

Supplemental Figures and Tables Captions

Table S1. Recombinant protein composition of human cardiac reconstituted thin filaments. All cTn and RTF utilized in this study were labeled with IAANS at T53C position with the two additional mutations C35S and C84S.

Table S2. Oligonucleotide sequences for sgRNA, ssODN and PCR primers used for hiPSC genome editing.

Table S3. List of 251 transcripts in the customized NanoString codeset.

Figure S1. Sequence alignment of the partial cTnT protein among different species. A multiple sequence alignment of the partial canonical cTnT protein from human (Uniprot ID: P45379), bovine (Uniprot ID: P13789), rabbit (Uniprot ID: P09741), rat (Uniprot ID: P50753), mouse (Uniprot ID: P50752), chicken (Uniprot ID: P02642), and zebrafish (Uniprot ID: Q90Y46) was performed using Clustal Omega. 179 residues are boxed in red and are highly conserved across different species. The human cTnT sequence shown here includes exon 5 which encodes 10 amino acids, which are boxed in blue. Exon 5 is not included in the splice variant normally expressed in the adult heart and so the residue numbers are shifted by ten in this figure.

Figure S2. Genome-editing to create *TNNT2* I79N^{+/-} hiPSC line. Panel A. Genome editing strategy map in which the relevant portion of the *TNNT2* nucleotide sequence is shown. Residue 79 is within exon 9 and the annealing position of the designed single guide RNA (sg RNA) is shown in orange and the PAM site (TGG) in pink. The donor DNA or single-stranded oligodeoxynucleotide (ssODN) spanned the region shown by the brown (36 bp PAM distal) and green (91 bp PAM proximal) arrows. Panel B. Sanger sequencing of relevant nucleotides. The gene edited nucleotide is shown by the red arrow in which the codon for Isoleucine (I) ATC was genome edited to AAC (Asparagine (N)) in one allele rendering a I79N heterozygous mutant. The blue arrow indicates the silent mutation (C->G) that was included in the sgRNA target sequence to prevent CRISPR from re-cutting the target sequence once the desired edit was introduced. The top 10 sites that scored high for off target or low for on-target were selected using “Custom Alt-R® CRISPR-Cas9 guide RNA tool” from IDT. The top 10 off target sites were then Sanger sequenced to ensure that the probability of off target mutations was minimized.

Figure S3. The expression of *TNNI3/TNNI1* in WT and I79N^{+/-} hiPSC-CMs.

Figure S4. Heat map and Principal Component Analysis (PCA) of data from NanoString multiplex gene expression analysis. Panel A, Heat map of unsupervised hierarchical clustering of all 251 genes. I79N^{+/-} mutation is clustering away from WT with low variability within group. Patterns of gene expression are evident through the effect of the mutation. Panel B PCA of normalized gene expression for I79N^{+/-} (blue) vs WT (black) hiPSC-CMs shows PC1 and PC2 account for 73% and 16% of the variance, respectively. In PCA, I79N^{+/-} samples are clustered away from WT.

Figure S5. NanoString multiplex gene expression functional analysis. Panels A and B. Top 5 functional pathways (i.e., enrichment term) based on adjusted p-value (< 0.05) across six gene set libraries. Enrichment analysis was conducted using the 45 upregulated and 23 downregulated differentially expressed genes (DEG) in I79N^{+/-} hiPSC-CMs compared to WT hiPSC-CMs. Panels C and D. Top 5 enriched Gene Ontology (GO) terms sorted by adjusted p-value. X-axis represents the combined score (combination of the p-value and the z-score). Y-axis refers to the names of pathways terms split by the gene set libraries. Data point size represents the Odds ratio which is the estimate from Fisher’s exact test calculated

through Enrichr (Chen et al., 2013; Kuleshov et al., 2016). Color gradient refers to the adjusted p-value of each term. GO_BP: Gene Ontology biological process, GO_CC: Gene Ontology cellular component, GO_MF: Gene Ontology molecular function, Hu_Phenotype: human phenotype, Jensen_Diseases: disease database from Jensen Lab, MGI_Phenotype: Mammalian Phenotype Ontology. I79N^{+/-} (n=3) to WT (n=4) TNNT2 hiPSC-CMs.

Table S1.

Construct	Gene Name	Description	Length	Molecular Weight (kDa)*	Extinction Coefficient (M ⁻¹ cm ⁻¹)*	Uniprot ID
# WT cTnC ^{T53C}	<i>TNNC1</i>	Full length TnC with three mutations (T53C, C35S, C84S) for fluorescence labeling specifically at T53C for functional studies.	161	18	4470	P63316
Troponin I and Troponin T Subunits						
WT cTnI	<i>TNNI3</i>	Full length WT cTnI.	211	24	9970	P19429
WT cTnT	<i>TNNT2</i>	Full length WT cTnT (isoform 3).	288	35	16960	P45379
I79N cTnT	<i>TNNT2</i>	Full length cTnT with I79N mutation.				
Other constructs						
Alpha MAS-Tm	<i>TPM1</i>	Full-length Tm with the additional tripeptide (Met-Ala-Ser) to the N-terminus to mimic acetylation process in native Tm and ensure its binding to actin filaments.	287	33	8940	P09493

Both the molecular weight and extinction coefficients were calculated using ProtParam¹.

