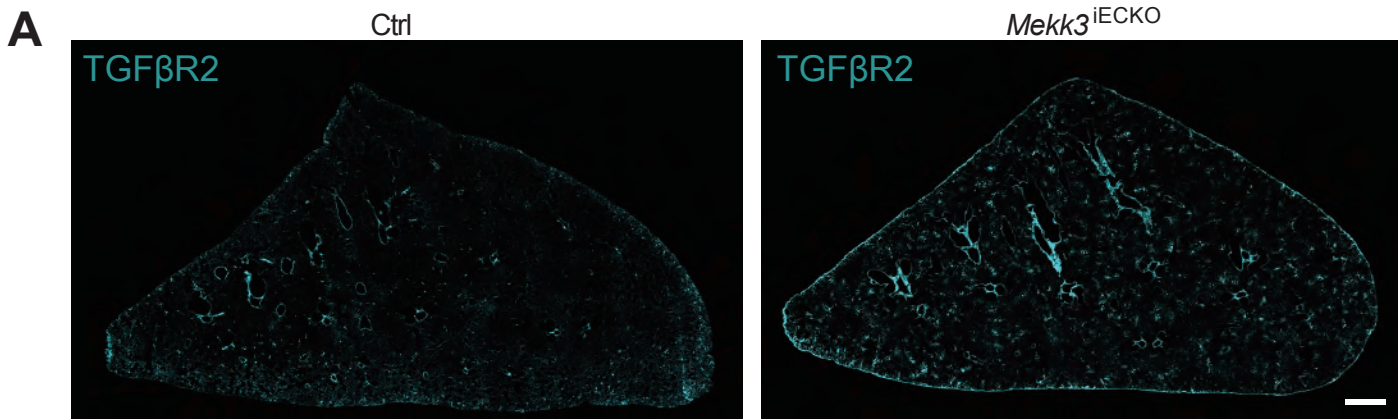


Figure S5

