Supplemental Figure 1



2 Figure S1. Gene expression analysis by RNA-Seq.

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3 (A) The distribution of samples was determined by the 1st and 2nd principal component generated 4 by principal component analysis (PCA). (B) Represented one of the dynamic expression patterns 5 of genes across Sham, MI (Day 1) and MI (Day 7), with (C) showing the functional analysis of 6 such genes by GO Biological Process based on hypergeometric distribution (the adjusted P-value 7 < 0.05).



Figure S2. Gating strategy for flow cytometric analysis.

Supplemental Figure 3





12 **Figure S3. The baseline of WT or** *Gsdmd*^{-/-} **mice leukocytes.**

Flow cytometric analysis (**A**) and quantification (**B**) of Cd11b⁺Ly6G⁺ neutrophils and Cd11b⁺Ly6C⁺ monocytes in blood from WT or *Gsdmd^{-/-}* mice at baseline without a MI surgery (n= 3-7). Data are mean \pm SD and were analyzed by unpaired two-tailed Student's *t* test (**B**). NS, not significant.

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Supplemental Figure 4







25	72 hours after AMI assessed by ELISA. The corresponding n values were indicated in the plot. (E)
26	Serum levels of LDH from WT or $Gsdmd^{-/-}$ mice at 24 hours after MI or a sham surgery. (F)
27	Serum levels of IL-1 β from WT or <i>Gsdmd</i> ^{-/-} mice at 72 hours after MI or a sham surgery. (G)
28	Representative immunoblotting images of protein levels of BM cells from WT or Gsdmd ^{-/-} mice
29	at 24 hours after MI or a sham surgery ($n = 2$ per group). (H-I) Representative immunoblotting
30	images of protein levels of neutrophils cells with or without BafA1 (200 nM) from the heart of WT
31	or $Gsdmd^{-/-}$ mice at 24 hours or 72 hours after MI (n = 2 - 3 per group). Data represent mean \pm SD
32	and were analyzed by unpaired two-tailed Student's t test (A, C and D) or One-way analysis of
33	variance followed by Tukey's multiple comparison test (E). NS, not significant; *, P<0.05; **,
34	<i>P</i> <0.01; ***, <i>P</i> <0.001.
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42 Figure S5. Bone marrow transplantation and functional analysis of mice.

(A) Immunoblotting for GSDMD validating the success of BMT experiment. (B-D) Echocardiography images (B) and M-mode quantification of ejection fraction (C) and fractional shortening (**D**) for WT or *Gsdmd*^{-/-} mice before or at 1 week after MI (baseline: WT \rightarrow WT, n = 5; *Gsdmd* KO \rightarrow WT, n = 3; WT \rightarrow *Gsdmd* KO, n=11; 1 week: WT \rightarrow WT, n = 3; *Gsdmd* KO \rightarrow WT, n = 3; WT \rightarrow Gsdmd KO, n=5). (E-F) Masson's Trichrome staining (E) and quantification (F) of fibrotic area of short-axis heart sections from WT \rightarrow WT (n = 3), Gsdmd KO \rightarrow WT (n = 3) or WT \rightarrow Gsdmd KO (n=7) mice at 1 week after MI (scale bar, 1 mm). Data are mean ±SD and were analyzed by One-way analysis of variance followed by Tukey's multiple comparison test (C, D and F). NS, not significant.

Supplementary Figure 6





(A) Flow cytometric quantification of Cd11b⁺Ly6G⁺ neutrophils in blood from WT or anti-ly6G treated mice at different time points. The corresponding n values were indicated in the plot. (B) Immunofluorescence imaging on heart sections from Isotype or Anti-ly6G mice at 24h after MI showing α -Actinin (gray), Ly6G (green), MPO (red) and DAPI (blue). Representative fields are presented (scale bar, 100 μ m). (C) Quantification of Ly6G and MPO positive area proportion in the field of heart sections from Isotype or Anti-ly6G mice, each value was averaged from the values of 5 fields of view from the same mouse (n = 4 per group). Data represent mean ± SD and were analyzed by One-way analysis of variance followed by Tukey's multiple comparison test (A)
or unpaired two-tailed Student's *t* test (C). NS, not significant; *, P<0.05; **, P<0.01; ***,
P<0.001; ****, P<0.0001.