All cTn and RTF utilized in this study were labeled with IAANS at T53C position with the two additional mutations C35S and C84S.

¹Gasteiger, E. et al. Protein Identification and Analysis Tools on the ExPASy Server. *The Proteomics Protocols Handbook*, 571 (2005).

Table S2. Oligonucleotide sequences for sgRNA, ssODN and PCR primers used for hiPSC genome editing

sgRNA	
I79N - F	5'-CACCGAGTCCACTCTCTCTCCATCG-3'
I79N - R	5'-AAACCGATGGAGAGAGAGTGGACTC-3'
ssODN	
I79N	5' AGTCCCTGGGTCCAGAATGGGGCTGATGCTGACTATTCCTCTCTCCAACAGGTCGTTTCATGCCCAACTTGGTGCCTCCAAGAATCC CGATGGAGAGAGAGTCTGACTTTGATGTAAGCGGTGGCTGT -3'
PCR primers for sequencing	
I79 - F	5'-GCTTCTTTGATTCCAAGTTGTGTG-3'
I79 - R	5'-CCCATCCCACCTATGCTC-3'

NanoString Custom Codeset

Table S3.

	Gene Name	Accession no.	Probe Position			Gene name	Accession no.	Probe Position
1	ABCC8	NM_000352.4	1761-1860		33	CALM3	NM_005184.2	1306-1405
2	ACADM	NM_000016.5	1290-1389		34	CALR	NM_004343.2	966-1065
3	ACTA1	NM_001100.3	46-145		35	CaMK2A	NM_171825.1	4081-4180
4	B actin	NM_001101.2	1011-1110		36	CaMK2B	NM_001220.3	366-465
5	ACTC1	NM_005159.4	2006-2105		37	CaMK2D	NM_001221.3	1906-2005
6	ACTN2	NM_001103.2	413-512		38	CaMK2G	NM_001222.2	234-333
7	ADRB1	NM_000684.1	796-895		39	CASQ2	NM_001232.3	598-697
8	ADRB2	NM_000024.3	1246-1345		40	CAV3	NM_001234.3	212-311
9	AKT	NM_0010144	1276-1375		41	CER1	NM_005454.2	589-688
10	ALDH1A1	NM_000689.3	12-111		42	CHRM2	NM_0010066	1171-1270
11	ALDH1A2	NM_003888.2	3131-3230		43	CKB	NM_001823.4	321-420
12	ALDH1A3	NM_000693.2	2281-2380		44	CKM	NM_001824.4	136-235
13	CD13	NM_001150.1	2671-2770		45	COL1A1	NM_000088.3	5211-5310
14	JCTN or	NM_032466.3	396-495		46	COL2A1	NM_001844.4	4746-4845
15	ATP1A2	NM_000702.3	4041-4140		47	COL3A1	NM_000090.3	181-280
16	ATP1A3	NM_152296.4	2626-2725		48	COL4A2	NM_001846.2	1151-1250
17	ATP2A2	NM_001681.3	1397-1496		49	COL4A3	NM_000091.4	1085-1184
18	ATP5F1	NM_001688.4	646-745		50	COL9A2	NM_001852.3	2414-2513
19	B2M	NM_004048.2	26-125		51	CPT1B	NM_004377.3	1655-1754
20	BMP10	NM_014482.1	441-540		52	CRHR2	NM_001883.2	236-335
21	BMP2	NM_001200.2	1516-1615		53	CS	NM_004077.2	741-840
22	BMP4	NM_001202.3	660-759		54	CTNNB1	NM_0010982	1816-1915
23	BRCA1	NM_007305.2	1276-1375		55	CXCR4	NM_003467.2	1336-1435
24	BTAF1	NM_003972.2	1697-1796		56	CYP26A1	NM_057157.2	1591-1690
25	BTK	NM_000061.1	571-670		57	CYP26B1	NM_019885.2	1806-1905
26	CACNA1C	NM_199460.2	4786-4885		58	DKK1	NM_012242.2	76-175
27	CACNA1D	NM_000720.2	5196-5295		59	DYNC111	NM_0011355	1183-1282
28	CACNA1G	NM_198397.1	1381-1480		60	DYNLL2	NM_080677.2	639-738
29	CACNA1H	NM_021098.2	3330-3429		61	EMILIN2	NM_032048.2	1446-1545
30	CACNA2D1	NM_000722.2	2909-3008		62	ERBB2	NM_0010058	1256-1355
31	CALM1	NM_006888.3	1684-1783		63	FGF17	NM_003867.2	280-379
32	CALM2	NM_001743.3	869-968		64	FGF19	NM_005117.2	787-886