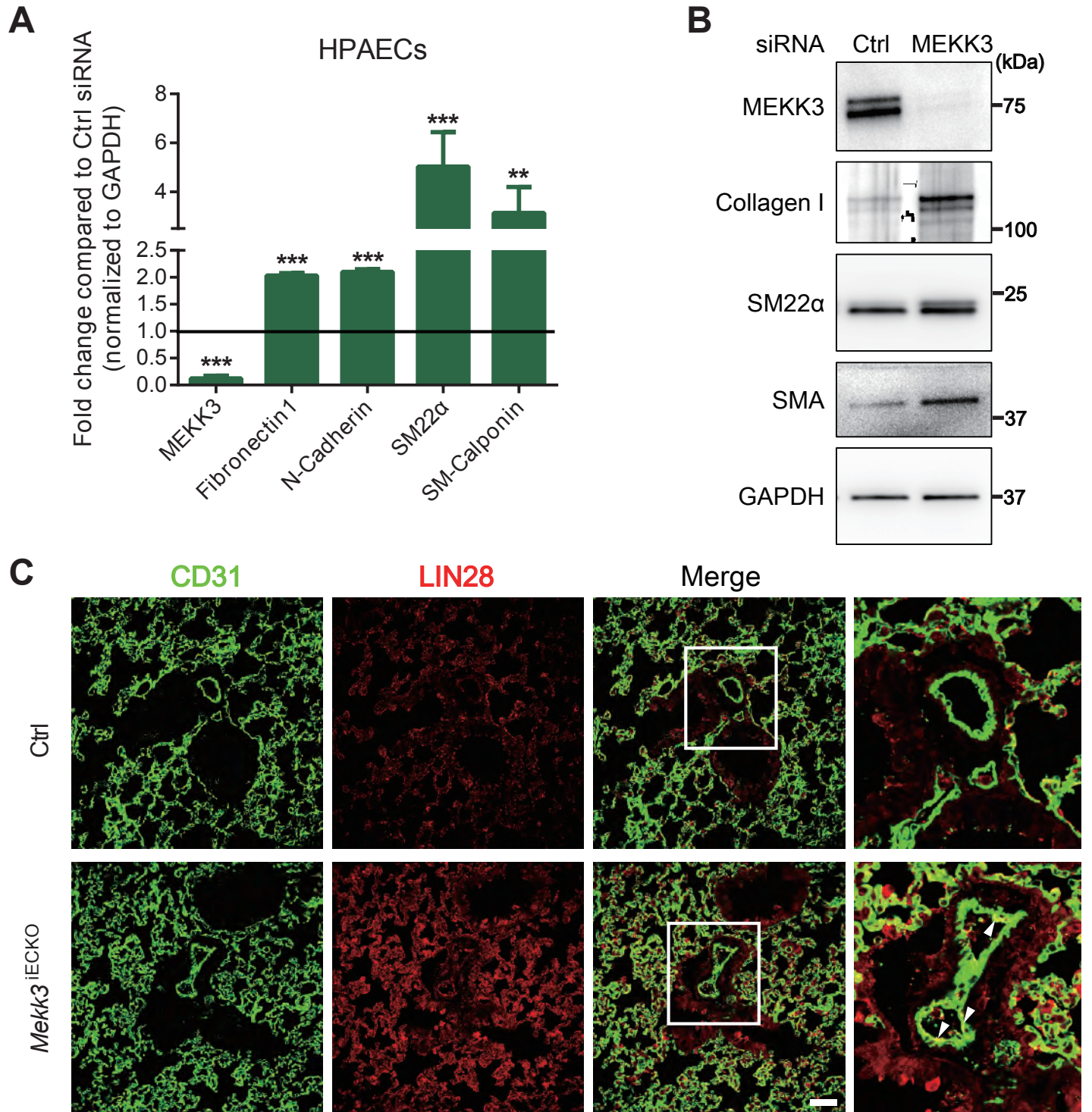


Figure S6

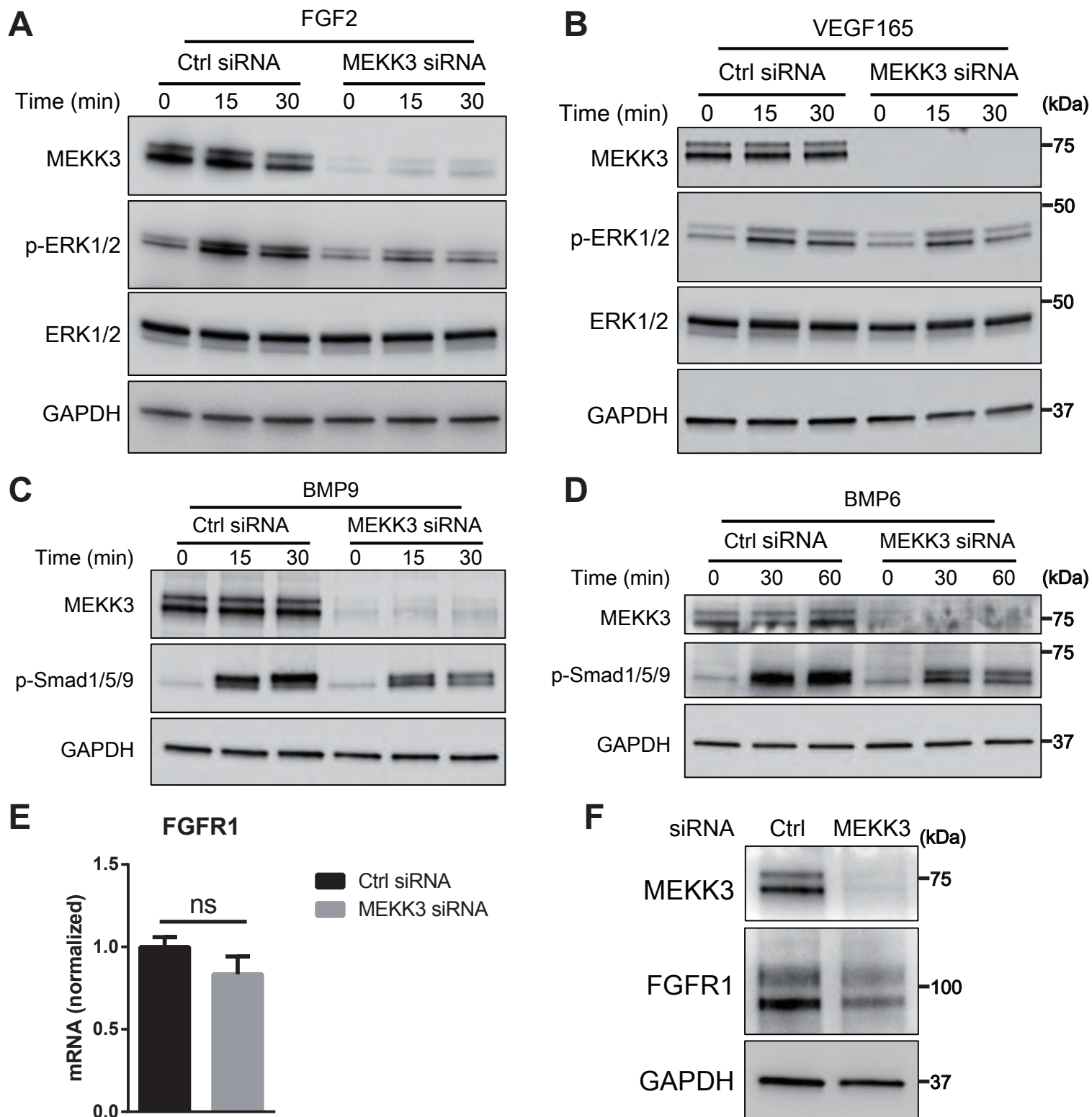