Supplementary Figure 7



76 Figure S7. Pharmacological inhibition of GSDMD reduces infarct size post MI.

(A) Schematic diagram showing the strategy of postoperative NSA administration to the mice. (B) Kaplan-Meier survival curves comparing post-MI survival of control (DMSO administration) mice (n = 13) to that of mice administrated with NSA (n = 17). Significance was determined by Mantel-Cox test. (C-D) Echocardiography images (C) and M-mode quantification (D) of ejection fraction (left) and fractional shortening (right) for control mice or mice with NSA administration before or at 1 week after MI. (baseline: DMSO, n = 12; NSA, = 10; 1 week: DMSO, n = 6; NSA, n

83 = 5). (E-F) Masson's Trichrome staining (E) and quantification of fibrotic area (F) of short-axis 84 heart sections from control mice or mice with NSA administration at 1 week after MI (DMSO, n = 85 6; NSA, n = 9) (scale bar, 1 mm). Data are mean \pm SD and were analyzed by One-way analysis of 86 variance followed by Tukey's multiple comparison test (**D**) or unpaired two-tailed Student's *t* test 87 (F). NS, not significant; *, P<0.05; **, *P*<0.01.

89 Supplemental Table 1: Patients' characteristics.

Demographics		n=234
Age (years)		63.97±11.991
Sex (Male/Fe	emale)	41/193
Clinical history		
Previous PCI	l, n(%)	5(2.1)
Previous CA	BG, n(%)	0(0)
Previous Stro	oke, n(%)	13(5.6)
Hypertensior	n, n(%)	111(47.4)
Diabetes, n(%	%)	41(17.5)
Dyslipidemia	a, n(%)	2(0.9)
Medications		
β-blockers, n	(%)	176(75.2)
Aspirin, n(%)	216(92.3)
Statin, n(%)		215(91.9)
ACEI/ARB,	n(%)	149(63.7)
ADP inhibito	ors, n(%)	217(92.7)

98 Supplemental Table 2: Key Resources and Reagents

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Anti-GSDMD	Abcam	Ab209845
Anti-CD68	Abcam	Ab53444
Anti-Sarcomeric Alpha Actinin	Abcam	Ab9465
Caspase-1+p10+p20	Abcam	Ab179515
Anti-mouse Caspase-1	Adipogen	AG-20B-0042-C100
Anti-mouse IL-1β	R&D Systems	AF-401-NA
MPO	R&D Systems	AF3667
Bcl-2	SantaCruz	Sc-7382
Anti-mouse NLRP3	Cell Signaling	1510S
SQSTM1/P62 Antibody	Cell Signaling	5114S
LC3 A/B Antibody	Cell Signaling	4108S
Caspase-3	Cell Signaling	9662S
HSP90	Cell Signaling	4877S
β-actin	Cell Signaling	4970S
β-tubulin	Cell Signaling	2146S
GAPDH	Cell Signaling	3683S
anti-Ly6G/Ly6C antibody	BioLegend	108436
CD45-BV605	BioLegend	103140
CD11b-AlexaFluor647	BioLegend	101218
Ly6G-FITC	BioLegend	127606
Ly6C-Perp-cy5.5	BioLegend	128012
Anti-Ly6G antibody	BioLegend	127632
Goat anti-mouse IgG, HRP	Invitrogen	31430
Goat anti-rabbit IgG, HRP	Invitrogen	31460
TEXAS-Red goat anti-rat IgG	Invitrogen	T2767
Alexa Fluor 488 goat anti-rat IgG	Invitrogen	A11034
Alexa Fluor 594 goat anti-mouse IgG	Invitrogen	A11032
Alexa Fluor 488 goat anti-rat IgG	Invitrogen	A11006
Alexa Fluor 555 donkey anti-goat IgG	Invitrogen	A21432
Alexa Fluor 488 goat anti-mouse IgG	Invitrogen	A11001
Alexa Fluor 647 goat anti-rat IgG	Invitrogen	A21247
CD11b microbeads UltraPure mouse	Miltenyi	130-126-725
Neutrophil separation kit	Miltenyi	130-097-658
Mouse chow	Changzhou SYSE Bio-Tec.Co.Ltd	PD450J
Bafilomycin A1	MCE	HY-100558