NanoString Custom Codeset

	Gene name	Accession number	Probe Position		Gene Name	Accession Number	Probe Position
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66	FGF4	NM_002007.2	689-788	98	HRC	NM_002152.2	367-466
67	FGF8	NM_033163.3	545-644	99	IPO8	NM_006390.2	861-960
68	PKBP1B or FKBP12.6	NM_000801.4	41-140	100	IRX4	NM_016358.2	359-458
69	FLNA	NM_001456.3	7336-7435	101	ISL1	NM_002202.2	1376-1475
70	FLNB	NM_001164317.1	2125-2224	102	ISL2	NM_145805.1	109-208
71	FOXA1	NM_004496.2	2466-2565	103	JAG1	NM_000214.2	916-1015
72	FOXF1	NM_001451.2	1761-1860	104	JAK1	NM_002227.1	286-385
73	FOXH1	NM_003923.2	696-795	105	JAK2	NM_004972.3	1465-1564
74	GAPDH	NM_001256799.1	387-486	106	JAK3	NM_000215.2	1716-1815
75	GATA2	NM_032638.3	1496-1595	107	JPH2	NM_020433.4	525-624
76	GATA4	NM_002052.3	2141-2240	108	KCNA4	NM_002233.3	657-756
77	GATA6	NM_005257.3	2131-2230	109	KCNA5	NM_002234.2	2331-2430
78	GJA1	NM_000165.3	706-805	110	KCND3	NM_004980.4	1011-1110
79	GJA5	NM_005266.5	1993-2092	111	KCNH2	NM_172057.2	781-880
80	GJC1	NM_001080383.1	811-910	112	KCNJ2	NM_000891.2	621-720
81	GNAI1	NM_002069.4	671-770	113	KCNJ3	NM_001260508.1	429-528
82	GNAI2	NM_002070.2	1001-1100	114	KCNJ5	NM_000890.3	631-730
83	GNAI3	NM_006496.1	1151-1250	115	KCNJ8	NM_004982.2	881-980
84	GNAO1	NM_020988.2	1423-1522	116	KCNK1	NM_002245.3	721-820
85	GNAS	NM_080425.1	1911-2010	117	KCNK3	XM_005264293.1	667-766
86	GREM2	NM_022469.3	3006-3105	118	KCNK5	NM_003740.3	3616-3715
87	GYPA	NM_002099.4	2312-2411	119	KCNN1	NM_002248.3	703-802
88	HAND1	NM_004821.2	1231-1330	120	KCNN2	NM_021614.2	1893-1992
89	HAND2	NM_021973.2	1822-1921	121	KCNN3	NM_002249.4	2636-2735
90	HAS2	NM_005328.2	961-1060	122	KCNQ1	NM_181798.1	836-935
91	HCN1	NM_021072.2	1451-1550	123	KDR	NM_002253.2	1421-1520
92	HCN2	NM_001194.3	2075-2174	124	KIF20A	NM_005733.2	1210-1309
93	HCN4	NM_005477.2	4911-5010	125	KIT	NM_000222.1	6-105
94	HEY1	NM_012258.3	586-685	126	KLF4**	NM_004235.4	1981-2080
95	HEY2	NM_012259.2	1576-1675	127	KMT2A	NM_005933.2	14001-14100
96	HIF1A	NM_001530.2	1986-2085	128	KMT2B	NM_014727.1	7846-7945

NanoString Custom Codeset

	Gene Name	Accession Number	Probe Position			Gene	Accession	Probe Position
129	KMT2C	NM_170606.2	11974-12073		161	NR2F1	NM_005654.4	3111-3210
130	KMT2D	NM_003482.3	6071-6170		162	NR2F2	NM_021005.2	1531-1630
131	LDHA	NM_001165414.1	1691-1790		163	ORAI1	NM_032790.3	911-1010
132	LEFTY1	NM_020997.2	1406-1505		164	PDGFRA	NM_006206.3	1926-2025
133	LIN28	NM_024674.4	1961-2060		165	PDK2	NM_002611.3	436-535
134	JNK3	NM_138981.2	451-550		166	PDK4	NM_002612.3	1676-1775
135	JNK1-alpha	NM_002750.2	946-1045		167	p110a	NM_006218.2	2446-2545
136	JNK2-alpha	XM_005265940.1	164-263		168	PITX2	NM_000325.5	1382-1481
137	MEF2c	NM_002397.3	2446-2545		169	PKP2	NM_001005242.2	1692-1791
138	MESP1	NM_018670.3	865-964		170	PLCB1	NM_182734.1	171-270
139	MIXL1	NM_031944.1	419-518		171	PLCB2	NM_004573.2	411-510
140	IP3R	NM_130385.2	3056-3155		172	PLCD1	NM_006225.3	2126-2225
141	MSX2	NM_002449.4	31-130		173	PLCD3	NM_133373.4	1308-1407
142	MYBPC3	NM_000256.3	3429-3528		174	PLCE1	NM_001165979.1	393-492
143	MYC**	NM_002467.3	1611-1710		175	PLCG1	NM_002660.2	2291-2390
144	MYH6	NM_002471.3	928-1027		176	PLCG2	NM_002661.2	526-625
145	MYH7	NM_000257.2	1917-2016		177	PLN	NM_002667.3	213-312
146	MYL2	NM_000432.3	721-820		178	POLR2A	NM_000937.2	3776-3875
147	MYL7	XM_005249817.2	339-438		179	POU3F4	NM_000307.3	78-177
148	MYO1E	NM_004998.2	4556-4655		180	POU5F1	NM_002701.4	1226-1325
149	NANOG**	NM_024865.2	1101-1200		181	PPIA	NM_021130.3	316-415
150	NCAM1	NM_000615.5	1621-1720		182	PPP1CA	NM_002708.3	506-605
151	NFATC4	NM_001136022.2	2297-2396		183	PPP2CA	NM_002715.2	1076-1175
152	NKX2-5	NM_004387.3	219-318		184	PPP3CB	NM_021132.2	271-370
153	NNT	NM_012343.3	2431-2530		185	PRKACA	NM_002730.3	401-500
154	NODAL	NM_018055.3	321-420		186	PRKACB	NM_182948.2	806-905
155	NOTCH1	NM_017617.3	8212-8311		187	PRKACG	NM_002732.2	776-875
156	NOTCH2	NM_024408.3	2843-2942		188	PRKAR1A	NM_212472.1	368-467
157	NOTCH3	NM_000435.2	1966-2065		189	PRKAR1B	NM_001164759.1	1113-1212
158	NOTCH4	NM_004557.3	6421-6520		190	PRKAR2A	NM_004157.2	479-578
159	NPPA	NM_006172.2	116-215		191	PRKAR2B	NM_002736.2	1351-1450
160	NPPB	NM_002521.2	497-596		192	PRKCA	NM_002737.2	681-780