Figure S7

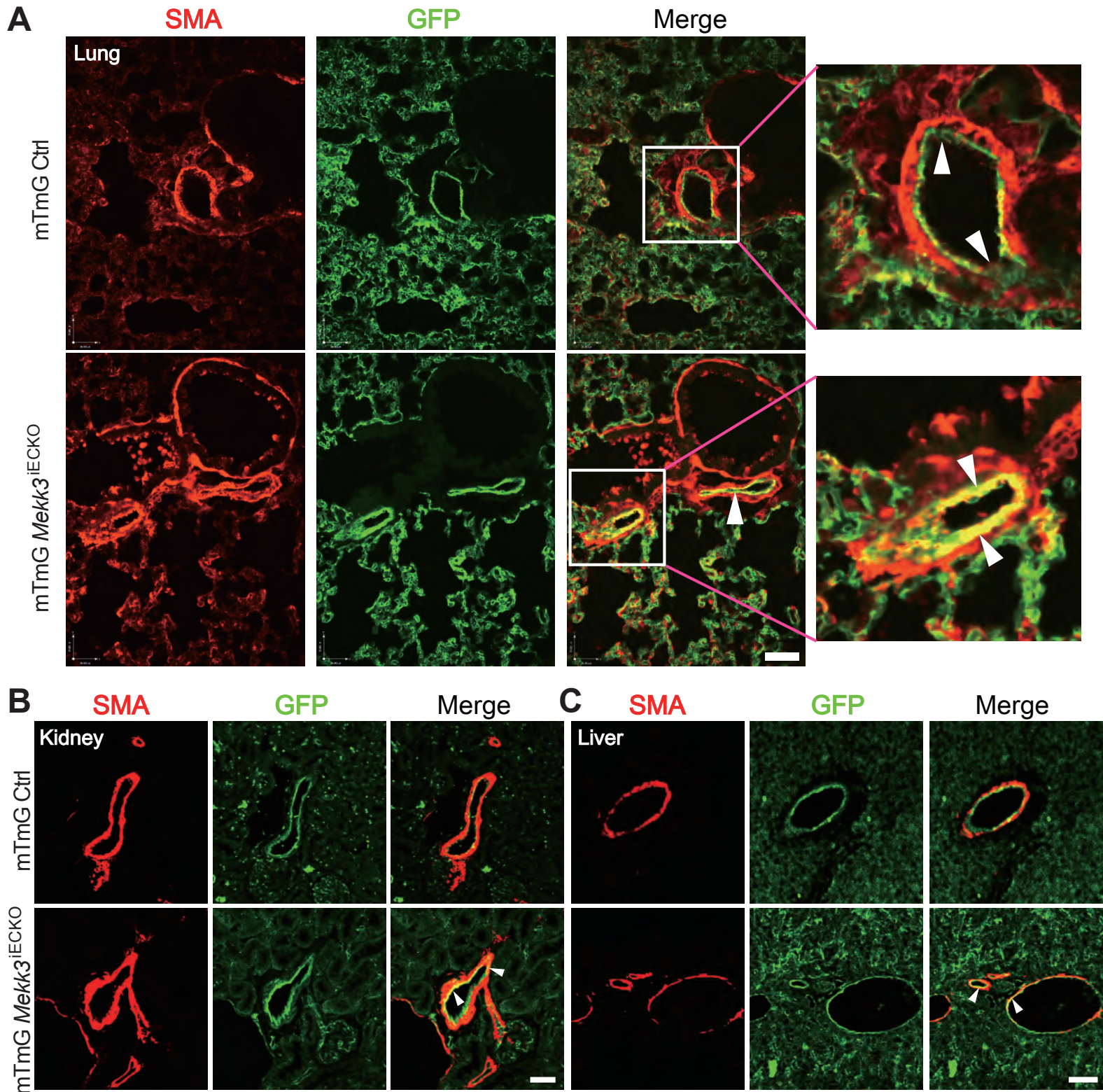