NanoString Custom Codeset

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194	PRKCG	NM_002739.3	446-545		226	TEC	NM_003215.1	1231-1330
195	PKG1	NM_006258.3	995-1094		227	CD90	NM_006288.2	136-235
196	PTGS2	NM_000963.1	496-595		228	TNNI1	NM_003281.3	976-1075
197	RARA	NM_001033603.1	1191-1290		229	TNNI3	NM_000363.4	226-325
198	RARB	NM_000965.3	2756-2855		230	TNNT2 exon 5	NM_000364.3	132-231
199	RARG	NM_000966.3	1541-1640		231	TNNT2	NM_001276346.1	797-896
200	RBM20	NM_001134363.1	6943-7042		232	TOP2A	NM_001067.3	3564-3663
201	ROR2	NM_004560.2	736-835		233	TPM1	NM_000366.5	843-942
202	RPS13	NM_001017.2	332-431		234	TRDN	NM_001251987.1	297-396
203	RYR2	NM_001035.2	1184-1283		235	TRH	NM_007117.2	1883-1982
204	SCN4A	NM_000334.4	6787-6886		236	TRPC1	NM_003304.4	991-1090
205	SCN5A	NM_198056.2	8341-8440		237	TRPC2	NR_002720.2	561-660
206	SGK1	NM_005627.2	1791-1890		238	TRPC3	NM_001130698.1	1479-1578
207	SHOX2	NM_003030.3	830-929		239	TRPC4	NM_001135956.1	2436-2535
208	SIRPA	NM_080792.2	3116-3215		240	TRPC5	NM_012471.2	3686-3785
209	SLC16A1	NM_003051.3	636-735		241	TRPC6	NM_004621.5	1001-1100
210	SLC2A1/GLUT1	NM_006516.2	2501-2600		242	TRPC7	NM_001167576.1	1119-1218
211	SLC2A4/GLUT4	NM_001042.2	2121-2220		243	TSHR	NM_001018036.2	736-835
212	SLC8A1	NM_021097.1	441-540		244	SSEA5	NM_001270483.1	1007-1106
213	SLN	NM_003063.2	376-475		245	TTN b	NM_003319.4	1805-1904
214	SOX2	NM_003106.2	152-251		246	TTN	XM_024453094.1	33791-33890
215	SPCS1	NM_014041.3	611-710		247	VCAM-1	NM_001078.3	2536-2635
216	STIM1	NM_003156.3	3101-3200		248	VEGFA	NM_001025366.1	1326-1425
217	TACR1	NM_015727.1	856-955		249	WNT3	NM_030753.3	1336-1435
218	TACR3	NM_001059.1	471-570		250	WNT3A	NM_033131.2	699-798
219	TBX1	NM_080646.1	541-640		251	ZFP42 (or REX1)	NM_174900.3	946-1045
220	TBX18	NM_001080508.2	545-644					
221	TBX2	NM_005994.3	936-1035					
222	TBX20	NM_020417.1	661-760					
223	TBX3	NM_005996.3	3276-3375					
224	TBX5	NM_080718.1	246-345					

Figure S1.