Figure S8

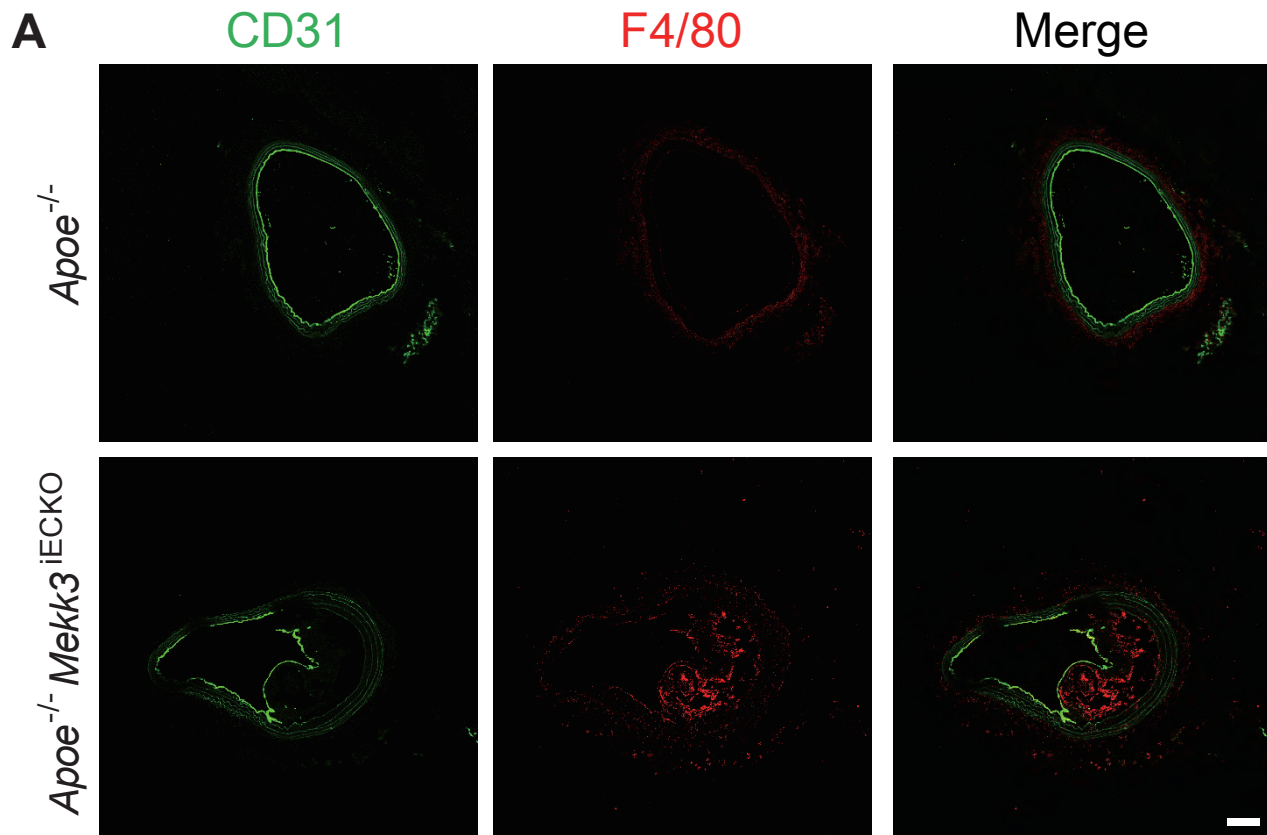
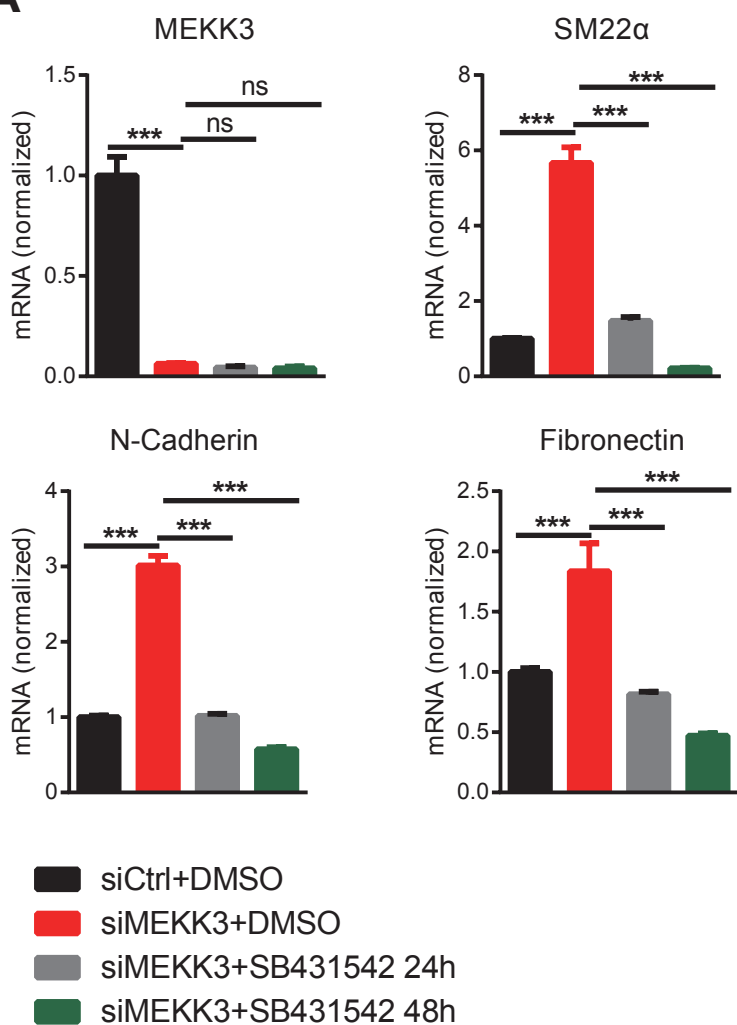
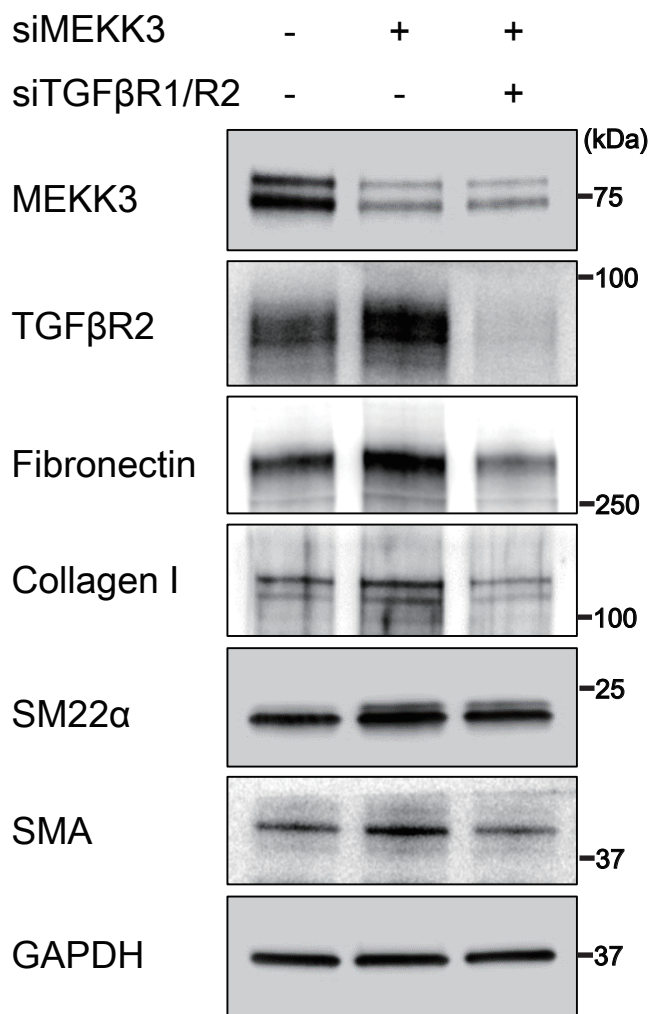