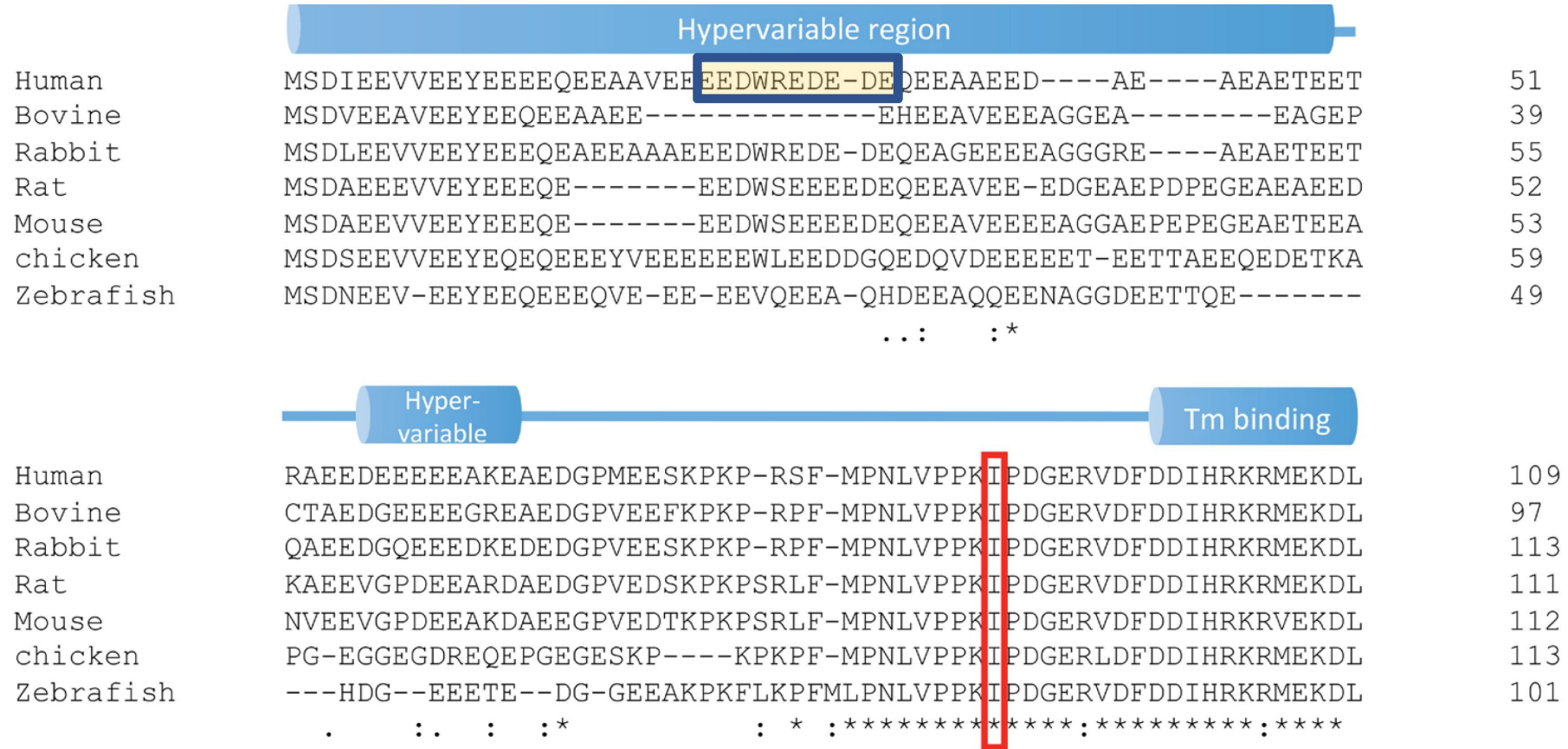
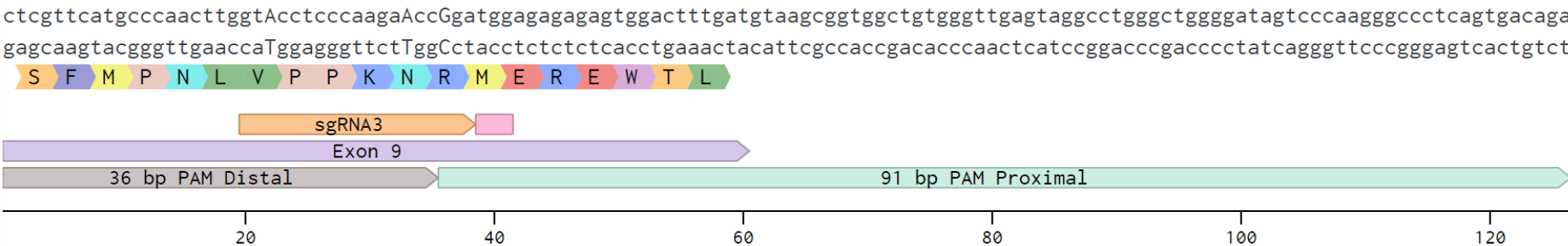


Figure S2.

A.



B.