Figure S9

A



B



C

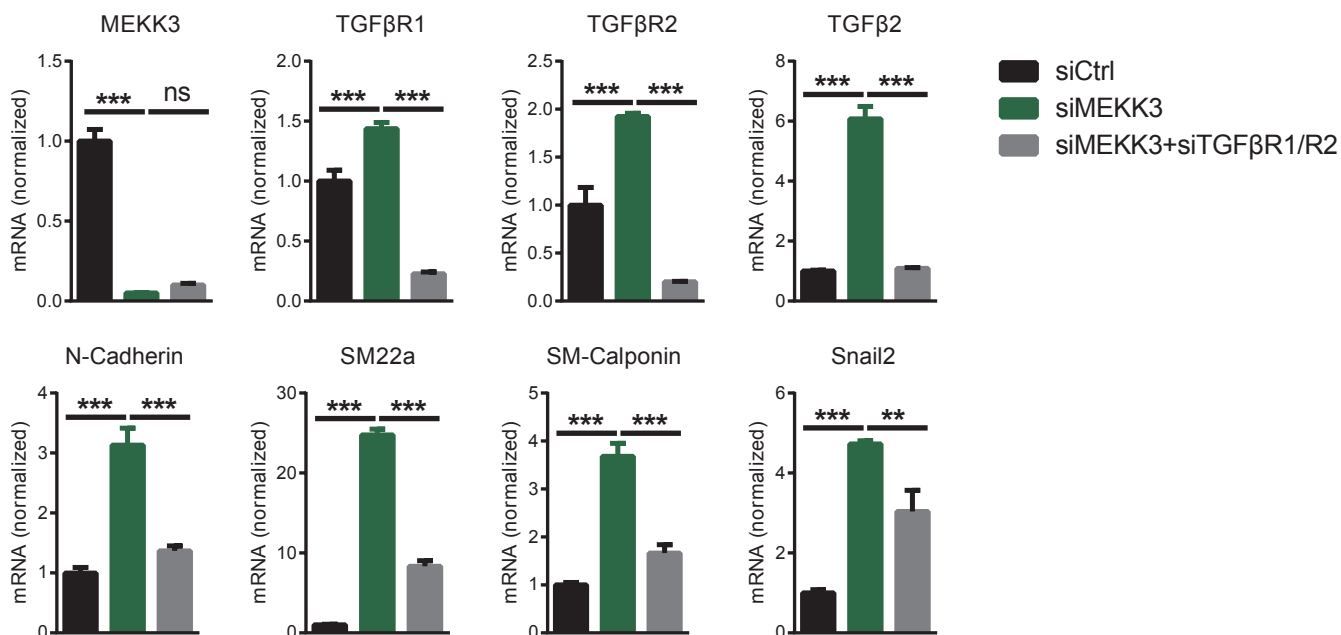
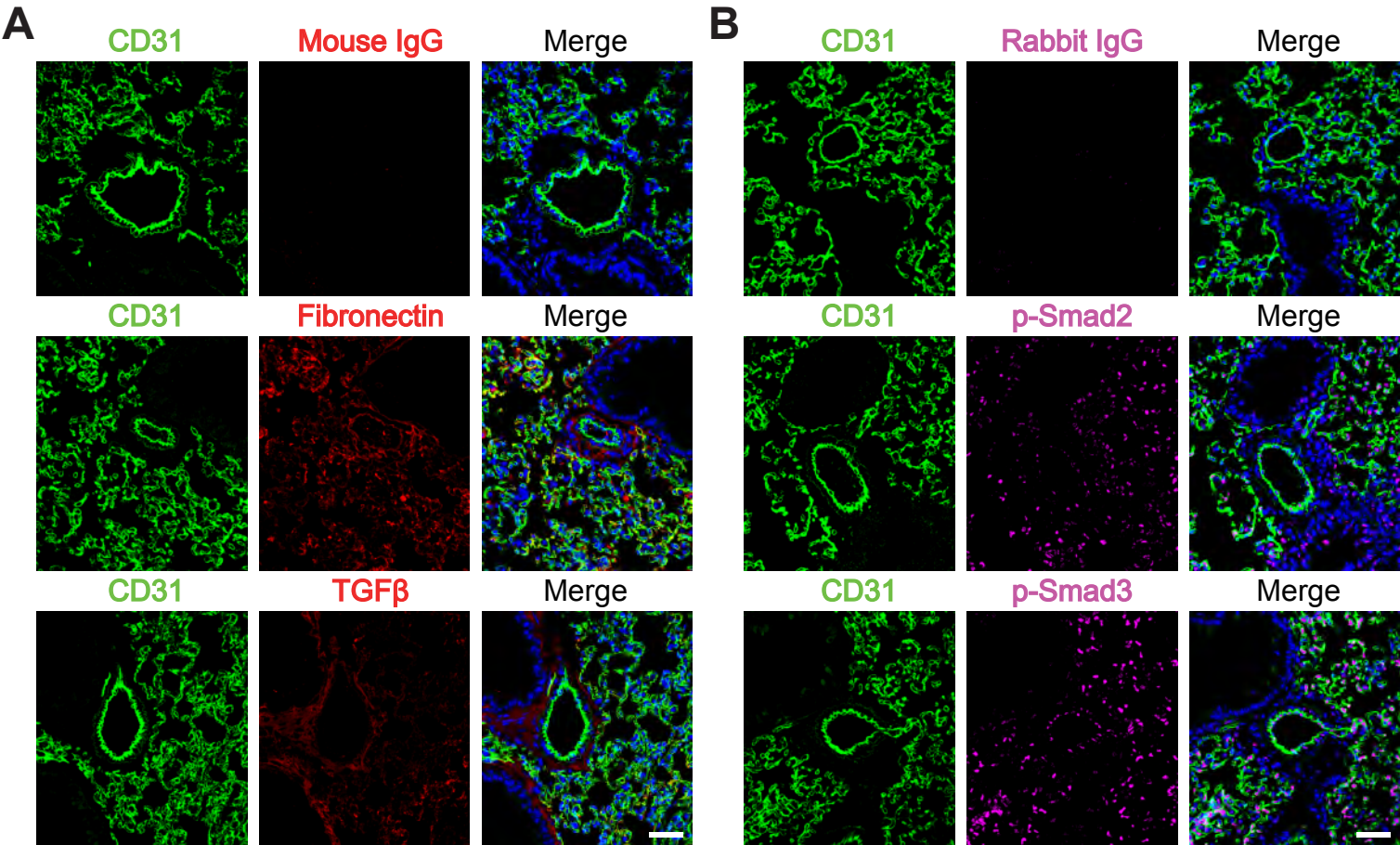


Figure S10



Supplemental Table 1. List of qPCR primers used.

Genes	Sequences (5' to 3')
hMEKK3-F	CAGACAGGAATACTCAGATCGGG
hMEKK3-R	TCTTCTGCCATCACTGTAGTCC
hGAPDH-F	GGAGCGAGATCCCTCCAAAAT
hGAPDH-R	GGCTGTTGTCATACTTCTCATGG
hFN1-F	GAGAATAAGCTGTACCATCGCAA
hFN1-R	CGACCACATAGGAAGTCCCAG
hCdh2-F	AGCCAACCTTAACTGAGGAGT
hCdh2-R	GGCAAGTTGATTGGAGGGATG
hNOS3-F	TGATGGCGAAGCGAGTGAAG
hNOS3-R	ACTCATCCATACACAGGACCC
hSnail2-F	TGTGACAAGGAATATGTGAGCC
hSnail2-R	TGAGCCCTCAGATTTGACCTG
hTGF β 1-F	GTACCTGAACCCGTGTTGCT
hTGF β 1-R	GTATCGCCAGGAATTGTTGC
hTGF β 2-F	ATGCGGCCTATTGCTTTAGA
hTGF β 2-R	GTTGGCATTGTACCCTTTGG
hTGF β 3-F	GCCTCAGTCTTTGGGATCTG
hTGF β 3-R	GTGTGAGCTGGGAAGAGAGG
hTGF β R1-F	CAGCTCTGGTTGGTGTGAGA
hTGF β R1-R	ATGTGAAGATGGGCAAGACC
hTGF β R2-F	TGAGTTCAACCTGGGAAACC
hTGF β R2-R	GGTTGATGTTGTTGGCACAC
hSM α -actin-F	CAAAGCCGGCCTTACAGAG
hSM α -actin-R	AGCCCAGCCAAGCACTG
hSM22 α -F	GATTTTGGACTGCACTTCGC
hSM22 α -R	GTCCGAACCCAGACACAAGT
hSM-calponin-F	CTGGCTGCAGCTTATTGATG
hSM-calponin-R	CTGAGAGAGTGGATCGAGGG
hLIN28a-F	AGCGCAGATCAAAAGGAGACA
hLIN28a-R	CCTCTCGAAAGTAGGTTGGCT
hLIN28b-F	CATCTCCATGATAAACCGAGAGG
hLIN28b-R	GTTACCCGTATTGACTCAAGGC
mGapdh-F	AGGTTCGGTGTGAACGGATTTG
mGapdh-R	TGTAGACCATGTAGTTGAGGTCA
mMekk3-F	GCCAATATCCTCCGAGACTCAGCTGGGAAT
mMekk3-R	CTTGAGAGCTCAGTACACTAGCTG
mNos3-F	TCAGCCATCACAGTGTTCCC
mNos3-R	ATAGCCCGCATAGCGTATCAG
mFn1-F	ATGTGGACCCCTCCTGATAGT

mFn1-R	GCCCAGTGATTTTCAGCAAAGG
mSnail2-F	TGGTCAAGAAACATTTCAACGCC
mSnail2-R	GGTGAGGATCTCTGGTTTTGGTA
mCdh2-F	AGCGCAGTCTTACCGAAGG
mCdh2-R	TCGCTGCTTTCATACTGAACTTT
mCol3a1-F	ACGTAGATGAATTGGGATGCAG
mCol3a1-R	GGGTTGGGGCAGTCTAGTG
mSm22a-F	CAACAAGGGTCCATCCTACGG
mSm22a-R	ATCTGGGCGGCCTACATCA
mTgfb2-F	TCGACATGGATCAGTTTATGCG
mTgfb2-R	CCCTGGTACTGTTGTAGATGGA
mTgfr1-F	CAGCTCCTCATCGTGTTGGTG
mTgfr1-R	GCACATACAAATGGCCTGTCTC
mTgfr2-F	CCGCTGCATATCGTCCTGTG
mTgfr2-R	AGTGGATGGATGGTCCTATTACA