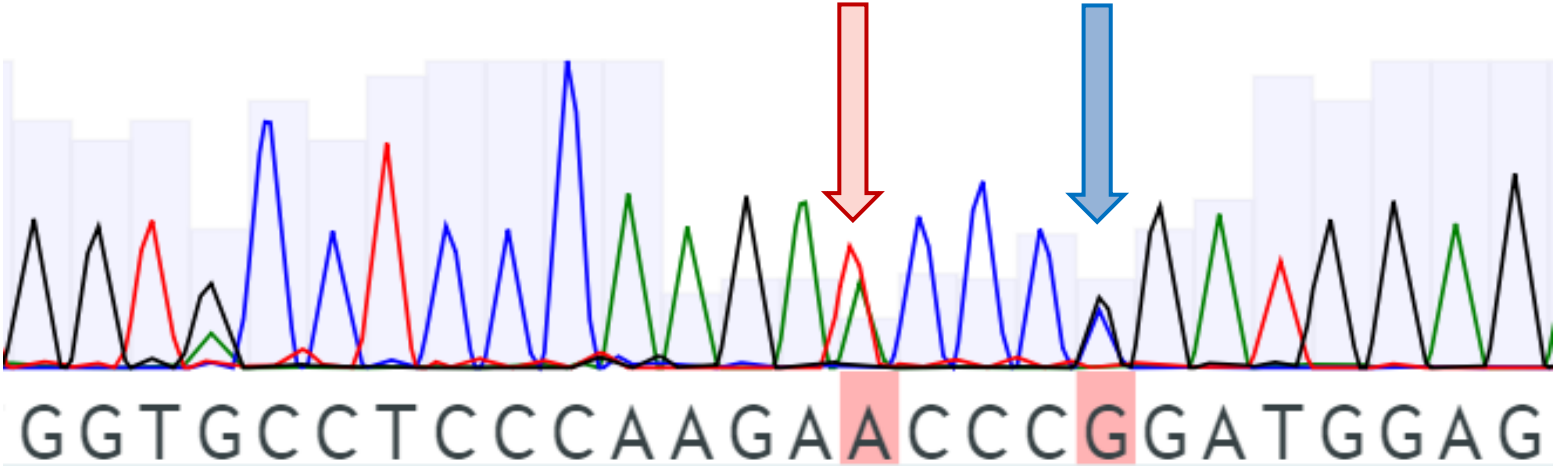


Figure S3.

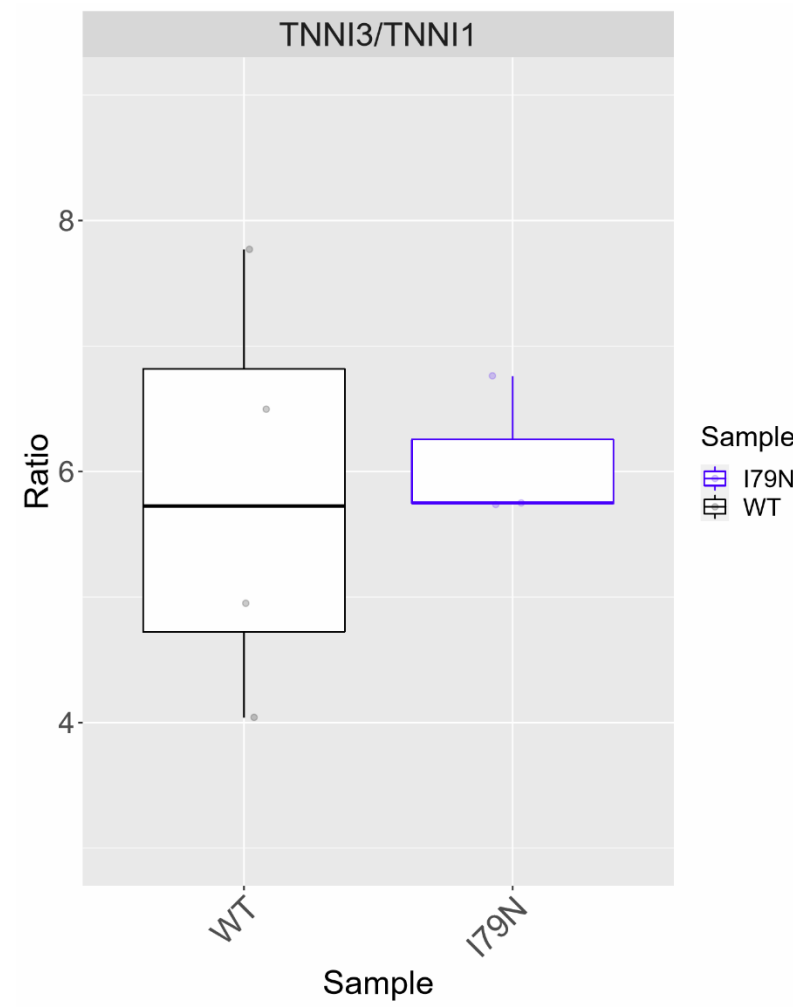
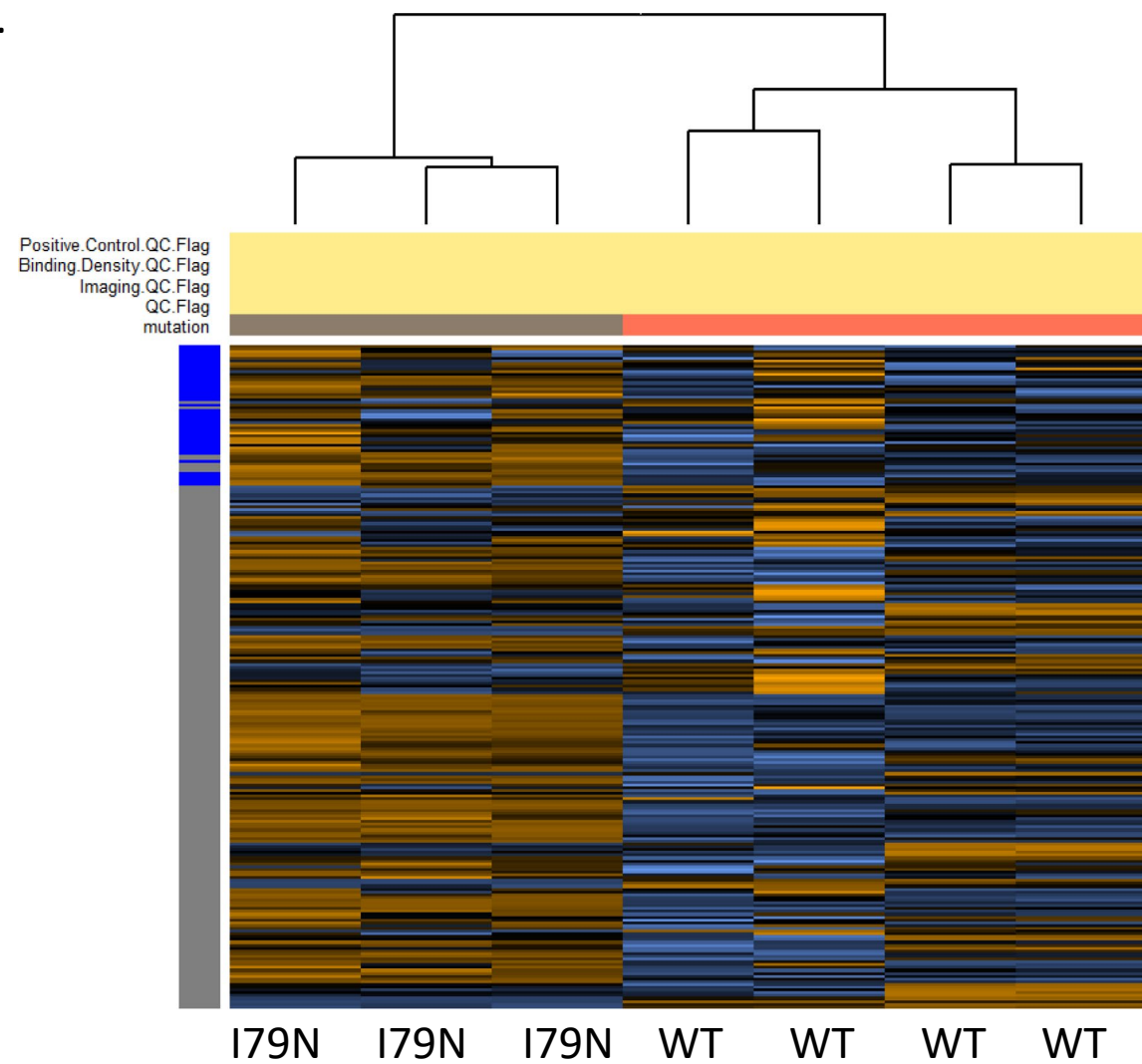


Figure S4.

A.



B.

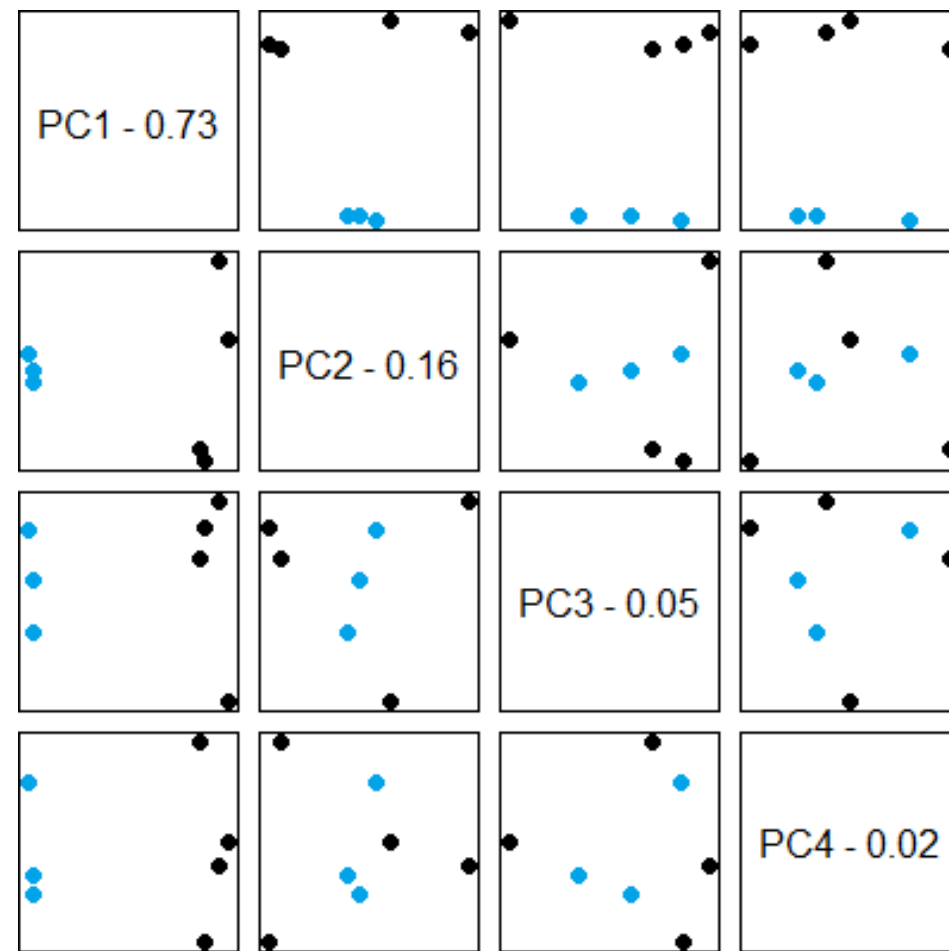
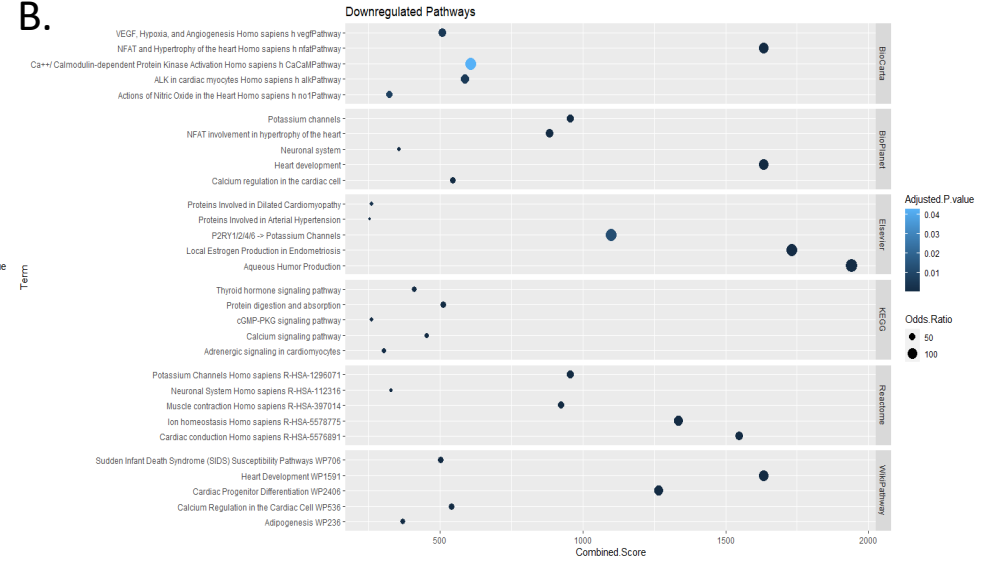


Figure S5

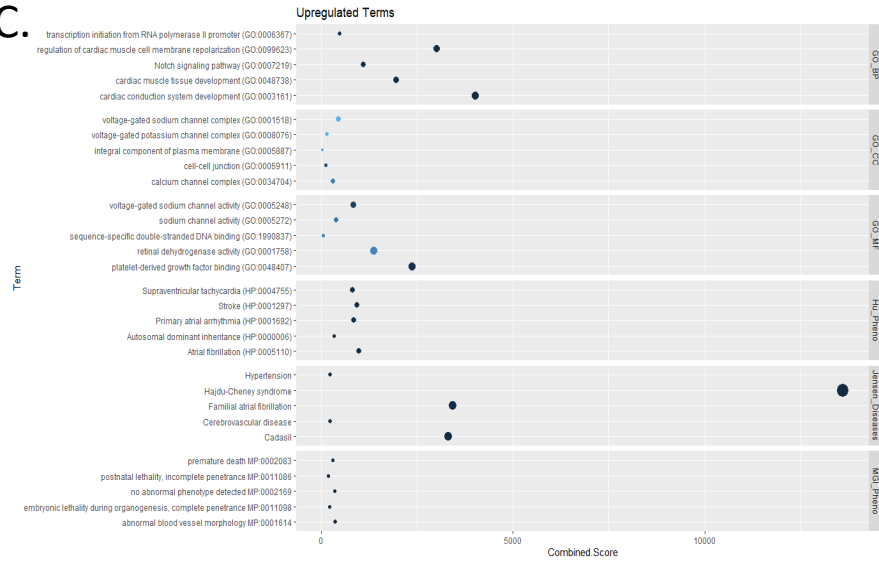
A.



B.



C.



D.

